

# Towards a framework for light-dosimetry studies: Methodological considerations

Steffen L Hartmeyer<sup>†</sup>, Forrest S Webler, Marilyne Andersen

Laboratory for Integrated Performance in Design, EPFL, Lausanne, Switzerland.

Received 18 February 2022; Revised 27 April 2022; Accepted 3 May 2022

Published online before print 19 July 2022, doi: 10.1177/14771535221103258

## Abstract

For field research of non-visual effects of light, accurate measurement of personal light exposure is required. A consensus framework for light-dosimetry could improve non-visual field research and ensure comparability between studies. Here we present a review of methodologies used in non-visual light-dosimetry studies published to date, focussing on considerations regarding the measurement and preparation of personal light exposure data. Overall, a large variability in the studies' methodologies is observed, highlighting the need for a consensus framework. We propose methodological considerations that should be included in such a framework and that can guide future studies.

Furthermore, we highlight important points that should be addressed in future research to ensure compatibility between different dosimetry studies. Taken together, this review effort underlines the importance of a systematic approach to light-dosimetry in order to harness all the power of integrative lighting research in real-life.

**Keywords:** Non-visual, Dosimetry, Lighting, Circadian, Field studies, Daylight

<sup>†</sup>Address for correspondence: Steffen Hartmeyer, Laboratory for Integrated Performance in Design (LIPID), École polytechnique fédérale de Lausanne (EPFL), LE 1 114, Station 18, 1015 Lausanne, Switzerland. Email: [steffen.hartmeyer@epfl.ch](mailto:steffen.hartmeyer@epfl.ch)

## 1. Introduction

Decades of research have shown that light has behavioural and physiological effects unrelated to human vision, mediated by a dedicated neural pathway. Through this pathway, light sets our biological clock,<sup>1</sup> directly affects various aspects of physiology and behaviour,<sup>2</sup> and may thereby be related to health and wellbeing.<sup>3</sup> Much of what is known about these non-visual effects of light has been established by extensive laboratory research, indicating that responses are modulated by different light exposure characteristics.<sup>4</sup> However, real-life light exposure resembles nothing like laboratory stimuli, but consists of complex patterns of light of different quantity and quality, as a result of moving

through our indoor and outdoor environments. Therefore, more field research is needed to complement and evaluate findings from controlled laboratory studies, and answer pressing questions pertaining to implications in applied contexts, such as architecture and lighting design, therapeutic applications, shift-work, transcontinental travel, and personal lifestyle.<sup>5</sup>

Due to the complexity of personal light exposure patterns and the uncontrolled nature of field research, it is crucial that results from different studies are comparable and repeatable, as well as transferable to applied contexts. This can be achieved by agreeing on standardised operating procedures for setting up and reporting lighting research studies.<sup>5</sup> A crucial aspect in non-visual lighting research is the accurate description of the lighting conditions under investigation. For experimental studies, guidelines for quantifying and reporting lighting conditions have recently been published.<sup>6-8</sup> However, these guidelines have only limited applicability for field research, where light exposure cannot be controlled. Thus, to date it is still unclear how to adequately measure, quantify, and analyse personal light exposure data with respect to non-visual responses.

Given the complexity of light exposure and its relationship with non-visual responses in real-life, accurate measurement of personal light exposure (i.e., dosimetry) is crucial. The dosimetry process – also called the dosimetry chain – consists of a series of steps (links), each of which has the potential to introduce additional uncertainty in the final research outcome.<sup>9</sup> Broadly, these links can be categorised as (1) selecting optical quantities, (2) calibrating dosimeters, (3) selecting a measurement setup, (4) processing the measured data, (5) calculating light exposure metrics, and (6) linking metrics to measured responses.<sup>9</sup> Importantly, different execution of each link may lead to substantial differences in the results, complicating their interpretation and comparison across studies. Therefore, agreement on standardised procedures for each link is urgently needed.<sup>9</sup>

In the present article, we set out to laying the groundwork for a consensus framework for non-visual light-dosimetry studies. To achieve this, we assembled a comprehensive set of dosimetry field studies published to date, which is taken as a basis for discussing methodological considerations for each link in the dosimetry chain. Herein, we review methodologies employed in previous dosimetry studies for measurement and preparation of personal light exposure data. Furthermore, we aim to identify crucial gaps in knowledge that need to be specifically addressed and clarified in future research.

As a brief note on nomenclature, throughout this article, the term *light exposure* is used to refer to the time series of light a person is exposed to and not to the quantity *luminous exposure*  $H_v$ . Furthermore, the term *light level* is used as a generic term where multiple light quantities are applicable within a given context (e.g., illuminance, alpha-opic irradiance etc).

## 2. Review method

### 2.1 Search strategy

To collect a comprehensive set of light-dosimetry field studies, a forward and backward citation search method was used, allowing for an efficient assembly of relevant studies within the same domain. Two of the first light-dosimetry studies in the context of chronobiology were chosen as a starting point,<sup>10,11</sup> from which eligible dosimetry studies published to date were identified by means of forward and backward citation search using the Web of Science citation database (forward and backward search) and the individual papers' reference list (forward search), within the period of January–March 2021.

### 2.2 Selection criteria

The main objective of this review was to identify light-dosimetry studies that covered the entire dosimetry chain from measurement to quantification and subsequent analysis of the collected light exposure data, the ultimate intent being to highlight methodological considerations in the dosimetry procedure and to identify metrics for the quantification of personal light exposure patterns. Therefore, studies were only eligible if personal light exposure data was measured with wearable light meters over a period of at least 24 hours and if the measured light data was included as a dependent or independent variable in the analysis. This criterion excluded studies that measured 24h-light exposure with static devices only (i.e., the device was not worn by the subject) or that measured light exposure but did not report any analyses of these measurements. Consequently, many intervention studies where personal light exposure was monitored but not analysed as a major dependent or independent variable were excluded, except for three studies. Specifically, the study by Peeters *et al.*<sup>12</sup> was included because personal light exposure was a primary dependent variable in the analysis, and Phillips *et al.*<sup>13</sup> and Zeitzer *et al.*<sup>14</sup> were included because multiple light exposure metrics were analysed.

In addition, to narrow the scope of this review, we primarily focused on dosimetry studies in the context of the non-visual effects of light; specifically, pertaining to sleep-wake regulation, circadian entrainment, and direct physiological and behavioural responses (e.g., alertness, cognitive performance, mood etc.). As a result, studies that measured personal light exposure in a different context (e.g., myopia, UV-light) were excluded, except for three myopia-related studies where the methodology added novel content to the review. Specifically, the studies by Alvarez and Wildsoet<sup>15</sup> and Ulaganathan *et al.*<sup>16</sup> were included because effects of sampling frequency on the accuracy of calculated light exposure metrics were investigated. Moreover, the study by Read *et al.*<sup>17</sup> was included because metrics are described that have not been used in any of the other studies already included in the review.

## 2.3 Final Set of Studies

In total, 104 studies were deemed eligible and formed the set of dosimetry field studies reviewed here. An overview of the selected studies<sup>10-113</sup> and study related characteristics is presented in the appendix (Table A1.1).

## 3. Methodological considerations

### 3.1 Dosimetry setup and measurement

#### 3.1.1 Dosimeter selection and optical quantities

Personal light exposure is usually measured with lightweight wearable devices (dosimeters; see Figure 1). A wide range of different technologies exists on the market, reflected in the variety of dosimeters employed by the reviewed dosimetry studies (see Table 1). The dosimeters can broadly be categorised into dedicated light meters or wrist-worn actigraphy devices with light sensors (note that most wrist-worn dosimeters can also be adapted to be worn elsewhere). Furthermore, devices differ in their spectral sensitivity and resolution. Most devices listed here measure photopic illuminance (e.g., *Actiwatch-L*), or spectral irradiance in the visible range across three channels (red, green, blue; e.g., *Actiwatch Spectrum*). Three devices include a sensor calibrated to approximate the circadian spectral sensitivity (i.e., LuxBlick, Daysimeter, Dimesimeter), while only two dosimeters were used that have a higher spectral resolution.

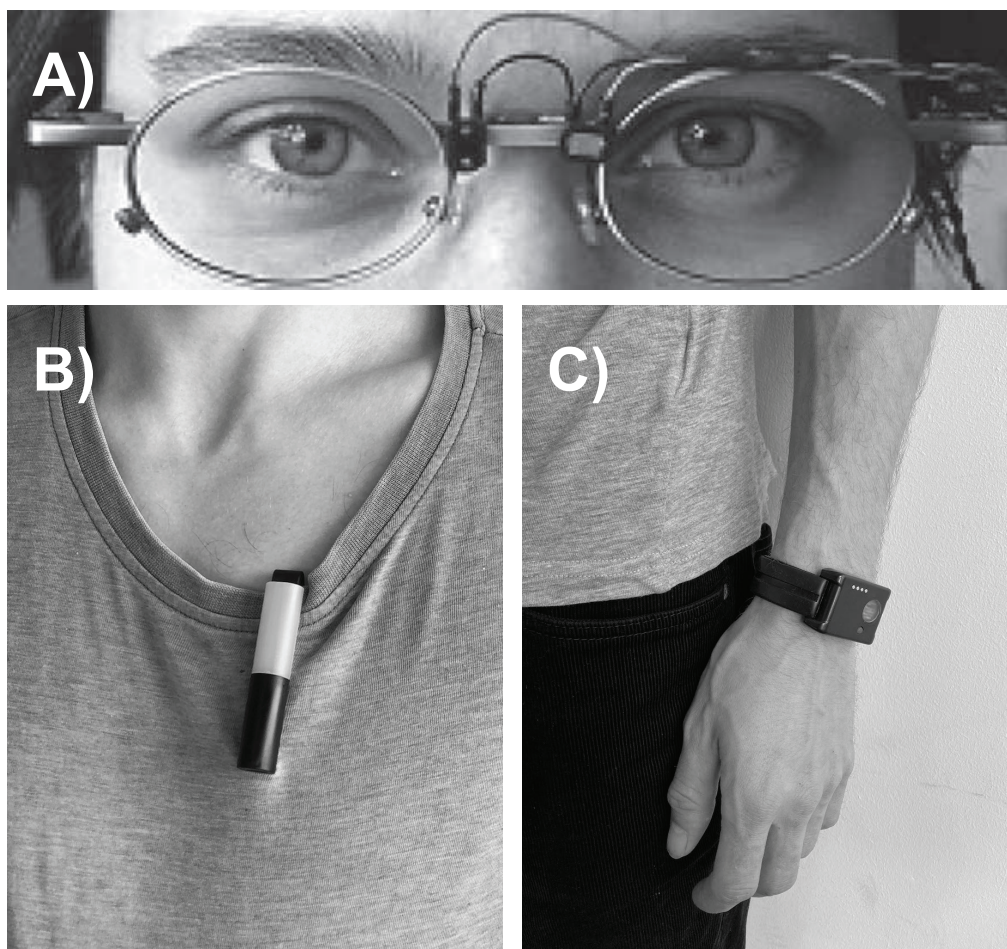
The optical quantities a dosimeter can measure is an important consideration for non-visual dosimetry, since the spectral composition of personal light patterns (i.e., the “relative spectral diet”, see Webler *et al.*<sup>115</sup>) can drastically vary over time. While historically, dosimeters have been developed to measure light for the visual system (i.e., photometric), we now know that the description of light in photometric terms is usually not appropriate when studying non-visual effects of light.<sup>116</sup> Consequently, new standards such as CIE S 026:2018<sup>117</sup> were developed, defining spectral sensitivity functions, quantities, and metrics to describe light for non-visual responses (i.e., alpha-opic quantities).

Currently the main limitation for the measurement of alpha-opic quantities is the limited availability of dosimetry devices that spectrally match the alpha-opic sensitivities. With the exception of devices used in two studies,<sup>19,26</sup> none of the dosimeters listed here match all alpha-opic sensitivities with sufficient accuracy.<sup>118</sup> Only one of the commercially available devices (i.e., the *Actiwatch Spectrum*) approximates the melanopic sensitivity function by a linear combination of photosensor outputs.<sup>118</sup> Note that the *ActTrust* dosimeter can be modified to provide a sufficient spectral match to the melanopic sensitivity curve as described previously<sup>119</sup>; however, no study in this review used this modified device. Given that the melanopic sensitivity curve matches the sensitivity of non-visual responses for a wide range of conditions, as suggested in a recent comprehensive review of several experimental studies,<sup>120</sup> dosimetry studies could use devices that match this curve (see<sup>66,69,81</sup>) until

sufficiently spectrally resolved devices become available. Such devices could contain broadband photosensors matching the alpha-opic sensitivities directly (i.e., analogous to the sensor in the *Daysimeter*); however, these sensors are then constrained by assumptions on the underlying neurophysiology. Therefore, devices from which (parts of) the visible spectrum can be sufficiently recovered in order to calculate alpha-opic quantities offer a significant advantage over the former type of devices, as further elaborated on in the Discussion section.

### 3.1.2 Dosimeter assessment and calibration

Light-dosimeters have been found to vary substantially in optical performance between and within models as assessed in a range of studies.<sup>118,121–123</sup> Many of the devices assessed in these specific performance assessment studies were used by the reviewed dosimetry studies. However, only some of the reviewed studies (N=26) report that dosimeters were validated and calibrated against an industry-standard light sensor (Figure 2A). Among the studies that provided a description of the validation and calibration procedure (N=19), a variety of methods were used: notably, validation was performed for different reference lighting conditions, including artificial light sources (N=13), simulated daylight (N=2), and/or under naturalistic indoor or outdoor conditions (N=6).



**Figure 1.** Example of different dosimeter types and positions: **A)** *LuxBlick* light-dosimeter for measurement at eyelevel (image retrieved from Hubalek *et al.*<sup>56</sup>), **B)** *Spectrace*<sup>141</sup> light-dosimeter worn at the chest, **C)** *ActTrust* wrist-worn actigraphy and light-dosimetry device.

The diversity of calibration methods used in dosimetry studies highlights the need for standardised assessment and calibration procedures. Typically, optical performance of photometers is assessed in terms of spectral sensitivity, directional response, and response linearity as defined in the standard ISO/CIE 19476:2014.<sup>124</sup> However, current standards are not readily applicable for the characterisation of light-dosimeters for non-visual effects research, therefore novel methods are needed.<sup>118</sup> Published optical performance metrics can help guiding the selection of an appropriate dosimeter for a given study. For non-visual effects research, devices that have good spectral sensitivity for matching the five  $\alpha$ -opic sensitivity functions should be preferred.<sup>117</sup> However, trade-offs between spectral and directional mismatch should be considered, since the arrangement of multiple photosensors may affect directional responses.<sup>118</sup> The impact of directional mismatch may also depend on where the device is worn and how the data is aggregated. That is, high movement at the wrist or aggregation over longer time periods may cancel out directional mismatches.<sup>125</sup> Moreover, the linear range and dynamic resolution that is required should be considered depending on the lighting conditions expected in the study.

The standard ISO/CIE 19476:2014<sup>124</sup> also describes calibration procedures for photometry devices; however, this standard may not be applicable to dosimeters that measure non-photometric quantities (e.g., spectral irradiance, alpha-opic irradiance). As a result, several methods have been described by individual studies.<sup>122,126–128</sup> An important aspect to consider during calibration is the selection of a calibration light source that matches the lighting conditions typically encountered during the study. For studies with various lighting conditions, it has been recommended to calibrate devices to an overcast sky at noon,<sup>122</sup> or by averaging across several light sources.<sup>129</sup> Note that ambient conditions (i.e., temperature and humidity) can also substantially impact sensor accuracy and, ideally, should be calibrated for accordingly. However, this calibration is difficult for the many available light-dosimeters that do not include sensors to measure ambient conditions.

### 3.1.3 Dosimeter position

For research on the non-visual effects of light, the amount of light reaching the eye (corneal light exposure) is of primary interest; therefore, the position of the dosimeter on the body is an important consideration. Among the reviewed studies, only very few measured light exposure at eye-level (N=9), whereas most measured at the wrist (N=67) and some at the chest (N=22; Figure 2B). Measurements at eye-level require a specific setup (e.g., *LuxBlick*; see Figure 1) and might be perceived as more obtrusive than at the wrist or chest,<sup>130</sup> which may explain the small number of studies measuring at this position. While both wrist and chest measurements may be less obtrusive, the large number of studies measuring at the wrist can be partly explained by the frequent use of actigraphy devices, allowing concomitant measurement of light exposure and sleep-wake activity within a single device. Only few studies measured actigraphy at the wrist and light at another position with separate devices (N=12), or with a single device that is transferred to the wrist for actigraphy during sleep (N=3).

**Table 1.** Overview of dosimeter models used in previous studies sorted by frequency of use.

Device Name	Manufacturer	Primary purpose	Output	N
Actiwatch-L	MiniMitter (now Philips Respironics)	Actigraphy	E <sub>v</sub>	25
Actillum	Ambulatory Monitoring Inc.	Actigraphy	E <sub>v</sub>	14
Actiwatch 2	Philips Respironics	Actigraphy	E <sub>v</sub>	12
Actiwatch Spectrum	Philips Respironics	Actigraphy	W, R, G, B	10
Daysimeter	Lighting Research Center	Light	E <sub>v</sub> , CS	6
HOBO Pendant	Onset Computer Co.	Light	E <sub>v</sub>	6
Actiwatch-L	CamNtech Ltd.	Actigraphy	E <sub>v</sub>	4
Dimesimeter	Lighting Research Center	Light	E <sub>v</sub> , CS	3
MotionWatch 8	CamNtech Ltd.	Actigraphy	E <sub>v</sub>	3
StowAway	Onset Computer Co.	Light	E <sub>v</sub>	3
Custom	various	various	E <sub>v</sub>	3
Actiwatch Plus	Philips Respironics	Actigraphy	W, R, G, B	2
GeneActiv	Activinsights Ltd.	Actigraphy	E <sub>v</sub>	2
Lightlog	LightlogProject	Light	W, R, G, B	2
Actiwatch RGB	CamNtech Ltd.	Actigraphy	W, R, G, B	1
ActTrust	Condor Instruments	Actigraphy	W, R, G, B	1
CSA AM	Computer Science Applications Inc.	Actigraphy	E <sub>v</sub>	1
Daqtometer 2.4	Daqtix GmbH	Actigraphy	E <sub>v</sub>	1
LightWatcher	Object-Tracker	Light	W, R, G, B, UV, IR	1
LuxBlick	Hubalek <i>et al.</i> <sup>56</sup>	Light	E <sub>v</sub> , E <sub>c</sub>	1
MotionLogger-L	CamNtech Ltd.	Actigraphy	E <sub>v</sub>	1
RaySeG	Eto <i>et al.</i> <sup>38</sup>	Light	R, G, B, UV	1
Sleepwatch-L	Ambulatory Monitoring Inc.	Actigraphy	E <sub>v</sub>	1
Custom	Adamsson <i>et al.</i> <sup>19</sup>	Light	Range 400-750 nm (50 nm res.)	1
Custom	Cain <i>et al.</i> <sup>26</sup>	Light	Range 340-780 nm (15 nm res.)	1

Note: E<sub>v</sub> = Illuminance, W = white light, R = long-wavelength irradiance, G = medium-wavelength irradiance, B = short-wavelength irradiance, UV = ultra-violet light, IR = infrared light, CS = Circadian Stimulus, E<sub>c</sub> = irradiance weighted by  $c(\lambda)$ <sup>114</sup>. This table does not list all available devices, but only devices that were used in the reviewed studies.

As most studies did not measure light exposure at eye-level, it is important to study to what extent corneal light exposure can be estimated from measurements at other positions. Yet surprisingly few studies have addressed this question. An early study that is frequently cited reported high correlation between measurements at the wrist and the forehead;<sup>10</sup> however, the validity of these findings is limited due to dosimeter saturation at higher illuminance levels. Furthermore, correlation does not show the amount and direction of deviations at different light levels. Another study in postsurgical in-hospital patients reported little average deviation (<10 lx) between wrist and eye-level measurements up to 5000 lx;<sup>131</sup> however, the findings may not be generalisable to normal living conditions. To our knowledge only one study examined different measurement positions under normal living conditions for an extended period, reporting little deviation between illuminance at eye-level and the chest, but large deviation for the wrist, which generally underestimated eye-level exposure, especially at higher illuminance levels.<sup>125</sup>

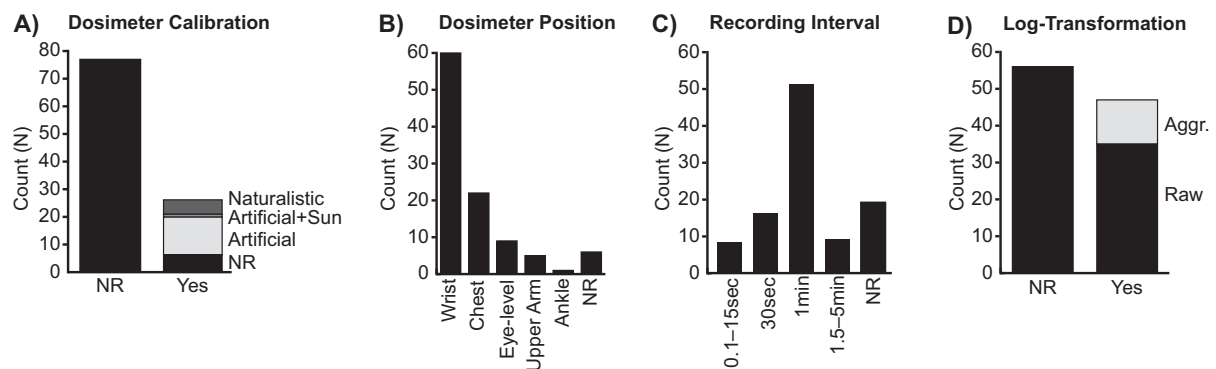
While this preliminary evidence suggests that measurements at the chest may be more accurate than at the wrist for estimating corneal light exposure, systematic variations in measurements at different positions should be considered. Aarts *et al.*<sup>123</sup> found that both wrist and chest measurements

deviated substantially more from eye-level measurements under indoor conditions compared to outdoor conditions, which was especially pronounced for wrist measurements. Moreover, under indoor conditions wrist measurements tended to overestimate and under outdoor conditions underestimate eye-level exposure, corroborating earlier studies. Additionally, under indoor conditions, systematic variations have been observed for different activities, body postures and gaze directions.<sup>132</sup> Interestingly, this study also found that the exposure threshold associated with phase shifts of dim light melatonin onset (DLMO) estimated from eye-level measurements was not associated with phase shifts when using the same threshold for wrist measurements, likely due to overestimation of eye-level exposure.

### 3.1.4 Recording interval

In light-dosimetry the recording interval or *epoch length* defines the rate at which individual light exposure measurements are recorded and therefore determines the temporal resolution and accuracy at which changes in light exposure can be captured. Among the studies under review, epoch lengths ranged from 100ms to 5min, with 1min and 30s being the most used (N=51 and N=16, respectively; Figure 2C). Note that these epochs indicate the rate at which samples are recorded but do not necessarily reflect the sampling rate of the photosensors, as some dosimeters record an aggregated value of several samples across the given interval.

Epoch lengths are usually selected to optimise battery and memory usage, since dosimeters are often employed continuously for several days in a row. However, while longer epochs may help to conserve battery power and reduce memory load, the accuracy with which light exposure is measured may be affected, resulting in a trade-off between battery/memory usage and accuracy. For example, it has been found that epochs of 3min or longer lead to a loss of accuracy relative to shorter epochs when calculating cumulative light exposure and time spent under bright light conditions.<sup>15,16</sup> On the other hand, while longer epochs may reduce accuracy, it is not yet known how much interpretability is gained by increasing recording rate.



**Figure 2.** Frequency of methods used across the reviewed studies: **A)** dosimeter calibration and, if applicable, calibration light sources, **B)** dosimeter positions, **C)** recording intervals, and **D)** log-transformation sequence (*Aggr.* = transformation of aggregated data, *Raw* = transformation of raw data). NR = not reported.



## 3.2 Data preparation

### 3.2.1 Data cleaning

Due to the uncontrolled nature of light-dosimetry, measured data may contain invalid or artifact data. Two major sources of invalid data are periods where the dosimeter was not worn or when the light sensors were obstructed. Invalid periods were identified by some studies (N=18) based on concomitant inactivity longer than a given timeframe, which ranged from 5–120min across studies. Some studies additionally examined the light data for smooth periods with little fluctuation, indicating invalid periods.<sup>31,69,81</sup> Furthermore, several methods for identifying measurement artifacts due to sensor obstruction have been reported. Some studies (N=13) identified artifacts as light measurements below a threshold value during day- or wake-time, ranging from 0–10 lx across studies. Other methods involved the identification of temporary drops in the data,<sup>26</sup> unusually high rate of change,<sup>74</sup> and outlier detection methods.<sup>105</sup> Moreover, some studies removed remaining noise in the data with smoothing methods, by using simple moving average (SMA) filters with different window sizes between 5–20min,<sup>32,79,85,104</sup> or local regression smoothing (LOESS).<sup>108</sup> The latter procedure preserves peaks in the signal better than a SMA filter, but it can be computationally expensive for large datasets. Note that smoothing methods can also be used as an analytical procedure; for example, to quantify light dose in time by mimicking non-visual response characteristics.<sup>81</sup> Beside smoothing, some studies (N=15) averaged the data into bins (e.g., hourly averages), which can also be considered a means to remove noise in the data. However, as with smoothing methods, it is important to consider logarithmic transformation when aggregating data, as discussed in the next section.

Although cleaning measured personal light exposure data is an important step in the dosimetry process, nearly no studies have systematically investigated what cleaning methods and parameters are most appropriate for these kinds of data. To our knowledge, only one study has examined identification thresholds for sensor obstruction, showing that dosimetry measurements in a very dim laboratory environment without coverage by clothing did not fall below 1lx.<sup>88</sup>

### 3.2.2 Logarithmic Transformation

Personal light exposure data can cover a large range of light levels over several orders of magnitude and usually follow a log-normal distribution.<sup>133</sup> Therefore, analyses may require logarithmic transformation of the data to ensure a normal distribution and enable interpretability. An important consideration when applying log-transformation is the sequence in which the data is transformed, aggregated, and analysed, particularly regarding the question whether the transformation should be applied *before* or *after* quantifying the “raw” light data. Amongst the reviewed studies, less than half applied a log-transformation, either before (N=35) or after (N=12) quantification (Figure 2D). A major rationale for transforming the data was to ensure normality for statistical analyses (N=10). Interestingly, four studies provide an explicit rationale for transforming raw data,<sup>65,92,108,109</sup> referring to

log-linearity observed in non-visual dose-response curves.<sup>134,135</sup> Contrastingly, Scheuermaier *et al.*<sup>88</sup> argue that log-transformation should be applied after aggregating the data, in order to account for brief episodes of very bright light.

Indeed, the sequence of log-transformation may substantially affect the results. Take for example the hypothetical light exposure pattern of an office worker between 12:00–13:00h, who during work is exposed to 200 lx and who goes outside for a lunch break at 12:30h, being exposed to 10000 lx. The calculated hourly means for this individual would be around  $10^{3.71}$  (~5100 lx) with log-transformation after aggregation, and  $10^{3.15}$  (~1400 lx) with log-transformation before aggregation. Realistically this difference could be even greater depending on the fluctuation of light levels. This potentially huge impact on aggregated light exposure makes it impossible to compare data such as mean hourly light exposure between studies that aggregated and transformed the data in a different sequence. Moreover, analyses of non-visual responses may be affected, such as when using linear models to examine acute effects of hourly light exposure. Note that although it can be argued that mean light exposure may not be the right metric to analyse non-visual effects, it is still the most widely used metric to describe and compare personal light exposure patterns across all studies reviewed here.

Logarithmic stimulus-response relationships are well established in psychophysics (i.e., *Fechner's law*), and non-visual responses to light are no exception: logarithmic relationships to light intensity have been described for circadian phase resetting,<sup>136</sup> melatonin suppression,<sup>134</sup> and acute alertness.<sup>135</sup> Some findings from cell recordings suggest that logarithmic encoding of light intensity may happen at the level of retinal ganglion cells, whose neuronal firing response is directly proportional to the detected amount of photons on a logarithmic scale.<sup>137–139</sup> However, no study has yet specifically addressed what these findings imply for the measurement of time-series light exposure data.

## 4. Discussion and recommendations

In this review we presented an overview of methods employed by previous dosimetry studies to measure and prepare personal light exposure data. Overall, a large variability in methodologies was observed across all studies. Personal light exposure was measured with a variety of dosimeters, at different positions on the body, and with different recording intervals. Very few studies measured spectrally resolved light exposure. Dosimetry devices were often not reported to be calibrated, and studies that calibrated the devices used a variety of different methods. Regarding data preparation, the few studies that report data-driven cleaning procedures used several different methods and parameters to identify invalid light exposure data. Discrepancy was also observed in the application and sequence of log-transformation of the light data. In the following, we briefly discuss methodological implications based on these findings and provide recommendations for future dosimetry studies, as well as important points that need to be addressed in future research.

## **4.1 A note on accuracy in dosimetry for non-visual effects of light**

As emphasized throughout this review, research of non-visual effects demands an accurate description of lighting conditions to improve the validity of research findings and ensure comparability and transferability of results across studies.<sup>7</sup> This holds especially for dosimetry, to avoid additional uncertainty in the presence of the plethora of confounders in field research. However, since an increase in accuracy often comes at the expense of factors such as practicality and cost, one of the central questions in dosimetry is how much accuracy is required at each dosimetry step, for analysing the relationship between measured non-visual responses and light exposure.

Given that dosimetry inherently refers to the study of the absorption of a physical quantity by the human body, it makes sense that dosimetry methods and metrics are biologically relevant. In theory, many of the methods discussed in this review could be based on neurophysiological mechanisms, for example, using smoothing methods that reflect temporal integration of the light signal. However, one of the central challenges in dosimetry is that our understanding of the mechanisms underlying signal encoding, adaptation, and photic integration in the non-visual system is still incomplete. These uncertainties make it difficult to define accuracy limits for spectral and temporal resolution, and dynamic range, which is further complicated by large inter-individual differences in light sensitivity.<sup>140</sup> In light of these uncertainties, instead of basing accuracy limits on assumptions of the underlying biology, it might be more appropriate to consider how the physical signal itself can be measured more accurately within the given technological and practical constraints, for instance, by using information theoretic approaches (see Section 4.3).

## **4.2 A proposal for guidelines and recommendations**

Based on the findings of this review we propose guidelines and recommendations for future dosimetry studies, summarised in Table 2. It is important to note that these recommendations are based on current knowledge, while several points demand further investigation (see Section 4.3). Therefore, it is crucial that methods and parameters used at each step in the dosimetry process are reported in necessary detail, either in the article itself or provided in the supplementary material. Insufficient reporting hinders transparency, reproducibility, and comparability, which is essential for research of non-visual effects in the field to be successful.

A crucial aspect in dosimetry is the selection of an appropriate dosimeter model, which depends on the context and aims of a given study. For research on the non-visual effects of light, devices that spectrally match the alpha-opic sensitivity curves should be preferred; however, given current limited availability of such devices, at least dosimeters that match the melanopic sensitivity curve should be used. In any case, we strongly recommend to always validate and calibrate dosimeters, and report a (reference to the) description of the calibration setup and procedure, including at least the calibration light sources used and which type(s) of calibration were performed (e.g., spectral, intensity, temperature calibration).

**Table 2.** Proposed guidelines and recommendations for dosimetry studies

Issue	Recommendation
Dosimeter Selection	<ul style="list-style-type: none"> <li>• Select dosimeters with a spectral resolution appropriate to the aim of the study: for research on non visual responses, ideally, select devices that spectrally match the alpha-opic sensitivities.</li> <li>• Consider optical performance characteristics in relation to study context and measurement setup.</li> </ul>
Calibration	<ul style="list-style-type: none"> <li>• Calibrate devices; Select reference light sources based on study context.</li> <li>• Report description of validation and calibration procedure, including the reference light sources used.</li> </ul>
Dosimeter Position	<ul style="list-style-type: none"> <li>• Position dosimeter at eye-level for precise measurements, otherwise at chest. Use dosimeter in tandem with wrist-actigraphy if needed.</li> <li>• Consider context dependent direction and amount of deviation from corneal exposure.</li> <li>• Consider obtrusiveness and practical issues; Carefully instruct participants to avoid sensor obstruction.</li> </ul>
Recording Interval	<ul style="list-style-type: none"> <li>• Select shortest epoch possible to achieve desired battery life.</li> <li>• Avoid epochs longer than 2min.</li> </ul>
Data Cleaning	<ul style="list-style-type: none"> <li>• Use data-driven methods to verify or replace subjective reports.</li> <li>• Use multiple variables to identify invalid periods (e.g., light exposure and activity).</li> <li>• Explain how and why given methods and parameters are selected.</li> </ul>
Log-Transformation	<ul style="list-style-type: none"> <li>• Consider the impact of log-transformation sequence; Compare analyses for different sequences.</li> <li>• Report geometric means alongside arithmetic means.</li> </ul>

Another important consideration are contextual factors, such as lighting conditions, environments, activities, and body positions expected during the study. These factors may determine the amount and direction of deviation from corneal exposure for different measurement positions, which can affect the analysis of non-visual responses. For example, using sensitivity thresholds based on corneal exposure for the analysis of light exposure measured at the wrist under predominantly indoor conditions may prevent detecting an effect, due to overestimation of corneal exposure. Similarly, contextual factors should be considered for selecting dosimeter models, calibration light sources, and data cleaning methods.

Furthermore, dosimetry methods should be selected to achieve the highest accuracy within the given study constraints, avoiding assumptions on the underlying biology. The latter is particularly important for data preparation, as it is still unclear how the non-visual system encodes and integrates light information over time. Therefore, we recommend evaluating a range of different methods and parameters during analyses and include a detailed description of the selected cleaning methods and transformation sequence. Moreover, alongside the arithmetic mean, the geometric mean and/or other measures of central tendency should be reported.

### 4.3 Future work

While the proposed recommendations given above are based on current knowledge and may form the basis of a framework for non-visual light dosimetry, we have identified several important points that need to be addressed in further research for such a framework to be able to become fully operational (see Table 3). Importantly, more research investigating systematic interactions between factors related to the study context (e.g., lighting conditions, activities etc.) and dosimetry methods are urgently needed, for example, to evaluate different dosimeter positions or improve data cleaning methods. On a long-term basis, more research studying underlying neurophysiological mechanisms such as photic

integration and sensitivity adaptation is required to inform dosimetry methods. Such research would be especially insightful regarding the sequence of logarithmic transformation, given the widespread use of this method in dosimetry data analysis.

To facilitate the development of a future consensus framework and standards for light-dosimetry, it is of interest to examine in how far current standards in light metrology can be recycled and adapted. For example, the CIE already proposes standards for the calibration and performance characterisation of photometers (ISO/CIE 19476:2014<sup>124</sup>). However, these standards are limited to photometric measurements, and may not be appropriate for the measurement of alpha-opic quantities as recommended in the new standard CIE S 026:2018<sup>117</sup>. Some suggestions for how these standards can be adapted have already been put forward by Price *et al.*<sup>118</sup>. Similarly, an effort for developing consensus reporting standards for light-intervention studies (ENLIGHT<sup>141</sup>) is currently under way, which could in future be expanded to light-dosimetry in general. Such reporting standards may also contain guidelines for reporting the intended use of light-dosimetry in intervention-studies, since intervention studies often make little use of collected dosimetry data.

In parallel, information theoretic approaches focussing on increasing measurement accuracy within the given technological and practical constraints should be explored. Specifically, novel data-driven methods may be used to increase spectral and temporal resolution of dosimeters, as described in a complementary paper<sup>142</sup> and briefly introduced in the following. Spectral and temporal resolution historically have been constrained by technological limits to size, weight, power, and cost of dosimeters. However, recent advances in sensor technology and signal processing methods, such as *compressed sensing*,<sup>143</sup> challenge conventional limitations. Such methods demonstrate that for compressible (i.e., non-random) signals, the amount of information encoded can be much smaller than prescribed under classic sampling theorems. Compressed sensing effectively finds the simplest representation of a signal by exploiting regularities in the structure and properties of encoded signals. In practice, sensors based this method can reduce the amount of encoded information at the source by a factor of 10,<sup>144</sup> enabling high spectral resolution at a small size and cost. Recent work on a compressed sensing dosimeter named *Spectrace*<sup>145</sup>, developed by the authors, has already demonstrated the ability to recover 5 nm resolution data over the visible range from 14 narrowband photodiodes,<sup>142</sup> all in an autonomous wearable form no larger than a USB stick. As with compressed spectral sensing, sparse representation and forecasting can also be used to construct adaptive sampling rates that respond to changes in the environment (e.g., sample more when the scene changes and less when it is stable). Such approaches offer low-power and compact hardware forms without information loss. Taken together, information theoretic approaches offer novel ways of increasing accuracy in dosimetry within the given constraints, while avoiding the uncertainty introduced by making assumptions on human biology.

**Table 3.** Important points to be addressed in further research

Outlook	Point to address
Short-term	<ul style="list-style-type: none"><li>• Evaluation of optimal reference light source(s) for dosimeter calibration.</li><li>• Development of standardised calibration protocols.</li><li>• Investigation of the effects of different lighting conditions and activities on measurements at different positions.</li><li>• Evaluation, validation, and development of new and existing data cleaning methods.</li><li>• Investigation of the effects of transformation sequence on dosimetry outcomes.</li><li>• Exploration of novel information theoretic approaches to increase measurement accuracy within constraints.</li></ul>
Long-term	<ul style="list-style-type: none"><li>• Investigate neurophysiological mechanisms to inform dosimetry methods and metrics.</li></ul>

## 5. Conclusion

With this article we discuss the status quo in dosimetry methodology to form a basis for further work towards a consensus framework for light-dosimetry studies. This review is the first to highlight the prevalent variability in methodologies employed by light-dosimetry studies during the past three decades, underscoring the need for standardised operating procedures to improve the comparability and repeatability of dosimetry studies. Moreover, by collecting this diverse set of methodologies, we were able to reveal important gaps in knowledge that need to be addressed in further research. At the same time, we show what methods are available for measuring and preparing personal light exposure, and what to consider and avoid when conducting light-dosimetry studies. With this work we would like to bring the topic of light-dosimetry to greater attention, with the hope to provide researchers with the required knowledge to perform high quality studies and inspire more research in this direction. Furthermore, we want to emphasise the importance of carefully considering each step in the dosimetry process as each methodological decision may substantially affect the final study results. In the forthcoming second part of this review, methods for quantifying and analysing personal light exposure with respect to the non-visual system will be discussed.

## Declaration of conflicting interests

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

## Funding

The authors disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This research was supported by the European Union's Horizon 2020 research and innovation programme Marie Skłodowska-Curie Innovative Training Networks (ITN), with the Grant No. 860613 (LIGHTCAP).

## References

1. Roenneberg T, Daan S, Merrow M. The art of entrainment. *Journal of Biological Rhythms* 2003; 18: 183–194.
2. Chellappa S, Steiner R, Blattner P, Oelhafen P, Götz T, Cajochen C. Non-visual effects of light on melatonin, alertness and cognitive performance: Can blue-enriched light keep us alert? *PLoS ONE* 2011; 6: e16429+.
3. Lunn RM, Blask DE, Coogan AN, Figueiro MG, Gorman MR, Hall JE, Hansen J, Nelson RJ, Panda S, Smolensky MH, Stevens RG, Turek FW, Vermeulen R, Carreón T, Caruso CC, Lawson CC, Thayer KA, Twery MJ, Ewens AD, Garner SC, Schwingl PJ, Boyd WA. Health consequences of electric lighting practices in the modern world: A report on the National Toxicology Program’s workshop on shift work at night, artificial light at night, and circadian disruption. *Science of The Total Environment* 2017; 607–608: 1073–1084.
4. Prayag AS, Münch M, Aeschbach D, Chellappa SL, Gronfier C. Light modulation of human clocks, wake, and sleep. *Clocks and Sleep* 2019; 1: 193–208.
5. Münch M, Wirz-Justice A, Brown SA, Kantermann T, Martiny K, Stefani O, Vetter C, Wright KP, Wulff K, Skene DJ. The role of daylight for humans: gaps in current knowledge. *Clocks and Sleep* 2020; 2: 61–85.
6. Knoop M, Broszio K, Diakite A, Liedtke C, Niedling M, Rothert I, Rudawski F, Weber N. methods to describe and measure lighting conditions in experiments on non-image-forming aspects. *LEUKOS* 2019; 15: 163–179.
7. Spitschan M, Stefani O, Blattner P, Gronfier C, Lockley SW, Lucas RJ. how to report light exposure in human chronobiology and sleep research experiments. *Clocks and Sleep* 2019; 1: 280–289.
8. CIE. *What to document and report in studies of ipRGC-influenced responses to light*. CIE TN 011:2020, Vienna, Austria, 2020.
9. Price LLA. Opinion: The dosimetry chain. *Lighting Research and Technology* 2015; 47: 896–896.
10. Okudaira N, Kripke DF, Webster JB. Naturalistic studies of human light exposure. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology* 1983; 245: R613–R615.
11. Savides TJ, Messin S, Senger C, Kripke DF. Natural light exposure of young adults. *Physiology and Behavior* 1986; 38: 571–574.
12. Peeters ST, Smolders KCHJ, de Kort YAW. What you set is (not) what you get: How a light intervention in the field translates to personal light exposure. *Building and Environment* 2020; 185: 107288.
13. Phillips AJK, Vidafar P, Burns AC, McGlashan EM, Anderson C, Rajaratnam SMW, Lockley SW, Cain SW. High sensitivity and interindividual variability in the response of the human circadian system to evening light. *Proceedings of the National Academy of Sciences* 2019; 116: 12019–12024.
14. Zeitzer JM, Friedman L, Yesavage JA. Effectiveness of evening phototherapy for insomnia is reduced by bright daytime light exposure. *Sleep Medicine* 2011; 12: 805–807.
15. Alvarez AA, Wildsoet CF. Quantifying light exposure patterns in young adult students. *Journal of Modern Optics* 2013; 60: 1200–1208.
16. Ulaganathan S, Read SA, Collins MJ, Vincent SJ. Measurement duration and frequency impact objective light exposure measures. *Optometry and Vision Science* 2017; 94: 588–597.
17. Read SA, Vincent SJ, Tan C-S, Ngo C, Collins MJ, Saw S-M. Patterns of daily outdoor light exposure in Australian and Singaporean children. *Translational Vision Science and Technology*; 2018; 7(3): 8.

18. aan het Rot M, Moskowitz DS, Young SN. Exposure to bright light is associated with positive social interaction and good mood over short time periods: A naturalistic study in mildly seasonal people. *Journal of Psychiatric Research* 2008; 42: 311–319.
19. Adamsson M, Laike T, Morita T. Annual variation in daily light exposure and circadian change of melatonin and cortisol concentrations at a northern latitude with large seasonal differences in photoperiod length. *Journal of Physiological Anthropology* 2016; 36: 6.
20. Akacem LD, Wright KP, LeBourgeois MK. Bedtime and evening light exposure influence circadian timing in preschool-age children: A field study. *Neurobiology of Sleep and Circadian Rhythms* 2016; 1: 27–31.
21. Asai Y, Obayashi K, Oume M, Ogura M, Takeuchi K, Yamagami Y, Tai Y, Kurumatani N, Saeki K. Farming habit, light exposure, physical activity, and depressive symptoms. A cross-sectional study of the HEIJO-KYO cohort. *Journal of Affective Disorders* 2018; 241: 235–240.
22. Auger RR, Burgess HJ, Dierkhising RA, Sharma RG, Slocumb NL. Light exposure among adolescents with delayed sleep phase disorder: A prospective cohort study. *Chronobiology International* 2011; 28: 911–920.
23. Böhmer MN, Valstar MJ, Aarts MPJ, Bindels PJE, Oppewal A, Someren EJW, Festen DAM. Shedding light on light exposure in elderly with intellectual disabilities. *Journal of Intellectual Disability Research* 2021; 65: 361–372.
24. Borugian MJ, Gallagher RP, Friesen MC, Switzer TF, Aronson KJ. Twenty-four-hour light exposure and melatonin levels among shift workers. *Journal of Occupational and Environmental Medicine* 2005; 47: 1268–1275.
25. Boubekri M, Cheung IN, Reid K. Impact of windows and daylight exposure on overall health and sleep quality of office workers: a case-control pilot study. *Journal of Clinical Sleep Medicine* 2014; 10: 603–611.
26. Cain SW, McGlashan EM, Vidafar P, Mustafavska J, Curran SPN, Wang X, Mohamed A, Kalavally V, Phillips AJK. Evening home lighting adversely impacts the circadian system and sleep. *Scientific Reports* 2020; 10: 19110.
27. Campbell SS, Kripke DF, Gillin JC, Hrubovcak JC. Exposure to light in healthy elderly subjects and alzheimer’s patients. *Physiology and Behavior* 1988; 42: 141–144.
28. Cole RJ, Kripke DF, Wisbey J, Mason WJ, Gruen W, Hauri PJ, Juarez S. Seasonal variation in human illumination exposure at two different latitudes. *Journal of Biological Rhythms* 1995; 10: 324–334.
29. Crowley SJ, Molina TA, Burgess HJ. A week in the life of full-time office workers: Work day and weekend light exposure in summer and winter. *Applied Ergonomics* 2015; 46: 193–200.
30. Darling AL, Hart KH, Arber S, Berry JL, Morgan PL, Middleton BA, Lanham-New S, Skene DJ. 25-Hydroxyvitamin D status, light exposure and sleep quality in UK dwelling South Asian and Caucasian postmenopausal women. *The Journal of Steroid Biochemistry and Molecular Biology* 2019; 189: 265–273.
31. Daugaard S, Garde AH, Bonde JPE, Christoffersen J, Hansen ÅM, Markvart J, Schlünssen V, Skene DJ, Vistisen HT, Kolstad HA. Night work, light exposure and melatonin on work days and days off. *Chronobiology International* 2017; 34: 942–955.
32. Daugaard S, Markvart J, Bonde JP, Christoffersen J, Garde AH, Hansen ÅM, Schlünssen V, Vestergaard JM, Vistisen HT, Kolstad HA. Light exposure during days with night, outdoor, and indoor work. *Annals of Work Exposures and Health* 2019; 63: 651–665.
33. Dumont M, Benhaberou-Brun D, Paquet J. Profile of 24-h Light exposure and circadian phase of melatonin secretion in night workers. *Journal of Biological Rhythms* 2001; 16: 502–511.
34. Dumont M, Lanctôt V, Cadieux-Viau R, Paquet J. Melatonin production and light exposure of rotating night workers. *Chronobiology International* 2012; 29: 203–210.
35. Emens JS, Yuhas K, Rough J, Kochar N, Peters D, Lewy AJ. Phase angle of entrainment in morning- and evening-types under naturalistic conditions. *Chronobiology International* 2009; 26: 474–493.



36. Esaki Y, Kitajima T, Obayashi K, Saeki K, Fujita K, Iwata N. Daytime light exposure in daily life and depressive symptoms in bipolar disorder: A cross-sectional analysis in the APPLE cohort. *Journal of Psychiatric Research* 2019; 116: 151–156.
37. Espiritu RC, Kripke DF, Ancoli-Israel S, Mowen MA, Mason WJ, Fell RL, Klauber MR, Kaplan OJ. Low illumination experienced by San Diego adults: Association with atypical depressive symptoms. *Biological Psychiatry* 1994; 35: 403–407.
38. Eto N, Okada K, Obana A, Okazaki S, Nishiwaki Y. Use of an eyeglass-type measuring device to assess exposure of the eye to light among urban office workers. *Toho Journal of Medicine* 2016; 2: 86–94.
39. Feigl B, Ojha G, Hides L, Zele AJ. Melanopsin-driven pupil response and light exposure in non-seasonal major depressive disorder. *Frontiers in Neurology* 2018; 9.
40. Figueiro MG, Rea MS. Evening daylight may cause adolescents to sleep less in spring than in winter. *Chronobiology International* 2010; 27: 1242–1258.
41. Figueiro MG, Hamner R, Higgins P, Hornick T, Rea MS. Field measurements of light exposures and circadian disruption in two populations of older adults. *Journal of Alzheimer's Disease: JAD* 2012; 31: 711–715.
42. Figueiro MG, Steverson B, Heerwagen J, Kampschroer K, Hunter CM, Gonzales K, Plitnick B, Rea MS. The impact of daytime light exposures on sleep and mood in office workers. *Sleep Health* 2017; 3: 204–215.
43. Flanagan SC, Cobice D, Richardson P, Sittlington JJ, Saunders KJ. Elevated melatonin levels found in young myopic adults are not attributable to a shift in circadian phase. *Investigative Ophthalmology and Visual Science* 2020; 61: 45–45.
44. Gibbs M, Hampton S, Morgan L, Arendt J. Predicting circadian response to abrupt phase shift: 6-sulphatoxymelatonin rhythms in rotating shift workers offshore. *Journal of Biological Rhythms* 2016; 22: 368–370.
45. Goulet G, Mongrain V, Desrosiers C, Paquet J, Dumont M. Daily light exposure in morning-type and evening-type individuals. *Journal of Biological Rhythms* 2007; 22: 151–158.
46. Grandner MA, Kripke DF, Langer RD. Light exposure is related to social and emotional functioning and to quality of life in older women. *Psychiatry Research* 2006; 143: 35–42.
47. Grundy A, Sanchez M, Richardson H, Tranmer J, Borugian M, Graham CH, Aronson KJ. Light intensity exposure, sleep duration, physical activity, and biomarkers of melatonin among rotating shift nurses. *Chronobiology International* 2009; 26: 1443–1461.
48. Grundy A, Tranmer J, Richardson H, Graham CH, Aronson KJ. The influence of light at night exposure on melatonin levels among Canadian rotating shift nurses. *Cancer Epidemiology, Biomarkers and Prevention* 2011; 20: 2404–2412.
49. Guillemette J, Hébert M, Paquet J, Dumont M. Natural bright light exposure in the summer and winter in subjects with and without complaints of seasonal mood variations. *Biological Psychiatry* 1998; 44: 622–628.
50. Gumenyuk V, Roth T, Drake CL. Circadian phase, sleepiness, and light exposure assessment in night workers with and without shift work disorder. *Chronobiology International* 2012; 29: 928–936.
51. Hall AL, Davies HW, Koehoorn M. Personal light-at-night exposures and components of variability in two common shift work industries: Uses and implications for future research. *Scandinavian Journal of Work, Environment and Health* 2018; 44: 80–87.
52. Harb F, Hidalgo MP, Martau B. Lack of exposure to natural light in the workspace is associated with physiological, sleep and depressive symptoms. *Chronobiology International* 2015; 32: 368–375.
53. Hebert M, Dumont M, Paquet J. Seasonal and diurnal patterns of human illumination under natural conditions. *Chronobiology International* 1998; 15: 59–70.
54. Heil DP, Mathis SR. Characterizing free-living light exposure using a wrist-worn light monitor. *Applied Ergonomics* 2002; 33: 357–363.

55. Higgins PA, Hornick TR, Figueiro MG. Rest-activity and light exposure patterns in the home setting: A methodological case study. *American Journal of Alzheimer's Disease and Other Dementias* 2010; 25: 353–361.
56. Hubalek S, Brink M, Schierz C. Office workers' daily exposure to light and its influence on sleep quality and mood. *Lighting Research and Technology* 2010; 42: 33–50.
57. Jean-Louis G, Kripke DF, Ancoli-Israel S, Klauber MR, Sepulveda RS, Mowen M-A, Assmus JD, Langer RD. Circadian sleep, illumination, and activity patterns in women: Influences of aging and time reference. *Physiology and Behavior* 2000; 68: 347–352.
58. Jean-Louis G, Kripke DF, Elliott JA, Zizi F, Wolintz AH, Lazzaro DR. Daily illumination exposure and melatonin: Influence of ophthalmic dysfunction and sleep duration. *Journal of Circadian Rhythms* 2005; 3: 13.
59. Joo EY, Abbott SM, Reid KJ, Wu D, Kang J, Wilson J, Zee PC. Timing of light exposure and activity in adults with delayed sleep-wake phase disorder. *Sleep Medicine* 2017; 32: 259–265.
60. Kawinska A, Dumont M, Selmaoui B, Paquet J, Carrier J. Are modifications of melatonin circadian rhythm in the middle years of life related to habitual patterns of light exposure? *Journal of Biological Rhythms* 2005; 20: 451–460.
61. Keller LK, Grünewald B, Vetter C, Roenneberg T, Schulte-Körne G. Not later, but longer: Sleep, chronotype and light exposure in adolescents with remitted depression compared to healthy controls. *European Child and Adolescent Psychiatry* 2017; 26: 1233–1244.
62. Kim SJ, Lim YC, Kwon HJ, Lee JH. Association of rest-activity and light exposure rhythms with sleep quality in insomnia patients. *Chronobiology International* 2020; 37: 403–413.
63. Koller M, Kundi M, Stidl H-G, Zidek T, Haider M. Personal light dosimetry in permanent night and day workers. *Chronobiology International* 1993; 10: 143–155.
64. Koller M, Härma M, Laitinen JT, Kundi M, Piegler B, Haider M. Different patterns of light exposure in relation to melatonin and cortisol rhythms and sleep of night workers. *Journal of Pineal Research* 1994; 16: 127–135.
65. Kripke DF, Jean-Louis G, Elliott JA, Klauber MR, Rex KM, Tuunainen A, Langer RD. Ethnicity, sleep, mood, and illumination in postmenopausal women. *BMC Psychiatry* 2004; 4: 8.
66. Lee EE, Amritwar A, Hong LE, Mohyuddin I, Brown T, Postolache TT. Daily and seasonal variation in light exposure among the Old Order Amish. *International Journal of Environmental Research and Public Health* 2020; 17: 4460.
67. Liu L, Marler MR, Parker BA, Jones V, Johnson S, Cohen-Zion M, Fiorentino L, Sadler GR, Ancoli-Israel S. The relationship between fatigue and light exposure during chemotherapy. *Supportive Care in Cancer* 2005; 13: 1010–1017.
68. Lowden A, Lemos NAM, Gonçalves BSB, Öztürk G, Louzada F, Pedrazzoli M, Moreno CR. Delayed sleep in winter related to natural daylight exposure among Arctic day workers. *Clocks and Sleep* 2018; 1: 105–116.
69. Maren SV der, Moderie C, Duclos C, Paquet J, Daneault V, Dumont M. Daily profiles of light exposure and evening use of light-emitting devices in young adults complaining of a delayed sleep schedule. *Journal of Biological Rhythms* 2018; 33: 192–202.
70. Martin J, Jeste DV, Caliguri MP, Patterson T, Heaton R, Ancoli-Israel S. Actigraphic estimates of circadian rhythms and sleep/wake in older schizophrenia patients. *Schizophrenia Research* 2001; 47: 77–86.
71. Martin JS, Hébert M, Ledoux É, Gaudreault M, Laberge L. Relationship of chronotype to sleep, light exposure, and work-related fatigue in student workers. *Chronobiology International* 2012; 29: 295–304.
72. Martin JS, Gaudreault MM, Perron M, Laberge L. Chronotype, light exposure, sleep, and daytime functioning in high school students attending morning or afternoon school shifts: An actigraphic study. *Journal of Biological Rhythms* 2016; 31: 205–217.

73. Martinez-Nicolas A, Ortiz-Tudela E, Madrid JA, Rol MA. Crosstalk between environmental light and internal time in humans. *Chronobiology International* 2011; 28: 617–629.
74. Martinez-Nicolas A, Madrid JA, Rol MA. Day–night contrast as source of health for the human circadian system. *Chronobiology International* 2014; 31: 382–393.
75. Nioi A, Roe J, Gow A, McNair D, Aspinall P. Seasonal differences in light exposure and the associations with health and well-being in older adults: An exploratory study. *Health Environments Research and Design Journal* 2017; 10: 64–79.
76. Obayashi K, Saeki K, Iwamoto J, Okamoto N, Tomioka K, Nezu S, Ikada Y, Kurumatani N. Positive effect of daylight exposure on nocturnal urinary melatonin excretion in the elderly: A cross-sectional analysis of the HEIJO-KYO study. *The Journal of Clinical Endocrinology and Metabolism* 2012; 97: 4166–4173.
77. Obayashi K, Saeki K, Iwamoto J, Okamoto N, Tomioka K, Nezu S, Ikada Y, Kurumatani N. Effect of exposure to evening light on sleep initiation in the elderly: A longitudinal analysis for repeated measurements in home settings. *Chronobiology International* 2014; 31: 461–467.
78. Papantoniou K, Pozo OJ, Espinosa A, Marcos J, Castaño-Vinyals G, Basagaña X, Ribas FC, Mirabent J, Martín J, Careny G, Martín CR, Middleton B, Skene DJ, Kogevinas M. Circadian variation of melatonin, light exposure, and diurnal preference in day and night shift workers of both sexes. *Cancer Epidemiology, Biomarkers and Prevention* 2014; 23: 1176–1186.
79. Pattinson CL, Allan AC, Staton SL, Thorpe KJ, Smith SS. Environmental Light Exposure Is Associated with Increased Body Mass in Children. *PLOS ONE* 2016; 11: e0143578.
80. Phillips AJK, Clerx WM, O’Brien CS, Sano A, Barger LK, Picard RW, Lockley SW, Klerman EB, Czeisler CA. Irregular sleep/wake patterns are associated with poorer academic performance and delayed circadian and sleep/wake timing. *Scientific Reports* 2017; 7: 3216.
81. Price LLA, Udovicic L, Khazova M. Circadian light exposures of shift working nurses. In: *Proceedings of the 29th Quadrennial Session of the CIE*, Washington DC, USA, 14-22 June 2019, paper no. PP30, pp.838–845. Vienna, CIE.
82. Rabstein S, Burek K, Lehnert M, Beine A, Vetter C, Harth V, Putzke S, Kantermann T, Walther J, Wang-Sattler R, Pallapies D, Brüning T, Behrens T. Differences in twenty-four-hour profiles of blue-light exposure between day and night shifts in female medical staff. *Science of The Total Environment* 2019; 653: 1025–1033.
83. Rea MS, Brons JA, Figueiro MG. Measurements of light at night (LAN) for a sample of female school teachers. *Chronobiology International* 2011; 28: 673–680.
84. Refinetti R. Chronotype variability and patterns of light exposure of a large cohort of United States residents. *The Yale Journal of Biology and Medicine* 2019; 92: 179–186.
85. Reid KJ, Santostasi G, Baron KG, Wilson J, Kang J, Zee PC. Timing and intensity of light correlate with body weight in adults. *PLOS ONE* 2014; 9: e92251.
86. Rufiange M, Beaulieu C, Lachapelle P, Dumont M. Circadian light sensitivity and rate of retinal dark adaptation in indoor and outdoor workers. *Journal of Biological Rhythms* 2007; 22: 454–457.
87. Ruiz FS, Bejjamini F, Beale AD, Gonçalves B da SB, Vartanian D, Taporoski TP, Middleton B, Krieger JE, Vallada H, Arendt J, Pereira AC, Knutson KL, Pedrazzoli M, Schantz M von. Early chronotype with advanced activity rhythms and dim light melatonin onset in a rural population. *Journal of Pineal Research* 2020; 69: e12675.
88. Scheuermaier K, Laffan AM, Duffy JF. Light exposure patterns in healthy older and young adults. *Journal of Biological Rhythms* 2010; 25: 113–122.
89. Silva A, Simón D, Pannunzio B, Casaravilla C, Díaz Á, Tassinio B. Chronotype-dependent changes in sleep habits associated with dim light melatonin onset in the antarctic summer. *Clocks and Sleep* 2019; 1: 352–366.
90. Smit AN, Broesch T, Siegel JM, Mistlberger RE. Sleep timing and duration in indigenous villages with and without electric lighting on Tanna Island, Vanuatu. *Scientific Reports* 2019; 9: 17278.

91. Shochat T, Martin J, Marler M, Ancoli-Israel S. Illumination levels in nursing home patients: effects on sleep and activity rhythms. *Journal of Sleep Research* 2000; 9: 373–379.
92. Shochat T, Santhi N, Herer P, Flavell SA, Skeldon AC, Dijk D-J. Sleep timing in late autumn and late spring associates with light exposure rather than sun time in college students. *Frontiers in Neuroscience* 2019; 13: 882.
93. Smolders KCHJ, de Kort YAW, van den Berg SM. Daytime light exposure and feelings of vitality: Results of a field study during regular weekdays. *Journal of Environmental Psychology* 2013; 36: 270–279.
94. Staples VSL, Archer SN, Arber S, Skene DJ. Daily light exposure profiles in older non-resident extreme morning and evening types. *Journal of Sleep Research* 2009; 18: 466–471.
95. Stone JE, Sletten TL, Magee M, Ganesan S, Mulhall MD, Collins A, Howard M, Lockley SW, Rajaratnam SMW. Temporal dynamics of circadian phase shifting response to consecutive night shifts in healthcare workers: role of light–dark exposure. *The Journal of Physiology* 2018; 596: 2381–2395.
96. Stothard ER, McHill AW, Depner CM, Birks BR, Moehlman TM, Ritchie HK, Guzzetti JR, Chinoy ED, LeBourgeois MK, Axelsson J, Wright KP. Circadian entrainment to the natural light-dark cycle across seasons and the weekend. *Current Biology* 2017; 27: 508–513.
97. Sun J-L, Wu S-C, Chang L-I, Chiou J-F, Chou P-L, Lin C-C. The Relationship between light exposure and sleep, fatigue, and depression in cancer outpatients: Test of the mediating effect. *Cancer Nursing* 2014; 37: 382–390.
98. te Lindert BHW, Itzhacki J, van der Meijden WP, Kringelbach ML, Mendoza J, Van Someren EJW. Bright environmental light ameliorates deficient subjective ‘liking’ in insomnia: An experience sampling study. *Sleep* 2018; 41: zsy022.
99. Thorne H, Hampton S, Morgan L, Skene DJ, Arendt J. Differences in sleep, light, and circadian phase in offshore 18:00–06:00 h and 19:00–07:00 h shift workers. *Chronobiology International* 2008; 25: 225–235.
100. Thorne HC, Jones KH, Peters SP, Archer SN, Dijk D-J. Daily and seasonal variation in the spectral composition of light exposure in humans. *Chronobiology International* 2009; 26: 854–866.
101. Tsai S-Y, Barnard KE, Lentz MJ, Thomas KA. Twenty-four hours light exposure experiences in postpartum women and their 2–10-week-old infants: An intensive within-subject design pilot study. *International Journal of Nursing Studies* 2009; 46: 181–188.
102. Tsai S-Y, Thomas KA, Lentz MJ, Barnard KE. Light is beneficial for infant circadian entrainment: an actigraphic study. *Journal of Advanced Nursing* 2012; 68: 1738–1747.
103. Tsuzuki K, Mori I, Sakoi T, Kurokawa Y. Effects of seasonal illumination and thermal environments on sleep in elderly men. *Building and Environment* 2015; 88: 82–88.
104. Ulset VS, Czajkowski NO, Staton S, Smith S, Pattinson C, Allen A, Thorpe K, Bekkhus M. Environmental light exposure, rest-activity rhythms, and symptoms of inattention and hyperactivity: An observational study of Australian preschoolers. *Journal of Environmental Psychology* 2021; 73: 101560.
105. van Duijnhoven J, Aarts M, Kort H. Personal lighting conditions of office workers: An exploratory field study. *Lighting Research and Technology* 2020; 1477153520976940.
106. Vinzio S, Ruellan A, Perrin A-E, Schlienger J-L, Goichot B. Actigraphic assessment of the circadian rest–activity rhythm in elderly patients hospitalized in an acute care unit. *Psychiatry and Clinical Neurosciences* 2003; 57: 53–58.
107. Wallace-Guy GM, Kripke DF, Jean-Louis G, Langer RD, Elliott JA, Tuunainen A. Evening light exposure: Implications for sleep and depression. *Journal of the American Geriatrics Society* 2002; 50: 738–739.
108. Wams EJ, Woelders T, Marring I, van Rosmalen L, Beersma DGM, Gordijn MCM, Hut RA. linking light exposure and subsequent sleep: A field polysomnography study in humans. *Sleep* 2017; 40: 12.

109. Wang EJ, Kripke DF, Stein MT, Parry BL. Measurement of illumination exposure in postpartum women. *BMC Psychiatry* 2003; 3: 5.
110. Wilson J, Reid KJ, Braun RI, Abbott SM, Zee PC. Habitual light exposure relative to circadian timing in delayed sleep-wake phase disorder. *Sleep* 2018; 41: 11.
111. Woelders T, Beersma DGM, Gordijn MCM, Hut RA, Wams EJ. Daily light exposure patterns reveal phase and period of the human circadian clock. *Journal of Biological Rhythms* 2017; 32: 274–286.
112. Wright KP, McHill AW, Birks BR, Griffin BR, Rusterholz T, Chinoy ED. Entrainment of the human circadian clock to the natural light-dark cycle. *Current Biology* 2013; 23: 1554–1558.
113. Youngstedt SD, Leung A, Kripke DF, Langer RD. Association of morning illumination and window covering with mood and sleep among post-menopausal women. *Sleep and Biological Rhythms* 2004; 2: 174–183.
114. Gall D. Circadiane Lichtgrößen und deren messtechnische Ermittlung. *Licht* 2002; 54: 1292–1297.
115. Webler FS, Spitschan M, Foster RG, Andersen M, Peirson SN. What is the ‘spectral diet’ of humans? *Current Opinion in Behavioral Sciences* 2019; 30: 80–86.
116. Lucas RJ, Peirson SN, Berson DM, Brown TM, Cooper HM, Czeisler CA, Figueiro MG, Gamlin PD, Lockley SW, O’Hagan JB, Price LLA, Provencio I, Skene DJ, Brainard GC. Measuring and using light in the melanopsin age. *Trends in Neurosciences* 2014; 37: 1–9.
117. CIE. *System for metrology of optical radiation for ipRGC-influenced responses to light*. CIE S 026/E:2018, Vienna, Austria, 2018.
118. Price LLA, Lyachev A, Khazova M. Optical performance characterization of light-logging actigraphy dosimeters. *JOSA A* 2017; 34: 545–557.
119. Price LLA, Lyachev A. Modification of a personal dosimetry device for logging melanopic irradiance. *Lighting Research and Technology* 2017; 49: 922–927.
120. Brown TM. Melanopic illuminance defines the magnitude of human circadian light responses under a wide range of conditions. *Journal of Pineal Research* 2020; 69: e12655.
121. Price LLA, Khazova M, O’Hagan JB. Performance assessment of commercial circadian personal exposure devices. *Lighting Research and Technology* 2012; 44: 17–26.
122. Markvart J, Hansen ÅM, Christoffersen J. Comparison and correction of the light sensor output from 48 wearable light exposure devices by using a side-by-side field calibration method. *LEUKOS* 2015; 11: 155–171.
123. Aarts MPJ, van Duijnhoven J, Aries MBC, Rosemann ALP. Performance of personally worn dosimeters to study non-image forming effects of light: Assessment methods. *Building and Environment* 2017; 117: 60–72.
124. CIE. *Characterization of the performance of illuminance meters and luminance meters*. ISO/CIE 19476:2014(E), Vienna, Austria, 2014.
125. Figueiro MG, Hamner R, Bierman A, Rea MS. Comparisons of three practical field devices used to measure personal light exposures and activity levels. *Lighting Research and Technology* 2013; 45: 421–434.
126. Joyce DS, Zele AJ, Feigl B, Adhikari P. The accuracy of artificial and natural light measurements by actigraphs. *Journal of Sleep Research* 2020; 29: e12963.
127. Mohamed A, Kalavally V, Cain SW, Phillips AJK, McGlashan EM, Tan CP. Wearable light spectral sensor optimized for measuring daily  $\alpha$ -opic light exposure. *Optics Express* 2021; 29: 27612–27627.
128. Stone JE, McGlashan EM, Facer-Childs ER, Cain SW, Phillips AJK. Accuracy of the GENEActiv device for measuring light exposure in sleep and circadian research. *Clocks and Sleep* 2020; 2: 143–152.

129. Udovičić L, Janßen M, Nowack D, Price LLA. Personenbezogene Lichtexpositionsmessungen in Feldstudien - Eine Handlungsanleitung zur Charakterisierung und Kalibrierung von Lichtexpositionsdetektoren. *Bundesanstalt für Arbeitsschutz und Arbeitsmedizin* 2016; 1.
130. van Duijnhoven J, Aarts MPJ, Aries MBC, Böhmer MN, Rosemann ALP. Recommendations for measuring non-image-forming effects of light: A practical method to apply on cognitive impaired and unaffected participants. *Technology and Health Care* 2017; 25: 171–186.
131. Jardim ACN, Pawley MDM, Cheeseman JF, Guesgen MJ, Steele CT, Warman GR. Validating the use of wrist-level light monitoring for in-hospital circadian studies. *Chronobiology International* 2011; 28: 834–840.
132. Yoshimura M, Kitamura S, Eto N, Hida A, Katsunuma R, Ayabe N, Motomura Y, Nishiwaki Y, Negishi K, Tsubota K, Mishima K. Relationship between indoor daytime light exposure and circadian phase response under laboratory free-living conditions. *Biological Rhythm Research* 2020; 0: 1–21.
133. Dobb R, Martial F, Elijah D, Storchi R, Brown TM, Lucas RJ. The impact of temporal modulations in irradiance under light adapted conditions on the mouse suprachiasmatic nuclei (SCN). *Scientific Reports* 2017; 7: 10582.
134. Zeitzer JM, Dijk D-J, Kronauer RE, Brown EN, Czeisler CA. Sensitivity of the human circadian pacemaker to nocturnal light: melatonin phase resetting and suppression. *The Journal of physiology* 2000; 526: 695–702.
135. Cajochen C, Zeitzer JM, Czeisler CA, Dijk DJ. Dose-response relationship for light intensity and ocular and electroencephalographic correlates of human alertness. *Behavioural Brain Research* 2000; 115: 75–83.
136. Boivin DB, Duffy JF, Kronauer RE, Czeisler CA. Dose-response relationships for resetting of human circadian clock by light. *Nature* 1996; 379: 540–542.
137. Dacey DM, Liao H-W, Peterson BB, Robinson FR, Smith VC, Pokorny J, Yau K-W, Gamlin PD. Melanopsin-expressing ganglion cells in primate retina signal colour and irradiance and project to the LGN. *Nature* 2005; 433: 749–754.
138. Milosavljevic N, Storchi R, Eleftheriou CG, Colins A, Petersen RS, Lucas RJ. Photoreceptive retinal ganglion cells control the information rate of the optic nerve. *Proceedings of the National Academy of Sciences* 2018; 115: E11817–E11826.
139. Wong KY. A retinal ganglion cell that can signal irradiance continuously for 10 hours. *Journal of Neuroscience* 2012; 32: 11478–11485.
140. Stone JE, McGlashan EM, Quin N, Skinner K, Stephenson JJ, Cain SW, Phillips AJK. The role of light sensitivity and intrinsic circadian period in predicting individual circadian timing. *Journal of Biological Rhythms* 2020; 35: 628–640.
141. Spitschan M, Najjar R, McGlashan E, Lok R, Kervezee L. ENLIGHT Initiative, [www.lightcat.group/enlight](http://www.lightcat.group/enlight) (2021, accessed 26 April 2022).
142. Webler FS, Spitschan M, Andersen M. Towards ‘Fourth Paradigm’ Spectral Sensing. *Sensors* 2022; 22: 2377.
143. Donoho DL. Compressed sensing. *IEEE Transactions on Information Theory* 2006; 52: 1289–1306.
144. August Y, Stern A. Compressive sensing spectrometry based on liquid crystal devices. *Optics Letters* 2013; 38: 4996–4999.
145. Webler FS, Chinazzo G, Andersen M. Towards a wearable sensor for spectrally-resolved personal light monitoring. *Journal of Physics: Conference Series* 2021; 2042: 012120.

# Appendix

**Table A1.1.** Overview of the dosimetry studies included in the review

Authors	N	Sample	Dosimeter	Position	Actigraphy setup	Recording interval	Calibration	Quantities	Log-transformation
aan het Rot <i>et al.</i> <sup>18</sup>	48	SAD patients	Actiwatch-L	Wrist	Single	2min	-	E <sub>v</sub>	-
Adamsson <i>et al.</i> <sup>19</sup>	15	Office-workers	Actiwatch-L, Custom	Wrist, Chest	Tandem	1min	Art. + Sun	E <sub>v</sub> , R, G, B, M	-
Akacem <i>et al.</i> <sup>20</sup>	21	Children	Dimesimeter	Chest	Tandem	1min	-	E <sub>v</sub>	-
Alvarez <i>et al.</i> <sup>15</sup>	27	Students	HOBO Pendant	Upper arm	None	10s	-	E <sub>v</sub>	-
Asai <i>et al.</i> <sup>21</sup>	1005	Eldery	Actiwatch 2	Wrist	Single	1min	-	E <sub>v</sub>	-
Auger <i>et al.</i> <sup>22</sup>	38	DSPD patients	Actiwatch-L	Wrist	Single	1min	-	E <sub>v</sub>	Aggr.
Böhmer <i>et al.</i> <sup>23</sup>	48	Elderly	HOBO Pendant	Chest	Tandem	1min	Nat.	E <sub>v</sub>	Raw
Bougian <i>et al.</i> <sup>24</sup>	17	Shift-workers	StowAway	Chest	None	30s	-	E <sub>v</sub>	-
Boubreki <i>et al.</i> <sup>25</sup>	21	Office-workers	Actiwatch-L	Wrist	Single	30s	-	E <sub>v</sub>	Raw
Cain <i>et al.</i> <sup>26</sup>	59	General pop.	Custom	Chest	Tandem	2min	Art. + Sun	E <sub>v</sub> , mEDI	Raw
Campbell <i>et al.</i> <sup>27</sup>	23	Eldery	Custom	Wrist	Single	1min	No descr.	E <sub>v</sub>	-
Cole <i>et al.</i> <sup>28</sup>	54	General pop.	Actillum	Wrist, Chest	Tandem	1min	Art.	E <sub>v</sub>	Raw
Crowley <i>et al.</i> <sup>29</sup>	14	Office-workers	Actiwatch Spectrum	Chest	Tandem	30s	-	E <sub>v</sub>	Raw
Darling <i>et al.</i> <sup>30</sup>	32	Postmenop. women	Actiwatch-L (CamNtech)	Wrist, Chest	Tandem	1min	-	E <sub>v</sub>	-
Daugaard <i>et al.</i> <sup>31</sup>	341	Shift-workers	Actiwatch Spectrum	Upper arm	Transfer	1min	Nat.	E <sub>v</sub>	-
Daugaard <i>et al.</i> <sup>32</sup>	485	Indoor-, Outdoor-, Shift-workers	Actiwatch Spectrum	Upper arm	Transfer	1min	Nat.	E <sub>v</sub>	Aggr.
Dumont <i>et al.</i> <sup>33</sup>	30	Shift-workers	Actillum	Wrist	Single	1min	Art.	E <sub>v</sub>	Raw
Dumont <i>et al.</i> <sup>34</sup>	13	Shift-workers	Actiwatch-L	Chest	None	1min	Art.	E <sub>v</sub>	Raw
Emens <i>et al.</i> <sup>35</sup>	66	General pop.	Actiwatch-L	Wrist	Single	30s	-	E <sub>v</sub>	Raw
Esaki <i>et al.</i> <sup>36</sup>	181	BD patients	Actiwatch Plus	Wrist	Single	1min	-	E <sub>v</sub>	-
Espiritu <i>et al.</i> <sup>37</sup>	150	General pop.	Actillum	Wrist	Single	-	-	E <sub>v</sub>	Raw
Eto <i>et al.</i> <sup>38</sup>	39	Office-workers	RaySeG	Eye	None	1ms	-	R, G, B	-
Feigl <i>et al.</i> <sup>39</sup>	21	General pop.	Geneactiv	-	Single	-	-	E <sub>v</sub>	Aggr.
Figueiro & Rea <sup>40</sup>	12	Adolescents	Daysimeter	Eye	None	30s	No descr.	E <sub>v</sub> , CLA	-
Figueiro <i>et al.</i> <sup>41</sup>	37	Eldery	Dimesimeter	Wrist	Single	-	-	CS	-
Figueiro <i>et al.</i> <sup>42</sup>	109	Office-workers	Daysimeter	Chest	Transfer	90s	No descr.	CS	-
Flanagan <i>et al.</i> <sup>43</sup>	51	General pop.	Actiwatch 2	Wrist	Single	30s	-	E <sub>v</sub>	-
Gibbs <i>et al.</i> <sup>44</sup>	23	Shift-workers	Actiwatch-L	-	Single	1min	-	E <sub>v</sub>	-
Goulet <i>et al.</i> <sup>45</sup>	19	General pop.	Actiwatch-L	Wrist	Single	1min	Art.	E <sub>v</sub>	Raw
Grandner <i>et al.</i> <sup>46</sup>	459	Postmenop. women	Actillum	Wrist	Single	1min	-	E <sub>v</sub>	Raw
Grundy <i>et al.</i> <sup>47</sup>	61	Shift-workers	StowAway	Chest	None	5min	-	E <sub>v</sub>	Aggr.
Grundy <i>et al.</i> <sup>48</sup>	123	Shift-workers	StowAway	Chest	None	1min	-	E <sub>v</sub>	Aggr.
Guillemette <i>et al.</i> <sup>49</sup>	19	General pop.	Actillum	Wrist, Chest	Tandem	1min	-	E <sub>v</sub>	-
Gumenyuk <i>et al.</i> <sup>50</sup>	10	Shift-workers	Actiwatch-L	Wrist	Single	-	-	E <sub>v</sub>	-

(Continued)

**Table A1.1. (Continued)**

Authors	N	Sample	Dosimeter	Position	Actigraphy setup	Recording interval	Calibration	Quantities	Log-transformation
Hall <i>et al.</i> <sup>51</sup>	102	Shift-workers	Daysimeter	Chest	None	1min	-	E <sub>v</sub>	-
Harb <i>et al.</i> <sup>52</sup>	20	Hospital staff	Actiwatch 2	Wrist	Single	1min	-	E <sub>v</sub>	-
Hebert <i>et al.</i> <sup>53</sup>	16	General pop.	Actillum	Wrist	Single	1min	-	E <sub>v</sub>	-
Heil & Mathis <sup>54</sup>	11	Hospital staff	CSA	Wrist	Single	1min	-	E <sub>v</sub>	-
Higgins <i>et al.</i> <sup>55</sup>	2	Eldery	Daysimeter, Sleepwatch-L	Eye, Wrist	Tandem	-	No descr.	E <sub>v</sub> , CS	-
Hubalek <i>et al.</i> <sup>56</sup>	23	Office-workers	LuxBlick	Eye	None	100ms	-	E <sub>v</sub> , E <sub>c</sub>	-
Jean-Louis <i>et al.</i> <sup>57</sup>	224	General pop.	Actillum	Wrist	Single	1min	-	E <sub>v</sub>	Raw
Jean-Louis <i>et al.</i> <sup>58</sup>	30	General pop.	Actiwatch-L	Wrist	Single	1min	-	E <sub>v</sub>	Raw
Joo <i>et al.</i> <sup>59</sup>	68	DSPD patients	Actiwatch-L	Wrist	Single	-	-	E <sub>v</sub>	Aggr.
Kawinska <i>et al.</i> <sup>60</sup>	37	General pop.	Actiwatch-L	Wrist	Single	1min	Art.	E <sub>v</sub>	Raw
Keller <i>et al.</i> <sup>61</sup>	38	Adolescents	Daqtometer 2.4	Wrist	Single	10s	-	E <sub>v</sub>	-
Kim <i>et al.</i> <sup>62</sup>	182	Insomnia patients	Actiwatch 2	-	Single	1min	-	E <sub>v</sub>	Raw
Koller <i>et al.</i> <sup>63</sup>	12	Shift-workers	Custom	Eye	None	15s	-	E <sub>v</sub>	-
Koller <i>et al.</i> <sup>64</sup>	12	Shift-workers	Custom	Eye	None	30s	-	E <sub>v</sub>	-
Kripke <i>et al.</i> <sup>65</sup>	459	Postmenop. women	Actillum	Wrist	Single	-	-	E <sub>v</sub>	Raw
Lee <i>et al.</i> <sup>66</sup>	33	Old Order Amish	Actiwatch Spectrum	Wrist	Single	1min	-	E <sub>v</sub> , R, G, B, mEDI	Raw
Liu <i>et al.</i> <sup>67</sup>	63	Cancer patients	Actillum	Wrist	Single	-	-	E <sub>v</sub>	-
Lowden <i>et al.</i> <sup>68</sup>	32	Office-workers	MotionWatch 8	Wrist	Single	1min	-	E <sub>v</sub>	Raw
Maren <i>et al.</i> <sup>69</sup>	28	Young adults	Actiwatch Spectrum	Upper arm	Tandem	1min	Art.	E <sub>v</sub> , B	Raw
Martin <i>et al.</i> <sup>70</sup>	28	Schizophrenia patients	Actillum	Wrist	Single	1min	-	E <sub>v</sub>	-
Martin <i>et al.</i> <sup>71</sup>	88	Students	Actiwatch-L	Wrist	Single	1min	Art.	E <sub>v</sub>	Raw
Martin <i>et al.</i> <sup>72</sup>	57	Students	Actiwatch-L	Wrist	Single	30s	Art.	E <sub>v</sub>	Raw
Martinez-Nicolas <i>et al.</i> <sup>73</sup>	88	Students	HOBO Pendant	Chest	Tandem	30s	Nat.	E <sub>v</sub>	Raw
Martinez-Nicolas <i>et al.</i> <sup>74</sup>	131	Students	HOBO Pendant	Chest	Tandem	30s	Nat.	E <sub>v</sub>	Raw
Nioi <i>et al.</i> <sup>75</sup>	20	Eldery	Actiwatch Spectrum	Chest	Tandem	15s	-	E <sub>v</sub> , B	-
Obayashi <i>et al.</i> <sup>76</sup>	192	Eldery	Actiwatch 2	Wrist	Single	1min	-	E <sub>v</sub>	Aggr.
Obayashi <i>et al.</i> <sup>77</sup>	192	Eldery	Actiwatch 2	Wrist	Single	1min	-	E <sub>v</sub>	Aggr.
Okudaira <i>et al.</i> <sup>10</sup>	10	General	Custom	Eye, Wrist	Tandem	5min	-	E <sub>v</sub>	-
Papantoniou <i>et al.</i> <sup>78</sup>	117	Shift-workers	HOBO Pendant	Chest	None	15s	-	E <sub>v</sub>	-
Pattinson <i>et al.</i> <sup>79</sup>	48	Children	Actiwatch 2	Wrist	Single	1min	-	E <sub>v</sub>	-
Peeters <i>et al.</i> <sup>12</sup>	20	Office-workers	Lightlog	Chest	None	3min	Art.	E <sub>v</sub>	Raw
Phillips <i>et al.</i> <sup>80</sup>	22	Students	MotionLogger-L	Wrist	Single	1min	Art.	E <sub>v</sub>	-
Phillips <i>et al.</i> <sup>13</sup>	56	General pop.	Actiwatch 2, Actiwatch-L, Actiwatch Spectrum, Actiwatch Plus	Wrist	Single	1min	-	E <sub>v</sub>	-
Price <i>et al.</i> <sup>81</sup>	40	Shift-workers	Actiwatch Spectrum	-	Single	-	Art.	mEDI, B	-
Rabstein <i>et al.</i> <sup>82</sup>	100	Shift-workers	LightWatcher	Upper arm	Single	10s	-	E <sub>v</sub> , B	Raw

(Continued)



**Table A1.1. (Continued)**

Authors	N	Sample	Dosimeter	Position	Actigraphy setup	Recording interval	Calibration	Quantities	Log-transformation
Rea <i>et al.</i> <sup>83</sup>	58	Teachers	Daysimeter	Eye	Single	-	No descr.	E <sub>v</sub> , CS	-
Read <i>et al.</i> <sup>17</sup>	112	Children	HOBO Pendant, Actiwatch 2	Chest, Wrist	None	5min, 30s	-	E <sub>v</sub>	-
Refinetti <i>et al.</i> <sup>84</sup>	1887	Hispanic, Latino	Actiwatch Spectrum	Wrist	Single	30s	-	E <sub>v</sub> , R, G, B	-
Reid <i>et al.</i> <sup>85</sup>	54	General pop.	Actiwatch-L	Wrist	Single	2min	-	E <sub>v</sub>	-
Rufange <i>et al.</i> <sup>86</sup>	25	Indoor-, Outdoor-workers	Actiwatch-L	-	Single	-	-	E <sub>v</sub>	Raw
Ruiz <i>et al.</i> <sup>87</sup>	76	General pop.	ActiTrust	Wrist	Single	1min	-	E <sub>v</sub>	-
Savides <i>et al.</i> <sup>11</sup>	10	General pop.	Custom	Eye, Wrist	None	5min	-	E <sub>v</sub>	-
Scheuermaier <i>et al.</i> <sup>88</sup>	44	General pop.	Actiwatch-L	Wrist	Single	1-2min	-	E <sub>v</sub>	Aggr.
Silva <i>et al.</i> <sup>89</sup>	12	Students	Geneactiv	Wrist	Single	1min	-	E <sub>v</sub>	-
Smit <i>et al.</i> <sup>90</sup>	91	Indigenous	Actiwatch 2	Wrist	Single	-	-	E <sub>v</sub>	-
Shochat <i>et al.</i> <sup>91</sup>	77	Eldery	Actillum	Wrist	Single	1min	-	E <sub>v</sub>	-
Shochat <i>et al.</i> <sup>92</sup>	19	Students	Actiwatch-L	Wrist	Single	1min	-	E <sub>v</sub>	Raw
Smolders <i>et al.</i> <sup>93</sup>	42	University staff, Students	Daysimeter	Eye	None	30s	-	E <sub>v</sub> , CS	Aggr.
Staples <i>et al.</i> <sup>94</sup>	23	Eldery	Actiwatch-L (CamNtech)	Wrist	Single	1min	-	E <sub>v</sub>	Raw
Stone <i>et al.</i> <sup>95</sup>	25	Shift-workers	Actiwatch Spectrum	Wrist	Single	1min	-	E <sub>v</sub>	Raw
Stoohard <i>et al.</i> <sup>96</sup>	5	General pop.	Actiwatch Spectrum	Wrist	Single	1min	-	E <sub>v</sub> , R, G, B	-
Sun <i>et al.</i> <sup>97</sup>	163	Cancer patients	Actiwatch-L	Wrist	Single	-	-	E <sub>v</sub>	-
te Lindert <i>et al.</i> <sup>98</sup>	35	Insomnia patients	Daysimeter	Chest	None	1min	-	CS	-
Thome <i>et al.</i> <sup>99</sup>	17	Shift-workers	Actiwatch-L (CamNtech)	-	Single	1min	-	E <sub>v</sub>	-
Thome <i>et al.</i> <sup>100</sup>	34	General pop.	Actiwatch-L (CamNtech)	Wrist	Single	2min	-	E <sub>v</sub> , R, G, B	-
Tsai <i>et al.</i> <sup>101</sup>	22	Mothers, Infants	Actiwatch-L	Wrist, Ankle	Tandem	30s	-	E <sub>v</sub>	Raw
Tsai <i>et al.</i> <sup>102</sup>	26	Infants	Actiwatch-L	Ankle	Single	30s	-	E <sub>v</sub>	Raw
Tsuzuki <i>et al.</i> <sup>103</sup>	8	Eldery	Actiwatch-L	Wrist	Single	1min	-	E <sub>v</sub>	-
Uleganathan <i>et al.</i> <sup>16</sup>	61	Adolescents, Children	Actiwatch 2	Wrist	Single	30s	-	E <sub>v</sub>	-
Ulset <i>et al.</i> <sup>104</sup>	49	Children	Actiwatch 2	Wrist	Single	1min	-	E <sub>v</sub>	Raw
van Duijnhoven <i>et al.</i> <sup>105</sup>	62	Office-workers	Lightlog	Chest	None	5min	Art.	E <sub>v</sub> , B, CCT	-
Vinzio <i>et al.</i> <sup>106</sup>	10	Eldery	Actiwatch-L (CamNtech)	Wrist	Single	-	-	E <sub>v</sub>	-
Wallace-Guy <i>et al.</i> <sup>107</sup>	154	Postmenop- women	Actillum	Wrist	Single	-	-	E <sub>v</sub>	-
Wans <i>et al.</i> <sup>108</sup>	20	General	MotionWatch 8	Wrist	Single	1min	-	E <sub>v</sub>	Raw
Wang <i>et al.</i> <sup>109</sup>	37	Postpart. women	Actillum	Wrist	Single	1min	-	E <sub>v</sub>	Raw
Wilson <i>et al.</i> <sup>110</sup>	24	DSPD patients	Actiwatch-L	Wrist	Single	-	-	E <sub>v</sub>	-
Woelders <i>et al.</i> <sup>111</sup>	20	General pop.	MotionWatch 8	Wrist	Single	1min	-	E <sub>v</sub>	Raw
Wright <i>et al.</i> <sup>112</sup>	8	General pop.	Actiwatch-L	Wrist	Single	-	-	E <sub>v</sub>	-
Youngsted <i>et al.</i> <sup>113</sup>	459	Postmenop. women	Actillum	Wrist	Single	-	No descr.	E <sub>v</sub>	Aggr.
Zeitler <i>et al.</i> <sup>114</sup>	31	Insomnia patients	Actiwatch-L	Wrist	Single	-	-	E <sub>v</sub>	-

Note: E<sub>v</sub> = Illuminance, mEDI = melanopic equivalent daylight illuminance, R = long-wavelength irradiance, G = medium-wavelength irradiance, B = short-wavelength irradiance, M = irradiance at 450-500nm, E<sub>c</sub> = Irradiance weighted by c(λ)<sup>114</sup>, CLA = Circadian Light, CS = Circadian Stimulus, CCT = correlated color temperature, Nat. = naturalistic, Art. = artificial, Aggr. = aggregated data. Empty cells with a hyphen indicate that no information was reported.