# Metabolic Messengers: bile acids

- 2 Alessia Perino<sup>1</sup> and Kristina Schoonjans<sup>1\*</sup>
- 3 <sup>1</sup>Institute of Bioengineering, Faculty of Life Science, Ecole Polytechnique Fédérale de
- 4 Lausanne, CH-1015 Lausanne, Switzerland.
- 5 \*To whom correspondence should be addressed: Kristina Schoonjans, EPFL, SV IBI
- 6 UPSCHOONJANS, Station 15, CH-1015 Lausanne, Switzerland. Tel: +41 21 693 18 91.
- 7 Email: <u>kristina.schoonjans@epfl.ch</u>
- 8 ORCID: 0000-0002-5434-3266 (A.P.); 0000-0003-1247-4265 (K.S.)

9

10

1

#### Abstract

- Bile acids (BAs) are amphipathic steroids whose production and diversity depend on both
- 12 host and microbial metabolism. These metabolites have emerged as biologically active
- 13 signaling molecules informing organs of nutrient availability. Their actions, through
- 14 activation of the dedicated BA receptors, FXR and TGR5, control the body's integrated
- 15 physiological metabolic responses. Alterations in BA abundance or signaling are associated
- 16 with multiple metabolic diseases including obesity, type 2 diabetes, non-alcoholic
- 17 steatohepatitis, and atherosclerosis. Consequently, modulation of the BA pool could be a
- 18 valid therapeutic approach, as demonstrated in preclinical and clinical models. Here we
- 19 provide a historical summary of the discovery of BAs and their receptors, as well as on the
- 20 role of BA signaling in the control of energy homeostasis.

21

22

#### History of bile and bile acids: from bodily fluids to hormones

Bile — the fluid in which bile acids (BAs) are stored — has been studied for centuries for its unique beneficial properties. While the first written record depicting bile as a therapeutic dates back to the ancient Egyptian period (Ebers Papyrus, 1550 B.C.)<sup>1</sup>, it came to prominence with Hippocrates (460-377 B.C.), who postulated bile as one of the four bodily fluids, the "humours". Humoral imbalances were the basis of all diseases and harmonizing the four humors was the main therapeutic approach of the time and one of the cornerstones of traditional Chinese medicine, which proposed the extraction of bile from different animals to treat multiple diseases<sup>2</sup>. However, the biochemical and molecular basis by which BAs govern health and disease was only addressed in the last 150 years (extensively reviewed elsewhere<sup>3–</sup> <sup>5</sup>) (Figure 1). BAs were isolated and purified in the second half of the 19<sup>th</sup> century, but the greatest achievements in the BA field were obtained in the 20<sup>th</sup> century. In the early 1930s, several laboratories began to elucidate the chemical structure of BAs. In the following years, other breakthroughs followed, including the development of the chromatographic separation of BAs, the discovery of cholesterol as substrate for primary BA synthesis in the liver, and the identification of secondary BAs as intestinal derivatives from primary BAs in both rats and humans. These studies were instrumental in the identification of the key intermediates, enzymes and mechanisms controlling BA biosynthesis<sup>3-5</sup>. Moreover, they considerably advanced our knowledge on BA diversity and BA recirculation, two processes that are described in the next sections. In the last three decades, several discoveries have led to a better understanding of the mechanisms of action of BAs. In 1999, three different research groups cloned and identified farnesoid X receptor (FXR; NR1H4) as the chief nuclear receptor responsive to BAs<sup>6-8</sup>. Shortly after, BAs were described as agonists of the G protein-coupled receptor (GPCR) Takeda G-protein receptor 5 (TGR5; GPBAR1)<sup>9,10</sup>. Another major advance involved the chemical modification of natural BAs into semi-synthetic analogs with enhanced selectivity

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

for a specific BA receptor<sup>11–14</sup>. These receptor-tailored molecules have moved the field forward, not only because of their utility as research tools to dissect the functions of FXR and TGR5, but also because they laid the foundation for biomedical discoveries and therapeutic opportunities. Among all the semi-synthetic BA derivatives, obeticholic acid was the first to reach the clinic as an FDA-approved FXR agonist for the treatment of primary biliary cholangitis in patients who are unresponsive to the hydrophilic BA specimen, ursodeoxycholic acid<sup>15</sup>. Of note, these semi-synthetic molecules, as well as BAs *per se*, can also trigger non-receptor-mediated effects<sup>16</sup>, which due to space constraints will not be described here.

## **Biology of BAs**

BAs are the main constituents of bile, which also contains cholesterol, phospholipids, bilirubin, fatty acids, vitamins and minerals<sup>17</sup>. Originally identified as amphipathic steroid metabolites facilitating intestinal absorption of lipids and fat-soluble vitamins, they are now recognized as true hormones capable of reaching virtually every organ of our body to fine-tune metabolic functions. BAs are initially synthesized from cholesterol in hepatocytes through two different well-characterized metabolic pathways, comprised of multiple enzymes (Figure 3)<sup>18</sup>. BAs can be synthesized through two distinct pathways. Under physiological conditions, the bulk of primary BAs is produced by the classical pathway initiated by cholesterol  $7\alpha$ -hydroxylase (CYP7A1). The alternative branch of BA production, which is dependent on sterol 27-hydroxylase (CYP27A1), contributes to BA synthesis to a minor extent, but becomes important in response to environmental stresses to mediate adaptive responses<sup>19</sup>. Together, these pathways contribute to the hepatic abundance of the primary BAs, chenodeoxycholic acid (CDCA) and cholic acid (CA), with oxysterol  $12\alpha$ -hydroxylase (CYP8B1) being the enzyme that determines the abundance of CA versus CDCA (Figure 3).

In rodents, CDCA is rapidly converted to the more hydrophilic muricholic acids (MCAs). Primary BAs are then conjugated in the liver to either taurine or glycine, mainly in mice and humans respectively, secreted into bile and released into the intestinal lumen after food ingestion. Once in the gut, BAs are bio-transformed into secondary BAs (deoxycholic acid (DCA), lithocholic acid (LCA),  $\omega$ MCA, and hyodeoxycholic acid (HDCA)) by the intestinal microbiota through different reactions including deconjugation,  $7\alpha$ -dehydroxylation,  $6\alpha$ -hydroxylation and epimerization<sup>20</sup>. When BAs reach the distal part of the small intestine, only a minor fraction transits through the colon and is excreted in the feces. The bulk of BAs is reabsorbed by the gut epithelium and returns to the liver via the enterohepatic circulation. During this process, BAs can spill over into the systemic circulation and signal energy availability to almost all organs through the binding and activation of dedicated BA receptors.

### **BA** receptors

BA receptors trigger genomic and non-genomic adaptive responses in target tissues following changes in BA pool size and/or composition. While several BA receptors have been identified over the last 3 decades<sup>21</sup>, FXR and TGR5 have been particularly well studied for their regulatory role in metabolic health and disease<sup>21–28</sup>, and their characterization has led to the notion that BA species bind to and activate receptors with distinct potencies *in vitro*. However, it is important to emphasize that the extent of receptor activation *in vivo* is largely determined by the total BA pool composition, composed of weak and strong agonists and antagonists.

FXR is abundant in the liver and intestine but also found in other metabolic tissues. Originally discovered as a farnesol receptor<sup>29</sup>, FXR represents the first nuclear receptor that confers BA responsiveness<sup>6–8</sup>. CDCA is a potent natural agonist for FXR and binding to the FXR-retinoic acid receptor α (RXRα) complex activates the transcription of its target

genes<sup>23</sup>. This process is further modulated by the association of coregulators of the FXR-

RXRα complex and by post-translational modifications of FXR itself, as described

elsewhere <sup>30,31</sup>.

While FXR coordinates many of the transcriptional programs elicited by BAs, TGR5

mediates the rapid non-genomic effects. TGR5 is a member of the Rhodopsin-like subfamily

of GPCRs and is modestly expressed in almost every tissue, with the exception of the

gallbladder epithelium where it is highly abundant<sup>32</sup>. Secondary BAs are the most potent

agonists for TGR5 and their conjugation to taurine or glycine further increases their potency<sup>9</sup>.

Binding of BAs to TGR5 triggers a Gas-adenylate cyclase-cAMP signaling cascade resulting

in the activation of multiple downstream targets<sup>21</sup>.

BAs are thus dual agonists for FXR and TGR5 and their relative abundance, along with their

different spatial expression pattern, will ultimately determine receptor activity and biological

response. As such, TGR5 and FXR often complement each other in multiple organs and are

required to orchestrate whole-body metabolism, as described in the following sections.

# Regulation of BA production and flow

Despite the multiple benefits of BAs described below, levels need to be maintained in place to avoid BA accumulation and toxicity. FXR is the master regulator of BA homeostasis and controls BA levels by acting on multiple target genes in enterohepatic organs. For instance, hepatic BA synthesis is controlled by feedback mechanisms involving the FXR-mediated induction of Fibroblast Growth Factor 15/19 (FGF15/19) in the small intestine<sup>33</sup> and the orphan nuclear receptor/transcriptional corepressor small heterodimer partner (SHP; NR0B2)<sup>34,35</sup> in the liver. In addition, FXR regulates the expression of dedicated BA transporters, thereby controlling bile formation and BA recycling<sup>36</sup>. A further level of complexity is given by the TGR5-dependent regulation of bile secretion and flow in

cholangiocytes and gallbladder epithelium<sup>37</sup>. Disruption of these control mechanisms results in altered BA size and composition, and can lead to the development of metabolic diseases<sup>23,24,38</sup> and cancer<sup>38</sup>.

126

127

128

129

130

131

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

147

123

124

125

### **BAs and microbiome**

BA homeostasis is tightly controlled by the gut microbiome through a complex interaction that leads to modulation of whole-body metabolism, as extensively described elsewhere<sup>39,40</sup>. As a direct consequence of their detergent properties, BAs act as antimicrobial molecules capable of altering the microbial ecology of the gut. Conversely, microbial metabolism of BAs increases the diversity of the BA pool<sup>41</sup>, as demonstrated by enrichment of primary conjugated BAs in germ-free (GF) and antibiotic-treated mice. These changes in BA composition impact FXR and TGR5 activation. For example, the absence of microbiota in GF mice has been associated with an increase in BAs acting as FXR antagonists, such as tauro-βMCA (TβMCA)<sup>42</sup>. Diet and pharmacological interventions also regulate FXR activation by modulating the levels of *Lactobacillus*, a bacterial strain expressing bile salt hydrolase, which catalyzes the deconjugation of BAs, including TβMCA<sup>24</sup>. More recently, the gut microbiome has been shown to be required for the production of low abundant phenylalanine- and tyrosine-conjugated CA derivatives, which were proposed to act as FXR agonists<sup>43</sup>. Further studies are warranted to explore the physiological relevance of this subset of conjugated BAs in humans. The role of the gut microbiome in the generation of TGR5 agonists has been well explored. In particular, 7α-dehydroxylating bacteria transform the primary BAs CA and CDCA into DCA and LCA respectively 44,45. Diet composition (fat and protein amount) and pharmacological interventions (antibiotics) strongly influence the availability of TGR5 endogenous agonists, in part by modulating the proportion of intestinal  $7\alpha$ -dehydroxylating bacteria<sup>46–49</sup>. A recent study also reported that a decrease in *Bacteroides vulgatus* and an increase in *Ruminococcus torques* abundance induces DCA<sup>50</sup>. Thus, changes in the activity or quantity of these bacterial species could greatly influence host metabolism via TGR5. While these studies are promising, further research will be needed to fully understand the metabolic benefits of microbiome therapeutics in the context of TGR5 and FXR signaling.

### Metabolic actions of BA signaling

Food intake is the main trigger for intestinal BA release, recirculation, and spillover into the systemic circulation. Consequently, BAs have emerged as postprandial messengers that fine-tune whole-body metabolism through the coordinated activation of FXR and TGR5 in multiple organs (Figure 2), as extensively reviewed elsewhere 21,23,28. In addition to its role in maintaining BA homeostasis, FXR regulates the expression of multiple genes involved in hepatic lipid, glucose and amino acid metabolism, thereby orchestrating the postprandial hepatic response. BAs also modulate whole-body glucose homeostasis, for instance by controlling the secretion of intestinal enteroendocrine hormones, including the insulinotropic and satiety-inducing hormone Glucagon-Like Peptide-1 (GLP-1). Furthermore, intestinal BA signaling is essential for nutrient and cholesterol absorption, as well as for maintaining intestinal integrity, regeneration and motility. Any alterations in these homeostatic control mechanisms may result in profound metabolic dysfunctions. Due to space constraints, this review will hereafter focus on the peripheral and central role of receptor-mediated BA signaling in energy homeostasis.

### **BA** signaling and energy homeostasis

- 171 The regulation of energy expenditure and food intake is a tightly controlled biological event,
- but can be compromised by chronic high-fat diet (HFD) feeding or by excess caloric intake.

Energy homeostasis involves complex inter-cellular and intra-organ communication between peripheral tissues and the central nervous system. BAs are established key regulators of the energy balance during diet-induced obesity (DIO) and orchestrate both peripheral and central responses by modulating TGR5 and FXR activity (Figure 3), as described in the next paragraphs.

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

197

173

174

175

176

177

BA-TGR5 signaling in peripheral organs. BAs have been identified as metabolites with potent body weight-reducing properties. In 2006, Watanabe et al. showed that supplementation of CA to a HFD prevented diet-induced increase in adipose mass, and reversed weight gain after obesity was already established<sup>51</sup>. This phenotype was proposed to result from the activation of the TGR5-deiodinase 2 (DIO2) pathway in murine brown adipose tissue (BAT) and in human skeletal muscle myoblasts<sup>51</sup>. The enzyme DIO2 is involved in the cellular production of bioactive thyroid hormone, and increased DIO2 activity has been associated with enhanced mitochondrial function, adaptive thermogenesis and energy expenditure<sup>52</sup>. Like for CA, feeding mice with CDCA also induced body weight loss<sup>53,54</sup>, and enhanced Uncoupling Protein 1 (UCP-1)<sup>54</sup>, one of the best studied mitochondrial uncouplers, and DIO2<sup>53</sup> expression in BAT. A similar CDCA-DIO2 mechanism was shown to increase energy expenditure in humans<sup>55</sup>. The use of TGR5-selective agonists and genetically modified mouse models further established that these anti-obesogenic effects are at least partially dependent on TGR5 activation 14,32,51,53,56-61. While some studies have corroborated these findings and observed increased energy expenditure after BA administration or TGR5 activation 14,51,55,59,61, others, on the contrary, did not 54,62. Similarly, while some studies report that BAs are not associated with energy metabolism in humans <sup>63,64</sup>, others support this association<sup>55,65</sup>. Differences in the experimental interventions, environment and genetics could explain these discrepancies. Indeed, the genetic background

and the endogenous BA pool size have been proposed as factors that likely contribute to the efficacy of the response<sup>62</sup>. These findings underscore the complexity of BAs in coordinating the energy balance and urge for additional studies to comprehensively evaluate the contribution of genetics and diet to this phenotype. Finally, other uncharacterized mechanisms may also contribute to the BA-mediated body weight maintenance and further research is needed to clarify this point in both mice and humans. Separately, recent studies have also supported the role of BA-TGR5 signaling in inducing muscle cell differentiation and hypertrophy, thereby improving muscle function<sup>66</sup> and increasing glucose utilization without affecting energy expenditure or physical activity<sup>67</sup>. On the other hand, endurance exercise was recently demonstrated to increase the circulating levels of LCA and DCA, which has been proposed to contribute to the increased energy expenditure that is seen through TGR5 activation<sup>68</sup>. In addition to their function in BAT and muscle, three independent studies demonstrated that TGR5 activators are potent triggers of beiging in subcutaneous fat (Figure 3)<sup>56,57,69</sup>. Although the contribution of this mechanism to the regulation of whole-body energy homeostasis is still under debate, activation of TGR5 enhanced lipolysis as well as mitochondrial biogenesis and function in DIO mice<sup>56,69</sup>. Interestingly, this TGR5-mediated adipocyte reprogramming is also triggered by cold exposure and is independent of adrenergic stimulation<sup>69</sup>, one of the common signaling pathways for beiging. It will be interesting to investigate whether physiological or pathological fluctuations in endogenous BAs are sufficiently potent to modulate the white-to-brown conversion of adipocytes. Cold exposure, a well-established approach to increase energy homeostasis, not only boosts mitochondrial activity, but also reshapes microbiota composition<sup>70</sup>. Transplantation of coldremodeled microbiota to GF mice improved both energy and glucose homeostasis 70 and was at least in part dependent on changes in the BA pool size and composition 71,72. Cold exposure

198

199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218

219

220

221

stimulated the expression of genes related to BA synthesis, mainly of the alternative pathway<sup>71</sup>, and increased the production of tauro-conjugated BAs<sup>71,72</sup> including the FXR antagonist  $T\beta MCA^{42}$ . On the other hand, deletion of TGR5 in the adipose tissue blunted cold-induced beiging of subcutaneous WAT<sup>69</sup>. Altogether, these changes in BA pool size/composition and their differential impact on TGR5 (upregulation) and FXR (downregulation) activity seem essential for adjusting whole-body energy homeostasis to sustained cold.

230

231

232

233

234

235

236

237

238

239

240

241

242

243

244

245

246

247

223

224

225

226

227

228

229

BA-TGR5 signaling in the control of central energy homeostasis. Although BAs emerged as satiety-inducing factors in 1968<sup>73</sup>, the physiological relevance and mechanisms underlying this action remained unexplored for decades. Several recent studies have provided insight into these aspects (Figure 3). While studying the brain regions that control food intake, Perino et al. discovered that BAs spike in the hypothalamus shortly after feeding to prime the onset of satiety by turning off the hypothalamic orexigenic Agouti-Related Peptide/Neuropeptide Y (AgRP/NPY) neurons<sup>74</sup>. BAs acutely prevent the release of hunger-stimulating AgRP and NPY neuropeptides during the first minutes following binding of TGR5 through Rho/ROCK/actin-remodeling, but then further sustain the repression by blunting their expression<sup>74</sup>. In line with previous studies conducted in mice fed a normal chow diet (CD)<sup>14,32,58</sup>, these homeostatic mechanisms coordinate the physiological transition between fasting and feeding. During HFD feeding, compensatory neuronal circuits are recruited to counteract DIO. In this context of obesity, another report revealed that chronic activation of central BA-TGR5 signaling not only led to reduced food intake, but also to increased energy expenditure<sup>75</sup> (Figure 3), adding an additional layer of complexity to the established knowledge of peripheral TGR5 activation 14,51,69,76-79. Mechanistically, chronic selective central TGR5

activation increased energy expenditure by engaging the sympathetic nervous system<sup>75</sup>. Of note, the enhanced sympathetic tone was not observed in genetic models of obesity<sup>75</sup>, suggesting that dietary lipid-induced stress, rather than obesity itself, activates these peripheral neuronal circuits. Several possibilities may account for this diet-specific phenotype. HFD feeding triggers significant alterations in hypothalamic structure and function, including hypothalamic inflammation<sup>80</sup> and endoplasmic reticulum stress<sup>81</sup>, and may even lead to partial damage of the neuronal projections<sup>80</sup>. However, HFD feeding also profoundly alters the size and composition of the BA pool<sup>49,82,83</sup>, which in turn may modulate hypothalamic BA signaling. More studies are needed to fully address these possibilities. The central anorexigenic action of BAs can be further enhanced by the regulation of fat preference in lingual taste bud cells<sup>59</sup>. These findings, which were shown to be TGR5dependent, may point to an additional mechanism to prevent obesity. TGR5 is also expressed in vagal afferent neurons and colocalizes with cholecystokinin A-receptor (CCK-AR)<sup>84</sup>. Both BAs and CCK are released after a meal, and inhibit food intake and body weight through activation of Pro-opiomelanocortin/Cocaine- and Amphetamine-Regulated Transcript (POMC/CART) hypothalamic neurons<sup>84</sup>, a well-established satiety-inducing neuronal population<sup>85</sup>. In addition, BA-TGR5 signaling coordinates enteroendocrine differentiation<sup>83,86,87</sup> and function, in particular in L cells, for which BAs and TGR5-specific BA derivatives act as secretagogues of GLP-1 and other incretin hormones 14,88,89. Of note, GLP-1 is a well-established regulator of energy homeostasis 90 and likely acts, together with the above described mechanisms, as an integrated response to amplify BA-induced satiety

270

271

272

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265

266

267

268

269

(Figure 3).

**BA-FXR signaling in peripheral organs.** Several groups have investigated the role of BA-FXR signaling in obesity and reported that FXR deficiency decreased body weight gain and

fat mass in both genetic and DIO mouse models (Figure 3)91,92. Deletion of intestinal FXR was sufficient to prevent weight gain 93-95. In contrast, liver-specific FXR knockout mice were not protected against DIO<sup>91</sup>. Despite the general consensus that FXR loss-of-function protects against obesity, the effects of its selective modulation by BAs or synthetic molecules are more controversial. Both FXR antagonists and agonists were recently shown to act as antiobesity molecules. In agreement with the phenotype observed in intestine-specific FXR knockout mice, selective inhibition of FXR signaling by administration of the synthetic Gly-MCA<sup>94</sup> or the natural TβMCA<sup>93,95</sup> protects against DIO by reducing ceramide levels and increasing beiging. However, administration of the gut-restricted agonist fexaramine also attenuates DIO in mice by increasing the thermogenic response in BAT and WAT<sup>96</sup>. In contrast, the function of the synthetic FXR agonist GW4064 in obese mice is less clear and findings diverge, most likely due to different experimental procedures<sup>51,97–99</sup>. The changes in microbiome and BA composition and/or the existence of off-target effects may also explain some of the discrepancies. Of interest, the gut microbiota controls weight gain in an FXRdependent manner 100 and changes in the level of secondary BAs, as was described with fexaramine administration 96,101, could activate TGR5 and contribute to differential modulation of whole-body energy metabolism. In addition, FXR activation induces Tgr5 expression in the intestine 102, which could also contribute to the complexity of BA signaling. In line with these findings, dual FXR/TGR5 agonists, such as INT-767, significantly prevent the development of obesity in wildtype mice<sup>79</sup>. However, other unknown factors may also weigh in, and further studies will be needed to clarify the apparent controversial actions of FXR-modulating molecules.

295

296

297

273

274

275

276

277

278

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

**BA-FXR** signaling in the control of central energy homeostasis. Although FXR expression has been reported in the brain and hypothalamus<sup>103</sup>, its exact function remains ill-

defined. Until recently, the direct role of FXR in the brain was thought to be marginal, as central activation of FXR by intracerebroventricular (icv) administration of GW4064 did not modulate energy metabolism<sup>104</sup>. Instead, the central action of FXR would indirectly depend on the induction of FGF15/19 in the intestine (Figure 3). Consistent with these findings, FGF19 can cross the blood brain barrier<sup>105</sup> and FGF receptors and β-Klotho, the obligate coreceptor mediating FGF15/19 signaling, are expressed in the hypothalamus<sup>106</sup>, supporting a central action of FGF15/19. Accordingly, activation of central FGF15/19 signaling reduces body weight<sup>107–109</sup> and food intake<sup>108,109</sup> in animal models of obesity. A recent report, however, showed that icv delivery of FXR agonists in the hypothalamus can decrease sympathetic tone and energy expenditure in chow diet-fed mice<sup>110</sup>. It will be interesting to evaluate the relative importance of each of these pathways in the context of whole-body energy homeostasis.

## BA metabolism and genetics in obesity

Decades of preclinical research have revealed how BAs counteract several processes that contribute to the onset of obesity and its associated comorbidities, and have led to the concept that BAs could be used as biomarkers for human disease progression. Lessons from mice should nevertheless be taken with caution, as BA metabolism is not perfectly conserved in evolution. For instance, humans lack enzymes that transform secondary back to primary BAs upon return to the liver, and cannot synthesize the endogenous FXR antagonist TβMCA, two features that fundamentally impact TGR5 and FXR signaling.

Monitoring BA levels in patients with obesity is important to understand how patients with metabolic syndrome could benefit from BA-based therapies. Although findings diverge across studies, fasting circulating BAs tend to augment, while the post-prandial increase of BAs seems to be blunted during obesity<sup>23,111</sup>. BAs are known to show large inter-individual

variations. This is not surprising as they represent several dozens of species, each with unique turn-over and properties. These aspects, together with their multi-compartment physiology and complex regulation by diet, genetics, microbiome and disease, render their investigation highly challenging. For long, human studies have failed to establish solid correlations between changes in BA entities and disease development. A major limitation has been the relatively small sample size of patients, since plasma BA concentrations and composition are notoriously known for their high inter-individual variability. Recent GWAS studies are starting to provide crucial insights into the contribution of genetics and the microbiome to individual BA species in human<sup>112,113</sup> and murine<sup>114</sup> obesity. With the identification of hundreds of quantitative trait loci, these *in silico* studies open up an exciting area of research in which the rigorous validation of these genetic correlations could reveal meaningful causality, and ideally lead to personalized therapeutic perspectives to modulate BA levels and, consequently, whole-body metabolism.

### BAs and bariatric surgery

To date, the most promising clinical approach for severe obesity is bariatric surgery (BS), including Roux-en-Y gastric bypass (RYGB), vertical sleeve gastrectomy (VSG) and bile diversion to the ileum (GB-IL). Following these surgical interventions, patients show elevated levels of BAs and incretin hormones in the systemic circulation, as well as gut microbial changes, factors that have been proposed to contribute to the metabolic benefits of BS<sup>115</sup>. The earliest report linking body weight loss to increased BA levels in BS patients dates back to 2009<sup>116</sup>, and more recent studies showed that this BA phenotype is sustainable up to at least 5 years after BS<sup>117</sup>. Others confirmed elevation in BA levels after RYGB but found no correlation with weight<sup>118</sup>. Interestingly, the BS-dependent increase in circulating BAs was independent from caloric restriction<sup>119</sup>. Despite the large number of clinical and

preclinical studies, there is still no unequivocal consensus on the molecular mechanisms underlying BS-induced body weight loss, especially in the context of BA signaling. For instance, a functional FXR signaling is required to maintain the VSG-mediated weight loss in obese mice<sup>120</sup>, but the beneficial effects of VSG were preserved when FXR was selectively deleted in the intestine or liver<sup>121</sup>, suggesting that FXR inactivation and VSG may enhance energy metabolism through yet uncharacterized mechanisms. Recent literature proposed that intestinal FGF15/19 is an important component of the metabolic responses to VSG<sup>122</sup>. In contrast, intestinal FXR appears to be sufficient to support GB-IL-dependent weight loss in obese mice<sup>123</sup>, whereas FXR signaling seems to be dispensable in RYGB<sup>124</sup>. Similarly, the role of TGR5 signaling in regulating energy metabolism after BS is debated. While some studies demonstrated that intact TGR5 signaling is required for weight loss after RYGB<sup>125</sup> and VSG<sup>78</sup>, others have suggested that RYGB and GB-IL decreased body weight independently of TGR5<sup>123,126</sup>. Altogether, these studies suggest divergent molecular mechanisms following different bariatric interventions.

### **Conclusion and future perspectives**

BAs are peripheral and central regulators of energy homeostasis, and alterations in the BA pool size and composition, or in their biological actions, may shift the balance toward obesity. Despite the extensive characterization of the functions of BAs and their receptors in both preclinical models and patients, multiple aspects of BA signaling remain unknown, and BA analogs or drugs targeting BA signaling are not yet available in the clinic for the treatment of obesity and eating disorders. Studies aimed at increasing the accessibility of BAs to the brain could open up exciting therapeutic perspectives. In addition, modulating the size and composition of the peripheral BA pool by altering the gut microbiota through a change in diet, exercise, cold exposure, antibiotics, probiotics or fecal transplantation could be a

- 373 promising strategy to modulate energy homeostasis. More studies, especially in humans, will
- be needed to validate these preclinical findings into effective therapies.

### Figure legends

Figure 1. Timeline of major discoveries in the BA field. The beneficial effects of bile in the treatment of multiple diseases were already known in ancient times, but the discovery of the chemistry and mechanisms of action of BAs have only been elucidated in the last 150 years, as illustrated in this timeline. B.C., before Christ; BAs, bile acids; UDCA, ursodeoxycholic acid; CDCA, chenodeoxycholic acid; PBC, primary biliary cholangitis; FXR, farnesoid X receptor; TGR5, Takeda G-protein receptor 5; FDA, food and drug administration.

**Figure 2. Target tissues and biological activity of BAs.** TGR5 and FXR coordinate the BA-mediated whole-body metabolic response by acting in multiple organs. The biological functions listed in this figure are non-exhaustive and the receptor indicated in parentheses represents the most studied one. Most observations on the biological action of the BA-TGR5 signaling have been made in mice, but are supported by *in vitro* studies in human cells and/or by clinical data. TGR5, Takeda G-protein receptor 5; HFD, high-fat diet; FXR, farnesoid X receptor.

**Figure 3. BA signaling controls whole-body energy homeostasis through direct and indirect mechanisms.** a) Genetic deletion of FXR (*Nr1h4*) or TGR5 (*Gpbar1*) in obese mice is sufficient to alter body weight gain. b) BAs activate FXR and TGR5 in enterocytes and enteroendocrine cells to indirectly modulate food intake through increased secretion of FGF15/19 and GLP-1, respectively. c) Primary BAs are produced in the liver through the classical or alternative pathways (main enzymes are illustrated in the box) and converted to secondary BAs in the intestine by the gut microbiome. Dashed arrow: not fully defined reaction as deduced from 127. d) BAs reach peripheral organs via the systemic circulation and orchestrate whole-body energy homeostasis by directly activating TGR5 in the brain, BAT

and WAT. Primary (yellow) and secondary (red) BAs can activate both TGR5 and FXR but with different potency (higher potency = more BAs of a color near the receptor). This figure represents an overview and is not exhaustive. Nr1h4, nuclear receptor subfamily 1 group H member 4; Gpbar1, G protein-coupled bile acid receptor 1; FXR, farnesoid X receptor; KO, knockout; TGR5, Takeda G-protein receptor 5; FGF15/19, fibroblast Growth Factor-15/19; EEC, enteroendocrine cell; GLP-1, glucagon-like peptide 1; BA, bile acid; CYP7A1, cholesterol 7α-hydroxylase; CYP8B1, sterol 12α-hydroxylase; CYP27A1, sterol 27-CYP7B1, hydroxylase; oxysterol 7α-hydroxylase; CA, cholic acid; CDCA, chenodeoxycholic acid; MCA, muricholic acids; BAT, brown adipose tissue; WAT, white adipose tissue. Created with BioRender.com.

410

411

412

413

414

415

400

401

402

403

404

405

406

407

408

409

- **Acknowledgements.** We thank Maroun Bou Sleiman for assistance with the figures. The authors were supported by the Swiss National Science Foundation (SNSF N° 310030\_189178 and CRSII5\_180317/1, the Kristian Gerhard Jebsen Foundation, and the Ecole Polytechnique Fédérale de Lausanne (EPFL) (to K.S.), CAIXA (N° HR17-00601) (to K.S.) and a postdoctoral fellowship from AXA Research Fund (to A.P.).
- 416 **Author contributions.** A.P. and K.S. wrote the article.
- 417 **Competing interests.** The authors declare no competing interests.
- 418 **Additional information.** Correspondence should be addressed to K.S.

419

- 421 **References**
- 422 1. Bryan, C.P. *The Papyrus Ebers*. (New York: Appleton, 1931).
- 423 2. Wang, D.Q.H. & Carey, M.C. Therapeutic uses of animal biles in traditional Chinese
- medicine: an ethnopharmacological, biophysical chemical and medicinal review. World
- 425 *J. Gastroenterol.* **20**, 9952–9975 (2014).
- 3. Russell, D.W. Fifty years of advances in bile acid synthesis and metabolism. *J. Lipid Res.*
- **50 Suppl**, S120-125 (2009).
- 428 4. Sjövall, J. Fifty years with bile acids and steroids in health and disease. *Lipids* **39**, 703–722
- 429 (2004).
- 430 5. Hofmann, A.F. & Hagey, L.R. Key discoveries in bile acid chemistry and biology and
- their clinical applications: history of the last eight decades. *J. Lipid Res.* **55**, 1553–1595
- 432 (2014).
- 6. Makishima, M. et al. Identification of a nuclear receptor for bile acids. Science 284, 1362–
- 434 1365 (1999).
- 435 7. Wang, H., Chen, J., Hollister, K., Sowers, L.C. & Forman, B.M. Endogenous bile acids are
- ligands for the nuclear receptor FXR/BAR. *Mol. Cell* **3**, 543–553 (1999).
- 8. Parks, D.J. et al. Bile acids: natural ligands for an orphan nuclear receptor. Science 284,
- 438 1365–1368 (1999).
- 9. Kawamata, Y. et al. A G protein-coupled receptor responsive to bile acids. J. Biol. Chem.
- **278**, 9435–9440 (2003).
- 441 10. Maruyama, T. et al. Identification of membrane-type receptor for bile acids (M-BAR).
- 442 *Biochem. Biophys. Res. Commun.* **298**, 714–719 (2002).
- 443 11. Fiorucci, S. et al. The nuclear receptor SHP mediates inhibition of hepatic stellate cells
- by FXR and protects against liver fibrosis. *Gastroenterology* **127**, 1497–1512 (2004).

- 445 12. Pellicciari, R. et al. Discovery of 6alpha-ethyl-23(S)-methylcholic acid (S-EMCA, INT-
- 446 777) as a potent and selective agonist for the TGR5 receptor, a novel target for
- 447 diabesity. J. Med. Chem. **52**, 7958–7961 (2009).
- 448 13. Pellicciari, R. et al. 6alpha-ethyl-chenodeoxycholic acid (6-ECDCA), a potent and
- selective FXR agonist endowed with anticholestatic activity. J. Med. Chem. 45, 3569–
- 450 3572 (2002).
- 451 14. Thomas, C. et al. TGR5-mediated bile acid sensing controls glucose homeostasis. Cell
- 452 *Metab.* **10**, 167–177 (2009).
- 453 15. Trauner, M. et al. Long-term efficacy and safety of obeticholic acid for patients with
- primary biliary cholangitis: 3-year results of an international open-label extension study.
- 455 *Lancet Gastroenterol. Hepatol.* **4**, 445–453 (2019).
- 456 16. Ahmad, T.R. & Haeusler, R.A. Bile acids in glucose metabolism and insulin signalling -
- mechanisms and research needs. *Nat. Rev. Endocrinol.* **15**, 701–712 (2019).
- 458 17. Boyer, J.L. Bile formation and secretion. *Compr. Physiol.* **3**, 1035–1078 (2013).
- 459 18. Russell, D.W. The enzymes, regulation, and genetics of bile acid synthesis. *Annu. Rev.*
- 460 *Biochem.* **72**, 137–174 (2003).
- 461 19. Jia, W., Wei, M., Rajani, C. & Zheng, X. Targeting the alternative bile acid synthetic
- pathway for metabolic diseases. *Protein Cell* **12**, 411–425 (2021).
- 463 20. Guzior, D.V. & Quinn, R.A. Review: microbial transformations of human bile acids.
- 464 *Microbiome* **9**, 140 (2021).
- 465 21. Perino, A., Demagny, H., Velazquez-Villegas, L.A. & Schoonjans, K. Molecular
- Physiology of Bile Acid Signaling in Health, Disease and Aging. *Physiol. Rev.* **101**,
- 467 683–731 (2020).

- 468 22. Arab, J.P., Karpen, S.J., Dawson, P.A., Arrese, M. & Trauner, M. Bile acids and
- nonalcoholic fatty liver disease: Molecular insights and therapeutic perspectives.
- 470 *Hepatology* **65**, 350–362 (2017).
- 471 23. Chávez-Talavera, O., Tailleux, A., Lefebvre, P. & Staels, B. Bile Acid Control of
- 472 Metabolism and Inflammation in Obesity, Type 2 Diabetes, Dyslipidemia, and
- Nonalcoholic Fatty Liver Disease. *Gastroenterology* **152**, 1679-1694.e3 (2017).
- 474 24. Gonzalez, F.J., Jiang, C. & Patterson, A.D. An Intestinal Microbiota-Farnesoid X
- 475 Receptor Axis Modulates Metabolic Disease. *Gastroenterology* **151**, 845–859 (2016).
- 476 25. Hegyi, P., Maléth, J., Walters, J.R., Hofmann, A.F. & Keely, S.J. Guts and Gall: Bile
- 477 Acids in Regulation of Intestinal Epithelial Function in Health and Disease. *Physiol*.
- 478 *Rev.* **98**, 1983–2023 (2018).
- 479 26. Kuipers, F., Bloks, V.W. & Groen, A.K. Beyond intestinal soap--bile acids in metabolic
- 480 control. Nat. Rev. Endocrinol. 10, 488–498 (2014).
- 481 27. Molinaro, A., Wahlström, A. & Marschall, H.U. Role of Bile Acids in Metabolic
- 482 Control. *Trends Endocrinol. Metab.* **29**, 31–41 (2018).
- 483 28. Perino, A. & Schoonjans, K. TGR5 and Immunometabolism: Insights from Physiology
- 484 and Pharmacology. *Trends Pharmacol. Sci.* **36**, 847–857 (2015).
- 485 29. Forman, B.M. et al. Identification of a nuclear receptor that is activated by farnesol
- 486 metabolites. *Cell* **81**, 687–693 (1995).
- 487 30. Appelman, M.D., van der Veen, S.W. & van Mil, S.W.C. Post-Translational
- 488 Modifications of FXR; Implications for Cholestasis and Obesity-Related Disorders.
- 489 Front. Endocrinol. **12**, 729828 (2021).
- 490 31. Kemper, J.K. Regulation of FXR transcriptional activity in health and disease:
- Emerging roles of FXR cofactors and post-translational modifications. *Biochim*.
- 492 *Biophys. Acta* **1812**, 842–850 (2011).

- 493 32. Vassileva, G. et al. Targeted deletion of Gpbar1 protects mice from cholesterol gallstone
- 494 formation. *Biochem. J.* **398**, 423–430 (2006).
- 495 33. Gadaleta, R.M. & Moschetta, A. Metabolic Messengers: fibroblast growth factor 15/19.
- 496 *Nat. Metab.* **1**, 588–594 (2019).
- 497 34. Goodwin, B. et al. A regulatory cascade of the nuclear receptors FXR, SHP-1, and
- 498 LRH-1 represses bile acid biosynthesis. *Mol. Cell* **6**, 517–526 (2000).
- 499 35. Lu, T.T. et al. Molecular basis for feedback regulation of bile acid synthesis by nuclear
- 500 receptors. *Mol. Cell* **6**, 507–515 (2000).
- 501 36. Halilbasic, E., Claudel, T. & Trauner, M. Bile acid transporters and regulatory nuclear
- receptors in the liver and beyond. *J. Hepatol.* **58**, 155–168 (2013).
- 503 37. Keitel, V. & Häussinger, D. Role of TGR5 (GPBAR1) in Liver Disease. Semin. Liver
- 504 Dis. 38, 333–339 (2018).
- 38. Sun, L., Cai, J. & Gonzalez, F.J. The role of farnesoid X receptor in metabolic diseases,
- and gastrointestinal and liver cancer. Nat. Rev. Gastroenterol. Hepatol. 18, 335–347
- 507 (2021).
- 39. Wahlström, A., Sayin, S.I., Marschall, H.U. & Bäckhed, F. Intestinal Crosstalk between
- Bile Acids and Microbiota and Its Impact on Host Metabolism. *Cell Metab.* **24**, 41–50
- 510 (2016).
- 511 40. Fiorucci, S. & Distrutti, E. Bile Acid-Activated Receptors, Intestinal Microbiota, and the
- Treatment of Metabolic Disorders. *Trends Mol. Med.* **21**, 702–714 (2015).
- 513 41. Winston, J.A. & Theriot, C.M. Diversification of host bile acids by members of the gut
- 514 microbiota. *Gut Microbes* **11**, 158–171 (2020).
- 515 42. Sayin, S.I. et al. Gut microbiota regulates bile acid metabolism by reducing the levels of
- tauro-beta-muricholic acid, a naturally occurring FXR antagonist. Cell Metab. 17, 225–
- 517 235 (2013).

- 518 43. Quinn, R.A. et al. Global chemical effects of the microbiome include new bile-acid
- 519 conjugations. *Nature* **579**, 123–129 (2020).
- 520 44. Funabashi, M. et al. A metabolic pathway for bile acid dehydroxylation by the gut
- 521 microbiome. *Nature* **582**, 566–570 (2020).
- 522 45. Marion, S. et al. In vitro and in vivo characterization of Clostridium scindens bile acid
- 523 transformations. *Gut Microbes* **10**, 481–503 (2019).
- 524 46. Fujisaka, S. et al. Antibiotic effects on gut microbiota and metabolism are host
- 525 dependent. J. Clin. Invest. 126, 4430–4443 (2016).
- 526 47. Watanabe, K. et al. Dietary soybean protein ameliorates high-fat diet-induced obesity by
- modifying the gut microbiota-dependent biotransformation of bile acids. *PloS One* **13**,
- 528 e0202083 (2018).
- 529 48. Devkota, S. & Chang, E.B. Interactions between Diet, Bile Acid Metabolism, Gut
- Microbiota, and Inflammatory Bowel Diseases. *Dig. Dis.* **33**, 351–356 (2015).
- 531 49. Just, S. et al. The gut microbiota drives the impact of bile acids and fat source in diet on
- mouse metabolism. *Microbiome* **6**, 134 (2018).
- 533 50. Wu, Q. et al. Intestinal hypoxia-inducible factor 2α regulates lactate levels to shape the
- gut microbiome and alter thermogenesis. *Cell Metab.* **33**, 1988-2003.e7 (2021).
- 535 51. Watanabe, M. et al. Bile acids induce energy expenditure by promoting intracellular
- thyroid hormone activation. *Nature* **439**, 484–489 (2006).
- 537 52. Bianco, A.C. & Kim, B.W. Deiodinases: implications of the local control of thyroid
- 538 hormone action. *J. Clin. Invest.* **116**, 2571–2579 (2006).
- 539 53. Chen, X. et al. Chenodeoxycholic acid attenuates high-fat diet-induced obesity and
- 540 hyperglycemia via the G protein-coupled bile acid receptor 1 and proliferator-activated
- receptor γ pathway. *Exp. Ther. Med.* **14**, 5305–5312 (2017).

- 542 54. Teodoro, J.S. et al. Enhancement of brown fat thermogenesis using chenodeoxycholic
- acid in mice. *Int. J. Obes.* **38**, 1027–1034 (2014).
- 544 55. Broeders, E.P.M. et al. The Bile Acid Chenodeoxycholic Acid Increases Human Brown
- 545 Adipose Tissue Activity. *Cell Metab.* **22**, 418–426 (2015).
- 546 56. Carino, A. et al. Gpbar1 agonism promotes a Pgc-1α-dependent browning of white
- adipose tissue and energy expenditure and reverses diet-induced steatohepatitis in mice.
- 548 *Sci. Rep.* **7**, 13689 (2017).
- 549 57. Carino, A. et al. Agonism for the bile acid receptor GPBAR1 reverses liver and vascular
- damage in a mouse model of steatohepatitis. FASEB J. 33, 2809–2822 (2019).
- 551 58. Maruyama, T. et al. Targeted disruption of G protein-coupled bile acid receptor 1
- 552 (Gpbar1/M-Bar) in mice. *J. Endocrinol.* **191**, 197–205 (2006).
- 553 59. Bensalem, A. et al. Bile acid receptor TGR5 is critically involved in preference for
- dietary lipids and obesity. *J. Nutr. Biochem.* **76**, 108298 (2020).
- 555 60. Briere, D.A. et al. Novel Small Molecule Agonist of TGR5 Possesses Anti-Diabetic
- Effects but Causes Gallbladder Filling in Mice. *PloS One* **10**, e0136873 (2015).
- 557 61. Zietak, M. & Kozak, L.P. Bile acids induce uncoupling protein 1-dependent
- thermogenesis and stimulate energy expenditure at thermoneutrality in mice. Am. J.
- 559 *Physiol. Endocrinol. Metab.* **310**, E346-354 (2016).
- 560 62. Fromme, T. et al. Bile acid supplementation decreases body mass gain in C57BL/6J but
- not 129S6/SvEvTac mice without increasing energy expenditure. Sci. Rep. 9, 131
- 562 (2019).
- 563 63. Brufau, G. et al. Plasma bile acids are not associated with energy metabolism in
- 564 humans. *Nutr. Metab.* **7**, 73 (2010).

- 565 64. van Nierop, F.S. et al. Differential effects of a 40-hour fast and bile acid
- supplementation on human GLP-1 and FGF19 responses. Am. J. Physiol. Endocrinol.
- 567 *Metab.* **317**, E494–E502 (2019).
- 568 65. Ockenga, J. et al. Plasma bile acids are associated with energy expenditure and thyroid
- 569 function in humans. *J. Clin. Endocrinol. Metab.* **97**, 535–542 (2012).
- 570 66. Sasaki, T. et al. The exercise-inducible bile acid receptor Tgr5 improves skeletal muscle
- function in mice. *J. Biol. Chem.* **293**, 10322–10332 (2018).
- 572 67. Sasaki, T. et al. Muscle-specific TGR5 overexpression improves glucose clearance in
- 573 glucose-intolerant mice. *J. Biol. Chem.* **296**, 100131 (2021).
- 574 68. Morville, T. et al. Divergent effects of resistance and endurance exercise on plasma bile
- acids, FGF19, and FGF21 in humans. *JCI Insight* **3**, 122737 (2018).
- 576 69. Velazquez-Villegas, L.A. et al. TGR5 signalling promotes mitochondrial fission and
- beige remodelling of white adipose tissue. *Nat. Commun.* **9**, 245 (2018).
- 578 70. Chevalier, C. et al. Gut Microbiota Orchestrates Energy Homeostasis during Cold. Cell
- **163**, 1360–1374 (2015).
- 580 71. Worthmann, A. et al. Cold-induced conversion of cholesterol to bile acids in mice
- shapes the gut microbiome and promotes adaptive thermogenesis. *Nat. Med.* 23, 839–
- 582 849 (2017).
- 583 72. Zietak, M. et al. Altered Microbiota Contributes to Reduced Diet-Induced Obesity upon
- 584 Cold Exposure. *Cell Metab.* **23**, 1216–1223 (2016).
- 585 73. Bray, G.A. & Gallagher, T.F. Suppression of appetite by bile acids. Lancet 1, 1066–
- 586 1067 (1968).
- 74. Perino, A. et al. Central anorexigenic actions of bile acids are mediated by TGR5. Nat.
- 588 *Metab.* **3**, 595–603 (2021).

- 589 75. Castellanos-Jankiewicz, A. et al. Hypothalamic bile acid-TGR5 signaling protects from
- 590 obesity. *Cell Metab.* **33**, 1483-1492.e10 (2021).
- 591 76. Watanabe, M. et al. Bile acid binding resin improves metabolic control through the
- induction of energy expenditure. *PloS One* **7**, e38286 (2012).
- 593 77. Chen, X. et al. Identification of miR-26a as a target gene of bile acid receptor GPBAR-
- 594 1/TGR5. *PloS One* **10**, e0131294 (2015).
- 595 78. Ding, L. et al. Vertical sleeve gastrectomy activates GPBAR-1/TGR5 to sustain weight
- loss, improve fatty liver, and remit insulin resistance in mice. *Hepatol. Baltim. Md* **64**,
- 597 760–773 (2016).
- 598 79. Jadhav, K. et al. Reversal of metabolic disorders by pharmacological activation of bile
- 599 acid receptors TGR5 and FXR. *Mol. Metab.* **9**, 131–140 (2018).
- 600 80. Thaler, J.P., Guyenet, S.J., Dorfman, M.D., Wisse, B.E. & Schwartz, M.W.
- Hypothalamic inflammation: marker or mechanism of obesity pathogenesis? *Diabetes*
- **602 62**, 2629–2634 (2013).
- 81. Contreras, C. et al. Reduction of Hypothalamic Endoplasmic Reticulum Stress Activates
- Browning of White Fat and Ameliorates Obesity. *Diabetes* **66**, 87–99 (2017).
- 605 82. Fu, T. et al. FXR Regulates Intestinal Cancer Stem Cell Proliferation. Cell 176, 1098-
- 606 1112.e18 (2019).
- 83. Sorrentino, G. et al. Bile Acids Signal via TGR5 to Activate Intestinal Stem Cells and
- Epithelial Regeneration. *Gastroenterology* **159**, 956-968.e8 (2020).
- 84. Wu, X. et al. Satiety induced by bile acids is mediated via vagal afferent pathways. JCI
- 610 *Insight* **5**, (2020).
- 611 85. Fan, W., Boston, B.A., Kesterson, R.A., Hruby, V.J. & Cone, R.D. Role of
- melanocortinergic neurons in feeding and the agouti obesity syndrome. *Nature* **385**,
- 613 165–168 (1997).

- 86. Lund, M.L. et al. L-Cell Differentiation Is Induced by Bile Acids Through GPBAR1
- and Paracrine GLP-1 and Serotonin Signaling. *Diabetes* **69**, 614–623 (2020).
- 87. Harach, T. et al. TGR5 potentiates GLP-1 secretion in response to anionic exchange
- 617 resins. Sci. Rep. 2, 430 (2012).
- 88. Brighton, C.A. et al. Bile Acids Trigger GLP-1 Release Predominantly by Accessing
- Basolaterally Located G Protein-Coupled Bile Acid Receptors. *Endocrinology* **156**,
- 620 3961–3970 (2015).
- 89. Kuhre, R.E. et al. Bile acids are important direct and indirect regulators of the secretion
- of appetite- and metabolism-regulating hormones from the gut and pancreas. *Mol.*
- 623 *Metab.* **11**, 84–95 (2018).
- 624 90. Gribble, F.M. & Reimann, F. Function and mechanisms of enteroendocrine cells and gut
- hormones in metabolism. *Nat. Rev. Endocrinol.* **15**, 226–237 (2019).
- 91. Prawitt, J. et al. Farnesoid X receptor deficiency improves glucose homeostasis in
- mouse models of obesity. *Diabetes* **60**, 1861–1871 (2011).
- 628 92. Zhang, Y. et al. Loss of FXR protects against diet-induced obesity and accelerates liver
- 629 carcinogenesis in ob/ob mice. Mol. Endocrinol. Baltim. Md 26, 272–280 (2012).
- 630 93. Li, F. et al. Microbiome remodelling leads to inhibition of intestinal farnesoid X
- receptor signalling and decreased obesity. *Nat. Commun.* **4**, 2384 (2013).
- 632 94. Jiang, C. et al. Intestine-selective farnesoid X receptor inhibition improves obesity-
- related metabolic dysfunction. *Nat. Commun.* **6**, 10166 (2015).
- 634 95. Xie, C. et al. An Intestinal Farnesoid X Receptor-Ceramide Signaling Axis Modulates
- Hepatic Gluconeogenesis in Mice. *Diabetes* **66**, 613–626 (2017).
- 636 96. Fang, S. et al. Intestinal FXR agonism promotes adipose tissue browning and reduces
- obesity and insulin resistance. *Nat. Med.* **21**, 159–165 (2015).

- 638 97. Cariou, B. et al. The farnesoid X receptor modulates adiposity and peripheral insulin
- 639 sensitivity in mice. *J. Biol. Chem.* **281**, 11039–11049 (2006).
- 98. Ma, Y., Huang, Y., Yan, L., Gao, M. & Liu, D. Synthetic FXR agonist GW4064
- prevents diet-induced hepatic steatosis and insulin resistance. *Pharm. Res.* **30**, 1447–
- 642 1457 (2013).
- 643 99. Watanabe, M. et al. Lowering bile acid pool size with a synthetic farnesoid X receptor
- 644 (FXR) agonist induces obesity and diabetes through reduced energy expenditure. *J. Biol.*
- 645 *Chem.* **286**, 26913–26920 (2011).
- 646 100. Parséus, A. et al. Microbiota-induced obesity requires farnesoid X receptor. Gut 66,
- 647 429–437 (2017).
- 648 101. Pathak, P. et al. Intestine farnesoid X receptor agonist and the gut microbiota activate G-
- protein bile acid receptor-1 signaling to improve metabolism. Hepatology 68, 1574–
- 650 1588 (2018).
- 651 102. Pathak, P. et al. Farnesoid X receptor induces Takeda G-protein receptor 5 cross-talk to
- regulate bile acid synthesis and hepatic metabolism. J. Biol. Chem. 292, 11055–11069
- 653 (2017).
- 654 103. Huang, C. et al. Identification of functional farnesoid X receptors in brain neurons.
- 655 FEBS Lett. **590**, 3233–3242 (2016).
- 656 104. Eggink, H.M. et al. Chronic infusion of taurolithocholate into the brain increases fat
- oxidation in mice. *J. Endocrinol.* **236**, 85–97 (2018).
- 658 105. Hsuchou, H., Pan, W. & Kastin, A.J. Fibroblast growth factor 19 entry into brain. *Fluids*
- 659 Barriers CNS 10, 32 (2013).
- 106. Hultman, K. et al. The central fibroblast growth factor receptor/beta klotho system:
- Comprehensive mapping in Mus musculus and comparisons to nonhuman primate and

- human samples using an automated in situ hybridization platform. J. Comp. Neurol.
- **527**, 2069–2085 (2019).
- 664 107. Lan, T. et al. FGF19, FGF21, and an FGFR1/β-Klotho-Activating Antibody Act on the
- Nervous System to Regulate Body Weight and Glycemia. *Cell Metab.* **26**, 709-718.e3
- 666 (2017).
- 108. Marcelin, G. et al. Central action of FGF19 reduces hypothalamic AGRP/NPY neuron
- activity and improves glucose metabolism. *Mol. Metab.* **3**, 19–28 (2014).
- 669 109. Ryan, K.K. et al. Fibroblast growth factor-19 action in the brain reduces food intake and
- body weight and improves glucose tolerance in male rats. *Endocrinology* **154**, 9–15
- 671 (2013).
- 672 110. Deckmyn, B. et al. Farnesoid X Receptor Activation in Brain Alters Brown Adipose
- Tissue Function via the Sympathetic System. Front. Mol. Neurosci. 14, 808603 (2021).
- 111. Chávez-Talavera, O., Haas, J., Grzych, G., Tailleux, A. & Staels, B. Bile acid alterations
- in nonalcoholic fatty liver disease, obesity, insulin resistance and type 2 diabetes: what
- do the human studies tell? *Curr. Opin. Lipidol.* **30**, 244–254 (2019).
- 677 112. Chen, L. et al. Genetic and Microbial Associations to Plasma and Fecal Bile Acids in
- Obesity Relate to Plasma Lipids and Liver Fat Content. *Cell Rep.* **33**, 108212 (2020).
- 679 113. Wang, D. et al. Characterization of gut microbial structural variations as determinants of
- human bile acid metabolism. *Cell Host Microbe* **29**, 1802-1814.e5 (2021).
- 681 114. Kemis, J.H. et al. Genetic determinants of gut microbiota composition and bile acid
- profiles in mice. *PLoS Genet.* **15**, e1008073 (2019).
- 683 115. Evers, S.S., Sandoval, D.A. & Seeley, R.J. The Physiology and Molecular
- Underpinnings of the Effects of Bariatric Surgery on Obesity and Diabetes. *Annu. Rev.*
- 685 *Physiol.* **79**, 313–334 (2017).

- 686 116. Patti, M.E. et al. Serum bile acids are higher in humans with prior gastric bypass:
- potential contribution to improved glucose and lipid metabolism. Obesity 17, 1671-
- 688 1677 (2009).
- 689 117. Risstad, H. et al. Bile acid profiles over 5 years after gastric bypass and duodenal
- switch: results from a randomized clinical trial. Surg. Obes. Relat. Dis. 13, 1544–1553
- 691 (2017).
- 692 118. Nemati, R. et al. Increased Bile Acids and FGF19 After Sleeve Gastrectomy and Roux-
- 693 en-Y Gastric Bypass Correlate with Improvement in Type 2 Diabetes in a Randomized
- 694 Trial. Obes. Surg. 28, 2672–2686 (2018).
- 695 119. Jahansouz, C. et al. Bile Acids Increase Independently From Hypocaloric Restriction
- 696 After Bariatric Surgery. *Ann. Surg.* **264**, 1022–1028 (2016).
- 697 120. Ryan, K.K. et al. FXR is a molecular target for the effects of vertical sleeve
- 698 gastrectomy. *Nature* **509**, 183–188 (2014).
- 699 121. Ding, L. et al. Vertical sleeve gastrectomy confers metabolic improvements by reducing
- intestinal bile acids and lipid absorption in mice. Proc. Natl. Acad. Sci. U. S. A. 118,
- 701 e2019388118 (2021).
- 702 122. Bozadjieva-Kramer, N. et al. Intestinal-derived FGF15 protects against deleterious
- effects of vertical sleeve gastrectomy in mice. *Nat. Commun.* **12**, 4768 (2021).
- 704 123. Albaugh, V. L. et al. Role of Bile Acids and GLP-1 in Mediating the Metabolic
- Improvements of Bariatric Surgery. *Gastroenterology* **156**, 1041-1051.e4 (2019).
- 706 124. Li, K. et al. Farnesoid X receptor contributes to body weight-independent improvements
- in glycemic control after Roux-en-Y gastric bypass surgery in diet-induced obese mice.
- 708 *Mol. Metab.* **37**, 100980 (2020).

- 709 125. Yang, H., Liu, H., Jiao, Y. & Qian, J. Roux-en-Y Gastrointestinal Bypass Promotes
- 710 Activation of TGR5 and Peptide YY. Endocr. Metab. Immune Disord. Drug Targets 20,
- 711 1262–1267 (2020).
- 712 126. Hao, Z. et al. Roux-en-Y Gastric Bypass Surgery-Induced Weight Loss and Metabolic
- 713 Improvements Are Similar in TGR5-Deficient and Wildtype Mice. Obes. Surg. 28,
- 714 3227–3236 (2018).

- 715 127. Ferrell, J.M., Boehme, S., Li, F. & Chiang, J.Y.L. Cholesterol 7α-hydroxylase-deficient
- mice are protected from high-fat/high-cholesterol diet-induced metabolic disorders. J.
- 717 *Lipid Res.* **57**, 1144–1154 (2016).





