



Diversity of Reactive Astroglia in CNS Pathology: Heterogeneity or Plasticity?

Aaron J. Moulson^{1,2†}, Jordan W. Squair³, Robin J. M. Franklin⁴, Wolfram Tetzlaff^{2,5,6} and Peggy Assinck^{4,7*†}

¹ Faculty of Medicine, University of British Columbia, Vancouver, BC, Canada, ² International Collaboration on Repair Discoveries (ICORD), Vancouver, BC, Canada, ³ Department of Clinical Neuroscience, Faculty of Life Sciences, Center for Neuroprosthetics and Brain Mind Institute, École Polytechnique Fédérale de Lausanne (EPFL), NeuroRestore, Lausanne University Hospital (CHUV), University of Lausanne (UNIL), Lausanne, Switzerland, ⁴ Wellcome Trust - MRC Cambridge Stem Cell Institute, University of Cambridge, Cambridge, United Kingdom, ⁵ Department of Zoology, University of British Columbia, Vancouver, BC, Canada, ⁶ Department of Surgery, University of British Columbia, Vancouver, BC, Canada, ⁷ Centre for Regenerative Medicine, Institute for Regeneration and Repair, University of Edinburgh, Edinburgh, United Kingdom

OPEN ACCESS

Edited by:

Dimitrios Davalos,
Case Western Reserve University,
United States

Reviewed by:

Shane A. Liddelow,
New York University, United States
Jessica L. Williams,
Cleveland Clinic, United States

*Correspondence:

Peggy Assinck
pa465@cam.ac.uk

[†] These authors have contributed
equally to this work and share first
authorship

Specialty section:

This article was submitted to
Non-Neuronal Cells,
a section of the journal
Frontiers in Cellular Neuroscience

Received: 30 April 2021

Accepted: 02 July 2021

Published: 26 July 2021

Citation:

Moulson AJ, Squair JW,
Franklin RJM, Tetzlaff W and
Assinck P (2021) Diversity of Reactive
Astroglia in CNS Pathology:
Heterogeneity or Plasticity?
Front. Cell. Neurosci. 15:703810.
doi: 10.3389/fncel.2021.703810

Astrocytes are essential for the development and homeostatic maintenance of the central nervous system (CNS). They are also critical players in the CNS injury response during which they undergo a process referred to as “reactive astroglia.” Diversity in astrocyte morphology and gene expression, as revealed by transcriptional analysis, is well-recognized and has been reported in several CNS pathologies, including ischemic stroke, CNS demyelination, and traumatic injury. This diversity appears unique to the specific pathology, with significant variance across temporal, topographical, age, and sex-specific variables. Despite this, there is limited functional data corroborating this diversity. Furthermore, as reactive astrocytes display significant environmental-dependent plasticity and fate-mapping data on astrocyte subsets in the adult CNS is limited, it remains unclear whether this diversity represents heterogeneity or plasticity. As astrocytes are important for neuronal survival and CNS function post-injury, establishing to what extent this diversity reflects distinct established heterogeneous astrocyte subpopulations vs. environmentally dependent plasticity within established astrocyte subsets will be critical for guiding therapeutic development. To that end, we review the current state of knowledge on astrocyte diversity in the context of three representative CNS pathologies: ischemic stroke, demyelination, and traumatic injury, with the goal of identifying key limitations in our current knowledge and suggesting future areas of research needed to address them. We suggest that the majority of identified astrocyte diversity in CNS pathologies to date represents plasticity in response to dynamically changing post-injury environments as opposed to heterogeneity, an important consideration for the understanding of disease pathogenesis and the development of therapeutic interventions.

Keywords: reactive astrocytes, heterogeneity, plasticity, single-cell RNA sequencing, ischemic stroke, CNS demyelination, traumatic brain injury (TBI), spinal cord injury (SCI)

INTRODUCTION

Astrocytes are critical for the functioning of the adult central nervous system (CNS) in health and disease with a myriad of well-documented roles encompassing the spectrum of physiologic functions from metabolic support to blood-brain-barrier (BBB) integrity to synapse regulation (for review see Volterra and Meldolesi, 2005; Abbott et al., 2006; Koehler et al., 2008; Sofroniew and Vinters, 2010a; Clarke and Barres, 2013; Bayraktar et al., 2015; Khakh and Sofroniew, 2015; Khakh and Deneen, 2019). There is growing consensus that astrocytes are highly plastic in response to environmental fluctuations, particularly in the dynamically changing environment of CNS pathological states. This has shifted our understanding of this ubiquitous glial cell population from that of a binary-population of fibrous and protoplasmic types to one with significant variation across multiple variables, including temporal, topographical, sex, and age, both within and across pathological states. Accordingly, there is significant interest in better understanding and further defining this potential diversity within the astrocyte population.

For the purposes of this review, we define diversity broadly as any distinguishable morphological, physiological, transcriptomic, proteomic, metabolic, or functional difference within the astrocyte population, whether transient or not. The development of technologies enabling detailed descriptions of these responses has led to an accumulation of evidence for diversity within the astrocyte population in the healthy CNS but also across many different models of disease/injury. Here, we briefly address the origin of astrocytes during development and what is currently known about the diversity of astrocytes in the healthy adult CNS prior to an in-depth exploration of diversity in CNS injury/disease, which is the focus of this review. Finally, we initiate a discussion on whether diversity should be sub-divided into more biologically meaningful categories, such as “plasticity” and “heterogeneity.” We propose that the use of these definitions provides a framework that will be important as more is discovered about astrocyte diversity in CNS pathologies with specific relevance for future therapeutic development.

ASTROCYTES IN DEVELOPMENT AND ASTROCYTE DIVERSITY IN ADULT HOMEOSTASIS

Astrocytes in the Developing CNS

At the foundation of our discussion on astrocyte diversity in the adult CNS is the multitude of studies on the developmental origin of astrocytes, which we only discuss in brief here (for more details see the following reviews: Bayraktar et al., 2015; Molofsky and Deneen, 2015). The majority of astrocytes originate from subventricular zone (SVZ) resident neuroepithelium-derived radial glial (RG) cells (Noctor et al., 2002; Anthony et al., 2004; Kriegstein and Alvarez-Buylla, 2009), with additional contributions from marginal zone progenitor cells in superficial cortical layers (Costa et al., 2007; Breunig et al., 2012). Importantly, embryonic astroglialogenesis accounts for only a

fraction of adult astrocytes, as the majority of murine gliogenesis occurs postnatally (Bandeira et al., 2009) through the symmetric division of differentiated astrocytes (Ge et al., 2012). Direct transformation of RG cells is also a documented source of astrocytes (Merkle et al., 2004; Ghashghaei et al., 2007). Furthermore, NG2 glia (also referred to as oligodendrocyte progenitor cells—OPCs) have been reported to generate a distinct sub-type of ventral forebrain astrocytes (Zhu et al., 2008, 2011; Huang et al., 2014; Nishiyama et al., 2015). Once generated, astrocyte progenitors disperse radially from their site of origin within the confines of a single column leading to the establishment of a diverse population (Magavi et al., 2012; Gao et al., 2014). In the spinal cord, patterning of astrocyte progenitors is initiated by dorsoventral gradients of secreted molecules that facilitate radial organization. For example, three neural tube progenitor domains give rise to three spatially distinct ventral white matter (WM) astrocytes clusters (Hochstim et al., 2008) which are then likely influenced by a combination of intrinsic and extrinsic factors (for review, see Ben Haim and Rowitch, 2017). More fate-mapping analysis is required to establish the extent to which this developmentally established diversity persists into the adult CNS and contributes to the observed adult astrocyte diversity, both in the healthy CNS and in pathological states (e.g., Tsai et al., 2012).

Diversity of Astrocytes in the Healthy Adult CNS

Beginning with Cajal’s descriptions of diverse morphologies amongst human and rodent astrocytes over 100 years ago, astrocyte diversity has been a recognized feature of the healthy adult CNS (Zhang and Barres, 2010; Boulay et al., 2017; Lin et al., 2017; Buosi et al., 2018; Matias et al., 2019). Early histologic description designated protoplasmic and fibrous astrocytes as unique subsets (Kölliker, 1889; Raff et al., 1984; Raff, 1989; Andriezen, 1893) based upon differences in location [WM vs. gray matter (GM)], cell body morphology, and interaction with neighboring neuronal structures (Bushong et al., 2002; Oberheim et al., 2012; Lundgaard et al., 2014). Transcriptional-based approaches have expanded upon this initial description (Cahoy et al., 2008; Batiuk et al., 2020; Bayraktar et al., 2020) with unique astrocytic gene profiles demonstrated across various brain regions (Chai et al., 2017; Morel et al., 2017; Duran et al., 2019; Bayraktar et al., 2020). Using fluorescence-assisted cell sorting (FACS) and immunohistochemical approaches, Lin et al. (2017) identified five distinct astrocyte populations in the mouse CNS, which displayed diverse synaptogenesis mechanisms. Similarly, using a transcription factor motif discovery approach, Lozzi et al. (2020) found region-specific astrocytic expression profiles in astrocyte populations from the olfactory bulb, hippocampus, cortex, and brainstem. Furthermore, astrocyte reporter mouse lines exposed molecular differences between different astrocyte populations within the adult cortex (Morel et al., 2019).

Importantly, these unique transcriptomic gene profiles are correlated with neural-circuit-based functional differences (Höft et al., 2014; Chai et al., 2017). Variable expression of key functional components in astrocytes has been noted, including

glutamate receptors, transporter proteins, and ion channels (Matthias et al., 2003; Isokawa and McKhann, 2005; Olsen et al., 2007), as well as in calcium (Ca^{2+}) signaling dynamics (Takata and Hirase, 2008), which is theorized to have a functional role in astrocyte-neuron communication (Bazargani and Attwell, 2016; Chai et al., 2017; Yu et al., 2018). Using multi-photon confocal imaging, Takata and Hirase (2008) demonstrated significant variance in astrocytic Ca^{2+} activity between cortical layer I and layers II/III. Diversity amongst astrocyte populations in different cortical layers has been identified in other studies as well (Lanjakornsiripan et al., 2018; Bayraktar et al., 2020). For example, single-cell RNA-seq (scRNA-seq) analysis identified five transcriptionally distinct clusters distributed amongst cortical layer I and III-V (Bayraktar et al., 2020) and quantification of astrocyte marker expression across cortical layers in the developing mouse brain revealed significant diversity across functionally distinct cortical areas (Batiuk et al., 2020).

Glial fibrillary acidic protein (GFAP)—a major component of intermediate filaments in astrocytes—is a widely used marker of astrocytes (Eng et al., 1971). Importantly, the basal level of GFAP in astrocytes in the healthy CNS is variable (Griemsmann et al., 2015; Ben Haim and Rowitch, 2017). For example, hippocampal astrocytes display higher GFAP expression than striatal astrocyte populations (Chai et al., 2017). Furthermore, GFAP expression is higher amongst spinal cord astrocytes compared to the brain (Yoon et al., 2017). Interestingly, astrocytic GFAP expression is modulated by various extrinsic stimuli, including global physical activity (Rodriguez et al., 2013), exposure to enriched environments (Rodriguez et al., 2013), and glucocorticoid treatment (O’Callaghan et al., 1991), suggesting that plasticity may play a role in shaping the regional diversity of GFAP expression observed in the healthy CNS. Interestingly, GFAP expression also fluctuates with circadian rhythms in the suprachiasmatic nucleus of the thalamus (Gerics et al., 2006). Also, GFAP expression amongst progenitor cells depends upon the developmental stage, highlighting more diversity in the expression of this marker (Cahoy et al., 2008; Kriegstein and Alvarez-Buylla, 2009; Roybon et al., 2013). While an important marker used to identify astrocytes, it is important to note that GFAP is not considered sufficient as an identifier of astrocyte populations, either in the healthy or injured CNS. A combination of multiple astrocyte markers is generally viewed as an improved approach [e.g., GFAP, aldehyde dehydrogenase-1 (Aldh1L1), and glutamine synthetase (GS); Serrano-Pozo et al., 2013]. Importantly, diversity in the expression of these additional markers is also seen (Anlauf and Derouiche, 2013; Waller et al., 2016). For example, diverse expression of Aldh1L1 amongst cortical astrocytes (Waller et al., 2016) and GS amongst entorhinal cortical astrocytes is observed (Anlauf and Derouiche, 2013).

As astrocytes are critical to the normal functioning of local neuronal populations (Chai et al., 2017; Matias et al., 2019), further characterization of the extent of this diversity (for review see Khakh and Sofroniew, 2015; Ben Haim and Rowitch, 2017; Khakh and Deneen, 2019), and the functional implications for neural circuit functioning (for review see, Nagai et al., 2021b) in the healthy CNS is vital. Furthermore,

it is imperative to establish a baseline (Tsai et al., 2012; Batiuk et al., 2020; Bayraktar et al., 2020) against which identified diversity in models of CNS insult can be interpreted in order to develop targeted interventions aimed at manipulating aberrant and/or pro-pathogenic responses (Batiuk et al., 2020; Sofroniew, 2020).

DIVERSITY IN THE CONTEXT OF REACTIVE ASTROGLIOSIS

Defining Reactive Astrogliosis

Various terms have been used to describe the range of astrocytic responses to CNS insult and/or environmental perturbation (Eddleston and Mucke, 1993; Anderson et al., 2014; Pekny and Pekna, 2014; Sofroniew, 2015). In line with a recently published consensus statement (Escartin et al., 2021), we define “reactive astrogliosis” as the process by which astrocytes change in response to pathology. This can include changes in transcriptional regulation, or biochemical, morphological, metabolic, and physiological remodeling potentially associated with functional adaptation to the post-injury environment. Reactive astrogliosis was long viewed as homogenous and functionally passive, consisting of a stereotyped set of changes driving the conversion of homeostatic astrocytes to a distinct phenotype—the “reactive astrocyte” (Eddleston and Mucke, 1993; Anderson et al., 2014; Pekny and Pekna, 2014; Sofroniew, 2015). However, current evidence challenges this view, instead pointing to the existence of remarkable diversity in terms of morphology and transcriptional profile in varied CNS disease states (Hamby and Sofroniew, 2010; Zhang and Barres, 2010; Oberheim et al., 2012; Anderson et al., 2014; Schitine et al., 2015; Yoon et al., 2017; Zeisel et al., 2018; Masuda et al., 2019; Matias et al., 2019; Valori et al., 2019; Escartin et al., 2021). This begs the question of how extensive this diversity really is (Cahoy et al., 2008; Ståhlberg et al., 2011; Yoon et al., 2017; Batiuk et al., 2020; Bayraktar et al., 2020).

Role of Astrocytes in CNS Disease

Reactive astrogliosis is observed in virtually all neurological conditions, including epilepsy (Steinhäuser et al., 2015), neoplastic disease (Priego et al., 2018; Heiland et al., 2019), demyelination (Woodruff and Franklin, 1999; Williams et al., 2007; Tassoni et al., 2019; Rawji et al., 2020), traumatic injury (Faulkner et al., 2004; Filous and Silver, 2016; Boghdadi et al., 2020a), neurodegeneration (Vargas et al., 2008; Ben Haim et al., 2015), and ischemic stroke (Zhao and Rempe, 2010; Zamanian et al., 2012; Rakers et al., 2019), as well as microbial CNS infections (Drögemüller et al., 2008; Soung and Klein, 2018; Geyer et al., 2019) and neurotoxin exposure (O’Callaghan et al., 2014; Wheeler et al., 2019). Reactive astrogliosis enables astrocytes to serve key roles in CNS pathological states, including metabolic support of vulnerable neurons, regulation of BBB permeability, remodeling of extracellular matrix (ECM), mobilizing progenitors, as well as immunomodulation, synaptic remodeling, and neurite outgrowth (Woodruff et al., 2004; Sofroniew and Vinters, 2010a; Anderson et al., 2014;

Pekny and Pekna, 2014; Escartin et al., 2019, 2021). This process is regulated by a wide range of factors, both intrinsic and extrinsic to the CNS, and mediated through various cell surface receptors and intracellular signaling pathways (Sofroniew and Vinters, 2010a; Burda and Sofroniew, 2014; Sofroniew, 2015, 2020). Manipulation of these components in models of CNS injury/disease alters functional and histologic outcomes, demonstrating the importance of reactive astrogliosis and of understanding the extent and nature of its diversity (Brambilla et al., 2005, 2009; Okada et al., 2006; Herrmann et al., 2008; Haroon et al., 2011; Spence et al., 2011; Bonne-Barkay et al., 2012; Wanner et al., 2013).

Reactive Astrogliosis and the Aged CNS

Reactive astrogliosis is also a prominent feature of physiological aging in rodents, non-human primates, and humans (Nichols et al., 1993; Kanaan et al., 2010; Cerbai et al., 2012; Rodríguez et al., 2014; Jyothi et al., 2015; Robillard et al., 2016). Previous studies have noted regional diversity in astrocyte morphology, GFAP expression, and cellular density (Nichols et al., 1993; David et al., 1997; Hayakawa et al., 2007; Lynch et al., 2010; Cerbai et al., 2012; Geoffroy et al., 2016; Rodríguez et al., 2016). Aged astrocytes also demonstrate altered responses to CNS injury. For example, aged astrocytes display increased GFAP upregulation following SCI as compared to younger controls (Geoffroy et al., 2016), however, the functional implications of this remain unclear (Matias et al., 2019; Sutherland and Geoffroy, 2020).

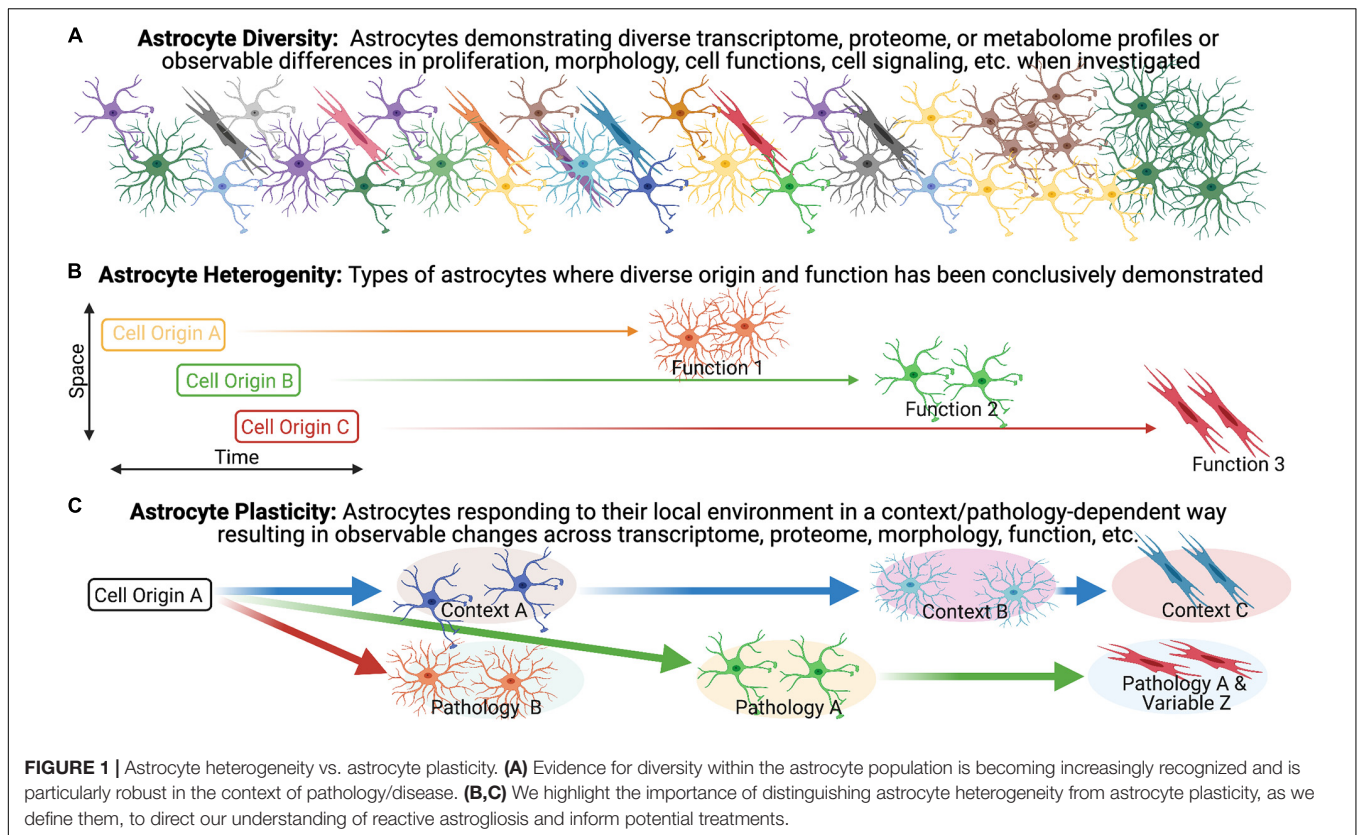
Transcriptional approaches have once again greatly expanded our characterization of astrocyte diversity in the aged CNS (Soreq et al., 2017; Boisvert et al., 2018; Clarke et al., 2018). These studies have collectively revealed that aged astrocytes adopt a more pro-inflammatory phenotype (Orre et al., 2014), consistent with the concept that physiological aging is characterized by chronic low-level inflammation (i.e., “inflamm-aging”; Franceschi et al., 2000). Aged astrocytes from distinct brain regions display unique transcriptional profiles in both murine and human brains (Soreq et al., 2017; Boisvert et al., 2018; Clarke et al., 2018). Boisvert et al. (2018) performed RNA-seq analysis on 4 month and 2-year-old astrocyte-ribotag mice, demonstrating significant upregulation of pro-inflammatory and synapse elimination-related genes and decreased expression of cholesterol synthetic enzymes in the aged mice, with significant regional diversity (Boisvert et al., 2018). Expanding on this, Clarke et al. (2018) demonstrated regional diversity amongst aged astrocytes isolated (using the Bac-Trap method) from the hippocampus, striatum, and cortex, with upregulated genes related to astrocyte reactivity, immune response, and synapse elimination. Soreq et al. (2017) extended these findings to post-mortem human tissue of patients ranging in age from 16 to 102 years, revealing significant regional diversity of astrocyte-specific genes (Clarke et al., 2018). Characterization of the astrocyte secretome during the aging process may help validate many of these observed transcriptomic changes (Rawji et al., 2020). Intriguing questions remain as to the functional relevance of these age-related changes and the role of astrocyte diversity in the spatial propensity of various age-related disease processes (Ben Haim et al., 2015; Rodríguez et al., 2016; Matias et al., 2019). Furthermore, comparison of the transcriptional

profiles of astrocytes across pathological states with those seen in the aged CNS may yield novel insights into disease pathogenesis, physiological aging, and the overlap between these states.

Reactive Astrogliosis Diversity: Plasticity or Heterogeneity?

We broadly define astrocyte diversity as any distinguishable morphological, physiological, transcriptomic, proteomic, metabolic, or functional difference within the astrocyte population, whether transient or not (**Figure 1A**). With this definition, and those that follow, we suggest that semantics are important, as a proper classification of diversity will likely lead to greater accuracy in the understanding of function and the directed development of therapeutic strategies. We therefore propose strict definitions to further classify reactive astrocyte diversity.

Heterogeneity has become a catch-all phrase that remains poorly defined, necessitating the need for clearer and unambiguous definitions. The term heterogeneity is derived from the Greek *heteros-* meaning “two, other, or different,” and the Latin *-genesis* meaning “origin or development” (Oxford English Dictionary). In distinguishing heterogeneity, we adhere to the definitions offered in a recent discussion of diversity in the oligodendrocyte lineage (Foerster et al., 2019), namely that heterogeneity implies distinct origin, as suggested in the definition, in combination with the demonstration of diverse functions (**Figure 1B**). In support of this, we look to other neural cell lineages. Firstly, heterogeneity amongst neurons is demonstrated by developmentally distinct neuronal subtypes with different transmission modes and firing patterns (i.e., function). A particularly relevant example of heterogeneity in the context of pathology lies within the microglia population. Dogma suggested that in mice, microglia progenitors arise at E7.5 from the yolk sac and then colonize the brain at E9.5 but mutant mice lead to the discovery that *Hoxb8* microglia (which express the *Hoxb8* transcription factor) represent a distinctive subpopulation of cells that are derived from a second wave which do not populate the brain until E12.5 (De et al., 2018). Although the non-*Hoxb8* microglia and *Hoxb8* microglia are very similar with few differentially expressed genes, they occupy distinct distributions in the post-natal mouse brain and demonstrate unique functional characteristics including their ability to participate in synaptic pruning and their response to injury. For example, the two microglia subpopulations are indistinguishable in their response to a stab wound injury in the acute phase (<30 min) but *Hoxb8* microglia demonstrated a greater tendency to accumulate at the injury epicenter at 7 days post-injury (dpi) compared to non-*Hoxb8* microglia (De et al., 2018). A similarly relevant example of heterogeneity in the context of pathology can be seen in the oligodendrocyte lineage. In the context of remyelination, OPCs arising from distinct ventral and dorsal domains during development have differential responses. For example, dorsally derived OPCs in the adult CNS demonstrate enhanced recruitment and differentiation into oligodendrocytes in response to demyelination as compared to their ventrally derived counter-parts (Crawford et al., 2016). Furthermore,



dorsally derived OPCs demonstrate increased susceptibility to the age-associated differentiation impairment observed in the context of demyelination (Crawford et al., 2016). These findings illustrate the influence of heterogeneous populations on disease-associated variables (e.g., aging) in pathological settings (Crawford et al., 2016).

In the absence of pathology, one example of astrocyte heterogeneity (as we define it) is the demonstration that postnatal region-restricted spinal cord astrocytes have unique functions. In spinal cord development, spatially distinct astrocytes are specified through a homeodomain transcriptional code from positionally distinct progenitor populations (Hochstim et al., 2008) and spatially distinct domains remain stable throughout life in both mouse brain and spinal cord (Tsai et al., 2012). Specifically within the spinal cord, spatially distinct astrocytes with unique origins were shown to express postnatal region-specific genes and the ventral population plays a distinct role in sensorimotor circuit formation (Molofsky et al., 2014). This region-specific expression pattern of genes was further demonstrated across the cortical and subcortical adult mouse brain. Furthermore, astrocytes from these brain regions exhibited region-matched astrocyte to neuron communication specific to their ability to promote neurite growth and synaptic activity *in vitro* (Morel et al., 2017). Therefore, these spatially distinct astrocytes are a prime example of heterogeneity, as we define it, within the uninjured astrocyte population due to the direct evidence underlying their distinct origin and functions.

In contrast to our heterogeneity definition, plasticity would manifest as malleable morphological and/or phenotypic profiles among cells of a common origin in response to changing environmental conditions (**Figure 1C**). An illustration of this concept can be seen in the macrophage lineage. Monocyte differentiation into effector phenotypes occurs in accordance with local microenvironmental signals, accounting for the diverse macrophage effector functions in various tissues, and in response to different insults (Mosser and Edwards, 2008; Italiani and Boraschi, 2014; Lavin et al., 2014; Xue et al., 2014; Kuznetsova et al., 2020). Applied to the astrocyte lineage, plasticity would imply the CNS is populated with a homogenous population of astrocytes that undergo specialization at their final location as directed by local environmental features. For example, in the healthy postnatal CNS, the functional maturation of cortical astroglia is modified by the loss of neuronal glutaminergic signaling (Morel et al., 2014). In the adult healthy CNS, sonic hedgehog released from local neurons plays an active role in regulating both astrocyte function and the astrocytes' molecular profile (Farmer et al., 2016) demonstrating that astrocytes can respond to cues from neurons that drive their properties/functions. We suggest that for distinct reactive astrocyte populations to be considered heterogeneous, definitive demonstration of distinct origins and functions need to be established to effectively exclude plasticity.

While we segregate plasticity and heterogeneity here for conceptual purposes, it is likely that there is a dynamic interplay between the two with varying contributions across multiple

disease variables. An example of this can be seen in the oligodendrocyte lineage. While the diversity of myelin internode length appears to be a function of the axon characteristic and not oligodendrocyte diversity (i.e., plasticity; Chong et al., 2012; Tomassy et al., 2014), diverse oligodendrocytes isolated from either the spinal cord or cortex form myelin sheaths of different lengths when provided artificial microfiber as a substrate for myelination (Bechler and Byrne, 2015), suggesting at least a degree of intrinsic determination (i.e., heterogeneity; Crawford et al., 2016). Importantly, diversity in internode length is reduced when oligodendrocytes were cultured with dorsal root ganglion neurons or brain slices, implicating both intrinsic (i.e., heterogeneity) and extrinsic factors (i.e., plasticity) in

determining the outcome. It is likely that a similar dynamic combination exists for astrocytes as well. Importantly, this is not just a semantic argument, as recognition of the contributions of plasticity to the observed diversity has the potential to reveal novel targets amenable to extrinsic manipulation *via* targeted therapeutic approaches across multiple aspects of disease pathology.

Tools to Better Understand Cell Diversity in Reactive Astrocyte Populations

scRNA-seq or single-nuclei RNA-seq (snRNA-seq) have enabled the direct quantification of single-cell or nuclei RNA complements at an increased resolution (Tang et al., 2009;

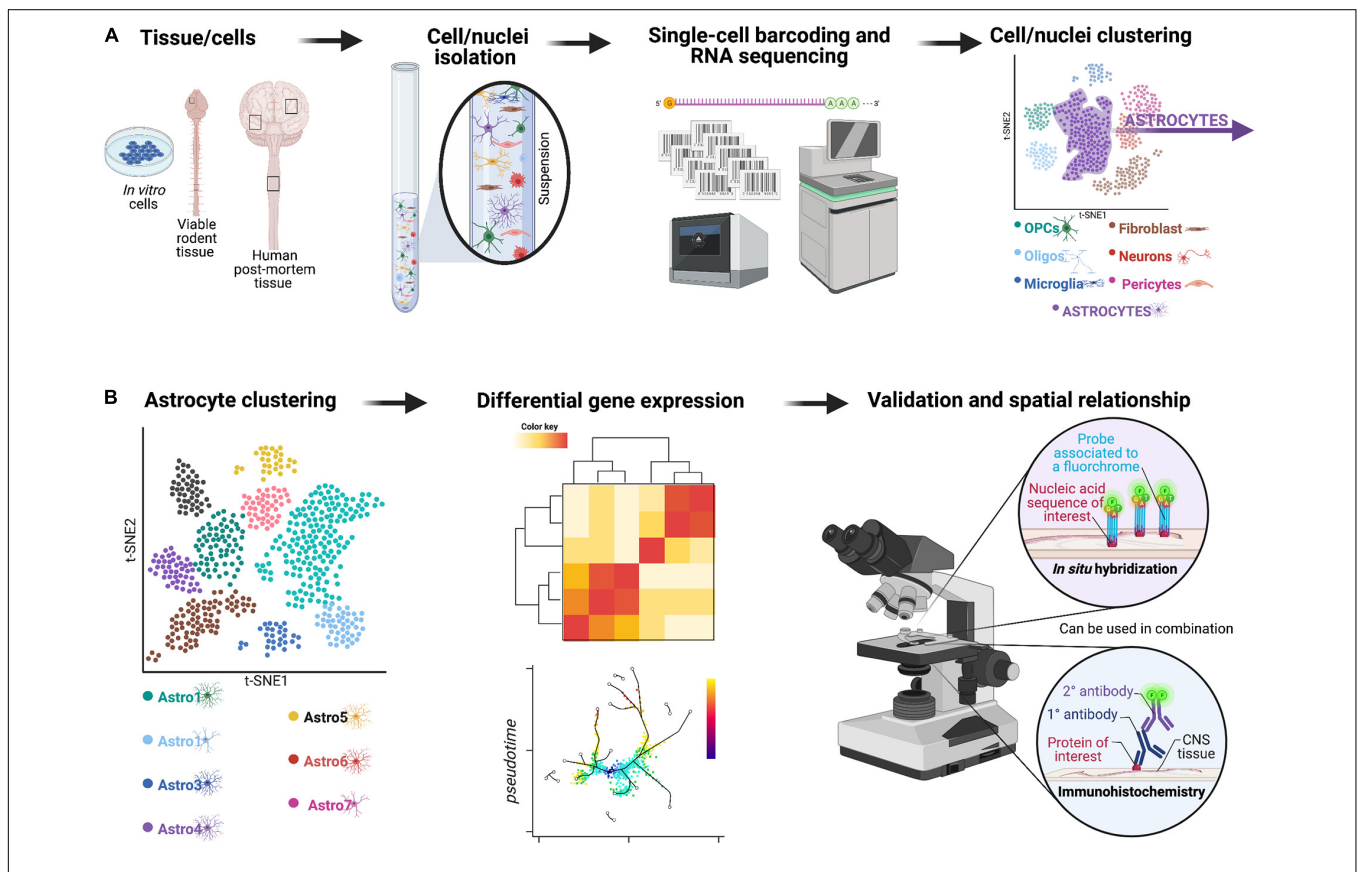


FIGURE 2 | Overview of a potential workflow featuring droplet-based sc/snRNA-seq approaches to investigate astrocyte diversity across different CNS pathologies. **(A)** In the past few years, a technological revolution in RNA-sequencing technology has made it possible to profile the entire transcriptome of individual cells on a massive scale—a technique known as single-cell RNA-sequencing or scRNA-seq (Svensson et al., 2018). Initially, scRNA-seq relied on manual cell picking (Van Gelder et al., 1990; Eberwine et al., 1992) or FACS-based sorting (Ramsköld et al., 2012; Shalek et al., 2013). Innovative analyses revealed a surprising degree of transcriptional heterogeneity in seemingly homogenous cell populations. Subsequent advances in microfluidic instrumentation (Shalek et al., 2014; Treutlein et al., 2014) and droplet-based methods (Klein et al., 2015; Macosko et al., 2015) have since driven experimental costs down significantly to now permit sequencing of tens to hundreds of thousands of cells in a single experiment (Cao et al., 2017; Schaum et al., 2018). The rapid pace of methodological and computational progress has fostered initiatives to profile the mRNA landscape within every single cell of various model organisms (Cao et al., 2017; Schaum et al., 2018) and, ultimately, in humans (Rozenblatt-Rosen et al., 2017). This framework now enables comprehensive interrogation of the molecular etiology of human disease at single-cell resolution (Stubbington et al., 2017; Cheung et al., 2019). For example, single-cell transcriptomics offers an opportunity to elucidate how individual types of cells coordinate their activity to drive pathophysiological processes, and how cell type-specific responses might be targeted to treat disease. Indeed, in only the past few years, scRNA-seq has been applied to asthma (Braga et al., 2019), inflammatory bowel disease (Martin et al., 2019; Parikh et al., 2019; Smillie et al., 2019), obesity (Svensson et al., 2018), Alzheimer's disease (Mathys et al., 2019), and TBI (Arneson et al., 2018), among other disorders. These technologies and analyses enable clustering of all viable cells/nuclei included in the original sample based on gene expression, and for example to identify astrocyte-like subpopulations can be isolated for further analyses. **(B)** The astrocyte-like subpopulations can be further clustered and examined using approaches, such as differential gene expression to yield important information about heterogeneous astrocyte populations.

Zeisel et al., 2018; Habib et al., 2020). Approaches, such as droplet-based sc/snRNA-seq approaches (Figure 2) have proven immensely powerful across various disease and injury models, revolutionizing our capacity for cellular characterization (Chen et al., 2018; Ding et al., 2020; Jäkel and Williams, 2020). Importantly, distinct clusters of astrocytes identified through these single-cell technologies need to then be validated to make sure results are not just noise and the spatial relationship needs to be re-established using *in situ* hybridization or immunohistochemistry alone or in combination. Use of these techniques also allow for analysis on human post-mortem tissue and *in vitro* culture systems (Grubman et al., 2019; Mathys et al., 2019) which has significantly enhanced our knowledge of species-specific differences in reactive astrogliosis, a critical hurdle for translation of pre-clinical findings in rodent model systems to human patients (Nichols et al., 1993; Oberheim et al., 2009, 2012; Soreq et al., 2017). When used alone these tools provide a powerful means of assessing astrocyte diversity but do not clearly distinguish between heterogeneity and plasticity unless they are combined with other complementary experiments, such as those looking at lineage-tracing. With that being said, recent papers highlight the potential of these single-cell technologies to yield information about lineage (Weinreb et al., 2020) as well as connectivity analysis (Clark et al., 2021) to be performed in high-throughput, with cell-type resolution. Furthermore, new techniques, such as sc-ATAC-seq will provide information about chromatin accessibility, which is likely to be an important determinant of astrocyte plasticity (Buenroostro et al., 2015). These new approaches will be influential in the task of determining whether reactive astrocyte clusters represent heterogeneity (with distinct origin and functions) or plasticity.

The Spectrum of Reactive Astrogliosis

Similar to the healthy CNS, transcriptional analysis has revealed multiple clusters of reactive astrocytes in various models of CNS insult (Adams and Gallo, 2018; Tassoni et al., 2019; Yang et al., 2020). Various schemes have been proposed to categorize this diversity, including the classification of reactive astrocytes as either proliferative border-forming or non-proliferative in models of CNS trauma (Sofroniew, 2020). The most well-known is the categorization of astrocytes as neurotoxic “A1” or neuroprotective “A2.” In rodent models, intraparenchymal lipopolysaccharide (LPS)-injection induces a neurotoxic astrocyte phenotype stimulated by microglia-derived factors (Liddelow et al., 2017), whereas cerebral ischemia induces astrocytes to adopt what was later coined an “A2” phenotype that appeared to be neuroprotective (Zamanian et al., 2012). Astrocytes resembling what was later referred to as “A1” phenotypes have been identified in various disease states, including models of amyotrophic lateral sclerosis (ALS; Sun et al., 2015), Alzheimer’s disease (Sekar et al., 2015; Wu et al., 2019), prion disease (Smith et al., 2020), glioblastoma (Heiland et al., 2019), Parkinson’s disease (Chen et al., 2009), and Huntington’s disease (Diaz-Castro et al., 2019; Al-Dalahmah et al., 2020). While these “A1” vs. “A2” distinctions are useful, they also likely represent an oversimplification of the much larger continuum of reactive astrocyte states that are present

in CNS pathologies. Indeed, the authors themselves stated that these two states were likely only a subset of many potential reactive states (Zamanian et al., 2012; Liddelow and Barres, 2017; Liddelow et al., 2017). In the recently published consensus statement, Escartin et al. (2021) highlight the shortcomings of using these binary divisions of reactive astrocytes, such as “A1” vs. “A2,” good vs. bad, or neurotoxic vs. neuroprotective and advocate for the assessment of multiple molecular and functional parameters moving forward (Escartin et al., 2021). Furthermore, a clear neurotoxic role for “A1” astrocytes is not always clearly demonstrated, highlighting the challenge with this dichotomous classification. For example, deletion of a subset of “A1” astrocytes accelerated neurodegenerative progression in a mouse model of prion disease (Hartmann et al., 2019), suggesting that these astrocyte states are considerably more nuanced. As will be discussed in the disease models below, transcriptional analysis has provided further evidence that astrocyte diversity exists along a spectrum of states likely driven by local microenvironments (Zhang and Barres, 2010; Anderson et al., 2014; Escartin et al., 2021). In this review, we still refer to “A1” vs. “A2” terminology for studies conducted in the past but adhere to the consensus put forward by Escartin et al. (2021) to avoid these terms in future research. Indeed, recent studies in models of ischemic stroke (Rakers et al., 2019; Androvic et al., 2020), demyelination (Yoon et al., 2017; Tassoni et al., 2019), and traumatic injury (Burda et al., 2016; Boghdadi et al., 2020b) have revealed substantial disease-specific, regional, and temporal reactive astrocyte diversity.

Potential Variables Shaping Diversity in the Context of Reactive Gliosis

There are many variables potentially influencing the diversity of reactive astrogliosis. Here, we review the current state of knowledge on astrocyte diversity in the context of three representative and clinically relevant CNS pathologies: ischemic stroke, demyelination, and traumatic injury across multiple injury-associated variables (e.g., temporal, topographical, sex, and age). Plasticity can be conceptualized as the diversity in a response to environmental factors. In pathological states this includes features of the post-injury environment (e.g., cytokines, inflammatory cells, etc.). In contrast, heterogeneity manifests as diversity that results from differential origins (either developmental and adult-derived) and functionality. Importantly, plasticity and heterogeneity are not mutually exclusive in our model and elucidating this relationship as it manifests in various conditions of CNS pathology is critical to understanding the contributions of astrocytes in the injured CNS and how these responses can be manipulated. As our understanding of the extent of astrocyte diversity develops, it will become ever more important to integrate and compare astrocyte responses across pathological states to be able to interpret and apply the substantive datasets gleaned from single-cell transcriptomic approaches. To that end, we expand our discussion beyond the confines of a singular pathology and aim to highlight key limitations of our current knowledge, propose areas for future research, and discuss the relevance of this

knowledge for therapeutic development. We specifically focus on the importance of differentiating the contributions of plasticity and heterogeneity to observed astrocyte diversity across multiple variables in CNS pathologies.

ISCHEMIC STROKE

Stroke is the primary cause of severe disability and a leading cause of death worldwide, associated with enormous socioeconomic burden (Cassidy and Cramer, 2017; Campbell et al., 2019; Campbell and Khatri, 2020). Accounting for 75–80% of all strokes (Cassidy and Cramer, 2017; Campbell et al., 2019; Campbell and Khatri, 2020), ischemic stroke results from the occlusion of a cerebral artery by a blood clot that either forms locally (i.e., thrombotic stroke) or more commonly, travels from another location, such as the heart or another proximal vessel (i.e., embolic stroke; Cassidy and Cramer, 2017; Campbell et al., 2019; Campbell and Khatri, 2020). Clinically, the extent of the resultant injury depends on several factors, including the severity and duration of ischemic injury and the quality of collateral blood flow to the affected perfusion territory (Bang et al., 2008; Maud et al., 2021). Despite recent advances in reperfusion techniques, therapeutic options remain limited and largely ineffective in attenuating the progressive neuronal loss and consequent functional impairment (Matei et al., 2021).

Various pre-clinical models of ischemic stroke have been employed (Figure 3A; Fluri et al., 2015; Rakers and Petzold, 2017; Sommer, 2017). Among these, the middle cerebral artery occlusion (MCAO) model is largely considered to most closely resemble human ischemic stroke (Longa et al., 1989; Rakers and Petzold, 2017; Sommer, 2017). Other frequently used models include direct mechanical occlusion of a cerebral vessel *via* clipping, ligation, or cauterization (Robinson et al., 1975; Chen et al., 1986; Brint et al., 1988; Hossmann, 2012), stereotactic administration of potent vasoconstrictors (e.g., endothelin-1) to induce vasospasm (Robinson and McCulloch, 1990; Sozmen et al., 2009; Roome et al., 2014; Fluri et al., 2015), and targeted activation of systemically administered photosensitive dye *via* transcranial illumination to induce localized thrombosis (i.e., the photothrombotic model; Watson et al., 1985; Kim et al., 2000; Kleinschnitz et al., 2008). Comparing tissue damage and astroglia responses across the range of pathology encompassed by these models will be informative. Another important consideration is the use of transient occlusive models, which mimic timely recanalization therapy (e.g., thrombolytics or endovascular thrombectomy; Sommer, 2017).

Role of Astrocytes in Ischemic Stroke

The peri-infarct region following ischemic stroke (i.e., the zone immediately surrounding the ischemic lesion; Figure 3B), is commonly segregated into a monocyte/macrophage-dense inner region directly bordering the lesion and an astrocyte-rich outer region (Schroeter et al., 1995; Pekny and Nilsson, 2005; Gliem et al., 2015). The peri-infarct reactive astrocytes that populate this outer region secrete a variety of pro-inflammatory cytokines, chemokines, and

matrix metalloproteinases that disrupt the BBB and recruit peripheral leukocytes, which are predominant contributors to secondary injury (Gliem et al., 2015; Yang and Rosenberg, 2015; Choudhury and Ding, 2016). Furthermore, astrocytes in this region demonstrate process elongation and polarization, as well as upregulation of several factors involved in ECM reorganization and reactive astrocyte clustering (Hirsch et al., 1994; Peng et al., 2013; Gliem et al., 2015). Using an RNA-seq approach in a MCAO rodent model, Rakers et al. (2019) demonstrated a significant upregulation of various neurotoxic genes associated with the “A2”-specific transcripts at 72 h post-stroke compared to control tissue. In total, >1,000 genes were differentially expressed after focal ischemia, including 38 transcription factors (Rakers et al., 2019), demonstrating the pronounced transcriptional changes that take place in response to ischemia. Importantly, many peri-infarct neurons survive the initial ischemic insult but undergo delayed degeneration due to progressive secondary damage to the surrounding spared tissue (Marchal et al., 1996; Liu et al., 2010; Chamorro et al., 2016). As astrocytes provide critical neuronal support and survive the ischemic insult in large numbers they represent an attractive target for therapeutic manipulation to promote, or at least mitigate, this neuronal loss and provide an increased substrate for repair (Zhao and Rempe, 2010; Liu and Chopp, 2016; Becerra-Calixto and Cardona-Gómez, 2017). As astrocytes are key players in both the pathogenesis of ischemic stroke and recovery process following insult, they are of significant interest. Clarification of the interplay between astrocyte plasticity and heterogeneity across multiple disease-associated variables (e.g., temporal, topographical, age-related, sex-related) promises to open novel therapeutic avenues aimed at mitigating stroke-associated morbidity and mortality.

Astrocyte Diversity in Ischemic Stroke

Transcriptomic studies have revealed astrocyte diversity in models of ischemic stroke (Zamanian et al., 2012; Rakers et al., 2019; Boghdadi et al., 2020a). For example, ischemic insult induces differential expression of a host of genes in diverse subsets of astrocytes, including genes involved in neuroinflammation, apoptosis, and transepithelial migration of leukocytes (Zamanian et al., 2012) as well as cell division and migration (Rakers et al., 2019). Application of snRNA-seq provided refinement of this diversity in an endothelin-1-induced ischemic stroke model in marmosets, revealing 19 discrete reactive astrocyte subsets in the primary visual cortex (Boghdadi et al., 2020a). Interestingly, these subsets were noted to express a mixture of “A1” and “A2” genes (Boghdadi et al., 2020a), further highlighting the limited utility of applying binary categorization to what is most likely a nuanced spectrum of activation states. Interestingly, certain astrocyte subsets in the peri-infarct region expressed Nogo-A (Boghdadi et al., 2020a), well-known as a robust neurite outgrowth inhibitor (Chen et al., 2000; GrandPre et al., 2000; Kim et al., 2004; Schwab, 2004). Astrocyte diversity in models of ischemic stroke most likely represents predominant contribution of plasticity in response to the changing post-injury environment superimposed upon a background of heterogeneity, manifested in the post-natal brain as regionally specific astrocyte subsets with variable responses

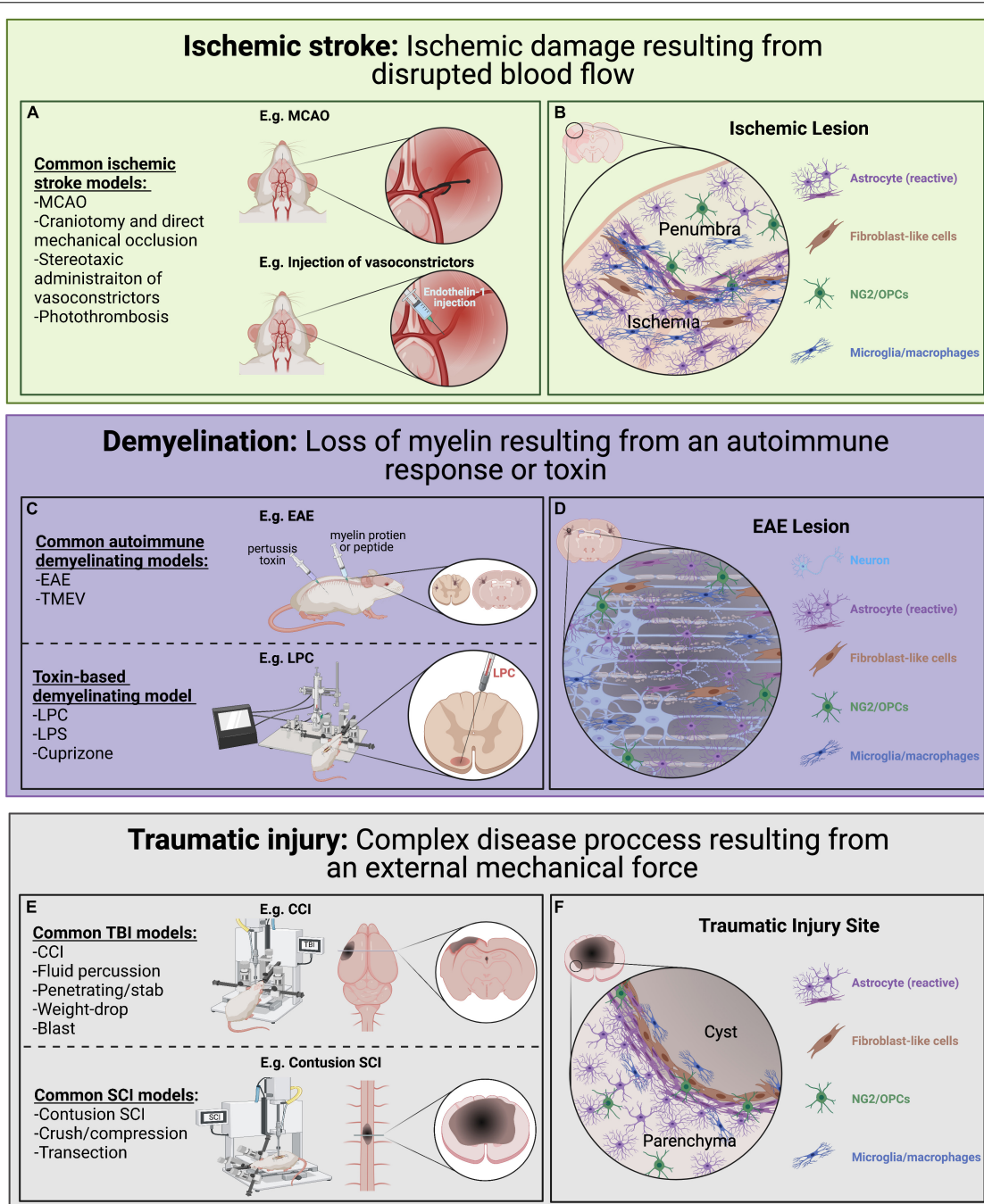


FIGURE 3 | Pre-clinical models of ischemic stroke, CNS demyelination, and traumatic injury used to look at astrocyte diversity. Each of these models have advantages and disadvantages with regards to modeling human disease and induce diverse astrocyte injury responses. **(A)** Common ischemic stroke models. Ischemic stroke results from disrupted blood flow leading to ischemic damage, cell death and associated loss of function. Common animal models of ischemic stroke involve the transient or sustained blockade of normal blood flow to an area of the brain through occlusion of a blood vessel (e.g., MCAO). **(B)** A simplified illustration of an ischemic lesion where the ischemic area is predominately populated by immune cells and reactive astrocytes with a slow gradient toward an inner cluster of microglia/macrophages and surrounded by an outer layer of astrocytes. **(C)** Common demyelination models can be initiated through either an autoimmunity-based or toxin-based route, each highlighting different pathological features and chosen based on the research questions being pursued. **(D)** A simplified illustration of an EAE induced lesion where there is a loss of oligodendrocytes and their myelin sheaths (beige) within the lesion. Demyelination lesions are often filled with immune cells, including microglia/macrophages, and NG2/OPCs which contribute to repair. In the situation where remyelination does not take place, axon degeneration can result. **(E)** Common traumatic injury models focusing on different mechanical injuries applied to the brain and spinal cord, each with its own complex secondary injury cascade. **(F)** A simplified illustration of a typical CNS traumatic injury where a pronounced secondary injury cascade often leads to the loss of tissue (sometimes forming a fluid-filled cyst) at injury epicenter. Generally, there is an inner accumulation of fibroblast-like cells closest to epicenter surrounded by densely packed reactive astrocytes which have an important role in protecting the parenchyma tissue.

to ischemic insult. Elucidating the relative contributions of these processes to observed diversity along several disease-associated variables will require fate-mapping and functional interrogation of astrocyte subsets to reveal the prospective therapeutic potential.

Potential for Astrocyte Plasticity Based on Disease-Associated Variables in Ischemic Stroke

In line with our definition, we propose that identified astrocyte diversity in ischemic stroke occurring along variables of time after injury (i.e., temporal), distance from lesion epicenter (i.e., topographical), age at onset, and sex of the individual, likely represent a greater contribution from plasticity than heterogeneity.

Astrocytes undergo dramatic morphological changes following ischemic insult that evolve over time (Lukaszevicz et al., 2002; Shannon et al., 2007; Benesova et al., 2009; Matyash and Kettenmann, 2009; Li et al., 2014). Using a murine photothrombosis model, Li et al. (2014) revealed increased GFAP expression by day 2 post-insult, with acquisition of stellate morphology and cellular hypertrophy by day 4, and dense astrocyte clustering by day 6. Interestingly, reactive astrocytes became less hypertrophic after day 6 post-insult with gradual lengthening of cellular processes by day 10, reflecting maturation of the reactive astrocyte clustering. Li et al. (2014) then went on to assess astrocyte proliferative dynamics, revealing a peak at day 3 post-insult, with a decline through day 14. Similarly, using an endothelin-1-induced vasospasm rat model, Mestriner et al. (2015) demonstrated significant increases at 30 days post-insult in astrocyte density and increased process ramification and length, as compared to uninjured controls. They also compared these dynamic changes across stroke types, using a time-matched post-hemorrhagic stroke model as comparison. Despite not revealing observable differences in several different GFAP-immunohistochemistry-based measurements, this cross-model comparison approach remains important as it enables isolation of intrinsic vs. extrinsic influences on astrocyte responses in the post-stroke brain (Mestriner et al., 2015). Use of higher resolution transcriptomic studies comparing reactive astrocyte responses over time and across different stroke types (e.g., ischemic, hemorrhagic) and vascular territories (e.g., middle cerebral artery, posterior cerebral artery, etc.) are likely to yield important information on reactive astrocyte diversity.

Astrocytic Ca^{2+} signaling dynamics also vary temporally in rodent models of ischemic stroke, in brain slice culture models of ischemia, and oxygen-glucose deprivation (OGD) models, both acutely and chronically (Duffy and MacVicar, 1996; Ding, 2013, 2014). For example, significant variability in latency to increased Ca^{2+} levels is observed in the initial minutes following ischemic onset in an OGD model (Duffy and MacVicar, 1996; Ding, 2013, 2014), suggesting that differing environmental factors may play a role. Furthermore, application of two-photon imaging in an *in vivo* photothrombosis murine model revealed reactive astrocyte subsets with dynamically changing amplitude and frequency of Ca^{2+} signaling, attributed to fluctuations in

extracellular glutamate and GABA levels (Ding et al., 2009), in keeping with our definition of plasticity. Diversity in astrocytic Ca^{2+} signaling can also be seen outside of the acute post-injury period (Winship and Murphy, 2008; Choudhury and Ding, 2016). Using a murine photothrombotic model, Winship and Murphy (2008) demonstrated a progressive increase in magnitude of penumbral astrocytic Ca^{2+} signaling for 2 months post-insult, driven by stimulation of neural circuits *via* limb stimulation. Once again this represents modulation of astrocyte diversity in response to environmental manipulation (e.g., neural circuit stimulation), thus is offered to represent plasticity. Ca^{2+} signaling diversity may have important functional consequences for the functioning of local neural circuits, thus representing a significant parameter to further characterize. As neural circuit remodeling is an important mechanism of functional recovery post-stroke, manifested by the relative success of physical therapy in stroke patients over the sub-acute to chronic period, it will be a priority to optimize environmental variables that promote supportive astrocyte phenotypes. Further elucidation of astrocyte diversity, especially plasticity vs. heterogeneity, in this context is an important aspect of understanding neural circuit remodeling and functional recovery in the post-stroke brain.

Significant astrocyte diversity is also seen in rodent models of ischemic stroke with respect to distance from the lesion site (i.e., proximal topographical variation). For example, in a rat MCAO model, GFAP immunostaining, process volume, diameter, length, and branching points were increased in close proximity to the cortical infarct border zone compared to distances slightly further from the ischemic area (Wagner et al., 2013). Using a photothrombotic murine stroke model, Li et al. (2014) demonstrated an outwards gradation of astrocyte proliferation from the lesion core in parallel with higher densities of GFAP+ reactive astrocytes. After ischemia due to endothelin-1-induced vasospasm in the primary visual cortex of marmosets, astrocyte subsets were shown to differ in their expression of multiple reactive astrocyte markers, immunomodulatory genes, and various cytokine pathways (Boghdadi et al., 2020a), as well as transcriptional regulators and cell surface receptors involved in cell-matrix adhesion and migration (Dzwonek and Wilczynski, 2015; Boghdadi et al., 2020a). Interestingly, a number of neuronal genes were also upregulated, including Growth Associated Protein 43 (*Gap-43*), which is involved in post-infarct plasticity and expressed by neurons after axotomy (Skene and Willard, 1981; Tetzlaff et al., 1991; Frey et al., 2000; Hung et al., 2016). Topographical diversity in astrocyte Ca^{2+} signaling is also seen, with reduced magnitude amongst penumbral astrocytes compared to the lesion core (Winship and Murphy, 2008). Moreover, restricted astrocyte Ca^{2+} signaling has been shown following single vessel occlusion photothrombosis (Zheng et al., 2013). As most work done to date examines proximal changes near the site of ischemia (i.e., within a singular brain region), this is most likely reflective of plasticity in response to gradients of injury-associated factors (e.g., cytokines, inflammatory cells, blood-borne elements) radiating out from the epicenter. One notable caveat is the observation of extensive changes amongst astrocytes occurring over significant distances (i.e., between different brain regions) post-stroke. For example,

in a MCAO rodent model, Rakers et al. (2019) demonstrated significant changes in gene expression in both the ipsilateral and contralateral hemispheres compared to non-injured control tissue. They demonstrated a 2- and 12-fold increase in *Gfap* and a 2- and 20-fold increase in vimentin (*Vim*) in the contralateral and ipsilateral hemispheres, respectively, compared to uninjured control tissue (Rakers et al., 2019). Diversity noted amongst such spatially separated regions in response to injury could represent a relatively greater contribution from heterogeneity as the astrocytes in these regions most likely have different origins. Despite this, plasticity is still likely a key driver of the observed diversity, albeit proportionally less than for changes observed in close proximity to the lesion.

As ischemic stroke is predominately a disease of older individuals, understanding age-related astrocyte diversity is critical for therapeutic development (Badan et al., 2003; Orre et al., 2014; Androvic et al., 2020). Physiologic aging enhances ischemia-induced astrocyte reactivity, resulting in exaggerated glial responses and accelerated formation of densely packed astrogliosis borders as compared to younger controls (Badan et al., 2003; Popa-Wagner and Badan, 2007). As this is likely to be a consequence of extrinsic/environmental factors, we propose that this age-related diversity represents plasticity, at least within the confines of a singular brain region. Using the MCAO model in 18 month old aged mice, Androvic et al. (2020) demonstrated a predominance of aged astrocytes toward a neurotoxic “A1” phenotype as compared to younger animals, which was associated with worse functional outcomes (Badan et al., 2003; Androvic et al., 2020). This neurotoxic predominance is thought to be largely driven by aged microglia, which tend to be more pro-inflammatory and reactive to insult (Orre et al., 2014; Androvic et al., 2020), consistent with plasticity in response to an altered environment. Using a targeted striatal infarction model in rats, Lively et al. (2011) demonstrated an age-related reduction in astrocyte-derived synaptic cleft-1 (SC1) expression, an ECM molecule associated with neural plasticity, a key aspect of recovery following stroke (Badan et al., 2003; Lively et al., 2011; Sohrabji et al., 2013; Cassidy and Cramer, 2017). Furthermore, ischemia-activated primary astrocyte cultures isolated from aged rats display reduced glutamate uptake compared to younger controls (Lewis et al., 2012). Impaired astrocyte-mediated buffering of glutamate levels in the post-stroke brain may contribute to the increased infarct volumes and worse functional outcomes seen in aged rodents following ischemic stroke (Popa-Wagner and Badan, 2007; Selvamani and Sohrabji, 2010), further suggesting an environmentally dependent effect. Further investigation with lineage tracing studies in the context of the aged CNS is an important area to elucidate potential contributions from heterogeneity to this observed diversity.

Although age-specific stroke incidence and mortality are higher in men, stroke-related morbidity is greater amongst women (Reeves et al., 2008; Petrea et al., 2009), making sex an important disease-associated variable to investigate astrocyte diversity. Adult female rodents demonstrate smaller infarct volumes than age-matched males (Hall et al., 1991; Alkayed et al., 1998; Manwani et al., 2011; Selvamani et al., 2014), an effect which reverses with advancing age (Manwani et al., 2013) suggesting

that the aged female brain is more susceptible to ischemic insult than its male counterpart (Chisholm and Sohrabji, 2016; McCullough et al., 2016). Importantly, these gross differences are underlain by sex-specific diversity amongst cellular populations, including astrocytes. Using a mouse MCAO model, Morrison and Filosa (2016) demonstrated sex-specific differences in frequency of astrocytic Ca^{2+} elevations as well as a more robust reactive astrocytic response in male mice as compared to age-matched females. Sex hormones appear to be a key player in this observed difference. Indeed, astrocyte-derived estradiol conveys neuroprotective and anti-inflammatory effects in rodent models of global ischemia (Zhang et al., 2014). Furthermore, cultured astrocytes isolated from female rodents are more resistant to *in vitro* ischemic insult and glucose deprivation, mediated in part by increased P450 expression and aromatase activity compared to male astrocytes, thus altering estrogen levels in the cells (Liu et al., 2007; Liu et al., 2008). Importantly, ischemia induces astrocyte-specific aromatase expression and activity *in vivo* as well (Carswell et al., 2005). Consistent with this, live imaging studies in a unilateral MCAO rodent model demonstrated significantly increased estrogen-dependent GFAP expression in adult female mice compared to adult males (Cordeau et al., 2008) and estrogen induces astrocytic expression of glutamate transporters GLT-1 and GLAST (Pawlak et al., 2005; Lee et al., 2009), suggesting that female astrocytes *in vivo* may be more effective at glutamate clearance than their male counterparts. Considering that much of this observed diversity is thought to be driven by sex hormones (e.g., estrogen, progesterone), we suggest that this astrocyte diversity is predominantly representative of plasticity, although we cannot exclude the potential of heterogeneity also being a contributing factor (McCullough et al., 2016). This is particularly interesting given the notable sex-specific difference in clinical outcomes (Reeves et al., 2008; Petrea et al., 2009) and the potential for modulation of hormone-responsive signaling pathways as a means of conveying neuroprotection, which are commonly targeted by therapeutics for malignancies (e.g., tamoxifen, trastuzumab; Jordan, 2003; Cameron et al., 2017; Shagufta and Ahmad, 2018; Xu et al., 2019). Adaptation of these therapeutics for the modulation of astrocytic responses in the post-stroke brain may be a viable approach for neuroprotection and/or post-stroke recovery. This requires detailed characterization of the extent of astrocyte diversity present following ischemic stroke, the functional implications of that diversity with respect to neural survival and function, and the identification of potentially malleable targets for therapeutics.

Potential for Astrocyte Heterogeneity Based on Disease-Associated Variables in Ischemic Stroke

In contrast to plasticity, heterogeneity as a contributor to astrocyte diversity in ischemic stroke represents potential diversity that exists amongst astrocyte populations with established differences in origin and function. According to this definition, we propose that this would be diversity observed between regionally specific astrocyte populations (i.e., diversity across brain regions). In the context of ischemic stroke, this could

be seen as diversity amongst astrocyte responses in diverse brain regions following a similar injury (i.e., infarction in different vascular territories). Mestriner et al. (2015) used endothelin-1-induced vasospasm rat model and compared ischemic lesions in the sensorimotor cortex and dorsolateral striatum and noted variation at 30 days post-insult in terms of GFAP-immunohistochemistry based measurements including cellular optimal density and primary process length (Mestriner et al., 2015). Using a MCAO murine model, Lukaszewicz et al. (2002) demonstrated differential functional responses of protoplasmic and fibrous astrocytes (thus effectively comparing WM and GM lesions), including differences in morphological response and cell death (Lukaszewicz et al., 2002), consistent with demonstration of differential ischemic sensitivities amongst astrocyte subsets in models of ischemia-reperfusion injury (Shannon et al., 2007). Importantly, it remains unclear whether protoplasmic and fibrous astrocytes have distinct origins considering that both can be derived from the postnatal SVZ during corticogenesis (Levison and Goldman, 1993; Parnavelas, 1999; Marshall and Goldman, 2002). Lineage tracing has shown that SVZ-derived multipotent neural stem cells (NSC) have the ability to migrate and contribute to the reactive astrocyte population in the context of multiple stroke models (Faiz et al., 2015) but it is unclear how different their function is from other reactive astrocytes in the region. Further direct lineage tracing experiments are required to establish these populations as truly distinct from an origin and functional perspective, and thus fitting of our offered definition of heterogeneity.

Due to limited high-quality fate-mapping studies, there is a scarcity of data on the extent of astrocyte heterogeneity in ischemic stroke. Furthermore, most data accumulated to date focus on astrocyte diversity as a function of changing environmental parameters, and therefore represent plasticity. Ischemic stroke has the potential to affect multiple vascular territories, and thus diverse populations of astrocytes. Determination of the impact of regionally specific astrocyte diversity to various post-stroke outcome measures is of great interest, both clinically and for understanding stroke pathophysiology. For example, hemorrhagic transformation of infarcted tissue (i.e., hemorrhage developing within infarcted tissue) is a devastating post-stroke complication caused by disruption of the BBB (Warach and Latour, 2004; Khatri et al., 2012; Sussman and Connolly, 2013), of which astrocytes are a key component. Identification of malleable aspects of astrocyte diversity that alter susceptibility of neurovascular degradation may be a viable approach to reduce occurrence in the post-stroke population. To that end, expanded study of reactive astrocyte diversity in ischemic stroke models affecting different vascular territories (e.g., middle cerebral artery, posterior cerebral artery, etc.) using combined fate-mapping and functional assays is warranted.

CNS DEMYELINATION

The most common demyelinating disorder is multiple sclerosis (MS), a complex immune disease characterized by inflammation,

primary demyelination (loss of myelin from an intact axon), and neuronal/axonal damage/degeneration (Molina-Gonzalez and Miron, 2019; Rawji et al., 2020). Various pre-clinical models have been employed to replicate the complexity of the MS disease process (Figure 3C; Mix et al., 2010; Baker and Amor, 2015; Lassmann and Bradl, 2017; Baecher-Allan et al., 2018). One of the main models is experimental autoimmune encephalitis (EAE; Constantinescu et al., 2011; Ben-Nun et al., 2014), which involves the administration of myelin peptides or CNS homogenate to induce an autoimmune-mediated demyelinating insult to the rodent CNS (Denic et al., 2011; van der Star et al., 2012; Baker and Amor, 2015). While EAE models the immunopathogenesis of acute MS lesions and has been instrumental to discover many disease modifying treatments (Plemel et al., 2017), it is of limited value for assessing the neurobiological aspects of remyelination (Owens, 2006; Haanstra et al., 2015; Procaccini et al., 2015; Plemel et al., 2017; Stimmer et al., 2018). Another immune-mediated model of MS is the use of Theiler's murine encephalomyelitis virus (TMEV) to induce spinal cord demyelinated lesions (Dal Canto and Lipton, 1977; Owens, 2006; Denic et al., 2011; McCarthy et al., 2012). Toxin-based models of demyelination have also been used widely in pre-clinical studies on neurobiological aspects of demyelinating disease. Most commonly employed are LPS (Liddel et al., 2017), cuprizone (Praet et al., 2014), lysophosphatidylcholine (lysolecithin; LPC; Blakemore and Franklin, 2008; Lassmann and Bradl, 2017), and ethidium bromide (EB) models (Woodruff and Franklin, 1999; McMurrin et al., 2019). LPS is a model of neuroinflammation initiated by the injection of a bacterial endotoxin and results in neurotoxic astrocyte phenotype (Liddel et al., 2017). Cuprizone is a copper chelating agent that is administered orally and induces acute CNS demyelination, particularly in the corpus callosum and cerebellar peduncles (Praet et al., 2014). LPC by contrast, is focally injected into white matter tracts, inducing damage to the myelin sheaths (Blakemore and Franklin, 2008; Lassmann and Bradl, 2017) and acting as a chemoattractant for monocytes, thus triggering a focal inflammatory response (Blakemore and Franklin, 2008; Lassmann and Bradl, 2017). Many toxin-based demyelination models represent a simpler system with predictable and reproducible spatiotemporal patterns to study the remyelination process (McMurrin et al., 2019) and have been used as tools to highlight both the permissive and inhibitory roles of astrocytes (reviewed by Rawji et al., 2020).

Role of Astrocytes in CNS Demyelination

Astrocytes are central to the demyelination response and the success of remyelination (Molina-Gonzalez and Miron, 2019; Rawji et al., 2020), as they release anti-inflammatory cytokines, proteins that modulate myelin regeneration (remyelination), actively maintain extracellular ionic and neurotransmitter concentrations, and provide critical neuronal support (Hinks and Franklin, 1999; Jessen et al., 2015; Liddel et al., 2017). Astrocytes also release pro-inflammatory cytokines, promote BBB permeability, and inhibit OPC maturation (Sisková et al., 2006; van Horsen et al., 2007; Lau et al., 2013; Stoffels et al., 2013). These contrasting functions are thought to represent

changing functional roles at different stages of the remyelination process (reviewed in Rawji et al., 2020). Importantly, astrocyte responses are often dependent on the type of lesion (Rao et al., 2019) where acute active lesions are filled with immune cells and hypertrophic astrocytes with increased GFAP expression and an associated upregulation of proinflammatory chemokines and cytokines (Williams et al., 2007; Franklin and Goldman, 2015). Alternatively, inactive lesions (Figure 3D) have less inflammation and the astrocytes typically are more densely packed with long thick processes in close proximity to extensively demyelinated axons (Franklin and Goldman, 2015). In the rodent cuprizone model, astrocytes have been shown to regulate the recruitment of microglia which then facilitates myelin debris removal, an imperative step for subsequent repair (Skripuletz et al., 2013). Astrocytes also play an important role in influencing the balance of OPC-derived oligodendrocyte vs. OPC-derived Schwann cell-mediated CNS remyelination (reviewed by Chen et al., 2021). The ability of astrocytes to take on different roles may reflect distinct sub-populations or dynamic changes in malleable phenotypes driven by spatiotemporal environmental fluctuations (e.g., microglia-derived factors, cytokines; Cerbai et al., 2012; Horstmann et al., 2016). The relative contributions of heterogeneity and plasticity to these dynamically changing roles remains to be elucidated, as well as the potential for manipulation of astrocytes to phenotypes that support a pro-remyelination lesion environment.

Astrocyte Diversity in CNS Demyelination

Similar to other CNS pathologies, transcriptional evidence has expanded our understanding of astrocyte diversity in CNS demyelination, including the use of bulk transcriptional analysis (Rothhammer et al., 2016, 2018; Itoh et al., 2018; Chao et al., 2019; Wheeler et al., 2019). For example, RNA-seq analysis of EAE and MS tissue identified several clusters of astrocytes with differential expression of *S100b*, *Gja1*, *Aldh1l1*, *Gfap*, and *Aqp4*, consistent with a spectrum of astrocyte transcriptional states in EAE (Wheeler et al., 2019). Furthermore, astrocytes in EAE and MS tissue were demonstrated to have variable expression of the transcription factor *Nrf2* and *Mafg* and *Mat2a* signaling, leading to DNA methylation and increased CNS pathology (Wheeler et al., 2020). These data identify epigenetic modifiers as potential therapeutic candidates to modulate pathology in MS. Importantly, as discussed below, there is a need to separate plasticity and heterogeneity in models of CNS demyelination to improve our understanding of factors that promote a pro-remyelination lesion environment and eventually guide therapeutic development. To that end, we propose here a framework for making that distinction using current evidence for astrocyte diversity in models of CNS demyelination as an example.

Potential for Astrocyte Plasticity Based on Disease-Associated Variables in CNS Demyelination

We argue that astrocyte diversity across variables, such as disease stage (early or late disease progression), lesion status

(status of demyelination or remyelination), age at disease onset, and sex of the individual are examples where plasticity is the predominant contributor to astrocyte diversity. In comparison to other CNS pathologies, MS and EAE have extensive spatiotemporal lesion variability and extensive immunological activity, complicating the interrogation of astrocyte diversity. For example, lesions present in the same individual at a given time can be at different stages (i.e., active, chronic active, inactive, remyelinated, etc.; Rao et al., 2019) and in different CNS regions (spanning WM and GM). Furthermore, analogous lesions can be present in individuals of different age and/or sex, complicating comparisons between spatiotemporally similar lesions. Given those complexities, we highlight the challenge in distinguishing plasticity and heterogeneity without clear fate-mapping studies to establish a baseline in regionally and temporally disparate CNS regions against which comparison of variability can be assessed.

Temporal evolution of astrocyte diversity in inflammatory demyelinated lesions has been noted. For example, in EAE models astrocyte cholesterol gene expression decreases late in the disease course in both the spinal cord and optic nerve (Itoh et al., 2018; Tassoni et al., 2019). Furthermore, astrocytic major histocompatibility complex-II (MHC-II) expression is markedly increased in early EAE brain and spinal cord lesions followed by a late decrease (Itoh et al., 2018), while optic nerve astrocytes in EAE optic neuritis show an early increase in MHC-II expression that persists through the disease course (Clarkson et al., 2015; Tassoni et al., 2019). Due to the role of MHC-II in immunomodulation, it is tempting to speculate about the functional relevance of this diversity, compounded by the fact that optic neuritis is commonly the initial presentation of MS in patients (Polman et al., 2005; Montalban et al., 2010; Amato et al., 2018). Diversity along the temporal component most likely represents response to a dynamic lesion environment, and thus plasticity. Despite this, it remains important to establish a baseline of heterogeneity, especially between CNS regions, as well as identify newly generated astrocytes in the context of demyelination injury as a potential source of heterogeneity.

Astrocyte diversity is also noted across lesion types in MS models (Rao et al., 2019). For example, acute lesions display hypertrophic astrocytes with increased expression of GFAP and several pro-inflammatory cytokines, chemokines, and remyelination-associated molecules (Woodruff et al., 2004; Williams et al., 2007; Franklin and Goldman, 2015; Rawji et al., 2020). By contrast, inactive lesions contain transcriptionally inactive astrocytes with small cell bodies and long filamentous processes that contribute to an increased density of reactive astrocytes (Franklin and Goldman, 2015; Ludwin et al., 2016). For example, acute lesions demonstrate reduced astrocytic connexin-43 (CX43) expression and disrupted CX43/CX47-mediated astrocyte-to-oligodendrocyte connections (Masaki et al., 2013). Loss of CX43 expression is associated with progressive MS disease, oligodendrocyte pathology, and astrocyte degeneration (Masaki et al., 2013) and patients with reduced CX43 expression have a more rapid clinical disease progression (Masaki et al., 2013). Furthermore, astrocytic chitinase 3-like 1 (CHI3L1) expression, associated with chronic inflammation and neurotoxicity (Cantó et al., 2012; Matute-Blanch et al., 2020),

is observed in chronic MS lesions but absent from other lesion types (Cantó et al., 2012). One potential driver of this diversity is microglial-derived factors (e.g., activin-A), which have been shown to modulate astrocyte activation in demyelinated lesions (Miron et al., 2013; Rawji et al., 2020), further consistent with plasticity as a predominant contributor.

Age is an important factor in the pathogenesis of MS, with clinical disease onset rare after the age of 50 (Tremlett et al., 2010; Amato et al., 2018; Baecher-Allan et al., 2018). Furthermore, age at onset appears to be a key prognostic factor for MS patients (Tremlett et al., 2010; Amato et al., 2018). Progressive MS is also correlated with increasing age (Koch et al., 2009; Tutuncu et al., 2013). Despite the multitude of factors that may be contributing to this observation, including age-related changes in hormones, immune function, neural and non-neural cell populations, it is tempting to speculate regarding the potential contribution of age-related diversity in astrocyte subpopulations to this disease susceptibility. Aged-related astrocyte diversity observed is thought to be at least in part secondary to the increased pro-inflammatory function of aged microglia (Kanaan et al., 2010; Cerbai et al., 2012; Jyothi et al., 2015), thus likely representing plasticity. Aged astrocytes are more reactive and respond to demyelination with a more pronounced cellular hypertrophy (Robillard et al., 2016; Boisvert et al., 2018; Clarke et al., 2018). Aged astrocytes were also demonstrated to have decreased expression of key cholesterol synthetic enzymes (Boisvert et al., 2018; Clarke et al., 2018), which may contribute to the reduced success of remyelination with age (Shields et al., 2000; van Wijngaarden and Franklin, 2013). Further study will help to elucidate the impact of age-related astrocytic changes to remyelination success, either directly through immunomodulatory effects or indirectly through interactions with other neural cell lineages populating the lesions, as well as reveal astrocyte features potentially amenable to therapeutic targeting.

MS affects 2–3 times more women than men (Whitacre et al., 1999), but men are more likely to have disease progression (Savettieri et al., 2004; Koch et al., 2009; Tomassini and Pozzilli, 2009; Shirani et al., 2012). Sex-specific diversity of astrocytes is notable in models of auto-immune demyelination (Chowen et al., 2017), reflecting this clinical observation. In EAE optic neuritis, optic nerve astrocytes in female mice display more robustly increased C3 expression (a component of the pro-inflammatory complement cascade; Stevens et al., 2007) and dampened upregulation of thombospondin-1 (*Thbs1*) when compared to their male counterparts (Tassoni et al., 2019), consistent with a more pro-inflammatory environment. Importantly, this is correlated with exaggerated retinal ganglion cell (RGC) and axonal loss in female mice (Tassoni et al., 2019), with a significant negative correlation between astrocytic expression of C3 and RGC density (Liddelow et al., 2017; Tassoni et al., 2019). This is particularly interesting given the female predominance of clinical MS, as well as the frequency of optic neuritis as a presentation in female MS patients (Voskuhl and Gold, 2012; Baecher-Allan et al., 2018; Voskuhl et al., 2018). Importantly, these sex-specific differences were not observed in spinal cord astrocytes in the same model (Tassoni et al., 2019), highlighting notable

regional diversity (Itoh et al., 2018). Further investigation to differentiate plasticity (e.g., hormone effects) from heterogeneity and to elucidate the functional relevance of this observed sex-specific diversity are important goals for future research (Chowen et al., 2017; Gilli et al., 2020).

Potential for Astrocyte Heterogeneity Based on Disease-Associated Variables in CNS Demyelination

In contrast to plasticity, we propose that astrocyte heterogeneity in models of CNS demyelination likely manifests as differences in regionally specific astrocyte populations in disparate CNS regions (e.g., brain regions, spinal cord, etc.). For example, scRNA-seq analysis of isolated murine astrocytes after EAE revealed distinct expression profiles for spinal cord, cerebellar, and hippocampal astrocytes (Itoh et al., 2018; Tassoni et al., 2019). Cholesterol synthesis pathways were significantly downregulated in WM-rich regions (e.g., spinal cord, cerebellum, optic nerve) due to altered ApoE-mediated cholesterol transport (Itoh et al., 2018; Tassoni et al., 2019). This is consistent with the downregulation of cholesterol synthesis observed in chronic demyelinated lesions in mouse MS models (Boisvert et al., 2018; Clarke et al., 2018), thought to be detrimental for remyelination given the lipid-rich nature of myelin sheaths (Itoh et al., 2018; Rawji et al., 2020). There are also notable differences between astrocytes that populate subcortical WM and GM lesions (Albert et al., 2007; Prins et al., 2015). Combined snRNA-seq and transcriptomic lesion mapping in MS tissue enabled mapping of dysregulated genes to either GM or WM astrocyte subpopulations (Schirmer et al., 2019) revealing that GM astrocytes in cortical lesions (i.e., GPC5+/SLC1A2+ cells) had decreased expression of genes involving glutamate and K⁺ homeostasis, whereas WM astrocytes in subcortical lesions (i.e., CD44+/LINC01088+ cells) displayed upregulated genes including *GFAP*, the transcription factors *BCL6* and *FOS*, and endothelin receptor B (Schirmer et al., 2019). Spinal cord astrocytes in LPC focal demyelination models were shown to have increased GFAP expression as compared to the cerebral cortex (Yoon et al., 2017). Molecular pathway analysis identified the antigen-presentation and interferon signaling pathways as specifically enriched in spinal cord and cerebellar astrocytes (Itoh et al., 2018). Antigen presentation by astrocytes has been previously shown in EAE models (Cornet et al., 2000; Constantinescu et al., 2005) and thought to be functionally important in astrocyte-mediated immunomodulation (Rawji et al., 2016, 2020). Regional diversity can also be seen in EAE optic neuritis models (Horstmann et al., 2016; Nolan et al., 2018; Tassoni et al., 2019). scRNA-seq revealed astrocyte-specific upregulation of the complement cascade and *Thbs1*, a gene involved in RGC synaptic plasticity and visual recovery following demyelination (Tassoni et al., 2019). Astrocytic *Thbs1* expression peaks early in disease (Tassoni et al., 2019), consistent with early astrocyte regulation of neural plasticity in demyelinated visual circuits. Optic nerve astrocytes also displayed reduced expression of cholesterol synthetic genes and increased expression of antigen presentation genes (Tassoni et al., 2019), similar to that seen

in spinal cord EAE lesions (Itoh et al., 2018). Diversity in gene expression was also noted between optic nerve and retinal astrocytes (Tassoni et al., 2019).

Regional diversity in EAE and MS is particularly interesting considering the multifocal nature of the disease (Baecher-Allan et al., 2018; Rawji et al., 2020). One can speculate that regional differences in astrocyte and other glial cell populations create an environment (e.g., ECM, cytokines, chemokines, etc.) more conducive to immune-mediated demyelination and/or for remyelination success. Astrocytes are known to exhibit regional specific responses to T-cell cytokines, leading to regionally specific neuroinflammatory responses in the hindbrain and spinal cord (Williams et al., 2020), suggesting that diverse astrocyte features may contribute to the anatomical propensity for demyelinated lesions to occur in certain locations (e.g., periventricular WM, spinal cord, optic nerve). Expression of pattern recognition receptors (PRRs) and interferon-induced genes, both at a basal level and in response to IFN induction, were higher in cerebellar than cortical astrocytes (Daniels et al., 2017), consistent with regionally specific astrocytic innate immune responses. Moreover, activation of the unfolded protein response alters the astrocytic secretome, generating a unique reactivity state and impairing synaptogenic function *in vitro* (Smith et al., 2020). Understanding the interplay between these environmental features and astrocytes is likely a fruitful area for therapeutic targeting, given the multitude of roles that astrocytes have in CNS demyelination. For example, GM lesions remyelinate more efficiently compared to WM (Albert et al., 2007; Bai et al., 2016; Strijbis et al., 2017), which could reflect, at least in part, contributions of regional specific astrocyte diversity. This will require detailed fate-mapping studies in models of CNS demyelination across lesion types, brain regions, and throughout the natural history of the condition.

TRAUMATIC INJURY

Traumatic CNS injuries remain the major causes of disability, premature death, and long-term neuropsychiatric impairment (Fitzharris et al., 2014; Gbd, 2016; National Spinal Cord Injury Statistical Center, 2021). Traumatic spinal cord injury (SCI) results from the application of an external force on the spine (e.g., motor vehicle accident, fall, sports-related injury, violent injury) damaging the underlying neural tissue in a variable manner (Ahuja et al., 2017; National Spinal Cord Injury Statistical Center, 2021). The primary insult results in massive damage to neural cells and triggers a complex cascade of secondary injury mechanisms that culminate in neuronal and glial cell death, ischemic injury, and inflammation (Alizadeh et al., 2019). This is generally followed by significant reorganization of spinal cord structure, including the formation of a densely packed astrogliosis border surrounding a cystic cavity at the lesion site (Sofroniew, 2009; Adams and Gallo, 2018; Alizadeh et al., 2019; Tran et al., 2021). Traumatic brain injury (TBI) is highly variable mechanistically with a wide range of causative insults and severities. Moreover, TBI has a complex post-injury course with numerous sequelae

that form part of a gradually evolving syndrome associated with chronic behavioral disturbances, seizure disorders, and protracted neurodegenerative diseases (Maas et al., 2008; Kovacs et al., 2014; Sharp et al., 2014). Reflecting the variable clinical presentation, TBI tissue pathology is highly inconsistent according to the type and severity of the injury and brain region involved (Sofroniew, 2009, 2015; Burda and Sofroniew, 2014). Due to the myriad and seemingly conflicting roles of astrocytes in the post-trauma environment, characterization of the contributions of heterogeneity and plasticity seems vital to the identification of targets for therapeutic manipulation both acutely for neuroprotective approaches and in the sub-acute/chronic period to enable for more effective axon regenerative approaches, which will continue to be the focus of extensive research efforts (Hilton et al., 2012; Cregg et al., 2014; Anderson et al., 2016; Geoffroy et al., 2016).

Several pre-clinical models of traumatic CNS injury have been utilized (Figure 3E; Jakeman et al., 2000; Metz et al., 2000; Dunham et al., 2010). Importantly, given the complexity of human traumatic injury, no one model alone can recreate all aspects of the injury process, therefore models are employed based on the study objectives and are thus limited in generalizability. In terms of injury mechanisms, SCI models can be broadly classified as contusion, crush, and transection (Hilton et al., 2013, 2019; Cheriyan et al., 2014), and recently emerging distraction and dislocation (Chen et al., 2016), with each mechanism having a unique tissue including glial response pattern. Contusion models are the most used due to their perceived clinical relevancy, although this relevancy has yet to be confirmed in a successful human trial of a neuroprotective treatment. The most common TBI models are controlled cortical impact (CCI) models and fluid percussion injury models, with the notable addition of models that mimic blast injuries or repeated mild TBI (i.e., concussion; Xiong et al., 2013). Differences in injury mechanism between the models and resultant secondary injury lead to differential astrocytic responses (Adams and Gallo, 2018; Boghdadi et al., 2020b), a consideration when attempting to generalize astrocyte responses across models.

Role of Astrocytes in Traumatic Injury

Astrocytes (Sofroniew, 2020), OPCs (Hackett et al., 2016), and fibroblasts (Goritz et al., 2011; Soderblom et al., 2013; Dias and Göritz, 2018) have all been shown to contribute to the densely packed accumulation of cells in close proximity to injury epicenter following SCI and TBI, with continual modulation by microglia and infiltrating innate and adaptive immune cells (Silver and Miller, 2004; Silver, 2016; O'Shea et al., 2017; Bradbury and Burnside, 2019). For the purposes of this review, we focus on data pertaining to the astrocytic component (Faulkner et al., 2004; Anderson et al., 2018; Gu et al., 2019; Yang et al., 2020) but recognize that reactive astrogliosis is often accompanied by clusters or intermingling fibroblast-like cells and OPCs (Faulkner et al., 2004; Anderson et al., 2018; Dias et al., 2018; Gu et al., 2019; Yang et al., 2020). In agreement with a recent suggestion to avoid using the term "glial scar" or "scar" (Sofroniew, 2020), which should be reserved for mesenchymal or stromal scar tissue, we will refer to the previously termed "glial scar" as

densely packed astrogliosis borders or densely packed reactive astrocytes. At the sub-acute/chronic traumatic injury epicenter (**Figure 3F**), there is a drastic loss of cells which overtime can result in the formation of a fluid-filled cyst surrounded by an inner layer of fibroblast-like cells and an outer layer of densely packed reactive astrocytes (Tran et al., 2021). Astrocytes are rapidly activated following traumatic injury culminating in the formation of a prominent densely packed astrogliosis border (Sofroniew, 2009; Adams and Gallo, 2018). Densely packed reactive astrocytes after TBI are partially regulated by monocyte invasion where reducing invasion using *CCR2*^{-/-} mice results in increased astrocyte proliferation but perhaps surprisingly, decreased GFAP+ scar area, ECM deposition, and lesion size compared to controls (Frik et al., 2018). While traditionally viewed as a barrier to axon regeneration and thus functional recovery, the post-traumatic densely packed astrogliosis border is also ascribed a beneficial function in the mitigation of further damage to the surrounding spared tissue (Adams and Gallo, 2018). For example, attenuating densely packed astrogliosis through signal transducer and activator of transcription 3 (*stat3*) deletion results in increased inflammation, lesion volume, and reduced motor recovery compared to controls (Herrmann et al., 2008). Importantly, the role of the densely packed astrogliosis border appears to be dynamic throughout the post-injury period, reflecting the multiplicity of functions ascribed to it (Anderson et al., 2016; Adams and Gallo, 2018).

Astrocyte Diversity in CNS Traumatic Injury

Diversity of reactive astrocytes following CNS trauma is thought to be driven largely by fluctuations in the inflammatory and cellular milieu of the post-injury environment (Hara et al., 2017; Adams and Gallo, 2018; Boghdadi et al., 2020b). RNA-seq performed on astrocytes 2 weeks following a spinal cord crush injury revealed differential expression of over 6,000 genes (Anderson et al., 2016), similar in magnitude with that observed post-ischemia (Zamanian et al., 2012; Anderson et al., 2016). Importantly, these reactive astrocytes demonstrate significant environmental-dependent plasticity (Hara et al., 2017; Boghdadi et al., 2020b). For example, integrin/N-cadherin-dependent collagen-1 signaling shapes astrocyte phenotypes in the post-injury spinal cord (Hara et al., 2017). Moreover, reactive astrocytes grafted into the spinal cord adopt an environmentally congruent phenotype (Hara et al., 2017). For example, reactive astrocytes grafted into the injured spinal cord retain the reactive phenotype, while those transplanted into the uninjured spinal cord revert phenotypically into resting astrocytes, confirmed by transcriptional analysis (Hara et al., 2017). After repetitive diffuse mild TBI, an atypical reactive astrocyte population was observed which featured a lack of GFAP expression, a downregulation of homeostatic proteins, and pronounced astrocyte coupling impairments, highlighting the importance of characterizing diversity across different types of injury (Shandra et al., 2019).

Extensive research has focused on manipulating the astrogliosis-associated deposition of growth-inhibitory ECM

molecules (e.g., chondroitin sulfate proteoglycans; CSPGs) in the context of traumatic injury (McKeon et al., 1995; Ponath et al., 2018) in an attempt to improve axonal regeneration (Bradbury et al., 2002; Cregg et al., 2014; Burda et al., 2016; Tran et al., 2018). In a rat model of TBI, CSPG expression is increased in a subset of peri-lesional astrocytes, peaking at 7 days post-injury, which is temporally correlated with a reduction in peri-neuronal net density and increased GAP-43+ neurons (Harris et al., 2009, 2010; Yi et al., 2012). This is consistent with the observed increase in neural plasticity, which is an important intrinsic mechanism of functional recovery following trauma. Additionally, subsets of astrocytes were observed to express the complement component C3 as early as 3 days post-injury with further increase through 28 days post-injury, mainly clustered around the lesion core (Qian et al., 2019). Whether these C3+ astrocytes represent neurotoxic or pro-inflammatory phenotypes akin to “A1” astrocytes (Liddel et al., 2017) remains to be determined. The potential role of these cells in axonal loss and retraction of transected axons from the lesion edge would be interesting to investigate as they remain key hurdles impeding regenerative and neuroprotective strategies in models of CNS trauma (Anderson et al., 2016; Geoffroy et al., 2016; Liddel et al., 2016; Hilton et al., 2017).

Potential for Astrocyte Plasticity Based on Disease-Associated Variables in Traumatic Injury

In the context of traumatic injury, we propose that astrocyte diversity across the disease-associated variables of time post-injury (i.e., temporal), distance from injury (i.e., topographical), age at which injury is sustained, and sex of the individual are examples where plasticity is likely to be predominant over heterogeneity. Extensive work has been performed to characterize reactive astrogliosis in the context of SCI (Filous and Silver, 2016; Silver, 2016; Bradbury and Burnside, 2019), focusing predominately on temporal changes in GFAP and CSPGs expression and morphological changes. Importantly, there are limited peer-reviewed transcriptomics data available to date. scRNA-seq was used to generate transcriptomics data based on temporal changes post-injury after mouse contusion SCI at 1, 3, and 7 dpi (Milich et al., 2021). Gene Ontology (GO) Enrichment Analysis for differentially expressed genes performed on astrocytes at 1 dpi were “translation” and “biosynthetic processes,” while by 3 dpi astrocytes were defined by “mitochondrial function” and “oxidative phosphorylation,” and at 7 dpi astrocytes were related to “lipid processing” (Milich et al., 2021). Despite numerous remaining questions regarding the extent and functional relevance of temporal astrocyte diversity following CNS traumatic injury, it appears probable that dynamic changes are evident following injury with shifting roles in the evolving injury and recovery processes, hence representing plasticity in response to environmental fluctuations. Further elucidation with fate-mapping studies and functional interrogation is key to revealing effective means of manipulating these processes to promote neuroprotection and recovery following injury.

As with other CNS pathologies, the degree of reactive astrogliosis in traumatic injury is highly dependent on the distance from the lesion, secondary to the gradation of pathology radiating outwards from the lesion epicenter, as reflected by parallel gradients of axonal injury, vascular disruption, ischemia, and inflammation (Wanner et al., 2013; Okada et al., 2018; Boghdadi et al., 2020b). GFAP expression, CSPG expression, astrocyte proliferation, and astrocyte density is highest directly adjacent the lesion, with a decreasing taper with increasing distance (White et al., 2009; Wanner et al., 2013). Importantly, in the injured spinal cord there appears to be diversity amongst locally intermingled astrocytes equidistant to the lesion, including variable expression of GFAP, NESTIN, and brain lipid-binding protein (BLBP; White et al., 2009) and only certain astrocyte subsets proliferate and/or polarize in response to a cortical stab injury (Bardehle et al., 2013), which are predominately juxta-vascular, reflecting the influence of blood-borne elements (Bardehle et al., 2013). These findings highlight the complexity and extent of astrocyte diversity that exists in CNS trauma, largely a reflection of the complex post-injury environment, in keeping with our definition of plasticity.

Despite the increasing prevalence of aged SCI patients, there is limited data on reactive astrocyte diversity in the aged CNS. One study examined astrocytes in young (i.e., 4 month old) and aged mice (i.e., 18-month-old) at 1, 3, and 7 days after a controlled cortical impact TBI (Early et al., 2020). Histological analysis demonstrated a significant increase in GFAP+ area in the aged mice compared to young, but notably only at 7 dpi (Early et al., 2020). Generally, transcriptomics data performed on these astrocyte populations found disproportionate changes in genes associated with reactive astrocytes in the aged mice after TBI compared to their young counterparts, as might be expected based on findings from other pathologies and normal aging process. Importantly, these profiles were not aligned with classic “A1/A2” phenotypes, once again highlighting the existence of a spectrum of reactive astrocyte subsets rather than a binary classification. Given the increasing importance of age in clinical presentation of traumatic CNS injury and the significant astrogliosis observed in animal models in the chronic SCI setting (months to years after injury; Silver and Miller, 2004), further work is needed to better characterize reactive astrogliosis diversity in the aged CNS and determine the functional relevance of these changes to disease outcomes (Burda et al., 2016; Okada et al., 2018).

Males are more commonly affected by CNS trauma than females (Cripps et al., 2011; Devivo, 2012; Gupte et al., 2019), attributed to increased risk-related behaviors. Despite this, there is limited data on sex-related astrocyte diversity in CNS trauma. In fact, because of the bladder problems associated with SCI models, 71% of pre-clinical SCI models are performed in female mice (Stewart et al., 2020), whose shorter urethra eases the stress of bladder expressions. In murine models of severe TBI, females display a larger GFAP+ area compared to males in the first week after injury which resolves by 30 dpi (Villapol et al., 2017). Moreover, chemokine C-C motif Ligand 2 (CCL2) expression was reduced after cortical injury in female astrocytes as compared to their male counterparts, potentially reflecting altered immune

cell recruitment (Acáz-Fonseca et al., 2015). In contrast, no sex-specific differences were noted in reactive astrocytes between male and female mice in a model of repetitive diffuse brain injury (Shandra et al., 2019), reflecting the importance of the specific model used. One of the main drivers of sex-specific differences is likely sex hormones, namely estrogen, which is in keeping with our definition of plasticity. The role of estrogens in astrocyte activation has been previously established (Arevalo et al., 2015) and recent data demonstrate that inhibiting estradiol synthesis promotes reactive astrogliosis in female mice after a controlled cortical impact TBI (Vierk et al., 2012), an effect not seen in males (Vierk et al., 2012). Therefore, the female CNS may be more sensitive to CNS trauma as compared to the male counterpart as was suggested previously (Vierk et al., 2012; Tabatadze et al., 2015; Bender et al., 2017). Further investigation of the influence of estrogens in models of CNS trauma is an important means of differentiation between hormonally driven effects (i.e., plasticity) and specific patterns of gene expression that are linked to sex chromosome codification (i.e., heterogeneity). 17 β -estradiol has been shown to have neuroprotective effects in pre-clinical models of SCI and TBI (Siriphorn et al., 2012; Day et al., 2017), and therefore targeted manipulation of estrogen signaling pathways may be a viable therapeutic target.

Potential for Astrocyte Heterogeneity Based on Disease-Associated Variables in Traumatic Injury

In contrast to plasticity, astrocyte heterogeneity in the context of traumatic injury may manifest as regionally variable responses to injuries at different locations across the neuraxis (i.e., brain vs. cervical spinal cord vs. thoracic spinal cord, etc.). Further characterization of astrocyte responses across models of CNS trauma utilizing fate-mapping and clonal analysis are important approaches to elucidate the extent of heterogeneity. For example, clonal analysis using the Star Track approach of proliferative astrocytes in a cortical stab injury model revealed a distinct progenitor cell origin as compared to non-proliferative astrocytes (Martín-López et al., 2013), consistent with our definition of heterogeneity. In the injured spinal cord, NSC-derived astrocytes more effectively constrain inflammation and the expansion of secondary damage (Sabelstrom et al., 2013), suggesting that distinct cellular origins may be a contributor to the diverse behavior of astrocytes observed following CNS trauma. Interestingly, a subset of reactive astrocytes in the post-trauma CNS displays certain characteristics of NSCs, including proliferative ability, multipotent lineage potential *in vitro*, and expression of several neurogenesis-associated genes (Buffo et al., 2008; Sirko et al., 2013). The potential for manipulation of this population has been demonstrated (Adams and Gallo, 2018; Magnusson et al., 2020; Zamboni et al., 2020) and represents a potentially attractive therapeutic approach to replace lost cell populations following trauma. Importantly, in models of CNS trauma there is also a potential contribution to observed diversity from astrocytes derived from other cells, including ependymal cells (Barnabé-Heider et al., 2010) and NG2 glia (Komitova et al., 2011; Hackett et al., 2016). For example, 2% of the astrocytes

after SCI were lineage-traced to FOXJ1 ependymal cells, which was dependent upon injury proximity to the central canal (Ren et al., 2017). Fate mapping approaches also revealed that NG2+ cells can give rise to a small percentage of densely packed GFAP+ astrocytes, both after a cortical stab injury (Komitova et al., 2011) and contusive SCI (Hackett et al., 2016). Distinct functionality and transcriptomic differences have not yet been established for these identified cell populations, representing important areas for further research.

As for ischemic stroke and demyelination, additional fate-mapping data in models of CNS trauma to establish the extent of heterogeneity is required. As CNS trauma can occur at various locations across the CNS, regional differences in the astrocyte populations may have a significant impact on injury outcome, as well as the success of functional recovery. For example, diverse astrocyte populations may be a key determinant of the extent of primary and secondary injury, considering the finding that elevated GFAP protein levels in cerebral spinal fluid after SCI are correlated with both baseline injury severity and poorer neurological outcomes (Skinnider et al., 2021a). Furthermore, differences in the local neuronal populations and the interplay between astrocytes and neurons in the post-injury environment may influence the success of neural circuit remodeling and thus functional recovery clinically. Understanding of the contribution of heterogeneity across brain regions, injury types (e.g., contusion, crush, etc.), and across stages of the disease process (e.g., acute, sub-acute, chronic) is therefore critical. This can be accomplished with performance of fate-mapping studies combined with high-resolution transcriptomic approaches and functional assays to interrogate the functional relevance of identified astrocyte diversity in diverse models of CNS trauma (e.g., SCI, TBI, concussion, etc.).

PERSPECTIVES ON ASTROCYTE DIVERSITY IN CNS DISEASE

Effective treatments are lacking for many neurological pathologies, including stroke, MS, and CNS trauma, resulting in significant morbidity and mortality. As astrocytes serve multiple vital roles in the post-insult CNS, they represent key players in disease pathogenesis, as well as promising targets for therapeutic manipulation. Characterization of the extent and functional relevance of astrocyte diversity in conditions of CNS insult is likely to aid in this endeavor: however, given the complexity of astrocyte reactivity (Lukovic et al., 2015; Anderson et al., 2016; Liddelov and Barres, 2016; Silver, 2016; Escartin et al., 2021) this will require a significant research effort. In this review, we summarize the current state of knowledge of astrocyte diversity in three representative and clinically relevant CNS pathologies; ischemic stroke, demyelination, and traumatic injury, highlighting the need for further characterization and functional interrogation of the contributions of heterogeneity vs. plasticity across temporal, topographical, age-specific, and sex-specific variables as a foundation to guide future development of therapeutic treatments to mitigate the impact of these conditions.

Areas of Future Research

Importantly, several knowledge gaps exist in the areas explored in this review. Most notably, there is a relative paucity of scRNA-seq studies characterizing astrocyte diversity in models of CNS trauma, as compared to demyelination and ischemic stroke. Moreover, as the vast majority of pre-clinical research has been performed in rodent models, increased use of non-human primate models or human tissue specimens is an important future avenue, as functional differences between species might pose a hurdle for clinical translation if not thoroughly addressed. Indeed, human astrocytes are known to be structurally more complex and diverse than rodent astrocytes (Oberheim et al., 2009; Zhang et al., 2016), as well as propagate Ca²⁺ waves approximately 4-fold faster than rodents in response to glutamate stimulation (Zhang et al., 2016), suggesting potential functional differences. Another critical area for future study is the influence of age on astrocyte diversity. This is particularly important not just for ischemic stroke, but for traumatic injury as well as older ages represent an increasing proportion of SCI and TBI, which is projected to increase in line with increasing age demographics in developed nations, and for progressive MS and the age-related decrease in remyelination efficiency. Considering that life expectancies of many of these clinical populations is increasing secondary to improved care and treatments, it will be essential to investigate astrocyte diversity after significant time periods after the onset of pathology (i.e., investigations in the 1–2 year post-injury chronic lesion sites within rodent models).

To meet these needs will require a combination of (1) increasing throughput of single-cell biology assays to achieve robust results across experimental conditions and biological replicates (Cao et al., 2020), (2) the application of multi-omic approaches to the context of CNS pathologies to avoid undue reliance on the functional interpretation of single-cell transcriptomes (Cao et al., 2018), and (3) bioinformatic methods development to aid biological investigators in differentiating cell type from cell state (Butler et al., 2018), to prioritize cell subsets most involved in the disease of interest (Skinnider et al., 2021b), and to avoid false positives that have the potential to drive research in unfruitful directions (Squair et al., 2021). Finally, these data will need to be met with stringent reporting standards to enable cross-disease investigations and to understand the conserved and differential responses of astrocytes across neurological disease.

In addition to conducting scRNA-seq-based characterization across multiple variables, it is increasingly important to further validate diverse astrocyte clusters in terms of origin and functionality, as most scRNA-seq analysis only provides a snapshot of gene expression within a specific context. Therefore, parallel assessments need to be performed to (1) validate mRNA and associated protein expression data, (2) establish the origin of identified clusters, and (3) assess functional differences. Furthermore, use of RNA-seq has the potential to identify cluster-specific, context-specific, or lineage-specific astrocyte markers across CNS states that will enable a more detailed interrogation of functionality and origin (e.g., with fate-mapping studies). New methods are also surfacing with the ability to predict ligand-targets links between interacting cells

(Browaeys et al., 2020) and using RABID-seq facilitates the simultaneous investigation of the transcriptome of specific cells interacting with your cell type of interest (Clark et al., 2021) in disease modeling which could further be a powerful tool. In addition, recent tools, such as transposase-accessible chromatin profiling (sc-ATAC-seq; Jia et al., 2018) have the potential to yield information about how chromatin accessibility dictates astrocyte diversity. New technology is also paving the way for sequencing both genomic DNA and mRNA in the same cell which could facilitate a direct comparison of genomic diversity and transcriptomic diversity (Dey et al., 2015). Multi-omics is allowing for increasingly complicated analysis with the potential to, for example, compare genomics, transcriptomics, proteomics, and metabolomics in the same cells. In addition, considering the importance of mitochondrial dynamics in astrocytes (Jackson and Robinson, 2018), future work looking at mitochondrial DNA as a potential source of genomic diversity will likely be important. As the definition of heterogeneity hangs on the demonstration of distinct functionality, detailed assessments across identified astrocyte clusters focusing on a wide range of function-related outcome measures is essential (e.g., Ca^{2+} signaling, neurotransmitter uptake/buffering, inter-cellular connectivity, neurotrophic factor production, etc.; for comprehensive list, see Escartin et al., 2021). Further characterizations of potential functional differences in how astrocytes respond to neuronal activity through G-protein-coupled receptor signaling *in vivo* will likely prove worthwhile (Nagai et al., 2021a). Further areas of research should focus on metabolic profiling to interrogate other potential functional differences between clusters of interest and the continued use of conditional knockout mice for loss-of-functions studies. The other requirement for demonstrating heterogeneity is cells with distinct origins which will require sound fate mapping follow-up studies. Recent sequencing methods are providing more insight specific to tracking clones of cells across time (Weinreb et al., 2020) which likely yields a powerful tool for demonstrating the potential district origin of cell clusters.

Astrocytes as Therapeutic Targets in CNS Disease

Astrocytes are promising therapeutic targets in several CNS diseases (Hamby and Sofroniew, 2010; Bradbury and Burnside, 2019; Valori et al., 2019). One potential approach is the use of viral vectors. Adeno-associated viruses (AAVs) demonstrate astrocyte-specific targeting in adult mice and non-human primates (Foust et al., 2009; Samaranch et al., 2012; Chan et al., 2017). Astrocyte-specific tropism may be increased through use of astrocyte-specific promoters (e.g., *Gfap*, *Aldh1L1*) and other viral capsid modifications (von Jonquieres et al., 2013; Meng et al., 2015; Vagner et al., 2016; Koh et al., 2017; Taschenberger et al., 2017). Lentiviral vectors pseudo-typed with glycoproteins from lymphocytic choriomeningitis virus (LCMV) or Moloney murine leukemia virus (MuLV) demonstrate astrocyte-specific targeting following intraparenchymal injection in rats (Cannon et al., 2011). Nanoparticles have also been used to target astrocytes (Zhou et al., 2018; Sharma et al., 2019), including the targeted regression of astrocyte-derived glioblastoma multiforme

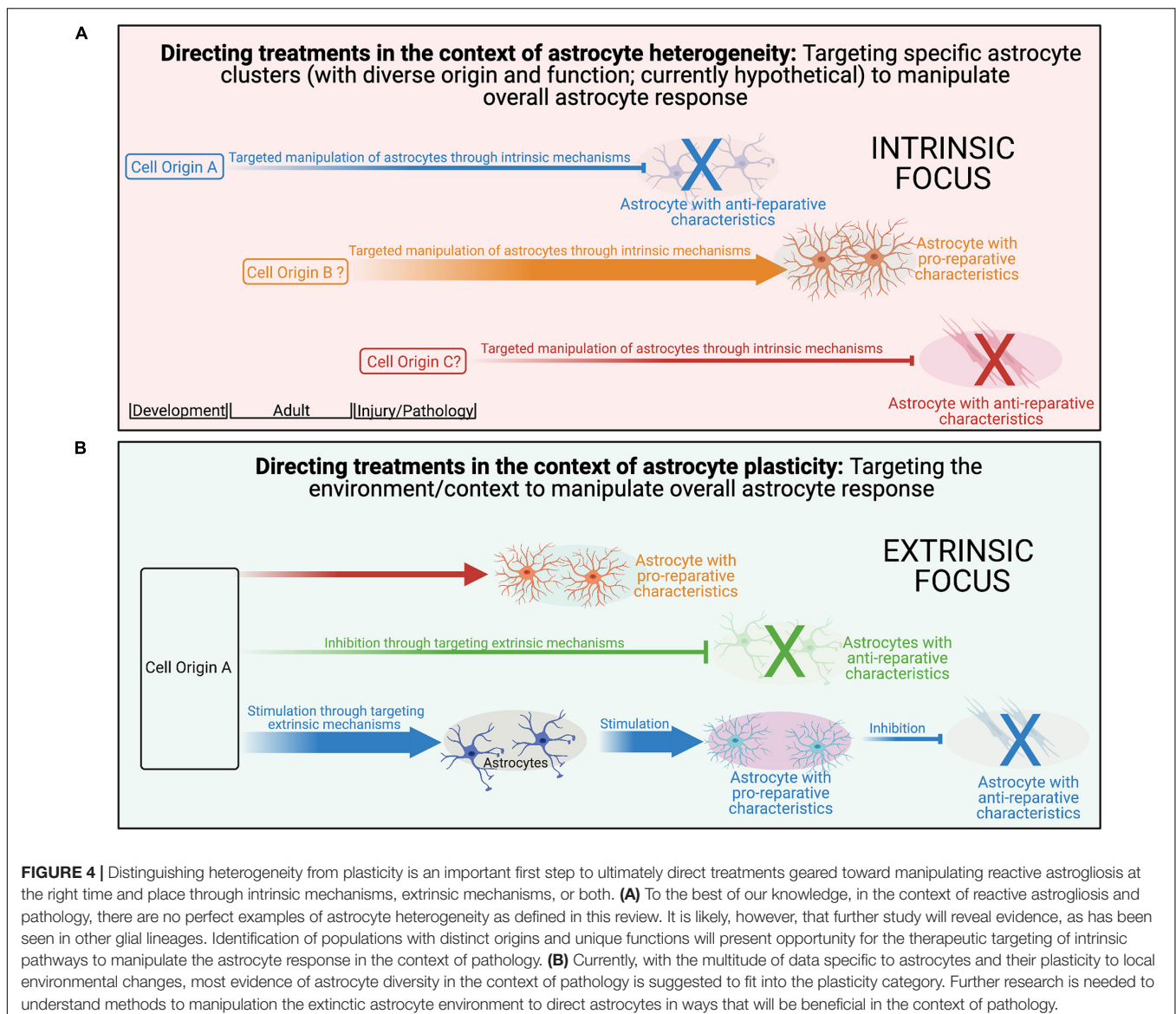
in mouse models (Jensen et al., 2013), inhibition of astrocyte-specific human immunodeficiency virus (HIV) replication *via* siRNA delivery (Gu et al., 2017), and astrocyte-specific delivery of mRNA *via* intraventricular administration (Tanaka et al., 2018). Alternatively, coupling of biologically active hydrophilic molecules (e.g., peptides, nucleic acids) to cell-permeable molecules can enable cell-specific delivery, as demonstrated in a mouse model of ALS (Martorana et al., 2012). Use of astrocyte-specific tropic factors, such as the phage AS1 homing peptide (Terashima et al., 2018) or herpes simplex virus type 1 (HSV-1) proteins (Valiante et al., 2015; Falanga et al., 2018) is a promising approach. Alternative approaches also include the use of Crispr-Cas9 gene-editing approach to specifically modify astrocytic genes (Huang and Nair, 2017; Kunze et al., 2018) and small molecules to modulate key signaling pathways (Zhang et al., 2015; Gao et al., 2017). As astrocytes are more resilient to the hostile post-injury environment in the CNS, direct cellular transplantation approaches may also be effective (Lepore et al., 2008; Izrael et al., 2018), although perhaps more limited in conditions, such as MS due to the disseminated nature of the lesions. Interestingly, dampening astrocyte reactivity (i.e., *via* STAT3 pathway inactivation) increases the number of OPC-derived Schwann cells in rodent demyelination lesions (Monteiro de Castro et al., 2015). In the context of the pronounced astrocyte loss seen in SCI, OPCs generate the majority of Schwann cells found in the contused lesion site (Assinck et al., 2017) and manipulating astrocytes to facilitate increased OPCs to differentiate into Schwann cells could be a promising alternative to invasive Schwann cell transplantation treatments which has been previously shown to promote repair and recovery in rat SCI models (Sparling et al., 2015; Assinck et al., 2020). Furthermore, astrocytes may represent an attractive source of cells for direct reprogramming to replace lost neurons or oligodendrocytes (Berninger et al., 2007; Heinrich et al., 2010). Forced expression of the transcription factors *Ngn2*, *Mash1*, or *Pax6* converts astrocytes to glutaminergic neuron-like cells in rodent models (Berninger et al., 2007), while overexpression of *Dlx2* or *Ascl1* converts them to GABAergic neuron-like cells (Heinrich et al., 2010; Liu et al., 2015). Further research is required to characterize the stability and functionality of these trans-differentiated “neurons,” but this is an intriguing approach. As transcriptional approaches and functional assessment further reveal the extent and relevance of astrocyte diversity in conditions of CNS disease there will undoubtedly be an expanded interest in astrocytes as therapeutic targets. Expansion of that knowledge is therefore critical to foster this development, specifically the identification of functionally relevant aspects of astrocyte diversity that are amendable to extrinsic manipulation (i.e., plasticity). This necessitates further research including expanded transcriptional studies, functional assays, and fate-mapping analyses to firmly establish and characterize this diversity.

Astrocyte Diversity: An Interplay of Plasticity and Heterogeneity

Using the framework proposed above, it remains unclear as to what proportion of the characterized astrocyte diversity in models of ischemic stroke, CNS demyelination, and traumatic

injury represent heterogeneity vs. plasticity. We propose that without unequivocal demonstration of functional differences and clear fate-mapping data on the origin of the identified astrocyte clusters, most of the identified diversity to date likely proportionally represents plasticity in response to a dynamically changing injury environment. This is a point that has been argued for diversity in the oligodendrocyte lineage (Foerster et al., 2019) and most likely holds true for astrocytes. It is likely that combinations of both heterogeneity and plasticity exist across a non-exhaustive number of variables including spatial, temporal, age, and sex, reflecting the superimposition of plasticity in response to fluctuating and complex environments upon a background of developmentally pre-determined variance. This has significant therapeutic implications, as our definition of heterogeneity defines a population of astrocytes with a unique origin and function that may be more amendable to intrinsic

manipulation approaches (Figure 4A). In contrast, plasticity implies a degree of malleability and thus would be amendable to extrinsic manipulation (Figure 4B). We propose here a simple model of intrinsic (e.g., viral targeting, DREADDs, optogenetics, transgenic manipulation, etc.) and extrinsic (e.g., manipulation of immune response, fibrotic scarring, ECM and stromal-derived matrix, exogenously administered cytokines, etc.) approaches for astrocyte-specific targeting, with the important caveat that extrinsic factors profoundly influence intrinsic mechanisms and no cell exists in complete isolation from their external environment, thus this distinction is not as clear-cut as presented here. Nonetheless, distinguishing heterogeneity from plasticity will yield a better understanding of the mechanisms of reactive astrogliosis and aspects of this response that can be targeted therapeutically. Given the significant interest in astrocytes as potential targets for therapeutic manipulation



(Zhang and Barres, 2010; Schitine et al., 2015; Matias et al., 2019) and the recent increase in tools to assess cell diversity, astrocyte diversity in CNS pathologies remains an exciting and promising field of future research.

CONCLUSION

Astrocytes display substantial diversity in ischemic stroke, CNS demyelination, and CNS traumatic injury, including temporal, topographical, sex-specific, and age-specific differences. Single-cell transcriptional approaches have significantly expanded our knowledge of the extent of this diversity, in the process altering our view of reactive astrocytes from a binary categorization to that of a spectrum of nuanced activation states involving hundreds of differentially expressed genes. Despite this, there are notable gaps in our knowledge, most notably whether this identified diversity represents heterogeneity or plasticity. As discussed throughout the review, the combination of transcriptomics approaches to further characterize the extent of diversity present with parallel functional assessments and expanded fate-mapping studies are critical areas of future study to fully develop our understanding of this diversity and to guide development of therapeutic approaches aimed at mitigating the morbidity and mortality of those afflicted with these conditions.

AUTHOR CONTRIBUTIONS

AJM, RJMF, WT, and PA contributed to the conception of review. AJM and PA contributed to design, original drafting,

and figure preparation of the review. AJM, JWS, RJMF, WT, and PA contributed to writing, editing, and approval of final manuscript. All authors contributed to the article and approved the submitted version.

FUNDING

JWS was supported by a Marie Skłodowska-Curie Individual Fellowship (MSCA-IF). RJMF was supported by the UK Multiple Sclerosis Society (MS50), the Adelson Medical Research Foundation, and a core support grant from the Wellcome Trust and MRC to the Wellcome Trust-Medical Research Council Cambridge Stem Cell Institute (203151/Z/16/Z). WT holds the John and Penny Ryan British Columbia Leadership Chair in Spinal Cord Injury Research and was funded by the Canadian Institute for Health Research (CIHR), MS Society of Canada, Craig H. Neilsen, Wings for Life, The US Department of Defense, and the Canadian Stem Cell Network. PA held a MS Society of Canada endMS Postdoctoral Fellowship and was currently supported by a Marie Skłodowska-Curie Individual Fellowship (MSCA-IF).

ACKNOWLEDGMENTS

We would like to thank Greg J. Duncan, Brett J. Hilton, Sarah Jäkel, Jason R. Plemel, Khalil S. Rawji, and Luise Seeker for their insightful discussions and/or valuable feedback on this review. The figures were created with BioRender.com.

REFERENCES

- Abbott, N. J., Ronnback, L., and Hansson, E. (2006). Astrocyte-endothelial interactions at the blood-brain barrier. *Nat. Rev. Neurosci.* 7:41. doi: 10.1038/nrn1824
- Acaz-Fonseca, E., Duran, J. C., Carrero, P., and Garcia-Segura, L. M. (2015). Sex differences in glia reactivity after cortical brain injury. *Glia* 63, 1966–1981. doi: 10.1002/glia.22867
- Adams, K. L., and Gallo, V. (2018). The diversity and disparity of the glial scar. *Nat. Neurosci.* 21, 9–15. doi: 10.1038/s41593-017-0033-9
- Ahuja, C. S., Wilson, J. R., Nori, S., Kotter, M. R. N., Druschel, C., Curt, A., et al. (2017). Traumatic spinal cord injury. *Nat. Rev. Dis. Primers* 3, 1–21. doi: 10.1038/nrdp.2017.18
- Albert, M., Antel, J., Brück, W., and Stadelmann, C. (2007). Extensive cortical remyelination in patients with chronic multiple sclerosis. *Brain Pathol.* 17, 129–138. doi: 10.1111/j.1750-3639.2006.00043.x
- Al-Dalahmah, O., Sosunov, A. A., Shaik, A., Ofori, K., Liu, Y., Vonsattel, J. P., et al. (2020). Single-nucleus RNA-seq identifies Huntington disease astrocyte states. *Acta Neuropathol. Commun.* 8, 1–21. doi: 10.1186/s40478-020-0880-6
- Alizadeh, A., Dyck, S. M., and Karimi-Abdolrezaee, S. (2019). Traumatic spinal cord injury: An overview of pathophysiology, models and acute injury mechanisms. *Front. Neurol.* 10:282. doi: 10.3389/fneur.2019.00282
- Alkayed, N. J., Harukuni, I., Kimes, A. S., London, E. D., Traystman, R. J., and Hurn, P. D. (1998). Gender-linked brain injury in experimental stroke. *Stroke* 29, 159–165.
- Amato, M. P., Derfuss, T., Hemmer, B., and Liblau, R. (2018). Environmental modifiable risk factors for multiple sclerosis: Report from the 2016 ECTRIMS focused workshop. *Multiple Sclerosis J.* 24, 590–603. doi: 10.1177/1352458516686847
- Anderson, M. A., Ao, Y., and Sofroniew, M. V. (2014). Heterogeneity of reactive astrocytes. *Neurosci. Lett.* 565, 23–29. doi: 10.1016/j.neulet.2013.12.030
- Anderson, M. A., Burda, J. E., Ren, Y., Ao, Y., O'Shea, T. M., Kawaguchi, R., et al. (2016). Astrocyte scar formation aids central nervous system axon regeneration. *Nature* 532, 195–200. doi: 10.1038/nature17623
- Anderson, M. A., O'Shea, T. M., Burda, J. E., Ao, Y., Barlately, S. L., Bernstein, A. M., et al. (2018). Required growth facilitators propel axon regeneration across complete spinal cord injury. *Nature* 561, 396–400. doi: 10.1038/s41586-018-0467-6
- Andriezen, W. L. (1893). The neuroglia elements in the human brain. *Br. Med. J.* 2:227. doi: 10.1136/bmj.2.1700.227
- Andrović, P., Kirdajova, D., Tureckova, J., and Zucha, D. (2020). Decoding the transcriptional response to ischemic stroke in young and aged mouse brain. *Cell Rep.* 31:107777. doi: 10.1016/j.celrep.2020.107777
- Anlauf, E., and Derouiche, A. (2013). Glutamine synthetase as an astrocytic marker: its cell type and vesicle localization. *Front. Endocrinol.* 4:144. doi: 10.3389/fendo.2013.00144
- Anthony, T. E., Klein, C., Fishell, G., and Heintz, N. (2004). Radial Glia serve as neuronal progenitors in all regions of the central nervous system. *Neuron* 41, 881–890. doi: 10.1016/S0896-6273(04)00140-0
- Arevalo, M.-A., Azcoitia, I., and Garcia-Segura, L. M. (2015). The neuroprotective actions of oestradiol and oestrogen receptors. *Nat. Rev. Neurosci.* 16, 17–29. doi: 10.1038/nrn3856

- Arneson, D., Zhang, G., Ying, Z., Zhuang, Y., Byun, H. R., Ahn, I. S., et al. (2018). Single cell molecular alterations reveal target cells and pathways of concussive brain injury. *Nat. Commun.* 9, 1–18. doi: 10.1038/s41467-018-06222-0
- Assinck, P., Duncan, G. J., Plemel, J. R., Lee, M. J., Stratton, J. A., Manesh, S. B., et al. (2017). Myelinogenic plasticity of oligodendrocyte precursor cells following spinal cord contusion injury. *J. Neurosci.* 37, 8635–8654. doi: 10.1523/JNEUROSCI.2409-16.2017
- Assinck, P., Sparling, J. S., Dworski, S., Duncan, G. J., Wu, D. L., Liu, J., et al. (2020). Transplantation of skin precursor-derived schwann cells yields better locomotor outcomes and reduces bladder pathology in rats with chronic spinal cord injury. *Stem Cell Rep.* 15, 140–155. doi: 10.1016/j.stemcr.2020.05.017
- Badan, I., Buchhold, B., Hamm, A., and Gratz, M. (2003). Accelerated Glial reactivity to stroke in aged rats correlates with reduced functional recovery. *J. Cerebral. Blood Flow Metabol.* 23:845. doi: 10.1097/01.WCB.0000071883.63724.A7
- Baecher-Allan, C., Kaskow, B. J., and Weiner, H. L. (2018). Multiple Sclerosis: mechanisms and immunotherapy. *Neuron* 97, 742–768. doi: 10.1016/j.neuron.2018.01.021
- Bai, C. B., Sun, S., Roholt, A., Benson, E., Edberg, D., Medicetty, S., et al. (2016). A mouse model for testing remyelinating therapies. *Exper. Neurol.* 283, 330–340. doi: 10.1016/j.expneurol.2016.06.033
- Baker, D., and Amor, S. (2015). Mouse models of multiple sclerosis: lost in translation? *Curr. Pharmaceut. Design* 21:2440. doi: 10.2174/1381612821666150316122706
- Bandeira, F., Lent, R., Herculano-Houzel, S., and Kaas, J. H. (2009). Changing numbers of neuronal and Non-Neuronal cells underlie postnatal brain growth in the rat. *Proc. Natl. Acad. Sci.* 106, 14108–14113. doi: 10.1073/pnas.0804650106
- Bang, O. Y., Saver, J. L., Buck, B. H., Alger, J. R., Starkman, S., Ovbiagele, B., et al. (2008). Impact of collateral flow on tissue fate in acute ischaemic stroke. *J. Neurol. Neurosurg. Psychiatry* 79, 625–629. doi: 10.1136/jnnp.2007.132100
- Bardehle, S., Krüger, M., Buggenthin, F., Schwausch, J., Ninkovic, J., Clevers, H., et al. (2013). Live imaging of astrocyte responses to acute injury reveals selective juxtavascular proliferation. *Nat. Neurosci.* 16, 580–586. doi: 10.1161/STROKEAHA.107.501460
- Barnabé-Heider, F., Göritz, C., Sabelström, H., Takebayashi, H., Pfrieger, F. W., Meletis, K., et al. (2010). Origin of new glial cells in intact and injured adult spinal cord. *Cell Stem Cell* 7, 470–482. doi: 10.1016/j.stem.2010.07.014
- Batiuk, M. Y., Martirosyan, A., Wahis, J., de Vin, F., Marneffe, C., Kusserow, C., et al. (2020). Identification of region-specific astrocyte subtypes at single cell resolution. *Nat. Commun.* 11:1220. doi: 10.1038/s41467-019-14198-8
- Bayraktar, O. A., Bartels, T., Holmqvist, S., Kleshchevnikov, V., Martirosyan, A., Polioudakis, D., et al. (2020). Astrocyte layers in the mammalian cerebral cortex revealed by a single-cell in situ transcriptomic map. *Nat. Neurosci.* 23, 500–509. doi: 10.1038/s41593-020-0602-1
- Bayraktar, O. A., Fuentealba, L. C., Alvarez-Buylla, A., and Rowitch, D. H. (2015). Astrocyte development and heterogeneity. *Cold Spring Harb. Perspect. Biol.* 7:a020362. doi: 10.1101/cshperspect.a020362
- Bazargani, N., and Attwell, D. (2016). Astrocyte calcium signaling: the third wave. *Nat. Neurosci.* 19, 182–189. doi: 10.1038/nn.4201
- Becerra-Calixto, A., and Cardona-Gómez, G. P. (2017). The role of astrocytes in neuroprotection after brain stroke: potential in cell therapy. *Front. Mol. Neurosci.* 10:88. doi: 10.3389/fnmol.2017.00088
- Bechler, M. E., and Byrne, L. (2015). CNS myelin sheath lengths are an intrinsic property of oligodendrocytes. *Curr. Biol.* 25, 2411–2416. doi: 10.1016/j.cub.2015.07.056
- Ben Haim, L., Carrillo-de Sauvage, M.-A., Ceyzériat, K., and Escartin, C. (2015). Elusive roles for reactive astrocytes in neurodegenerative diseases. *Front. Cell. Neurosci.* 9:278. doi: 10.3389/fncel.2015.00278
- Ben Haim, L., and Rowitch, D. H. (2017). Functional diversity of astrocytes in neural circuit regulation. *Nat. Rev. Neurosci.* 18:31. doi: 10.1038/nrn.2016.159
- Bender, R. A., Zhou, L., Vierk, R., Brandt, N., Keller, A., Gee, C. E., et al. (2017). Sex-dependent regulation of aromatase-mediated synaptic plasticity in the basolateral amygdala. *J. Neurosci.* 37, 1532–1545. doi: 10.1523/JNEUROSCI.1532-16.2016
- Benesova, J., Hock, M., Butenko, O., Prajerova, I., Anderova, M., and Chvatal, A. (2009). Quantification of astrocyte volume changes during ischemia in situ reveals two populations of astrocytes in the cortex of GFAP/EGFP mice. *J. Neurosci. Res.* 87, 96–111. doi: 10.1002/jnr.21828
- Ben-Nun, A., Kaushansky, N., Kawakami, N., and Krishnamoorthy, G. (2014). From classic to spontaneous and humanized models of multiple sclerosis: Impact on understanding pathogenesis and drug development. *J. Autoimmun.* 54, 33–50. doi: 10.1016/j.jaut.2014.06.004
- Berninger, B., Costa, M. R., Koch, U., Schroeder, T., Sutor, B., Grothe, B., et al. (2007). Functional properties of neurons derived from in vitro reprogrammed postnatal astroglia. *J. Neurosci.* 27, 8654–8664. doi: 10.1523/JNEUROSCI.1615-07.2007
- Blakemore, W. F., and Franklin, R. J. M. (2008). Remyelination in experimental models of toxin-induced demyelination. *Curr. Topics Microbiol. Immunol.* 318:193. doi: 10.1016/j.ejphar.2015.03.042
- Boghaddi, A. G., Spurrier, J., Teo, L., Li, M., Skarica, M., Cao, B., et al. (2020a). Primate-specific response of astrocytes to stroke limits peripheral macrophage infiltration. *bioRxiv* 2020:083501. doi: 10.1101/2020.05.08.083501
- Boghaddi, A. G., Teo, L., and Bourne, J. A. (2020b). The neuroprotective role of reactive astrocytes after central nervous system injury. *J. Neurotrauma* 37, 681–691. doi: 10.1089/neu.2019.6938
- Nagai, J., Yu, X., Papouin, T., Cheong, E., Freeman, M. R., Monk, K. R., et al. (2021b). Behaviorally consequential astrocytic regulation of neural circuits. *Neuron* 109, 576–596. doi: 10.1016/j.neuron.2020.12.008
- Nagai, J., Bellafard, A., Qu, Z., Yu, X., Ollivier, M., Gangwani, M. R., et al. (2021b). Specific and behaviorally consequential astrocyte G_q GPCR signaling attenuation in vivo with iβARK. *Neuron* S0896-6273, 376–377. doi: 10.1016/j.neuron.2021.05.023
- Boisvert, M. M., Erikson, G. A., Shokhirev, M. N., and Allen, N. J. (2018). The aging astrocyte transcriptome from multiple regions of the mouse brain. *Cell Rep.* 22, 269–285. doi: 10.1016/j.celrep.2017.12.039
- Bonneh-Barkay, D., Wang, G., LaFramboise, W. A., Wiley, C. A., and Bissel, S. J. (2012). Exacerbation of experimental autoimmune encephalomyelitis in the absence of breast regression Protein 39/Chitinase 3-Like 1. *J. Neuropathol. Exper. Neurol.* 71, 948–958. doi: 10.1097/NEN.0b013e31826aeae7
- Boulay, A.-C., Saubaméa, B., Adam, N., Chasseigneaux, S., Mazaré, N., Gilbert, A., et al. (2017). Translation in astrocyte distal processes sets molecular heterogeneity at the gliovascular interface. *Cell. Discov.* 3:17005. doi: 10.1038/celldisc.2017.5
- Bradbury, E. J., and Burnside, E. R. (2019). Moving beyond the glial scar for spinal cord repair. *Nat. Commun.* 10:3879. doi: 10.1038/s41467-019-11707-7
- Bradbury, E. J., Moon, L. D. F., Popat, R. J., King, V. R., Bennett, G. S., Patel, P. N., et al. (2002). Chondroitinase ABC promotes functional recovery after spinal cord injury. *Nature* 416, 636–640. doi: 10.1038/416636a
- Braga, F. A. V., Kar, G., Berg, M., Carpaij, O. A., Polanski, K., Simon, L. M., et al. (2019). A cellular census of human lungs identifies novel cell states in health and in asthma. *Nat. Med.* 25, 1153–1163. doi: 10.1038/s41591-019-0468-5
- Brambilla, R., Bracchi-Ricard, V., Hu, W.-H., and Frydel, B. (2005). Inhibition of astroglial nuclear factor κB reduces inflammation and improves functional recovery after spinal cord injury. *J. Exper. Med.* 202, 145–156. doi: 10.1084/jem.20041918
- Brambilla, R., Persaud, T., Hu, X., Karmally, S., Shestopalov, V. I., Dvorianchikova, G., et al. (2009). Transgenic inhibition of Astroglial NF-κB improves functional outcome in experimental autoimmune encephalomyelitis by suppressing chronic central nervous system inflammation. *J. Immunol.* 182, 2628–2640. doi: 10.4049/jimmunol.0802954
- Breunig, J. J., Gate, D., Levy, R., Rodriguez, J., Kim, G. B., Danielpour, M., et al. (2012). Rapid genetic targeting of pial surface neural progenitors and immature neurons by neonatal electroporation. *Neural. Dev.* 7:26. doi: 10.1186/1749-8104-7-26
- Brint, S., Jacewicz, M., Kiessling, M., Tanabe, J., and Pulsinelli, W. (1988). Focal brain ischemia in the rat: methods for reproducible neocortical infarction using tandem occlusion of the distal middle cerebral and ipsilateral common carotid arteries. *J. Cerebral. Blood Flow Metabol.* 8, 474–485. doi: 10.1038/jcbfm.1988.88
- Browaeys, R., Saelens, W., and Saeys, Y. (2020). NicheNet: modeling intercellular communication by linking ligands to target genes. *Nat. Methods* 17, 159–162. doi: 10.1038/s41592-019-0667-5

- Buenrostro, J. D., Wu, B., Litzenburger, U. M., Ruff, D., Gonzales, M. L., Snyder, M. P., et al. (2015). Single-cell chromatin accessibility reveals principles of regulatory variation. *Nature* 523, 486–490. doi: 10.1038/nature14590
- Buffo, A., Rite, I., Tripathi, P., and Lepier, A. (2008). Origin and progeny of reactive gliosis: A source of multipotent cells in the injured brain. *Proc. Natl. Acad. Sci.* 105, 3581–3586. doi: 10.1073/pnas.0709002105
- Buosi, A. S., Matias, I., Araujo, A. P. B., Batista, C., and Gomes, F. C. A. (2018). Heterogeneity in synaptogenic profile of astrocytes from different brain regions. *Mol. Neurobiol.* 55, 751–762. doi: 10.1007/s12035-016-0343-z
- Burda, J. E., Bernstein, A. M., and Sofroniew, M. V. (2016). Astrocyte roles in traumatic brain injury. *Exper. Neurol.* 275, 305–315. doi: 10.1016/j.expneurol.2015.03.020
- Burda, J. E., and Sofroniew, M. V. (2014). Reactive gliosis and the multicellular response to CNS damage and disease. *Neuron* 81, 229–248. doi: 10.1016/j.neuron.2013.12.034
- Bushong, E. A., Martone, M. E., Jones, Y. Z., and Ellisman, M. H. (2002). Protoplasmic astrocytes in CA1 stratum radiatum occupy separate anatomical domains. *J. Neurosci.* 22, 183–192. doi: 10.1523/JNEUROSCI.22-01-00183.2002
- Butler, A., Hoffman, P., Smibert, P., Papalexis, E., and Satija, R. (2018). Integrating single-cell transcriptomic data across different conditions, technologies, and species. *Nat. Biotechnol.* 36, 411–420. doi: 10.1038/nbt.4096
- Cahoy, J. D., Emery, B., Kaushal, A., Foo, L. C., Zamanian, J. L., Christopherson, K. S., et al. (2008). A transcriptome database for astrocytes, neurons, and Oligodendrocytes: A new resource for understanding brain development and function. *J. Neurosci.* 28, 264–278. doi: 10.1523/JNEUROSCI.4178-07.2008
- Cameron, D., Piccart-Gebhart, M. J., Gelber, R. D., Procter, M., Goldhirsch, A., de Azambuja, E., et al. (2017). 11 years' follow-up of trastuzumab after adjuvant chemotherapy in HER2-positive early breast cancer: final analysis of the HERceptin Adjuvant (HERA) trial. *Lancet* 389, 1195–1205. doi: 10.1016/S0140-6736(16)32616-2
- Campbell, B. C. V., De Silva, D. A., Macleod, M. R., Coutts, S. B., Schwamm, L. H., Davis, S. M., et al. (2019). Ischaemic stroke. *Nat. Rev. Dis. Primers* 5:70. doi: 10.1038/s41572-019-0118-8
- Campbell, B. C. V., and Khatri, P. (2020). Stroke. *Lancet* 396, 129–142. doi: 10.1016/S0140-6736(20)31179-X
- Cannon, J. R., Sew, T., Montero, L., Burton, E. A., and Greenamyre, J. T. (2011). Pseudotype-dependent lentiviral transduction of astrocytes or neurons in the rat substantia nigra. *Exper. Neurol.* 228, 41–52. doi: 10.1016/j.expneurol.2010.10.016
- Cantó, E., Reverter, F., Morcillo-Suárez, C., Matesanz, F., Fernández, O., Izquierdo, G., et al. (2012). Chitinase 3-like 1 plasma levels are increased in patients with progressive forms of multiple sclerosis. *Multiple Sclerosis* 18, 983–990. doi: 10.1177/1352458511433063
- Cao, J., Cusanovich, D. A., Ramani, V., Aghamirzaie, D., Pliner, H. A., Hill, A. J., et al. (2018). Joint profiling of chromatin accessibility and gene expression in thousands of single cells. *Science* 361, 1380–1385. doi: 10.1126/science.aau0730
- Cao, J., O'Day, D. R., Pliner, H. A., Kingsley, P. D., Deng, M., Daza, R. M., et al. (2020). A human cell atlas of fetal gene expression. *Science* 370:eaba7721. doi: 10.1126/science.aba7721
- Cao, J., Packer, J. S., Ramani, V., Cusanovich, D. A., Huynh, C., Daza, R., et al. (2017). Comprehensive single-cell transcriptional profiling of a multicellular organism. *Science* 357, 661–667. doi: 10.1126/science.aam8940
- Carswell, H. V., Dominiczak, A. F., Garcia-Segura, L. M., Harada, N., Hutchison, J. B., and Macrae, I. M. (2005). Brain aromatase expression after experimental stroke: topography and time course. *J. Steroid Biochem. Mol. Biol.* 96, 89–91. doi: 10.1016/j.jsmb.2005.02.016
- Cassidy, J. M., and Cramer, S. C. (2017). Spontaneous and therapeutic-induced mechanisms of functional recovery after stroke. *Transl. Stroke Res.* 8, 33–46. doi: 10.1007/s12975-016-0467-5
- Cerbai, F., Lana, D., Nosi, D., and Petkova-Kirova, P. (2012). The neuron-astrocyte-microglia triad in normal brain ageing and in a model of neuroinflammation in the rat hippocampus. *PLoS One* 7:e45250. doi: 10.1371/journal.pone.0045250
- Chai, H., Diaz-Castro, B., Shigetomi, E., and Monte, E. (2017). Neural circuit-specialized astrocytes: transcriptomic, proteomic, morphological, and functional evidence. *Neuron* 95, 531–549. doi: 10.1016/j.neuron.2017.06.029
- Chamorro, Á, Dirnagl, U., Urra, X., and Planas, A. M. (2016). Neuroprotection in acute stroke: targeting excitotoxicity, oxidative and nitrosative stress, and inflammation. *Lancet Neurol.* 15, 869–881. doi: 10.1016/S1474-4422(16)00114-9
- Chan, K. Y., Jang, M. J., Yoo, B. B., Greenbaum, A., Ravi, N., Wu, W.-L., et al. (2017). Engineered AAVs for efficient noninvasive gene delivery to the central and peripheral nervous systems. *Nat. Neurosci.* 20, 1172–1179. doi: 10.1038/nn.4593
- Chao, C.-C., Gutiérrez-Vázquez, C., Rothhammer, V., Mayo, L., Wheeler, M. A., Tjon, E. C., et al. (2019). Metabolic control of astrocyte pathogenic activity via cPLA2-MAVS. *Cell* 179, 1483–1498. doi: 10.1016/j.cell.2019.11.016
- Chen, C. Z., Neumann, B., Förster, S., and Franklin, R. J. M. (2021). Schwann cell remyelination of the central nervous system: why does it happen and what are the benefits? *Open Biol.* 11:200352. doi: 10.1098/rsob.200352
- Chen, K., Liu, J., Assinck, P., Bhatnagar, T., Strejiger, F., Zhu, Q., et al. (2016). Differential histopathological and behavioral outcomes eight weeks after rat spinal cord injury by contusion, dislocation, and distraction mechanisms. *J. Neurotrauma* 33, 1667–1684. doi: 10.1089/neu.2015.4218
- Chen, M. S., Huber, A. B., van der Haar, M. E., and Frank, M. (2000). Nogo-A is a myelin-associated neurite outgrowth inhibitor and an antigen for monoclonal antibody IN-1. *Nature* 403:434. doi: 10.1038/35000219
- Chen, P. C., Vargas, M. R., Pani, A. K., and Smeyne, R. J. (2009). Nrf2-Mediated neuroprotection in the MPTP mouse model of Parkinson's disease: critical role for the astrocyte. *Proc. Natl. Acad. Sci.* 106, 2933–2938. doi: 10.1073/pnas.0813361106
- Chen, S. T., Hsu, C. Y., Hogan, E. L., Maricq, H., and Balentine, J. D. (1986). A model of focal ischemic stroke in the rat: reproducible extensive cortical infarction. *Stroke* 17, 738–743. doi: 10.1161/01.STR.17.4.738
- Chen, X., Teichmann, S. A., and Meyer, K. B. (2018). From tissues to cell types and back: single-cell gene expression analysis of tissue architecture. *Ann. Rev. Biomed. Data Sci.* 1, 29–51. doi: 10.1146/annurev-biodatasci-080917-013452
- Cheriyian, T., Ryan, D. J., Weinreb, J. H., Cheriyian, J., Paul, J. C., Lafage, V., et al. (2014). Spinal cord injury models: a review. *Spinal Cord* 52, 588–595. doi: 10.1038/sc.2014.91
- Cheung, P., Khatri, P., Utz, P. J., and Kuo, A. J. (2019). Single-cell technologies—studying rheumatic diseases one cell at a time. *Nat. Rev. Rheumatol.* 15, 340–354. doi: 10.1038/s41584-019-0220-z
- Chisholm, N. C., and Sohrabji, F. (2016). Astrocytic response to cerebral ischemia is influenced by sex differences and impaired by aging. *Neurobiol. Dis.* 85, 245–253. doi: 10.1016/j.nbd.2015.03.028
- Chong, S. C., Rosenberg, S. S., Fancy, S. P., Zhao, C., Shen, Y.-A. A., Hahn, A. T., et al. (2012). Neurite outgrowth inhibitor Nogo-A establishes spatial segregation and extent of oligodendrocyte myelination. *Proc. Natl. Acad. Sci.* 109, 1299–1304. doi: 10.1073/pnas.1113540109
- Choudhury, G. R., and Ding, S. (2016). Reactive astrocytes and therapeutic potential in focal ischemic stroke. *Neurobiol. Dis.* 85, 234–244. doi: 10.1016/j.nbd.2015.05.003
- Chowen, J. A., Argente-Arízón, P., Freire-Regatillo, A., and Argente, J. (2017). Sex differences in the neuroendocrine control of metabolism and the implication of astrocytes. *Front. Neuroendocrinol.* 48, 3–12. doi: 10.1016/j.yfrne.2017.05.003
- Clark, I. C., Gutiérrez-Vázquez, C., Wheeler, M. A., Li, Z., Rothhammer, V., Linnerbauer, M., et al. (2021). Barcoded viral tracing of single-cell interactions in central nervous system inflammation. *Science* 372:eabf1230. doi: 10.1126/science.abf1230
- Clarke, L. E., and Barres, B. A. (2013). Emerging roles of astrocytes in neural circuit development. *Nat. Rev. Neurosci.* 14, 311–321. doi: 10.1038/nrn3484
- Clarke, L. E., Liddel, S. A., Chakraborty, C., and Münch, A. E. (2018). Normal aging induces A1-like astrocyte reactivity. *Proc. Natl. Acad. Sci.* 115, E1896–E1905. doi: 10.1073/pnas.1800165115
- Clarkson, B. D., Walker, A., Harris, M. G., Rayasam, A., Sandor, M., and Fabry, Z. (2015). CCR2-Dependent dendritic cell accumulation in the central nervous system during early effector experimental autoimmune encephalomyelitis is essential for Effector T cell restimulation in situ and disease progression. *J. Immunol.* 194, 531–541. doi: 10.4049/jimmunol.1401320
- Constantinescu, C. S., Farooqi, N., O'Brien, K., and Gran, B. (2011). Experimental autoimmune encephalomyelitis (EAE) as a model for multiple sclerosis (MS). *Br. J. Pharmacol.* 164, 1079–1106. doi: 10.1111/j.1476-5381.2011.01302.x

- Constantinescu, C. S., Tani, M., Ransohoff, R. M., Wysocka, M., Hilliard, B., Fujioka, T., et al. (2005). Astrocytes as antigen-presenting cells: expression of IL-12/IL-23. *J. Neurochem.* 95, 331–340. doi: 10.1111/j.1471-4159.2005.03368.x
- Cordeau, P. Jr., Lalancette-Hébert, M., Weng, Y. C., and Kriz, J. (2008). Live imaging of neuroinflammation reveals sex and estrogen effects on astrocyte response to ischemic injury. *Stroke* 39, 935–942.
- Cornet, A., Bettelli, E., Oukka, M., Cambouris, C., Avellana-Adalid, V., Kosmatopoulos, K., et al. (2000). Role of astrocytes in antigen presentation and naive T-cell activation. *J. Neuroimmunol.* 106, 69–77. doi: 10.1016/s0165-5728(99)00215-5
- Costa, M. R., Kessar, N., Richardson, W. D., and Gotz, M. (2007). The marginal Zone/Layer I as a novel niche for neurogenesis and gliogenesis in developing cerebral cortex. *J. Neurosci.* 27:11376. doi: 10.1523/JNEUROSCI.2418-07.2007
- Crawford, A. H., Tripathi, R. B., Richardson, W. D., and Franklin, R. J. M. (2016). Developmental origin of oligodendrocyte lineage cells determines response to demyelination and susceptibility to age-associated functional decline. *Cell Rep.* 15, 761–773. doi: 10.1016/j.celrep.2016.03.069
- Cregg, J. M., DePaul, M. A., Filous, A. R., Lang, B. T., Tran, A., and Silver, J. (2014). Functional regeneration beyond the glial scar. *Exper. Neurol.* 253, 197–207. doi: 10.1016/j.expneurol.2013.12.024
- Cripps, R. A., Lee, B. B., Wing, P., Weerts, E., Mackay, J., and Brown, D. (2011). A global map for traumatic spinal cord injury epidemiology: towards a living data repository for injury prevention. *Spinal Cord* 49, 493–501. doi: 10.1038/sc.2010.146
- Dal Canto, M. C., and Lipton, H. L. (1977). Multiple sclerosis. Animal model: Theiler's virus infection in mice. *Am. J. Pathol.* 88:497.
- Daniels, B. P., Ujjavarapu, H., Durrant, D. M., Williams, J. L., Green, R. R., White, J. P., et al. (2017). Regional astrocyte IFN signaling restricts pathogenesis during neurotropic viral infection. *J. Clin. Invest.* 127, 843–856. doi: 10.1172/JCI88720
- David, J. P., Ghazali, F., Fallet-Bianco, C., Watzet, A., Delaine, S., Boniface, B., et al. (1997). Glial reaction in the hippocampal formation is highly correlated with aging in human brain. *Neurosci. Lett.* 235, 53–56. doi: 10.1016/s0304-3940(97)00708-8
- Day, N. L., Carle, M. S., and Floyd, C. L. (2017). Post-injury administration of a combination of memantine and 17 β -estradiol is protective in a rat model of traumatic brain injury. *Neurochem. Int.* 111, 57–68. doi: 10.1016/j.neuint.2017.04.018
- De, S., Van Deren, D., Peden, E., Hockin, M., Boulet, A., Titen, S., et al. (2018). Two distinct ontogenies confer heterogeneity to mouse brain microglia. *Development* 145:dev152306. doi: 10.1242/dev.152306
- Denic, A., Johnson, A. J., Bieber, A. J., Warrington, A. E., Rodriguez, M., and Pirko, I. (2011). The relevance of animal models in multiple sclerosis research. *Pathophysiology* 18, 21–29.
- Devivo, M. J. (2012). Epidemiology of traumatic spinal cord injury: trends and future implications. *Spinal Cord* 50, 365–372. doi: 10.1038/sc.2011.178
- Dey, S. S., Kester, L., Spanjaard, B., Bienko, M., and Van Oudenaarden, A. (2015). Integrated genome and transcriptome sequencing of the same cell. *Nat. Biotechnol.* 33, 285–289. doi: 10.1038/nbt.3129
- Dias, D. O., and Göritz, C. (2018). Fibrotic scarring following lesions to the central nervous system. *Matrix Biol.* 68–69, 561–570. doi: 10.1016/j.matbio.2018.02.009
- Dias, D. O., Kim, H., Holl, D., Werne Solnestam, B., Lundeberg, J., Carlén, M., et al. (2018). Reducing pericyte-derived scarring promotes recovery after spinal cord injury. *Cell* 173, 153–165. doi: 10.1016/j.cell.2018.02.004
- Diaz-Castro, B., Gangwani, M. R., Yu, X., Coppola, G., and Khakh, B. S. (2019). Astrocyte molecular signatures in Huntington's disease. *Sci. Transl. Med.* 11:eaaw8546. doi: 10.1126/scitranslmed.aaw8546
- Ding, J., Adiconis, X., Simmons, S. K., Kowalczyk, M. S., Hession, C. C., Marjanovic, N. D., et al. (2020). Systematic comparison of single-cell and single-nucleus RNA-sequencing methods. *Nat. Biotechnol.* 38, 737–746. doi: 10.1038/s41587-020-0465-8
- Ding, S. (2013). In vivo astrocytic Ca2 signaling in health and brain disorders. *Future Neurol.* 8:529. doi: 10.2217/fnl.13.38
- Ding, S. (2014). "Ca2 Signaling in Astrocytes and its Role in Ischemic Stroke," in *Glutamate and ATP at the Interface of Metabolism and Signaling in the Brain Advances in Neurobiology*. New York: Springer International Publishing, 189–211. doi: 10.1007/978-3-319-08894-5_10
- Ding, S., Wang, T., Cui, W., and Haydon, P. G. (2009). Photothrombosis ischemia stimulates a sustained astrocytic Ca2 signaling in vivo. *Glia* 57, 767–776. doi: 10.1002/glia.20804
- Drögemüller, K., Helmuth, U., Brunn, A., Sakowicz-Burkiewicz, M., Gutmann, D. H., Mueller, W., et al. (2008). Astrocyte gp130 expression is critical for the control of toxoplasma encephalitis. *J. Immunol.* 181, 2683–2693. doi: 10.4049/jimmunol.181.4.2683
- Duffy, S., and MacVicar, B. A. (1996). In vitro ischemia promotes calcium influx and intracellular calcium release in hippocampal astrocytes. *J. Neurosci.* 16, 71–81. doi: 10.1523/JNEUROSCI.16-01-00071.1996
- Dunham, K. A., Siriphorn, A., Chompoopong, S., and Floyd, C. L. (2010). Characterization of a graded cervical hemiconfusion spinal cord injury model in adult male rats. *J. Neurotrauma* 27, 2091–2106. doi: 10.1089/neu.2010.1424
- Duran, R. C.-D., Wang, C.-Y., Zheng, H., Deneen, B., and Wu, J. Q. (2019). Brain region-specific gene signatures revealed by distinct astrocyte subpopulations unveil links to glioma and neurodegenerative diseases. *Environ. Neurosci.* 6:2019. doi: 10.1523/ENEURO.0288-18.2019
- Dzwonek, J., and Wilczynski, G. M. (2015). CD44: molecular interactions, signaling and functions in the nervous system. *Front. Cell. Neurosci.* 9:175. doi: 10.3389/fncel.2015.00175
- Early, A. N., Gorman, A. A., Van Eldik, L. J., Bachstetter, A. D., and Morganti, J. M. (2020). Effects of advanced age upon astrocyte-specific responses to acute traumatic brain injury in mice. *J. Neuroinflamm.* 17:115. doi: 10.1186/s12974-020-01800-w
- Eberwine, J., Yeh, H., Miyashiro, K., Cao, Y., Nair, S., Finnell, R., et al. (1992). Analysis of gene expression in single live neurons. *Proc. Natl. Acad. Sci.* 89, 3010–3014. doi: 10.1073/pnas.89.7.3010
- Eddleston, M., and Mucke, L. (1993). Molecular profile of reactive astrocytes—Implications for their role in neurologic disease. *Neuroscience* 54, 15–36. doi: 10.1016/0306-4522(93)90380-X
- Eng, L., Vanderhaeghen, J. J., Bignami, A., and Gerstl, B. (1971). An acidic protein isolated from fibrous astrocytes. *Brain Res.* 28, 351–354. doi: 10.1016/0006-8993(71)90668-8
- Escartin, C., Galea, E., Lakatos, A., O'Callaghan, J. P., Petzold, G. C., Serrano-Pozo, A., et al. (2021). Reactive astrocyte nomenclature, definitions, and future directions. *Nat. Neurosci.* 24, 312–325. doi: 10.1038/s41593-020-00783-4
- Escartin, C., Guillemaud, O., and Carrillo-de Sauvage, M.-A. (2019). Questions and (some) answers on reactive astrocytes. *Glia* 67, 2221–2247. doi: 10.1002/glia.23687
- Faiz, M., Sachewsky, N., Gascón, S., Bang, K. W. A., Morshead, C. M., and Nagy, A. (2015). Adult neural stem cells from the subventricular zone give rise to reactive Astrocytes in the cortex after stroke. *Cell Stem Cell* 17, 624–634. doi: 10.1016/j.stem.2015.08.002
- Falanga, A., Iachetta, G., Lombardi, L., Perillo, E., Lombardi, A., Morelli, G., et al. (2018). Enhanced uptake of gH625 by blood brain barrier compared to liver in vivo: characterization of the mechanism by an in vitro model and implications for delivery. *Sci. Rep.* 8, 1–13. doi: 10.1038/s41598-018-32095-w
- Farmer, W. T., Abrahamsson, T., Chierzi, S., Lui, C., Zaelzer, C., Jones, E. V., et al. (2016). Neurons diversify astrocytes in the adult brain through sonic hedgehog signaling. *Science* 351, 849–854. doi: 10.1126/science.aab3103
- Faulkner, J. R., Herrmann, J. E., Woo, M. J., and Tansey, K. E. (2004). Reactive astrocytes protect tissue and preserve function after spinal cord injury. *J. Neurosci.* 24:2143. doi: 10.1523/JNEUROSCI.3547-03.2004
- Filous, A. R., and Silver, J. (2016). Targeting astrocytes in CNS injury and disease: A translational research approach. *Prog. Neurobiol.* 144, 173–187. doi: 10.1016/j.pneurobio.2016.03.009
- Fitzharris, M., Cripps, R. A., and Lee, B. B. (2014). Estimating the global incidence of traumatic spinal cord injury. *Spinal Cord* 52, 117–122. doi: 10.1038/sc.2013.135
- Fluri, F., Schuhmann, M. K., and Kleinschnitz, C. (2015). Animal models of ischemic stroke and their application in clinical research. *Drug Design Dev. Ther.* 9:3445. doi: 10.2147/DDDT.S56071
- Foerster, S., Hill, M. F. E., and Franklin, R. J. M. (2019). Diversity in the oligodendrocyte lineage: Plasticity or heterogeneity? *Glia* 67, 1797–1805. doi: 10.1002/glia.23607
- Foust, K. D., Nurre, E., Montgomery, C. L., Hernandez, A., Chan, C. M., and Kaspar, B. K. (2009). Intravascular AAV9 preferentially targets neonatal neurons and adult astrocytes. *Nat. Biotechnol.* 27, 59–65. doi: 10.1038/nbt.1515

- Franceschi, C., Bonafè, M., Valensin, S., Olivieri, F., De Luca, M., Ottaviani, E., et al. (2000). Inflamm-aging: an evolutionary perspective on immunosenescence. *Ann. N.Y. Acad. Sci.* 908, 244–254. doi: 10.1111/j.1749-6632.2000.tb06651.x
- Franklin, R. J. M., and Goldman, S. A. (2015). Glia disease and repair—remyelination. *Cold Spring Harb. Perspect. Biol.* 7:a020594. doi: 10.1101/cshperspect.a020594
- Frey, D., Laux, T., Xu, L., Schneider, C., and Caroni, P. (2000). Shared and unique roles of CAP23 and GAP43 in actin regulation, neurite outgrowth, and anatomical plasticity. *J. Cell Biol.* 149, 1443–1454.
- Frik, J., Merl-Pham, J., Plesnila, N., Mattugini, N., Kjell, J., Kraska, J., et al. (2018). Cross-talk between monocyte invasion and astrocyte proliferation regulates scarring in brain injury. *EMBO Rep.* 19:e45294. doi: 10.15252/embr.201745294
- Gao, L., Guan, W., Wang, M., Wang, H., Yu, J., Liu, Q., et al. (2017). Direct generation of human neuronal cells from adult astrocytes by small molecules. *Stem Cell Rep.* 8, 538–547. doi: 10.1016/j.stemcr.2017.01.014
- Gao, P., Postiglione, M. P., Krieger, T. G., Hernandez, L., Wang, C., Han, Z., et al. (2014). Deterministic progenitor behavior and unitary production of neurons in the neocortex. *Cell* 159, 775–788. doi: 10.1016/j.cell.2014.10.027
- Gbd. (2016). Global, regional, and national burden of traumatic brain injury and spinal cord injury, 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016. *Lancet Neurol.* 18, 56–87. doi: 10.1016/S1474-4422(18)30415-0
- Ge, W. P., Miyawaki, A., Gage, F. H., Jan, Y. N., and Jan, L. Y. (2012). Local generation of glia is a major astrocyte source in postnatal cortex. *Nature* 484, 376–380. doi: 10.1038/nature10959
- Geoffroy, C. G., Hilton, B. J., Tetzlaff, W., and Zheng, B. (2016). Evidence for an age-dependent decline in axon regeneration in the adult mammalian central nervous system. *Cell Rep.* 15, 238–246. doi: 10.1016/j.celrep.2016.03.028
- Geric, B., Szalay, F., and Hajós, F. (2006). Glial fibrillary acidic protein immunoreactivity in the rat suprachiasmatic nucleus: circadian changes and their seasonal dependence. *J. Anat.* 209, 231–237. doi: 10.1111/j.1469-7580.2006.00593.x
- Geyer, S., Jacobs, M., and Hsu, N.-J. (2019). Immunity against bacterial infection of the central nervous system: An astrocyte perspective. *Front. Mol. Neurosci.* 12:57. doi: 10.3389/fnmol.2019.00057
- Ghashghaei, H. T., Weimer, J. M., Schmid, R. S., Yokota, Y., McCarthy, K. D., Popko, B., et al. (2007). Reinduction of ErbB2 in astrocytes promotes radial glial progenitor identity in adult cerebral cortex. *Genes Dev.* 21, 3258–3271. doi: 10.1101/gad.1580407
- Gilli, F., DiSano, K. D., and Pachner, A. R. (2020). SeXX matters in multiple sclerosis. *Front. Neurol.* 11:616. doi: 10.3389/fneur.2020.00616
- Gliem, M., Krammes, K., Liaw, L., van Rooijen, N., Hartung, H.-P., and Jander, S. (2015). Macrophage-derived osteopontin induces reactive astrocyte polarization and promotes re-establishment of the blood brain barrier after ischemic stroke. *Glia* 63, 2198–2207. doi: 10.1002/glia.22885
- Goritz, C., Dias, D. O., Tomilin, N., Barbacid, M., Shupliakov, O., and Frisen, J. (2011). A pericyte origin of spinal cord scar tissue. *Science* 333, 238–242. doi: 10.1126/science.1203165
- GrandPre, T., Nakamura, F., Vartanian, T., and Strittmatter, S. M. (2000). Identification of the Nogo inhibitor of axon regeneration as a reticulon protein. *Nature* 403:439. doi: 10.1038/35000226
- Griemsmann, S., Höft, S. P., Bedner, P., Zhang, J., Von Staden, E., Beinhauer, A., et al. (2015). Characterization of panglial gap junction networks in the thalamus, neocortex, and hippocampus reveals a unique population of glial cells. *Cerebral Cortex* 25, 3420–3433. doi: 10.1093/cercor/bhu157
- Grubman, A., Chew, G., Ouyang, J. F., Sun, G., Choo, X. Y., McLean, C., et al. (2019). A single-cell atlas of entorhinal cortex from individuals with Alzheimer's disease reveals cell-type-specific gene expression regulation. *Nat. Neurosci.* 22, 2087–2097. doi: 10.1038/s41593-019-0539-4
- Gu, J., Al-Bayati, K., and Ho, E. A. (2017). Development of antibody-modified chitosan nanoparticles for the targeted delivery of siRNA across the blood-brain barrier as a strategy for inhibiting HIV replication in astrocytes. *Drug Delivery Transl. Res.* 7, 497–506. doi: 10.1007/s13346-017-0368-5
- Gu, Y., Cheng, X., Huang, X., and Yuan, Y. (2019). Conditional ablation of reactive astrocytes to dissect their roles in spinal cord injury and repair. *Brain Behav. Immun.* 80, 394–405. doi: 10.1016/j.bbi.2019.04.016
- Gupte, R. P., Brooks, W. M., Vukas, R. R., Pierce, J. D., and Harris, J. L. (2019). Sex differences in traumatic brain injury: What we know and what we should know. *J. Neurotrauma* 36, 3063–3091. doi: 10.1089/neu.2018.6171
- Haanstra, K. G., Dijkman, K., Bashir, N., Bauer, J., Mary, C., Poirier, N., et al. (2015). Selective blockade of CD28-mediated T cell costimulation protects rhesus monkeys against acute fatal experimental autoimmune encephalomyelitis. *J. Immunol.* 194, 1454–1466. doi: 10.4049/jimmunol.1402563
- Habib, N., McCabe, C., Medina, S., Varshavsky, M., Kitsberg, D., Dvir-Szternfeld, R., et al. (2020). Disease-associated astrocytes in Alzheimer's disease and aging. *Nat. Neurosci.* 23, 701–706. doi: 10.1038/s41593-020-0624-8
- Hackett, A. R., Lee, D.-H., Dawood, A., Rodriguez, M., Funk, L., Tsoulfas, P., et al. (2016). STAT3 and SOCS3 regulate NG2 cell proliferation and differentiation after contusive spinal cord injury. *Neurobiol. Dis.* 89, 10–22. doi: 10.1016/j.nbd.2016.01.017
- Hall, E. D., Pazara, K. E., and Linseman, K. L. (1991). Sex differences in postischemic neuronal necrosis in gerbils. *J. Cerebral Blood Flow Metab.* 11, 292–298. doi: 10.1038/jcbfm.1991.61
- Hamby, M. E., and Sofroniew, M. V. (2010). Reactive astrocytes as therapeutic targets for CNS disorders. *Neurotherapeutics* 7, 494–506. doi: 10.1016/j.nurt.2010.07.003
- Hara, M., Kobayakawa, K., Ohkawa, Y., Kumamaru, H., Yokota, K., Saito, T., et al. (2017). Interaction of reactive astrocytes with type I collagen induces astrocytic scar formation through the integrin-N-cadherin pathway after spinal cord injury. *Nat. Med.* 23, 818–828. doi: 10.1038/nm.4354
- Haroon, F., Drögemüller, K., Händel, U., Brunn, A., Reinhold, D., Nishanth, G., et al. (2011). Gp130-dependent astrocytic survival is critical for the control of autoimmune central nervous system inflammation. *J. Immunol.* 186, 6521–6531. doi: 10.4049/jimmunol.1001135
- Harris, N. G., Carmichael, S. T., Hovda, D. A., and Sutton, R. L. (2009). Traumatic brain injury results in disparate regions of chondroitin sulfate proteoglycan expression that are temporally limited. *J. Neurosci. Res.* 87, 2937–2950. doi: 10.1002/jnr.22115
- Harris, N. G., Mironova, Y. A., Hovda, D. A., and Sutton, R. L. (2010). Pericontusion axon sprouting is spatially and temporally consistent with a growth-permissive environment after traumatic brain injury. *J. Neuropathol. Exper. Neurol.* 69, 139–154. doi: 10.1097/NEN.0b013e3181cb5bee
- Hartmann, K., Sepulveda-Falla, D., Rose, I. V. L., Madore, C., Muth, C., Matschke, J., et al. (2019). Complement 3+ astrocytes are highly abundant in prion diseases, but their abolishment led to an accelerated disease course and early dysregulation of microglia. *Acta Neuropathol. Commun.* 7:83. doi: 10.1186/s40478-019-0735-1
- Hayakawa, N., Kato, H., and Araki, T. (2007). Age-related changes of astrocytes, oligodendrocytes and microglia in the mouse hippocampal CA1 sector. *Mechan. Ageing Dev.* 128, 311–316. doi: 10.1016/j.mad.2007.01.005
- Heiland, H. D., Ravi, V. M., Behringer, S. P., Frenking, J. H., Wurm, J., Joseph, K., et al. (2019). Tumor-associated reactive astrocytes aid the evolution of immunosuppressive environment in glioblastoma. *Nat. Commun.* 10:2541. doi: 10.1038/s41467-019-10493-6
- Heinrich, C., Blum, R., Gascón, S., Masserdotti, G., Tripathi, P., Sánchez, R., et al. (2010). Directing astroglia from the cerebral cortex into subtype specific functional neurons. *PLoS Biol.* 8:e1000373. doi: 10.1371/journal.pbio.1000373
- Herrmann, J. E., Imura, T., Song, B., Qi, J., Ao, Y., Nguyen, T. K., et al. (2008). STAT3 is a critical regulator of astroglia and scar formation after spinal cord injury. *J. Neurosci.* 28, 7231–7243. doi: 10.1523/JNEUROSCI.1709-08.2008
- Hilton, B. J., Assinck, P., Duncan, G. J., Lu, D., Lo, S., and Tetzlaff, W. (2013). Dorsolateral funiculus lesioning of the mouse cervical spinal cord at C4 but Not at C6 results in sustained forelimb motor deficits. *J. Neurotrauma* 30, 1070–1083. doi: 10.1089/neu.2012.2734
- Hilton, B. J., Blanquie, O., Tedeschi, A., and Bradke, F. (2019). High-resolution 3D imaging and analysis of axon regeneration in unsectioned spinal cord with or without tissue clearing. *Nat. Protocols* 14, 1235–1260. doi: 10.1038/s41596-019-0140-z
- Hilton, B. J., Lang, B. T., and Cregg, J. M. (2012). Keratan sulfate proteoglycans in plasticity and recovery after spinal cord injury. *J. Neurosci.* 32, 4331–4333. doi: 10.1523/JNEUROSCI.0333-12.2012
- Hilton, B. J., Moulson, A. J., and Tetzlaff, W. (2017). Neuroprotection and secondary damage following spinal cord injury: concepts and methods. *Neurosci. Lett.* 652, 3–10. doi: 10.1016/j.neulet.2016.12.004

- Hinks, G. L., and Franklin, R. J. (1999). Distinctive patterns of PDGF-A, FGF-2, IGF-I, and TGF-beta1 gene expression during remyelination of experimentally-induced spinal cord demyelination. *Mol. Cell. Neurosci.* 14, 153–168. doi: 10.1006/mcne.1999.0771
- Hirsch, E., Gullberg, D., Balzac, F., Altruda, F., Silengo, L., and Tarone, G. (1994). αv integrin subunit is predominantly located in nervous tissue and skeletal muscle during mouse development. *Dev. Dynamics* 201, 108–120. doi: 10.1002/aja.1002010203
- Hochstim, C., Deneen, B., Lukaszewicz, A., Zhou, Q., and Anderson, D. J. (2008). Identification of positionally distinct astrocyte subtypes whose identities are specified by a homeodomain code. *Cell* 133, 510–522. doi: 10.1016/j.cell.2008.02.046
- Höft, S., Griemsmann, S., Seifert, G., and Steinhäuser, C. (2014). Heterogeneity in expression of functional ionotropic glutamate and GABA receptors in astrocytes across brain regions: insights from the thalamus. *Philos. Trans. R. Soc. B Biol. Sci.* 369:20130602. doi: 10.1098/rstb.2013.0602
- Horstmann, L., Kuehn, S., Pedreiturria, X., Haak, K., Pfarrer, C., Dick, H. B., et al. (2016). Microglia response in retina and optic nerve in chronic experimental autoimmune encephalomyelitis. *J. Neuroimmunol.* 298, 32–41. doi: 10.1016/j.jneuroim.2016.06.008
- Hossmann, K.-A. (2012). The two pathophysiologicals of focal brain ischemia: implications for translational stroke research. *J. Cerebral. Blood Flow Metabol.* 32, 1310–1316. doi: 10.1038/jcbfm.2011.186
- Huang, W., Zhao, N., Bai, X., Karram, K., Trotter, J., Goebels, S., et al. (2014). Novel NG2-CreERT2 knock-in mice demonstrate heterogeneous differentiation potential of NG2 glia during development: Differentiation in NG2-CreERT2 Knock-In Mice. *Glia* 62, 896–913. doi: 10.1002/glia.22648
- Huang, Z., and Nair, M. (2017). A CRISPR/Cas9 guidance RNA screen platform for HIV provirus disruption and HIV/AIDS gene therapy in astrocytes. *Sci. Rep.* 7, 1–12. doi: 10.1038/s41598-017-06269-x
- Hung, C. C., Lin, C. H., Chang, H., Wang, C.-Y., Lin, S.-H., Hsu, P.-C., et al. (2016). Astrocytic GAP43 Induced by the TLR4/NF- κ B/STAT3 axis attenuates astrogliosis-mediated microglial activation and Neurotoxicity. *J. Neurosci.* 36, 2027–2043. doi: 10.1523/JNEUROSCI.3457-15.2016
- Isokawa, M., and McKhann, G. M. (2005). Electrophysiological and morphological characterization of dentate astrocytes in the hippocampus. *J. Neurobiol.* 65, 125–134. doi: 10.1002/neu.20186
- Italiani, P., and Boraschi, D. (2014). From monocytes to M1/M2 Macrophages: Phenotypical vs. Functional differentiation. *Front. Immunol.* 5:514. doi: 10.3389/fimmu.2014.00514
- Itoh, N., Itoh, Y., Tassoni, A., Ren, E., Kaito, M., Ohno, A., et al. (2018). Cell-specific and region-specific transcriptomics in the multiple sclerosis model: Focus on astrocytes. *PNAS* 115, E302–E309. doi: 10.1073/pnas.1716032115
- Izrael, M., Slutsky, S. G., Admoni, T., Cohen, L., Granit, A., Hasson, A., et al. (2018). Safety and efficacy of human embryonic stem cell-derived astrocytes following intrathecal transplantation in SOD1 G93A and NSG animal models. *Stem Cell Res. Ther.* 9, 1–17. doi: 10.1186/s13287-018-0890-5
- Jackson, J. G., and Robinson, M. B. (2018). Regulation of mitochondrial dynamics in astrocytes: Mechanisms, consequences, and unknowns. *Glia* 66, 1213–1234. doi: 10.1002/glia.23252
- Jäkel, S., and Williams, A. (2020). What have advances in transcriptomic technologies taught us about human white matter pathologies? *Front. Cell. Neurosci.* 14:238. doi: 10.3389/fncel.2020.00238
- Jakeman, L. B., Guan, Z., Wei, P., Ponnappan, R., Dzwonczyk, R., Popovich, P. G., et al. (2000). Traumatic spinal cord injury produced by controlled contusion in mouse. *J. Neurotrauma* 17, 299–319. doi: 10.1089/neu.2000.17.299
- Jensen, S. A., Day, E. S., Ko, C. H., Hurley, L. A., Luciano, J. P., Kouri, F. M., et al. (2013). Spherical nucleic acid nanoparticle conjugates as an RNAi-based therapy for glioblastoma. *Sci. Transl. Med.* 5:209ra152. doi: 10.1126/scitranslmed.3006839
- Jessen, N. A., Munk, A. S. F., Lundgaard, I., and Nedergaard, M. (2015). The glymphatic system: A Beginner's guide. *Neurochem. Res.* 40, 2583–2599. doi: 10.1007/s11064-015-1581-6
- Jia, G., Preussner, J., Chen, X., Guenther, S., Yuan, X., Yekelchik, M., et al. (2018). Single cell RNA-seq and ATAC-seq analysis of cardiac progenitor cell transition states and lineage settlement. *Nat. Commun.* 9, 1–17. doi: 10.1038/s41467-018-07307-6
- Jordan, V. C. (2003). Tamoxifen: a most unlikely pioneering medicine. *Nat. Rev. Drug Discov.* 2, 205–213. doi: 10.1038/nrd1031
- Jyothi, H. J., Vidyadhara, D. J., Mahadevan, A., and Philip, M. (2015). Aging causes morphological alterations in astrocytes and microglia in human substantia nigra pars compacta. *Neurobiol. Aging* 36, 3321–3333. doi: 10.1016/j.neurobiolaging.2015.08.024
- Kanaan, N. M., Kordower, J. H., and Collier, T. J. (2010). Age-related changes in glial cells of dopamine midbrain subregions in rhesus monkeys. *Neurobiol. Aging* 31, 937–952. doi: 10.1016/j.neurobiolaging.2008.07.006
- Khakh, B. S., and Deneen, B. (2019). The emerging nature of astrocyte diversity. *Annu. Rev. Neurosci.* 42, 187–207. doi: 10.1146/annurev-neuro-070918-050443
- Khakh, B. S., and Sofroniew, M. V. (2015). Diversity of astrocyte functions and phenotypes in neural circuits. *Nat. Neurosci.* 18, 942–953. doi: 10.1038/nn.4043
- Khatiri, R., McKinney, A. M., Swenson, B., and Janardhan, V. (2012). Blood-brain barrier, reperfusion injury, and hemorrhagic transformation in acute ischemic stroke. *Neurology* 79, S52–S57. doi: 10.1212/WNL.0b013e3182697e70
- Kim, G. W., Sugawara, T., and Chan, P. H. (2000). Involvement of oxidative stress and caspase-3 in cortical infarction after photothrombotic ischemia in mice. *J. Cerebral. Blood Flow Metabol.* 20, 1690–1701. doi: 10.1097/00004647-200012000-00008
- Kim, J.-E., Liu, B. P., Park, J. H., and Strittmatter, S. M. (2004). Nogo-66 receptor prevents raphespinal and rubrospinal axon regeneration and limits functional recovery from spinal cord injury. *Neuron* 44, 439–451. doi: 10.1016/j.neuron.2004.10.015
- Klein, A. M., Mazutis, L., Akartuna, I., Tallapragada, N., Veres, A., Li, V., et al. (2015). Droplet barcoding for single-cell transcriptomics applied to embryonic stem cells. *Cell* 161, 1187–1201. doi: 10.1016/j.cell.2015.04.044
- Kleinschnitz, C., Braeuninger, S., Pham, M., Austinat, M., Nölte, I., Renné, T., et al. (2008). Blocking of platelets or intrinsic coagulation pathway-driven thrombosis does not prevent cerebral infarctions induced by photothrombosis. *Stroke* 39, 1262–1268. doi: 10.1161/STROKEAHA.107.496448
- Koch, M., Kingwell, E., Rieckmann, P., and Tremlett, H. (2009). The natural history of primary progressive multiple sclerosis. *Neurology* 73, 1996–2002. doi: 10.1212/WNL.0b013e318c5b47f
- Koehler, R. C., Roman, R. J., and Harder, D. R. (2008). Astrocytes and the regulation of cerebral blood flow. *Trends Neurosci.* 32, 160–169. doi: 10.1016/j.tins.2008.11.005
- Koh, W., Park, Y. M., Lee, S. E., and Lee, C. J. (2017). AAV-mediated astrocyte-specific gene expression under human ALDH1L1 promoter in mouse thalamus. *Exper. Neurobiol.* 26:350. doi: 10.5607/en.2017.26.6.350
- Kölliker, A. (1889). *Handbuch der Gewebelehre des Menschen*. Germany: Wilhelm Engelmann.
- Komitova, M., Serwanski, D. R., Lu, Q. R., and Nishiyama, A. (2011). NG2 cells are not a major source of reactive astrocytes after neocortical stab wound injury. *Glia* 59, 800–809. doi: 10.1002/glia.21152
- Kovacs, S. K., Leonessa, F., and Ling, G. S. (2014). Blast TBI models, neuropathology, and implications for seizure risk. *Front. Neurol.* 5:47. doi: 10.3389/fneur.2014.00047
- Kriegstein, A., and Alvarez-Buylla, A. (2009). The glial nature of embryonic and adult neural stem cells. *Ann. Rev. Neurosci.* 32, 149–184. doi: 10.1146/annurev-neuro.051508.135600
- Kunze, C., Börner, K., Kienle, E., Orschmann, T., Rusha, E., Schneider, M., et al. (2018). Synthetic AAV/CRISPR vectors for blocking HIV-1 expression in persistently infected astrocytes. *Glia* 66, 413–427. doi: 10.1002/glia.23254
- Kuznetsova, T., Prange, K. H. M., Glass, C. K., and de Winther, M. P. J. (2020). Transcriptional and epigenetic regulation of macrophages in atherosclerosis. *Nat. Rev. Cardiol.* 17, 216–228. doi: 10.1038/s41569-019-0265-3
- Lanjakornsiripan, D., Pior, B.-J., Kawaguchi, D., Furutachi, S., Tahara, T., Katsuyama, Y., et al. (2018). Layer-specific morphological and molecular differences in neocortical astrocytes and their dependence on neuronal layers. *Nat. Commun.* 9, 1–15. doi: 10.1038/s41467-018-03940-3
- Lassmann, H., and Bradl, M. (2017). Multiple sclerosis: experimental models and reality. *Acta Neuropathol.* 133, 223–244. doi: 10.1007/s00401-016-1631-4
- Lau, L. W., Cua, R., Keough, M. B., Haylock-Jacobs, S., and Yong, V. W. (2013). Pathophysiology of the brain extracellular matrix: a new target for remyelination. *Nat. Rev. Neurosci.* 14, 722–729. doi: 10.1038/nrn3550

- Lavin, Y., Winter, D., Blecher-Gonen, R., David, E., Keren-Shaul, H., Merad, M., et al. (2014). Tissue-resident macrophage enhancer landscapes are shaped by the local microenvironment. *Cell* 159, 1312–1326. doi: 10.1016/j.cell.2014.11.018
- Lee, E.-S. Y., Sidoryk, M., Jiang, H., Yin, Z., and Aschner, M. (2009). Estrogen and tamoxifen reverse manganese-induced glutamate transporter impairment in astrocytes. *J. Neurochem.* 110, 530–544. doi: 10.1111/j.1471-4159.2009.06105.x
- Lepore, A. C., Rauck, B., Dejea, C., Pardo, A. C., Rao, M. S., Rothstein, J. D., et al. (2008). Focal transplantation-based astrocyte replacement is neuroprotective in a model of motor neuron disease. *Nat. Neurosci.* 11, 1294–1301. doi: 10.1038/nn.2210
- Levison, S. W., and Goldman, J. E. (1993). Both oligodendrocytes and astrocytes develop from progenitors in the subventricular zone of postnatal rat forebrain. *Neuron* 10, 201–212. doi: 10.1016/0896-6273(93)90311-e
- Lewis, D. K., Thomas, K. T., Selvamani, A., and Sohrabji, F. (2012). Age-related severity of focal ischemia in female rats is associated with impaired astrocyte function. *Neurobiol. Aging* 33:1123.e1–16. doi: 10.1016/j.neurobiolaging.2011.11.007
- Li, H., Zhang, N., Lin, H.-Y., and Yu, Y. (2014). Histological, cellular and behavioral assessments of stroke outcomes after photothrombosis-induced ischemia in adult mice. *BMC Neurosci.* 15:58. doi: 10.1186/1471-2202-15-58
- Liddelow, S. A., and Barres, B. A. (2016). Regeneration: Not everything is scary about a glial scar. *Nature* 532, 182–183. doi: 10.1038/nature17318
- Liddelow, S. A., and Barres, B. A. (2017). Reactive astrocytes: production, function, and therapeutic potential. *Immunity* 46, 957–967. doi: 10.1016/j.immuni.2017.06.006
- Liddelow, S. A., Guttenplan, K. A., Clarke, L. E., Bennett, F. C., Bohlen, C. J., Schirmer, L., et al. (2017). Neurotoxic reactive astrocytes are induced by activated microglia. *Nature* 541, 481–487. doi: 10.1038/nature21029
- Lin, C.-C. J., Yu, K., Hatcher, A., Huang, T.-W., Lee, H. K., Carlson, J., et al. (2017). Identification of diverse astrocyte populations and their malignant analogs. *Nat. Neurosci.* 20, 396–406. doi: 10.1038/nn.4493
- Liu, M., Hurn, P. D., Roselli, C. E., and Alkayed, N. J. (2007). Role of P450 aromatase in sex-specific astrocytic cell death. *J. Cerebral. Blood Flow Metabol.* 27, 135–141. doi: 10.1038/sj.jcbfm.9600331
- Liu, M., Oyarzabal, E. A., Yang, R., Murphy, S. J., and Hurn, P. D. (2008). A novel method for assessing sex-specific and genotype-specific response to injury in astrocyte culture. *J. Neurosci. Methods* 171, 214–217. doi: 10.1016/j.jneumeth.2008.03.002
- Liu, S., Levine, S. R., and Winn, H. R. (2010). Targeting ischemic penumbra: part I - from pathophysiology to therapeutic strategy. *J. Exper. Stroke Transl. Med.* 3, 47–55. doi: 10.6030/1939-067x-3.1.47
- Liu, Y., Miao, Q., Yuan, J., Zhang, P., Li, S., Rao, Z., et al. (2015). Ascl1 converts dorsal midbrain astrocytes into functional neurons in vivo. *J. Neurosci.* 35, 9336–9355. doi: 10.1523/JNEUROSCI.3975-14.2015
- Liu, Z., and Chopp, M. (2016). Astrocytes, therapeutic targets for neuroprotection and neurorestoration in ischemic stroke. *Prog. Neurobiol.* 144, 103–120. doi: 10.1016/j.pneurobio.2015.09.008
- Lively, S., Moxon-Emre, I., and Schlichter, L. C. (2011). SC1/hevin and reactive gliosis after transient ischemic stroke in young and aged rats. *J. Neuropathol. Exper. Neurol.* 70, 913–929. doi: 10.1097/NEN.0b013e318231151e
- Longa, E. Z., Weinstein, P. R., Carlson, S., and Cummins, R. (1989). Reversible middle cerebral artery occlusion without craniectomy in rats. *Stroke* 20, 84–91. doi: 10.1161/01.str.20.1.84
- Lozzi, B., Huang, T.-W., Sardar, D., Huang, A. Y.-S., and Deneen, B. (2020). Regionally distinct astrocytes display unique transcription factor profiles in the adult brain. *Front. Neurosci.* 14:61. doi: 10.3389/fnins.2020.00061
- Ludwin, S. K., Rao, V. T., Moore, C. S., and Antel, J. P. (2016). Astrocytes in multiple sclerosis. *Multiple Sclerosis J.* 22, 1114–1124. doi: 10.1177/1352458516643396
- Lukaszevicz, A.-C., Sampaio, N., Guégan, C., and Benchoua, A. (2002). High sensitivity of protoplasmic cortical astroglia to focal ischemia. *J. Cerebral. Blood Flow Metabol.* 22:289. doi: 10.1097/00004647-200203000-00006
- Lukovic, D., Stojkovic, M., Moreno-Manzano, V., Jendelova, P., Sykova, E., Bhattacharya, S. S., et al. (2015). Concise review: reactive astrocytes and stem cells in spinal cord injury: good guys or bad guys? *Stem Cells* 33, 1036–1041. doi: 10.1002/stem.1959
- Lundgaard, I., Osório, M. J., Kress, B. T., Sanggaard, S., and Nedergaard, M. (2014). White matter astrocytes in health and disease. *Neuroscience* 276, 161–173. doi: 10.1016/j.neuroscience.2013.10.050
- Lynch, A. M., Murphy, K. J., Deighan, B. F., O'Reilly, J.-A., Gun'ko, Y. K., Cowley, T. R., et al. (2010). The impact of glial activation in the aging brain. *Aging Dis.* 1:262.
- Maas, A. I. R., Stocchetti, N., and Bullock, R. (2008). Moderate and severe traumatic brain injury in adults. *Lancet Neurol.* 7, 728–741. doi: 10.1016/S1474-4422(08)70164-9
- Macosko, E. Z., Basu, A., Satija, R., Nemes, J., Shekhar, K., Goldman, M., et al. (2015). Highly parallel genome-wide expression profiling of individual cells using nanoliter droplets. *Cell* 161, 1202–1214. doi: 10.1016/j.cell.2015.05.002
- Magavi, S., Friedmann, D., Banks, G., Stolfi, A., and Lois, C. (2012). Coincident generation of pyramidal neurons and protoplasmic astrocytes in neocortical columns. *J. Neurosci.* 32, 4762–4772. doi: 10.1523/JNEUROSCI.3560-11.2012
- Magnusson, J. P., Zamboni, M., Santopolo, G., Mold, J. E., Barrientos-Somarrivas, M., Talavera-Lopez, C., et al. (2020). Activation of a neural stem cell transcriptional program in parenchymal astrocytes. *elife* 9:e59733. doi: 10.7554/eLife.59733
- Manwani, B., Liu, F., Scranton, V., Hammond, M. D., Sansing, L. H., and McCullough, L. D. (2013). Differential effects of aging and sex on stroke induced inflammation across the lifespan. *Exper. Neurol.* 249, 120–131. doi: 10.1016/j.expneurol.2013.08.011
- Manwani, B., Liu, F., Xu, Y., Persky, R., Li, J., and McCullough, L. D. (2011). Functional recovery in aging mice after experimental stroke. *Brain Behav. Immun.* 25, 1689–1700. doi: 10.1016/j.bbi.2011.06.015
- Marchal, G., Beaudouin, V., Rioux, P., de la Sayette, V., Le Doze, F., Viader, F., et al. (1996). Prolonged persistence of substantial volumes of potentially viable brain tissue after stroke: a correlative PET-CT study with voxel-based data analysis. *Stroke* 27, 599–606. doi: 10.1161/01.str.27.4.599
- Marshall, C. A., and Goldman, J. E. (2002). Subpallial dlx2-expressing cells give rise to astrocytes and oligodendrocytes in the cerebral cortex and white matter. *J. Neurosci.* 22, 9821–9830. doi: 10.1523/JNEUROSCI.22-22-09821.2002single-cell
- Martin, J. C., Chang, C., Boschetti, G., Ungaro, R., Giri, M., Grout, J. A., et al. (2019). Single-cell analysis of Crohn's disease lesions identifies a pathogenic cellular module associated with resistance to anti-TNF therapy. *Cell* 178, 1493–1508. doi: 10.1016/j.cell.2019.08.008
- Martín-López, E., García-Marques, J., Núñez-Llaves, R., and López-Mascaraque, L. (2013). Clonal astrocytic response to cortical injury. *PLoS One* 8:e74039. doi: 10.1371/journal.pone.0074039
- Martorana, F., Brambilla, L., Valori, C. F., Bergamaschi, C., Roncoroni, C., Aronica, E., et al. (2012). The BH4 domain of Bcl-XL rescues astrocyte degeneration in amyotrophic lateral sclerosis by modulating intracellular calcium signals. *Hum. Mol. Genet.* 21, 826–840. doi: 10.1093/hmg/ddr513
- Masaki, K., Suzuki, S. O., Matsushita, T., and Matsuoka, T. (2013). Connexin 43 astrocytopathy linked to rapidly progressive multiple sclerosis and neuromyelitis optica. *PLoS One* 8:e72919. doi: 10.1371/journal.pone.0072919
- Masuda, T., Sankowski, R., Staszewski, O., Böttcher, C., Amann, L., Scheiwe, C., et al. (2019). Spatial and temporal heterogeneity of mouse and human microglia at single-cell resolution. *Nature* 566, 388–392. doi: 10.1038/s41586-019-0924-x
- Matei, N., Camara, J., and Zhang, J. H. (2021). The next step in the treatment of stroke. *Front. Neurol.* 11:582605. doi: 10.3389/fneur.2020.582605
- Mathys, H., Davila-Velderrain, J., Peng, Z., Gao, F., Mohammadi, S., Young, J. Z., et al. (2019). Single-cell transcriptomic analysis of Alzheimer's disease. *Nature* 570, 332–337. doi: 10.1038/s41586-019-1195-2
- Matias, I., Morgado, J., and Gomes, F. C. A. (2019). Astrocyte heterogeneity: impact to brain aging and disease. *Front. Aging Neurosci.* 11:59. doi: 10.3389/fnagi.2019.00059
- Matthias, K., Kirchoff, F., Seifert, G., Hüttmann, K., Matyash, M., Kettenmann, H., et al. (2003). Segregated expression of AMPA-type glutamate receptors and glutamate transporters defines distinct astrocyte populations in the mouse hippocampus. *J. Neurosci.* 23, 1750–1758. doi: 10.1523/JNEUROSCI.23-05-01750.2003
- Matute-Blanch, C., Calvo-Barreiro, L., Carballo-Carbajal, I., Gonzalo, R., Sanchez, A., Vila, M., et al. (2020). Chitinase 3-like 1 is neurotoxic in primary cultured neurons. *Sci. Rep.* 10, 1–6. doi: 10.1038/s41598-020-64093-2

- Matyash, V., and Kettenmann, H. (2009). Heterogeneity in astrocyte morphology and physiology. *Brain Res. Rev.* 63, 2–10. doi: 10.1016/j.brainresrev.2009.12.001
- Maud, A., Rodriguez, G. J., Vellipuram, A., Sheriff, F., Ghatali, M., Gupta, V., et al. (2021). Impact of early blood pressure lowering in patients presenting with acute ischemic stroke. *Curr. Cardiol. Rep.* 23:63. doi: 10.1007/s11886-021-01497-0
- McCarthy, D. P., Richards, M. H., and Miller, S. D. (2012). Mouse models of multiple sclerosis: experimental autoimmune encephalomyelitis and Theiler's virus-induced demyelinating disease. *Method Mol. Biol.* 900, 381–401. doi: 10.1007/978-1-4939-9072-6_21
- McCullough, L. D., Mirza, M. A., Xu, Y., Bentivegna, K., Steffens, E. B., Ritzel, R., et al. (2016). Stroke sensitivity in the aged: sex chromosome complement vs. gonadal hormones. *Aging* 8, 1432–1441. doi: 10.18632/aging.100997
- McKeon, R. J., Höke, A., and Silver, J. (1995). Injury-induced proteoglycans inhibit the potential for laminin-mediated axon growth on astrocytic scars. *Exper. Neurol.* 136, 32–43. doi: 10.1006/exnr.1995.1081
- McMurrin, C. E., Zhao, C., and Franklin, R. J. M. (2019). Toxin-based models to investigate demyelination and remyelination. *Methods Mol. Biol.* 1936, 377–396. doi: 10.1007/978-1-4939-9072-6_21
- Meng, X., Yang, F., Ouyang, T., Liu, B., Wu, C., and Jiang, W. (2015). Specific gene expression in mouse cortical astrocytes is mediated by a 1740bp-GFAP promoter-driven combined adeno-associated virus2/5/7/8/9. *Neurosci. Lett.* 593, 45–50. doi: 10.1016/j.neulet.2015.03.022
- Merkle, F. T., Tramontin, A. D., García-Verdugo, J. M., and Alvarez-Buylla, A. (2004). Radial glia give rise to adult neural stem cells in the subventricular zone. *Proc. Natl. Acad. Sci.* 101, 17528–17532. doi: 10.1073/pnas.0407893101
- Mestriner, R. G., Saur, L., Bagatini, P. B., and Baptista, P. P. A. (2015). Astrocyte morphology after ischemic and hemorrhagic experimental stroke has no influence on the different recovery patterns. *Behav. Brain Res.* 278, 257–261. doi: 10.1016/j.bbr.2014.10.005
- Metz, G. A., Curt, A., van de Meent, H., Klusman, I., Schwab, M. E., and Dietz, V. (2000). Validation of the weight-drop contusion model in rats: a comparative study of human spinal cord injury. *J. Neurotrauma* 17, 1–17. doi: 10.1089/neu.2000.17.1
- Milich, L. M., Choi, J., Ryan, C., Yahn, S. L., Tsoulfas, P., and Lee, J. K. (2021). Single cell analysis of the cellular heterogeneity and interactions in the injured mouse spinal cord. *J. Exper. Med.* 218:e20210040. doi: 10.1084/jem.20210040
- Miron, V. E., Boyd, A., Zhao, J.-W., Yuen, T. J., Ruckh, J. M., Shadrach, J. L., et al. (2013). M2 microglia and macrophages drive oligodendrocyte differentiation during CNS remyelination. *Nat. Neurosci.* 16, 1211–1218. doi: 10.1038/nn.3469
- Mix, E., Meyer-Rienecker, H., Hartung, H.-P., and Zettl, U. K. (2010). Animal models of multiple sclerosis—Potentials and limitations. *Prog. Neurobiol.* 92, 386–404. doi: 10.1016/j.pneurobio.2010.06.005
- Molina-Gonzalez, I., and Miron, V. E. (2019). Astrocytes in myelination and remyelination. *Neurosci. Lett.* 713:134532. doi: 10.1016/j.neulet.2019.134532
- Molofsky, A. V., and Deneen, B. (2015). Astrocyte development: A guide for the perplexed. *Glia* 63, 1320–1329. doi: 10.1002/glia.22836
- Molofsky, A. V., Kelley, K. W., Tsai, H.-H., Redmond, S. A., Chang, S. M., Madireddy, L., et al. (2014). Astrocyte-encoded positional cues maintain sensorimotor circuit integrity. *Nature* 509, 189–194. doi: 10.1038/nature13161
- Montalban, X., Tintore, M., Swanton, J., Barkhof, F., Fazekas, F., Filippi, M., et al. (2010). MRI criteria for MS in patients with clinically isolated syndromes. *Neurology* 74, 427–434. doi: 10.1212/WNL.0b013e3181cec45c
- Monteiro de Castro, G., Deja, N. A., Ma, D., Zhao, C., and Franklin, R. J. M. (2015). Astrocyte activation via Stat3 signaling determines the balance of oligodendrocyte versus schwann cell remyelination. *Am. J. Pathol.* 185, 2431–2440. doi: 10.1016/j.ajpath.2015.05.011
- Morel, L., Chiang, M. S. R., Higashimori, H., Shoneye, T., Iyer, L. K., Yelick, J., et al. (2017). Molecular and functional properties of regional astrocytes in the adult brain. *J. Neurosci.* 37, 8706–8717. doi: 10.1523/JNEUROSCI.3956-16.2017
- Morel, L., Higashimori, H., Tolman, M., and Yang, Y. (2014). VGLUT1+ neuronal glutamatergic signaling regulates postnatal developmental maturation of cortical protoplasmic astroglia. *J. Neurosci.* 34, 10950–10962. doi: 10.1523/JNEUROSCI.1167-14.2014
- Morel, L., Men, Y., Chiang, M. S. R., Tian, Y., Jin, S., Yelick, J., et al. (2019). Intracortical astrocyte subpopulations defined by astrocyte reporter Mice in the adult brain. *Glia* 67, 171–181. doi: 10.1002/glia.23545
- Morrison, H. W., and Filosa, J. A. (2016). Sex differences in astrocyte and microglia responses immediately following middle cerebral artery occlusion in adult mice. *Neuroscience* 339, 85–99. doi: 10.1016/j.neuroscience.2016.09.047
- Mosser, D. M., and Edwards, J. P. (2008). Exploring the full spectrum of macrophage activation. *Nat. Rev. Immunol.* 8, 958–969. doi: 10.1038/nri2448
- National Spinal Cord Injury Statistical Center. (2021). *Facts and Figures at a Glance*. Available Online at: <https://www.nscisc.uab.edu/> [Accessed June 16, 2021]
- Nichols, N. R., Day, J. R., Laping, N. J., and Johnson, S. A. (1993). GFAP mRNA increases with age in rat and human brain. *Neurobiol. Aging* 14, 421–429. doi: 10.1016/0197-4580(93)90100-P
- Nishiyama, A., Boshans, L., Goncalves, C. M., and Wegrzyn, J. (2015). Lineage, fate, and fate potential of NG2-glia. *Brain Res.* 1638, 116–128. doi: 10.1016/j.brainres.2015.08.013
- Noctor, S. C., Flint, A. C., Weissman, T. A., and Wong, W. S. (2002). Dividing precursor cells of the embryonic cortical ventricular zone have morphological and molecular characteristics of radial Glia. *J. Neurosci.* 22, 3161–3173. doi: 10.1523/JNEUROSCI.22-08-03161.2002
- Nolan, R. C., Galetta, S. L., Frohman, T. C., Frohman, E. M., Calabresi, P. A., Castrillo-Viguera, C., et al. (2018). Optimal intereye difference thresholds in retinal nerve fiber layer thickness for predicting a unilateral optic nerve lesion in multiple sclerosis. *J. Neuro-Ophthalmol.* 38, 451–458. doi: 10.1097/WNO.0000000000000629
- Oberheim, N. A., Goldman, S. A., and Nedergaard, M. (2012). Heterogeneity of astrocytic form and function. *Methods Mol. Biol.* 814, 23–45. doi: 10.1007/978-1-61779-452-0_3
- Oberheim, N. A., Takano, T., Han, X., He, W., Lin, J. H. C., Wang, F., et al. (2009). Uniquely hominid features of adult human astrocytes. *J. Neurosci.* 29, 3276–3287. doi: 10.1523/JNEUROSCI.4707-08.2009
- O'Callaghan, J. P., Brinton, R. E., and McEwen, B. S. (1991). Glucocorticoids regulate the synthesis of glial fibrillary acid protein in intact and adrenalectomized rats but do not affect its expression following brain injury. *J. Neurochem.* 57, 860–869. doi: 10.1111/j.1471-4159.1991.tb08230.x
- O'Callaghan, J. P., Kelly, K. A., VanGilder, R. L., Sofroniew, M. V., and Miller, D. B. (2014). Early activation of STAT3 regulates reactive astroglia induced by diverse forms of neurotoxicity. *PLoS One* 9:e102003. doi: 10.1371/journal.pone.0102003
- Okada, S., Hara, M., Kobayakawa, K., and Matsumoto, Y. (2018). Astrocyte reactivity and astroglia after spinal cord injury. *Neurosci. Res.* 126, 39–43. doi: 10.1016/j.neures.2017.10.004
- Okada, S., Nakamura, M., Katoh, H., and Miyao, T. (2006). Conditional ablation of Stat3 or Socs3 discloses a dual role for reactive astrocytes after spinal cord injury. *Nat. Med.* 12:829. doi: 10.1038/nm1425
- Olsen, M. L., Campbell, S. L., and Sontheimer, H. (2007). Differential distribution of Kir4. 1 in spinal cord astrocytes suggests regional differences in K+ homeostasis. *J. Neurophysiol.* 98, 786–793. doi: 10.1152/jn.00340.2007
- Orre, M., Kamphuis, W., Osborn, L. M., and Melief, J. (2014). Acute isolation and transcriptome characterization of cortical astrocytes and microglia from young and aged mice. *Neurobiol. Aging* 35, 1–14. doi: 10.1016/j.neurobiolaging.2013.07.008
- O'Shea, T. M., Burda, J. E., and Sofroniew, M. V. (2017). Cell biology of spinal cord injury and repair. *J. Clin. Invest.* 127, 3259–3270. doi: 10.1172/JCI90608
- Owens, T. (2006). Animal models for multiple sclerosis. *Adv. Neurol.* 98:77.
- Parikh, K., Antanaviciute, A., Fawcner-Corbett, D., Jagielowicz, M., Alicino, A., Lagerholm, C., et al. (2019). Colonic epithelial cell diversity in health and inflammatory bowel disease. *Nature* 567, 49–55. doi: 10.1038/s41586-019-0992-y
- Parnavelas, J. G. (1999). Glial cell lineages in the rat cerebral cortex. *Exper. Neurol.* 156, 418–429. doi: 10.1006/exnr.1999.7044
- Pawlak, J., Brito, V., Küppers, E., and Beyer, C. (2005). Regulation of glutamate transporter GLAST and GLT-1 expression in astrocytes by estrogen. *Mol. Brain Res.* 138, 1–7. doi: 10.1016/j.molbrainres.2004.10.043
- Pekny, M., and Nilsson, M. (2005). Astrocyte activation and reactive gliosis. *Glia* 50, 427–434. doi: 10.1002/glia.20207
- Pekny, M., and Pekna, M. (2014). Astrocyte reactivity and reactive astroglia: costs and benefits. *Physiol. Rev.* 94, 1077–1098. doi: 10.1152/physrev.00041.2013

- Peng, H., Ong, Y. M., Shah, W. A., Holland, P. C., and Carbonetto, S. (2013). Integrins regulate centrosome integrity and astrocyte polarization following a wound. *Dev. Neurobiol.* 73, 333–353. doi: 10.1002/dneu.22055
- Petrea, R. E., Beiser, A. S., Seshadri, S., Kelly-Hayes, M., Kase, C. S., and Wolf, P. A. (2009). Gender differences in stroke incidence and poststroke disability in the Framingham heart study. *Stroke* 40, 1032–1037. doi: 10.1161/01.str.29.1.159
- Plemel, J. R., Liu, W.-Q., and Yong, V. W. (2017). Remyelination therapies: a new direction and challenge in multiple sclerosis. *Nat. Rev. Drug Discov.* 16:617. doi: 10.1038/nrd.2017.115
- Polman, C. H., Reingold, S. C., Edan, G., and Filippi, M. (2005). Diagnostic criteria for multiple sclerosis: 2005 revisions to the “McDonald Criteria”. *Ann. Neurol.* 58, 840–846. doi: 10.1002/ana.20703
- Ponath, G., Park, C., and Pitt, D. (2018). The role of astrocytes in multiple sclerosis. *Front. Immunol.* 9:217. doi: 10.3389/fimmu.2018.00217
- Popa-Wagner, A., and Badan, I. (2007). Accelerated infarct development, cytogenesis and apoptosis following transient cerebral ischemia in aged rats. *Acta Neuropathol.* 113, 277–293. doi: 10.1007/s00401-006-0164-7
- Praet, J., Guglielmetti, C., Berneman, Z., and Van der Linden, A. (2014). Cellular and molecular neuropathology of the cuprizone mouse model: Clinical relevance for multiple sclerosis. *Neurosci. Biobehav. Rev.* 47:485. doi: 10.1016/j.neubiorev.2014.10.004
- Priego, N., Zhu, L., Monteiro, C., Mulders, M., Wasilewski, D., Bindeman, W., et al. (2018). STAT3 labels a subpopulation of reactive astrocytes required for brain metastasis. *Nat. Med.* 24, 1024–1035. doi: 10.1038/s41591-018-0044-4
- Prins, M., Schul, E., Geurts, J., van der Valk, P., Drukarch, B., and van Dam, A.-M. (2015). Pathological differences between white and grey matter multiple sclerosis lesions. *Ann. N.Y. Acad. Sci.* 1351, 99–113. doi: 10.1111/nyas.12841
- Procaccini, C., De Rosa, V., Pucino, V., Formisano, L., and Matarese, G. (2015). Animal models of multiple sclerosis. *Eur. J. Pharmacol.* 759, 182–191.
- Qian, D., Li, L., Rong, Y., and Liu, W. (2019). Blocking Notch signal pathway suppresses the activation of neurotoxic A1 astrocytes after spinal cord injury. *Cell Cycle* 18, 3010–3029. doi: 10.1080/15384101.2019.1667189
- Raff, M. C. (1989). Glial cell diversification in the rat optic nerve. *Science* 243, 1450–1455. doi: 10.1126/science.2648568
- Raff, M. C., Abney, E. R., and Miller, R. H. (1984). Two glial cell lineages diverge prenatally in rat optic nerve. *Dev. Biol.* 106, 53–60. doi: 10.1016/0012-1606(84)90060-5
- Rakers, C., and Petzold, G. C. (2017). Astrocytic calcium release mediates peri-infarct depolarizations in a rodent stroke model. *J. Clin. Invest.* 127, 511–516. doi: 10.1172/JCI89354
- Rakers, C., Schleif, M., Blank, N., and Matusšková, H. (2019). Stroke target identification guided by astrocyte transcriptome analysis. *Glia* 67, 619–633. doi: 10.1002/glia.23544
- Ramsköld, D., Luo, S., Wang, Y.-C., Li, R., Deng, Q., Faridani, O. R., et al. (2012). Full-length mRNA-Seq from single-cell levels of RNA and individual circulating tumor cells. *Nat. Biotechnol.* 30, 777–782. doi: 10.1038/nbt.2282
- Rao, V. T. S., Fuh, S.-C., Karamchandani, J. R., and Woulfe, J. M. J. (2019). Astrocytes in the pathogenesis of multiple sclerosis: An in situ MicroRNA study. *J. Neuropathol. Exper. Neurol.* 78, 1130–1146. doi: 10.1093/jnen/nlz098
- Rawji, K. S., Gonzalez Martinez, G. A., Sharma, A., and Franklin, R. J. M. (2020). The role of astrocytes in remyelination. *Trends Neurosci.* 43, 596–607. doi: 10.1016/j.tins.2020.05.006
- Rawji, K. S., Mishra, M. K., and Yong, V. W. (2016). Regenerative capacity of macrophages for remyelination. *Front. Cell. Dev. Biol.* 4:47. doi: 10.3389/fcell.2016.00047
- Reeves, M. J., Bushnell, C. D., Howard, G., Gargano, J. W., Duncan, P. W., Lynch, G., et al. (2008). Sex differences in stroke: epidemiology, clinical presentation, medical care, and outcomes. *Lancet Neurol.* 7, 915–926. doi: 10.1016/S1474-4422(08)70193-5
- Ren, Y., Ao, Y., O’Shea, T. M., Burda, J. E., Bernstein, A. M., Brumm, A. J., et al. (2017). Ependymal cell contribution to scar formation after spinal cord injury is minimal, local and dependent on direct ependymal injury. *Sci. Rep.* 7:41122. doi: 10.1038/srep41122
- Robillard, K. N., Lee, K. M., Chiu, K. B., and MacLean, A. G. (2016). Glial cell morphological and density changes through the lifespan of rhesus macaques. *Brain Behav. Immun.* 55, 60–69. doi: 10.1016/j.bbi.2016.01.006
- Robinson, M. J., and McCulloch, J. (1990). Contractile responses to endothelin in feline cortical vessels in situ. *J. Cerebral. Blood Flow Metabol.* 10, 285–289. doi: 10.1038/jcbfm.1990.46
- Robinson, R. G., Shoemaker, W. J., Schlumpf, M., Valk, T., and Bloom, F. E. (1975). Effect of experimental cerebral infarction in rat brain on catecholamines and behaviour. *Nature* 255, 332–334. doi: 10.1038/255332a0
- Rodríguez, J. J., Parpura, V., Zorec, R., and Verkhratsky, A. (2016). Astrocytes in physiological aging and Alzheimer’s disease. *Neuroscience* 323, 170–182. doi: 10.1016/j.neuroscience.2015.01.007
- Rodríguez, J. J., Terzieva, S., Olabarria, M., Lanza, R. G., and Verkhratsky, A. (2013). Enriched environment and physical activity reverse astroglial degeneration in the hippocampus of AD transgenic mice. *Cell Death Dis.* 4:678. doi: 10.1038/cddis.2013.194
- Rodríguez, J. J., Yeh, C.-Y., Terzieva, S., Olabarria, M., Kulijewicz-Nawrot, M., and Verkhratsky, A. (2014). Complex and region-specific changes in astroglial markers in the aging brain. *Neurobiol. Aging* 35, 15–23. doi: 10.1016/j.neurobiolaging.2013.07.002
- Roome, R. B., Bartlett, R. F., Jeffers, M., Xiong, J., Corbett, D., and Vanderluit, J. L. (2014). A reproducible Endothelin-1 model of forelimb motor cortex stroke in the mouse. *J. Neurosci. Methods* 233, 34–44. doi: 10.1016/j.jneumeth.2014.05.014
- Rothhammer, V., Borucki, D. M., Tjon, E. C., and Takenaka, M. C. (2018). Microglial control of astrocytes in response to microbial metabolites. *Nature* 557, 724–728. doi: 10.1038/s41586-018-0119-x
- Rothhammer, V., Mascalfroni, I. D., Bunse, L., Takenaka, M. C., Kenison, J. E., Mayo, L., et al. (2016). Type I interferons and microbial metabolites of tryptophan modulate astrocyte activity and central nervous system inflammation via the aryl hydrocarbon receptor. *Nat. Med.* 22, 586–597. doi: 10.1038/nm.4106
- Roybon, L., Lamas, N. J., Garcia-Diaz, A., Yang, E. J., Sattler, R., Jackson-Lewis, V., et al. (2013). Human stem cell-derived spinal cord astrocytes with defined mature or reactive phenotypes. *Cell Rep.* 4, 1035–1048. doi: 10.1016/j.celrep.2013.06.021
- Rozenblatt-Rosen, O., Stubbington, M. J., Regev, A., and Teichmann, S. A. (2017). The human cell Atlas: from vision to reality. *Nat. News* 550:451. doi: 10.1038/550451a
- Sabelstrom, H., Stenudd, M., Reu, P., Dias, D. O., Elfineh, M., Zdunek, S., et al. (2013). Resident neural stem cells restrict tissue damage and neuronal loss after spinal cord injury in mice. *Science* 342, 637–640. doi: 10.1126/science.1242576
- Samaranch, L., Salegio, E. A., San Sebastian, W., Kells, A. P., Foust, K. D., Bringas, J. R., et al. (2012). Adeno-associated virus serotype 9 transduction in the central nervous system of nonhuman primates. *Hum. Gene Ther.* 23, 382–389. doi: 10.1089/hum.2011.200
- Savettieri, G., Messina, D., Andreoli, V., Bonavita, S., Caltagirone, C., Cittadella, R., et al. (2004). Gender-related effect of clinical and genetic variables on the cognitive impairment in multiple sclerosis. *J. Neurol.* 251, 1208–1214. doi: 10.1007/s00415-004-0508-y
- Schaum, N., Karkanas, J., Neff, N. F., May, A. P., Quake, S. R., Wyss-Coray, T., et al. (2018). Single-cell transcriptomics of 20 mouse organs creates a Tabula Muris: The tabula muris consortium. *Nature* 562:367. doi: 10.1038/s41586-018-0590-4
- Schirmer, L., Velmeshev, D., Holmqvist, S., and Kaufmann, M. (2019). Neuronal vulnerability and multilineage diversity in multiple sclerosis. *Nature* 573, 75–82. doi: 10.1038/s41586-019-1404-z
- Schitine, C., Nogaroli, L., Costa, M. R., and Hedin-Pereira, C. (2015). Astrocyte heterogeneity in the brain: from development to disease. *Front. Cell. Neurosci.* 9:76. doi: 10.3389/fncel.2015.00076
- Schroeter, M., Schiene, K., Kraemer, M., Hagemann, G., Weigel, H., Eysel, U. T., et al. (1995). Astroglial responses in photochemically induced focal ischemia of the rat cortex. *Exper. Brain Res.* 106, 1–6. doi: 10.1007/BF00241351
- Schwab, M. E. (2004). Nogo and axon regeneration. *Curr. Opin. Neurobiol.* 14, 118–124. doi: 10.1016/j.conb.2004.01.004
- Sekar, S., McDonald, J., Cuyugan, L., and Aldrich, J. (2015). Alzheimer’s disease is associated with altered expression of genes involved in immune response and mitochondrial processes in astrocytes. *Neurobiol. Aging* 36, 583–591. doi: 10.1016/j.neurobiolaging.2014.09.027
- Selvamani, A., and Sohrabji, F. (2010). Reproductive age modulates the impact of focal ischemia on the forebrain as well as the effects of estrogen treatment

- in female rats. *Neurobiol. Aging* 31, 1618–1628. doi: 10.1016/j.neurobiolaging.2008.08.014
- Selvamani, A., Williams, M. H., Miranda, R. C., and Sohrabji, F. (2014). Circulating miRNA profiles provide a biomarker for severity of stroke outcomes associated with age and sex in a rat model. *Clin. Sci.* 127, 77–89.
- Serrano-Pozo, A., Gómez-Isla, T., Growdon, J. H., Frosch, M. P., and Hyman, B. T. (2013). A phenotypic change but not proliferation underlies glial responses in Alzheimer disease. *Am. J. Pathol.* 182, 2332–2344. doi: 10.1016/j.ajpath.2013.02.031
- Shagufu, and Ahmad, I. (2018). Tamoxifen a pioneering drug: An update on the therapeutic potential of tamoxifen derivatives. *Eur. J. Med. Chem.* 143, 515–531. doi: 10.1016/j.ejmech.2017.11.056
- Shalek, A. K., Satija, R., Adiconis, X., Gertner, R. S., Gaublot, J. T., Raychowdhury, R., et al. (2013). Single-cell transcriptomics reveals bimodality in expression and splicing in immune cells. *Nature* 498, 236–240. doi: 10.1038/nature12172
- Shalek, A. K., Satija, R., Shuga, J., Trombetta, J. J., Gennert, D., Lu, D., et al. (2014). Single-cell RNA-seq reveals dynamic paracrine control of cellular variation. *Nature* 510, 363–369. doi: 10.1038/nature13437
- Shandra, O., Winemiller, A. R., Heithoff, B. P., Munoz-Ballester, C., George, K. K., Benko, M. J., et al. (2019). Repetitive diffuse mild traumatic brain injury causes an atypical astrocyte response and spontaneous recurrent seizures. *J. Neurosci.* 39, 1944–1963. doi: 10.1523/JNEUROSCI.1067-18.2018
- Shannon, C., Salter, M., and Fern, R. (2007). GFP imaging of live astrocytes: regional differences in the effects of ischaemia upon astrocytes. *J. Anat.* 210, 684–692. doi: 10.1111/j.1469-7580.2007.00731.x
- Sharma, G., Sharma, A. R., Lee, S.-S., Bhattacharya, M., Nam, J.-S., and Chakraborty, C. (2019). Advances in nanocarriers enabled brain targeted drug delivery across blood brain barrier. *Int. J. Pharm.* 559, 360–372. doi: 10.1016/j.ijpharm.2019.01.056
- Sharp, D. J., Scott, G., and Leech, R. (2014). Network dysfunction after traumatic brain injury. *Nat. Rev. Neurol.* 10:156. doi: 10.1038/nrneuro.2014.15
- Shields, S. A., Gilson, J. M., Blakemore, W., and Franklin, R. J. (2000). Remyelination occurs as extensively but more slowly in old rats compared to young rats following flitoxin-induced CNS demyelination. *Glia* 29:102. doi: 10.1002/(sici)1098-1136(200001)29:1<102::aid-glia12<3.0.co;2-1
- Shirani, A., Zhao, Y., Kingwell, E., Rieckmann, P., and Tremlett, H. (2012). Temporal trends of disability progression in multiple sclerosis: findings from British Columbia, Canada (1975–2009). *Multiple Sclerosis J.* 18, 442–450. doi: 10.1177/1352458511422097
- Silver, J. (2016). The glial scar is more than just astrocytes. *Exper. Neurol.* 286, 147–149. doi: 10.1016/j.expneurol.2016.06.018
- Silver, J., and Miller, J. H. (2004). Regeneration beyond the glial scar. *Nat. Rev. Neurosci.* 5, 146–156. doi: 10.1038/nrn1326
- Siriphorn, A., Dunham, K. A., Chompoonong, S., and Floyd, C. L. (2012). Postinjury administration of 17 β -estradiol induces protection in the gray and white matter with associated functional recovery after cervical spinal cord injury in male rats. *J. Compar. Neurol.* 520, 2630–2646. doi: 10.1002/cne.23056
- Sirko, S., Behrendt, G., Johansson, P. A., and Tripathi, P. (2013). Reactive Glia in the injured brain acquire stem cell properties in response to sonic hedgehog. *Cell Stem Cell* 12, 426–439. doi: 10.1016/j.stem.2013.01.019
- Sisková, Z., Baron, W., de Vries, H., and Hoekstra, D. (2006). Fibronectin impedes “myelin” sheet-directed flow in oligodendrocytes: a role for a beta 1 integrin-mediated PKC signaling pathway in vesicular trafficking. *Mol. Cell. Neurosci.* 33, 150–159. doi: 10.1016/j.mcn.2006.07.001
- Skene, J. H., and Willard, M. (1981). Axonally transported proteins associated with axon growth in rabbit central and peripheral nervous systems. *J. Cell Biol.* 89, 96–103. doi: 10.1083/jcb.89.1.96
- Skinnder, M. A., Rogalski, J., Tigchelaar, S., Manouchehri, N., Prudova, A., Jackson, A. M., et al. (2021a). Proteomic portraits reveal evolutionarily conserved and divergent responses to spinal cord injury. *bioRxiv* 2021:428528. doi: 10.1101/2021.01.27.428528
- Skinnder, M. A., Squair, J. W., Kathe, C., Anderson, M. A., Gautier, M., Matson, K. J. E., et al. (2021b). Cell type prioritization in single-cell data. *Nat. Biotechnol.* 39, 30–34. doi: 10.1038/s41587-020-0605-1
- Skipuletz, T., Hackstette, D., Bauer, K., Gudi, V., Pul, R., Voss, E., et al. (2013). Astrocytes regulate myelin clearance through recruitment of microglia during cuprizone-induced demyelination. *Brain* 136, 147–167. doi: 10.1093/brain/aww262
- Smillie, C. S., Biton, M., Ordovas-Montanes, J., Sullivan, K. M., Burgin, G., Graham, D. B., et al. (2019). Intra- and inter-cellular rewiring of the human colon during ulcerative colitis. *Cell* 178, 714–730. doi: 10.1016/j.cell.2019.06.029
- Smith, H. L., Freeman, O. J., Butcher, A. J., and Holmqvist, S. (2020). Astrocyte unfolded protein response induces a specific reactivity state that causes non-cell-autonomous neuronal degeneration. *Neuron* 105, 855–866. doi: 10.1016/j.neuron.2019.12.014
- Soderblom, C., Luo, X., Blumenthal, E., Bray, E., Lyapichev, K., Ramos, J., et al. (2013). Perivascular fibroblasts form the fibrotic scar after contusive spinal cord injury. *J. Neurosci.* 33, 13882–13887. doi: 10.1523/JNEUROSCI.2524-13.2013
- Sofroniew, M. V. (2009). Molecular dissection of reactive astroglia and glial scar formation. *Trends Neurosci.* 32, 638–647. doi: 10.1016/j.tins.2009.08.002
- Sofroniew, M. V. (2015). Astroglia. *Cold Spring Harbor Perspect. Biol.* 7:a020420. doi: 10.1101/cshperspect.a020420
- Sofroniew, M. V. (2020). Astrocyte reactivity: subtypes, states, and functions in CNS innate immunity. *Trends Immunol.* 41, 758–770. doi: 10.1016/j.it.2020.07.004
- Sofroniew, M. V., and Vinters, H. V. (2010a). Astrocytes: biology and pathology. *Acta Neuropathol.* 119, 7–35. doi: 10.1007/s00401-009-0619-8
- Sohrabji, F., Bake, S., and Lewis, D. K. (2013). Age-related changes in brain support cells: Implications for stroke severity. *Neurochem. Int.* 63, 291–301. doi: 10.1016/j.neuint.2013.06.013
- Sommer, C. J. (2017). Ischemic stroke: experimental models and reality. *Acta Neuropathol.* 133, 245–261. doi: 10.1007/s00401-017-1667-0
- Soreq, L., Rose, J., Soreq, E., and Hardy, J. (2017). Major shifts in glial regional identity are a transcriptional hallmark of human brain aging. *Cell Rep.* 8, 557–570. doi: 10.1016/j.celrep.2016.12.011
- Soung, A., and Klein, R. S. (2018). Viral encephalitis and neurologic diseases: focus on astrocytes. *Trends Mol. Med.* 24, 950–962. doi: 10.1016/j.molmed.2018.09.001
- Sozmen, E. G., Kolekar, A., Havton, L. A., and Carmichael, S. T. (2009). A white matter stroke model in the mouse: axonal damage, progenitor responses and MRI correlates. *J. Neurosci. Methods* 180, 261–272. doi: 10.1016/j.jneumeth.2009.03.017
- Sparling, J. S., Bretzner, F., Biernaskie, J., Assinck, P., Jiang, Y., Arisato, H., et al. (2015). Schwann cells generated from neonatal skin-derived precursors or neonatal peripheral nerve improve functional recovery after acute transplantation into the partially injured cervical spinal cord of the rat. *J. Neurosci.* 35, 6714–6730. doi: 10.1523/JNEUROSCI.1070-14.2015
- Spence, R. D., Hamby, M. E., Umeda, E., Itoh, N., Du, S., Wisdom, A. J., et al. (2011). Neuroprotection mediated through estrogen receptor- in astrocytes. *Proc. Natl. Acad. Sci.* 108, 8867–8872. doi: 10.1073/pnas.1103833108
- Squair, J. W., Gautier, M., Kathe, C., Anderson, M. A., James, N. D., Hutson, T. H., et al. (2021). Confronting false discoveries in single-cell differential expression. *bioRxiv* 2021:435024. doi: 10.1101/2021.03.12.435024
- Stählberg, A., Andersson, D., Aurelius, J., Faiz, M., Pekna, M., Kubista, M., et al. (2011). Defining cell populations with single-cell gene expression profiling: correlations and identification of astrocyte subpopulations. *Nucleic Acids Res.* 39:e24. doi: 10.1093/nar/gkq1182
- Steinhäuser, C., Grunnet, M., and Carmignoto, G. (2015). Crucial role of astrocytes in temporal lobe epilepsy. *Neuroscience* 323, 157–169. doi: 10.1016/j.neuroscience.2014.12.047
- Stevens, B., Allen, N. J., Vazquez, L. E., and Howell, G. R. (2007). The classical complement cascade mediates CNS synapse elimination. *Cell* 131, 1164–1178. doi: 10.1016/j.cell.2007.10.036
- Stewart, A. N., MacLean, S. M., Stromberg, A. J., Whelan, J. P., Bailey, W. M., Gensel, J. C., et al. (2020). Considerations for studying sex as a biological variable in spinal cord injury. *Front. Neurol.* 11:802. doi: 10.3389/fneur.2020.00802
- Stimmer, L., Fovet, C.-M., and Serguera, C. (2018). Experimental models of autoimmune demyelinating diseases in nonhuman primates. *Veter. Pathol.* 55, 27–41. doi: 10.1177/0300985817712794
- Stoffels, J. M. J., de Jonge, J. C., Stancic, M., Nomden, A., van Strien, M. E., Ma, D., et al. (2013). Fibronectin aggregation in multiple sclerosis lesions impairs remyelination. *Brain* 136, 116–131. doi: 10.1093/brain/aww313

- Strijbis, E. M. M., Kooi, E.-J., van der Valk, P., and Geurts, J. J. G. (2017). Cortical remyelination is heterogeneous in multiple sclerosis. *J. Neuropathol. Exper. Neurol.* 76, 390–401. doi: 10.1093/jnen/nlx023
- Stubington, M. J., Rozenblatt-Rosen, O., Regev, A., and Teichmann, S. A. (2017). Single-cell transcriptomics to explore the immune system in health and disease. *Science* 358, 58–63. doi: 10.1126/science.aan6828
- Sun, S., Sun, Y., Ling, S.-C., Ferraiuolo, L., McAlonis-Downes, M., Zou, Y., et al. (2015). Translational profiling identifies a cascade of damage initiated in motor neurons and spreading to glia in mutant SOD1-mediated ALS. *Proc. Natl. Acad. Sci. U S A.* 112, E6993–E7002. doi: 10.1073/pnas.1520639112
- Sussman, E. S., and Connolly, E. S. M. D. Jr. (2013). Hemorrhagic transformation: A review of the rate of hemorrhage in the major clinical trials of acute ischemic stroke. *Front. Neurol.* 4:69. doi: 10.3389/fneur.2013.00069
- Sutherland, T. C., and Geoffroy, C. G. (2020). The influence of neuron-extrinsic factors and aging on injury progression and axonal repair in the central nervous system. *Front. Cell. Dev. Biol.* 8:190. doi: 10.3389/fcell.2020.00190
- Svensson, V., Vento-Tormo, R., and Teichmann, S. A. (2018). Exponential scaling of single-cell RNA-seq in the past decade. *Nat. Protocols* 13, 599–604. doi: 10.1038/nprot.2017.149
- Tabatadze, N., Huang, G., May, R. M., Jain, A., and Woolley, C. S. (2015). Sex differences in molecular signaling at inhibitory synapses in the hippocampus. *J. Neurosci.* 35, 11252–11265. doi: 10.1523/JNEUROSCI.1067-15.2015
- Takata, N., and Hirase, H. (2008). Cortical layer 1 and layer 2/3 astrocytes exhibit distinct calcium dynamics in vivo. *PLoS One* 3:e2525. doi: 10.1371/journal.pone.0002525
- Tanaka, H., Nakatani, T., Furihata, T., Tange, K., Nakai, Y., Yoshioka, H., et al. (2018). In Vivo introduction of mRNA encapsulated in lipid nanoparticles to brain neuronal cells and astrocytes via intracerebroventricular administration. *Mol. Pharmacol.* 15, 2060–2067. doi: 10.1021/acs.molpharmaceut.7b01084
- Tang, F., Barbacioru, C., Wang, Y., Nordman, E., Lee, C., Xu, N., et al. (2009). mRNA-Seq whole-transcriptome analysis of a single cell. *Nat. Methods* 6, 377–382. doi: 10.1038/nmeth.1315
- Taschenberger, G., Tereshchenko, J., and Kügler, S. (2017). A microRNA124 target sequence restores astrocyte specificity of gfaABC1D-driven transgene expression in AAV-mediated gene transfer. *Mol. Ther. Nucleic Acids* 8, 13–25. doi: 10.1016/j.omtn.2017.03.009
- Tassoni, A., Farkhondeh, V., Itoh, Y., and Itoh, N. (2019). The astrocyte transcriptome in EAE optic neuritis shows complement activation and reveals a sex difference in astrocytic C3 expression. *Sci. Rep.* 9, 1–12. doi: 10.1038/s41598-019-46232-6
- Terashima, T., Ogawa, N., Nakae, Y., Sato, T., Katagi, M., Okano, J., et al. (2018). Gene therapy for neuropathic pain through siRNA-IRF5 gene delivery with homing peptides to microglia. *Mol. Ther. Nucleic Acids* 11, 203–215. doi: 10.1016/j.omtn.2018.02.007
- Tetzlaff, W., Alexander, S. W., Miller, F. D., and Bisby, M. A. (1991). Response of facial and rubrospinal neurons to axotomy: changes in mRNA expression for cytoskeletal proteins and GAP-43. *J. Neurosci.* 11, 2528–2544.
- Tomassini, V., and Pozzilli, C. (2009). Sex hormones, brain damage and clinical course of Multiple Sclerosis. *J. Neurol. Sci.* 286, 35–39. doi: 10.1016/j.jns.2009.04.014
- Tomassy, G. S., Berger, D. R., Chen, H.-H., Kasthuri, N., Hayworth, K. J., Vercelli, A., et al. (2014). Distinct profiles of myelin distribution along single axons of pyramidal neurons in the neocortex. *Science* 344, 319–324. doi: 10.1126/science.1249766
- Tran, A. P., Warren, P. M., and Silver, J. (2018). The biology of regeneration failure and success after spinal cord injury. *Physiol. Rev.* 98, 881–917. doi: 10.1152/physrev.00017.2017
- Tran, A. P., Warren, P. M., and Silver, J. (2021). New insights into glial scar formation after spinal cord injury. *Cell Tissue Res.* doi: 10.1007/s00441-021-03477-w
- Tremlett, H., Zhao, Y., Rieckmann, P., and Hutchinson, M. (2010). New perspectives in the natural history of multiple sclerosis. *Neurology* 74, 2004–2015. doi: 10.1212/WNL.0b013e3181e3973f
- Treutlein, B., Brownfield, D. G., Wu, A. R., Neff, N. F., Mantalas, G. L., Espinoza, F. H., et al. (2014). Reconstructing lineage hierarchies of the distal lung epithelium using single-cell RNA-seq. *Nature* 509, 371–375. doi: 10.1038/nature13173
- Tsai, H.-H., Li, H., Fuentealba, L. C., Molofsky, A. V., Taveira-Marques, R., Zhuang, H., et al. (2012). Regional astrocyte allocation regulates CNS synaptogenesis and repair. *Science* 337, 358–362. doi: 10.1126/science.1222381
- Tutuncu, M., Tang, J., Zeid, N. A., Kale, N., Crusan, D. J., Atkinson, E. J., et al. (2013). Onset of progressive phase is an age-dependent clinical milestone in multiple sclerosis. *Multiple Sclerosis J.* 19, 188–198. doi: 10.1177/1352458512451510
- Vagner, T., Dvorzhak, A., Wójtowicz, A. M., Harms, C., and Grantyn, R. (2016). Systemic application of AAV vectors targeting GFAP-expressing astrocytes in Z-Q175-KI Huntington's disease mice. *Mol. Cell. Neurosci.* 77, 76–86. doi: 10.1016/j.mcn.2016.10.007
- Valiante, S., Falanga, A., Cigliano, L., Iachetta, G., Busiello, R. A., La Marca, V., et al. (2015). Peptide gh625 enters into neuron and astrocyte cell lines and crosses the blood-brain barrier in rats. *Int. J. Nanomed.* 10:1885. doi: 10.2147/IJN.S77734
- Valori, C. F., Guidotti, G., Brambilla, L., and Rossi, D. (2019). Astrocytes: emerging therapeutic targets in neurological disorders. *Trends Mol. Med.* 25, 750–759. doi: 10.1016/j.molmed.2019.04.010
- van der Star, B. J., Vogel, D. Y. S., Kipp, M., and Puentes, F. (2012). In vitro and in vivo models of multiple sclerosis. *CNS Neurol. Disord. Drug Targets* 11:570. doi: 10.2174/187152712801661284
- Van Gelder, R. N., von Zastrow, M. E., Yool, A., Dement, W. C., Barchas, J. D., and Eberwine, J. H. (1990). Amplified RNA synthesized from limited quantities of heterogeneous cDNA. *Proc. Natl. Acad. Sci.* 87, 1663–1667. doi: 10.1073/pnas.87.5.1663
- van Horssen, J., Dijkstra, C. D., and de Vries, H. E. (2007). The extracellular matrix in multiple sclerosis pathology. *J. Neurochem.* 103, 1293–1301. doi: 10.1111/j.1471-4159.2007.04897.x
- van Wijngaarden, P., and Franklin, R. J. (2013). Ageing stem and progenitor cells: implications for rejuvenation of the central nervous system. *Development* 140, 2562–2575. doi: 10.1242/dev.092262
- Vargas, M. R., Johnson, D. A., Sirkis, D. W., and Messing, A. (2008). Nrf2 activation in astrocytes protects against neurodegeneration in mouse models of familial amyotrophic lateral sclerosis. *J. Neurosci.* 28:13574. doi: 10.1523/JNEUROSCI.4099-08.2008
- Vierk, R., Glassmeier, G., Zhou, L., Brandt, N., Fester, L., Dudzinski, D., et al. (2012). Aromatase inhibition abolishes LTP generation in female but not in male mice. *J. Neurosci.* 32, 8116–8126. doi: 10.1523/JNEUROSCI.5319-11.2012
- Villapol, S., Loane, D. J., and Burns, M. P. (2017). Sexual dimorphism in the inflammatory response to traumatic brain injury. *Glia* 65, 1423–1438. doi: 10.1002/glia.23171
- Volterra, A., and Meldolesi, J. (2005). Astrocytes, from brain glue to communication elements: the revolution continues. *Nat. Rev. Neurosci.* 6, 626–640. doi: 10.1038/nrn1722
- von Jonquieres, G., Mersmann, N., Klugmann, C. B., Harasta, A. E., Lutz, B., Teahan, O., et al. (2013). Glial promoter selectivity following AAV-delivery to the immature brain. *PLoS One* 8:e065646. doi: 10.1371/journal.pone.0065646
- Voskuhl, R. R., and Gold, S. M. (2012). Sex-related factors in multiple sclerosis susceptibility and progression. *Nat. Rev. Neurol.* 8:255. doi: 10.1038/nrneuro.2012.43
- Voskuhl, R. R., Sawalha, A. H., and Itoh, Y. (2018). Sex chromosome contributions to sex differences in multiple sclerosis susceptibility and progression. *Multiple Sclerosis* 24, 22–31. doi: 10.1177/1352458517737394
- Wagner, D.-C., Scheibe, J., Glocke, I., and Weise, G. (2013). Object-based analysis of astroglial reaction and astrocyte subtype morphology after ischemic brain injury. *Acta Neurobiol. Exper.* 73:79.
- Waller, R., Woodroffe, M. N., Wharton, S. B., and Ince, P. G. (2016). Gene expression profiling of the astrocyte transcriptome in multiple sclerosis normal appearing white matter reveals a neuroprotective role. *J. Neuroimmunol.* 299, 139–146. doi: 10.1016/j.jneuroim.2016.09.010
- Wanner, I. B., Anderson, M. A., Song, B., Levine, J., Fernandez, A., Gray-Thompson, Z., et al. (2013). Glial scar borders are formed by newly proliferated, elongated astrocytes that interact to corral inflammatory and fibrotic cells via STAT3-dependent mechanisms after spinal cord injury. *J. Neurosci.* 33, 12870–12886. doi: 10.1523/JNEUROSCI.12121-13.2013
- Warach, S., and Latour, L. L. (2004). Evidence of reperfusion injury, exacerbated by thrombolytic therapy, in human focal brain ischemia using a novel imaging marker of early blood-brain barrier disruption. *Stroke* 35, 2659–2661. doi: 10.1161/01.STR.0000144051.32131.09

- Watson, B. D., Dietrich, W. D., Busto, R., Wachtel, M. S., and Ginsberg, M. D. (1985). Induction of reproducible brain infarction by photochemically initiated thrombosis. *Ann. Neurol.* 17, 497–504. doi: 10.1002/ana.410170513
- Weinreb, C., Rodriguez-Fraticelli, A., Camargo, F. D., and Klein, A. M. (2020). Lineage tracing on transcriptional landscapes links state to fate during differentiation. *Science* 367:eaaw3381. doi: 10.1126/science.aaw3381
- Wheeler, M. A., Clark, I. C., Tjon, E. C., Li, Z., Zandee, S. E. J., Couturier, C. P., et al. (2020). MAFG-driven astrocytes promote CNS inflammation. *Nature* 578, 593–599. doi: 10.1038/s41586-020-1999-0
- Wheeler, M. A., Jaronen, M., Covacu, R., Zandee, S. E. J., Scalisi, G., Rothhammer, V., et al. (2019). Environmental control of astrocyte pathogenic activities in CNS inflammation. *Cell* 176, 581–596. doi: 10.1016/j.cell.2018.12.012
- Whitacre, C. C., Reingold, S. C., O'Looney, P. A., Blankenhorn, E., Brinley, F., Collier, E., et al. (1999). A gender gap in autoimmunity: Task force on gender, multiple sclerosis and autoimmunity. *Science* 283, 1277–1278. doi: 10.1126/science.283.5406.1277
- White, R. E., McTigue, D. M., and Jakeman, L. B. (2009). Regional heterogeneity in astrocyte responses following contusive spinal cord injury in mice. *J. Compar. Neurol.* 518, 1370–1390. doi: 10.1002/cne.22282
- Williams, A., Piaton, G., and Lubetzki, C. (2007). Astrocytes—Friends or foes in multiple sclerosis? *Glia* 55, 1300–1312. doi: 10.1002/glia.20546
- Williams, J. L., Manivasagam, S., Smith, B. C., Sim, J., Vollmer, L. L., Daniels, B. P., et al. (2020). Astrocyte-T cell crosstalk regulates region-specific neuroinflammation. *Glia* 68, 1361–1374. doi: 10.1002/glia.23783
- Winship, I. R., and Murphy, T. H. (2008). In Vivo calcium imaging reveals functional rewiring of single somatosensory neurons after stroke. *J. Neurosci.* 28, 6592–6606. doi: 10.1523/JNEUROSCI.0622-08.2008
- Woodruff, R. H., and Franklin, R. J. M. (1999). Demyelination and remyelination of the caudal cerebellar peduncle of adult rats following stereotaxic injections of lysolecithin, ethidium bromide, and complement/anti-galactocerebroside: A comparative study. *Glia* 25, 216–228. doi: 10.1002/(SICI)1098-1136(19990201)25:3<216::AID-GLIA2<3.0.CO;2-L
- Woodruff, R. H., Fruttiger, M., Richardson, W. D., and Franklin, R. J. M. (2004). Platelet-derived growth factor regulates oligodendrocyte progenitor numbers in adult CNS and their response following CNS demyelination. *Mol. Cell. Neurosci.* 25, 252–262. doi: 10.1016/j.mcn.2003.10.014
- Wu, T., Dejanovic, B., Gandham, V. D., and Gogineni, A. (2019). Complement C3 is activated in human AD brain and is required for neurodegeneration in mouse models of amyloidosis and tauopathy. *Cell Rep.* 28, 2111–2123. doi: 10.1016/j.celrep.2019.07.060
- Xiong, Y., Mahmood, A., and Chopp, M. (2013). Animal models of traumatic brain injury. *Nat. Rev. Neurosci.* 14, 128–142. doi: 10.1038/nrn3407
- Xu, Z., Guo, D., Jiang, Z., Tong, R., Jiang, P., Bai, L., et al. (2019). Novel HER2-targeting antibody-drug conjugates of trastuzumab beyond T-DM1 in Breast Cancer: Trastuzumab Deruxtecan(DS-8201a) and (Vic-)Trastuzumab Duocarmazine (SYD985). *Eur. J. Med. Chem.* 183:111682. doi: 10.1016/j.ejmech.2019.111682
- Xue, J., Schmidt, S. V., Sander, J., Draffehn, A., Krebs, W., Quester, I., et al. (2014). Transcriptome-based network analysis reveals a spectrum model of human macrophage activation. *Immunity* 40, 274–288. doi: 10.1016/j.immuni.2014.01.006
- Yang, T., Dai, Y., Chen, G., and Cui, S. (2020). Dissecting the dual role of the glial scar and scar-forming astrocytes in spinal cord injury. *Front. Cell. Neurosci.* 14:78. doi: 10.3389/fncel.2020.00078
- Yang, Y., and Rosenberg, G. A. (2015). Matrix metalloproteinases as therapeutic targets for stroke. *Brain Res.* 1623, 30–38. doi: 10.1016/j.brainres.2015.04.024
- Yi, J.-H., Katagiri, Y., Susarla, B., Figge, D., Symes, A. J., and Geller, H. M. (2012). Alterations in sulfated chondroitin glycosaminoglycans following controlled cortical impact injury in mice. *J. Compar. Neurol.* 520, 3295–3313. doi: 10.1002/cne.23156
- Yoon, H., Walters, G., Paulsen, A. R., and Scarisbrick, I. A. (2017). Astrocyte heterogeneity across the brain and spinal cord occurs developmentally, in adulthood and in response to demyelination. *PLoS One* 12:e0180697. doi: 10.1371/journal.pone.0180697
- Yu, X., Taylor, A. M., Nagai, J., Golshani, P., Evans, C. J., Coppola, G., et al. (2018). Reducing astrocyte calcium signaling in vivo alters striatal microcircuits and causes repetitive behavior. *Neuron* 99, 1170–1187. doi: 10.1016/j.neuron.2018.08.015
- Zamanian, J. L., Xu, L., Foo, L. C., Nouri, N., Zhou, L., Giffard, R. G., et al. (2012). Genomic analysis of reactive astrogliosis. *J. Neurosci.* 32, 6391–6410. doi: 10.1523/JNEUROSCI.6221-11.2012
- Zamboni, M., Llorens-Bobadilla, E., Magnusson, J. P., and Frisén, J. (2020). A widespread neurogenic potential of neocortical astrocytes is induced by injury. *Cell Stem Cell* 27, 605–617. doi: 10.1016/j.stem.2020.07.006
- Zeisel, A., Hochgerner, H., Lönnerberg, P., Johnsson, A., Memic, F., van der Zwan, J., et al. (2018). Molecular architecture of the mouse nervous system. *Cell* 174, 999–1014. doi: 10.1016/j.cell.2018.06.021
- Zhang, L., Yin, J.-C., Yeh, H., Ma, N.-X., Lee, G., Chen, X. A., et al. (2015). Small molecules efficiently reprogram human astroglial cells into functional neurons. *Cell Stem Cell* 17, 735–747. doi: 10.1016/j.stem.2015.09.012
- Zhang, Q.-G., Wang, R., Tang, H., Dong, Y., Chan, A., Sareddy, G. R., et al. (2014). Brain-derived estrogen exerts anti-inflammatory and neuroprotective actions in the rat hippocampus. *Mol. Cell. Endocrinol.* 389, 84–91. doi: 10.1016/j.mce.2013.12.019
- Zhang, Y., and Barres, B. A. (2010). Astrocyte heterogeneity: an underappreciated topic in neurobiology. *Curr. Opin. Neurobiol.* 20, 588–594. doi: 10.1016/j.conb.2010.06.005
- Zhang, Y., Sloan, S. A., Clarke, L. E., and Caneda, C. (2016). Purification and characterization of progenitor and mature human astrocytes reveals transcriptional and functional differences with mouse. *Neuron* 89, 37–53. doi: 10.1016/j.neuron.2015.11.013
- Zhao, Y., and Rempe, D. A. (2010). Targeting astrocytes for stroke therapy. *Neurotherapeutics* 7, 439–451. doi: 10.1016/j.nurt.2010.07.004
- Zheng, W., Watts, L. T., Holstein, D. M., Wewer, J., and Lechleiter, J. D. (2013). P2Y₁ R-Initiated, IP₃ R-dependent stimulation of astrocyte mitochondrial metabolism reduces and partially reverses ischemic neuronal damage in mouse. *J. Cerebral Blood Flow Metabol.* 33, 600–611. doi: 10.1038/jcbfm.2012.214
- Zhou, Y., Peng, Z., Seven, E. S., and Leblanc, R. M. (2018). Crossing the blood-brain barrier with nanoparticles. *J. Control. Release* 270, 290–303. doi: 10.1016/j.jconrel.2017.12.015
- Zhu, X., Bergles, D. E., and Nishiyama, A. (2008). NG2 cells generate both oligodendrocytes and gray matter astrocytes. *Development* 135:145. doi: 10.1242/dev.004895
- Zhu, X., Hill, R. A., Dietrich, D., Komitova, M., Suzuki, R., and Nishiyama, A. (2011). Age-dependent fate and lineage restriction of single NG2 cells. *Development* 138, 745–753. doi: 10.1242/dev.047951

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Moulson, Squair, Franklin, Tetzlaff and Assinck. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.