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1. General information

A. Chemicals and Reagents

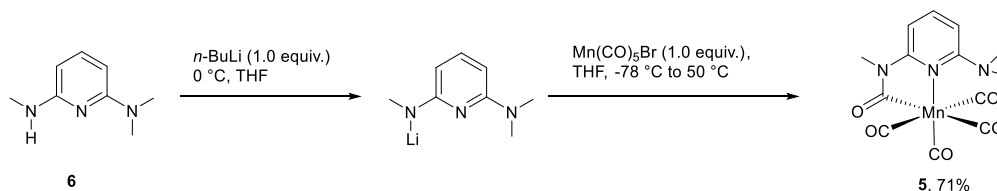
All manipulations were carried out under an inert N₂(g) atmosphere using glovebox techniques. Solvents were purified using a two-column solid-state purification system (Innovative Technology, NJ, USA) and transferred to the glovebox without exposure to air. Deuterated solvents were purchased from Cambridge Isotope Laboratories, Inc. and Gute Chemie, and were degassed and stored over activated 3Å molecular sieves. Compounds **9**^[1, 2], **11**^[3], **12**^[4], **13**^[5] was synthesized according to literature. All other reagents were purchased from commercial sources. Liquid compounds were degassed by standard freeze-pump-thaw procedures prior to use.

B. Physical Methods

The ¹H, ¹⁹F and ¹³C spectra were recorded on a Bruker Avance 400 spectrometer. The chemical shifts (δ) are given in parts per million relative to solvent peaks (CDCl₃ δ 7.26ppm ¹H NMR and 77.16 ppm in ¹³C NMR, CD₃CN, 1.94 ppm in ¹H NMR and 1.32 in ¹³C NMR, CD₂Cl₂, 5.32 ppm in ¹H NMR and 53.84 in ¹³C NMR, THF-*d*₈, 1.72 ppm in ¹H NMR and 25.31 in ¹³C NMR). IR spectra of the complexes were recorded as solution samples on a Varian 800 FT-IR spectrometer. Elemental analyses were performed on a Carlo Erba EA 1110 CHN instrument at EPFL. X-ray diffraction studies were carried out in the EPFL Crystallographic Facility. Data collections were performed at low temperature using four-circle kappa diffractometers equipped with CCD detectors. Data were reduced and then corrected for absorption. Solution, refinement and geometrical calculations for all crystal structures were performed by SHELXTL.^[6, 7]

2. Experimental details

2.1 Synthesis of complex 5



Scheme 1 Synthesis of complex 5.

Compound **6** (600 mg, 1.0 equiv.) was dissolved in 25 mL dry THF in a Schlenk flask. To this solution, *n*-BuLi (1.0 equiv.) was added dropwise at 0 °C and the solution was further stirred for 30 min at 0 °C. In another Schlenk flask a THF solution of Mn(CO)₅Br (1.10 g, 1.0 equiv.) was cooled to -78 °C. The solution of deprotonated **6** was then added dropwise to the Mn(CO)₅Br solution at -78 °C. The resulting mixture was allowed to slowly warm to room temperature and was further heated to 50 °C. After stirring at 50 °C overnight, the mixture was cooled to room temperature. The THF solvent was removed. The residue was further purified by silica gel chromatography in glovebox using ethyl acetate/hexane as eluent. Yield 71%. Single crystal suitable for X-ray test was obtained via layer diffusion of pentane to a THF solution of complex at -22 °C.

2.2 General procedure for hydrogenation of aldehydes, ketones and imines.

Substrate (1.0 equiv., 1 mmol for aldehydes and ketones, 0.2 mmol for imines), *N*-methyl pyrrolidine (0.5 equiv.), complex **5** (0.02 equiv.) and dry THF (1 mL for aldehydes and ketone, 0.5 mL for imines) were added to a 2 mL tube. The tube was then put into a 50 mL autoclave. After addition of 50 bar H₂ gas, the autoclave was heated to 100 °C for 24 h. Products were isolated and purified through a short silica gel chromatography using ethyl acetate/hexane as eluent.

2.3 General procedure for hydrogenation of methenyl-H₄MPT⁺ mimic substrates

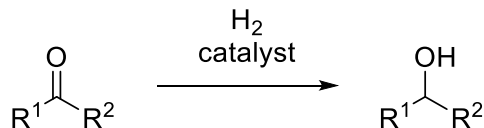
Substrate (1.0 equiv., 0.1 mmol), *N*-methyl pyrrolidine (5 equiv.), complex **5** (0.025 equiv.) and 3 mL dry dioxane were added to a 10 mL vial. The vial was then put into a 250 mL autoclave. After addition of 50 bar H₂ gas, the autoclave was heated to 80 °C for 24 h. Yield of **10a** and **10b** was determined by ¹H NMR using 1,3,5-trimethoxybenzene as internal standard. **10c-e** were isolated and purified through a silica gel chromatography using ethyl acetate/hexane as eluent.

2.4 General procedure for asymmetric relay hydrogenation

Substrate (1.0 equiv., 0.1 mmol), **13** (0.02 mmol, 0.2 equiv.), La(OTf)₃ (0.02 mmol, 0.2 equiv.), complex **5** (0.1 equiv.) and 1 mL dry CHCl₃ were added to a 2 mL tube. The tube was then put into a 50 mL autoclave. After addition of 50 bar H₂ gas, the autoclave was heated to 80 °C for 48 h. Products were isolated and purified through preparative TLC. Ee was determined by chiral HPLC using OD-H column.

2.5 Reaction condition optimization

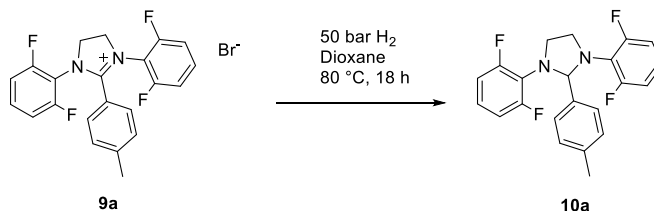
Table S1 Optimization of reaction conditions for hydrogenation of benzaldehyde and acetophenone.



	substrate	Catalyst (mol%)	Base (mol%)	solvent	T (°C)	Time	P (bar)	Yield (%)
1	benzaldehyde	4 (5)	MP(25)	THF	50	16 h	50	7
2	benzaldehyde	4 (5)	MP(25)	THF	80	16 h	50	100
3	benzaldehyde	4 (5)	MP(25)	CH ₃ CN	80	16 h	50	72
4	benzaldehyde	4 (5)	MP(25)	MeOH	80	16 h	50	84
5	benzaldehyde	4 (1)	MP(20)	THF	80	16 h	50	100
6	benzaldehyde	4 (1)	MP(20)	THF	80	16 h	30	98
7	benzaldehyde	4 (1)	MP(20)	THF	80	16 h	10	88
8	benzaldehyde	4 (1)	MP(20)	THF	80	16 h	1	1
9	acetophenone	4 (2)	MP(50)	THF	100	24 h	50	45
10	acetophenone	5 (2)	MP(50)	THF	100	24 h	50	91

MP = *N*-methyl pyrrolidine

Table S2 Optimization of reaction conditions for hydrogenation of **9a**



	catalyst	Base	Concentration	Yield
1	5 (20 mol%)	MP(1.0 equiv.)	0.50	25%
2	5 (20 mol%)	MP(2.0 equiv.)	0.50	53%
3	5 (20 mol%)	MP(5.0 equiv.)	0.50	67%
4	5 (10 mol%)	MP(5.0 equiv.)	0.50	62%
5	5 (10 mol%)	MP(5.0 equiv.)	0.25	87%
6	5 (10 mol%)	MP(5.0 equiv.)	0.17	90%
7	5 (5.0 mol%)	MP(5.0 equiv.)	0.17	94%
8	5 (2.5 mol%)	MP(5.0 equiv.)	0.17	92%
9	4 (2.5 mol%)	MP(5.0 equiv.)	0.17	43%
10	11 (2.5 mol%)	MP(5.0 equiv.)	0.17	0%
		KOtBu(10 mol%)		
11	12 (2.5 mol%)	MP(5.0 equiv.)	0.17	0%
		KOtBu(10 mol%)		

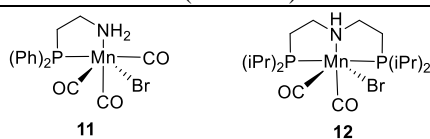
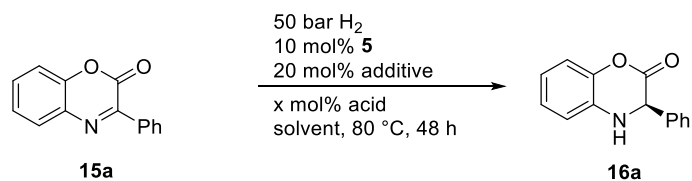
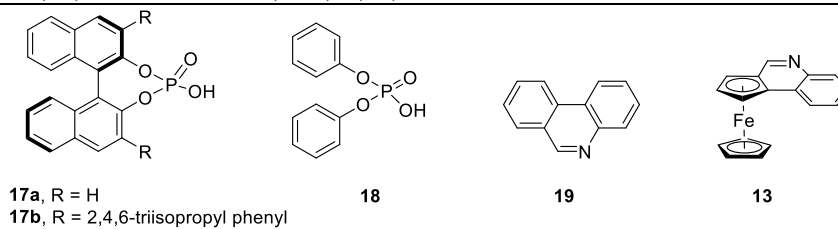


Table S3 Optimization of reaction conditions for asymmetric hydrogenation of **13a** catalyzed by **5**.

	Additive (%)	Acid (%)	solvent	Yield	ee
1	--	17a (10)	dioxane	<2% ^a	--
2	19 (20)	17a (10)	dioxane	70% ^a	--
3	19 (20)	17b (5)	dioxane	<2% ^{a,c}	--
4	19 (20)	17b (5)	toluene	36% ^{a,c}	70%
5	19 (20)	17b (5)	mesitylene	81% ^{b,c}	80%
6	13 (20)	18 (20)	mesitylene	<5%	--
7	13 (20)	18 (20)	CHCl ₃	46%	94%
8	13 (20)	Sm(OTf) ₃ (20)	CHCl ₃	<5%	--
9	13 (20)	La(OTf) ₃ (20)	CHCl ₃	63%	96%



^a18 h of reaction time; ^b72 h of reaction time; ^c60 °C

3. Characterization of complex 5

^1H NMR (400 MHz, THF- d_8) δ 7.77 (t, $J = 8.0$ Hz, 1H), 6.82 (d, $J = 7.7$ Hz, 1H), 6.75 (d, $J = 8.2$ Hz, 1H), 3.15 (s, 3H), 2.74 (s, 6H).

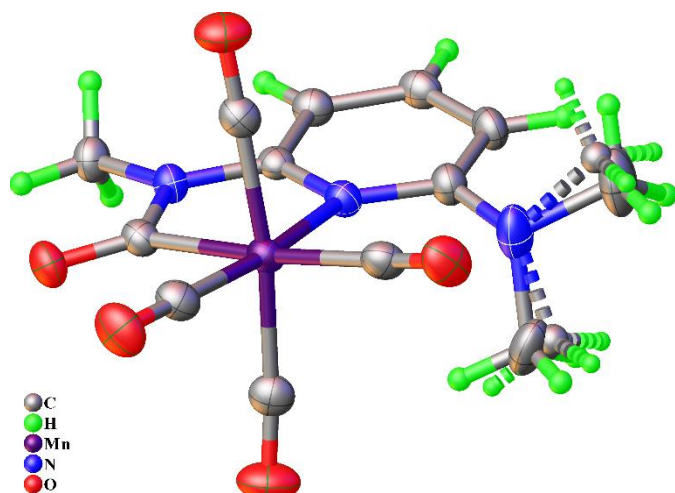
^{13}C NMR (101 MHz, THF) δ 218.26, 216.55, 214.90, 213.62, 168.47, 161.40, 141.41, 109.20, 105.54, 44.29, 27.46.

HRMS (APCI/QTOF) m/z : $[\text{M} + \text{Na}]^+$ Calcd for $\text{C}_{13}\text{H}_{12}\text{MnN}_3\text{NaO}_5^+$ 368.0050; Found 368.0049

IR: $\nu(\text{cm}^{-1})$ 1952 (s, terminal CO), 1975 (s, terminal CO), and 2072 (s, terminal CO)

Anal. Calcd for $\text{C}_{11}\text{H}_7\text{MnN}_2\text{O}_5$: C, 45.2; H, 3.5; N, 12.2. Found: C, 45.1; H, 3.5; N, 12.0.

Crystal structure



Experimental. Single clear pale yellow plate-shaped crystals of complex **5** were obtained by recrystallisation from THF/pentane at $-22\text{ }^{\circ}\text{C}$. A suitable crystal of $0.54 \times 0.24 \times 0.16\text{ mm}^3$ was selected and mounted on a suitable support on a SuperNova, Dual, Cu at zero, Atlas diffractometer. The crystal was kept at a steady $T = 140.00(10)\text{ K}$ during data collection. The structure was solved with the **ShelXT** ^[6] structure solution program using the dual solution method and by using **Olex2** ^[8] as the graphical interface. The model was refined with version 2018/3 of **ShelXL** ^[6] using full matrix least squares on $|F|^2$ minimisation.

Crystal Data. $\text{C}_{13}\text{H}_{12}\text{MnN}_3\text{O}_5$, $M_r = 345.20$, monoclinic, $P2_1/c$ (No. 14), $a = 13.2766(3)\text{ \AA}$, $b = 6.48128(12)\text{ \AA}$, $c = 17.8426(4)\text{ \AA}$, $\beta = 103.307(2)^{\circ}$, $\alpha = \gamma = 90^{\circ}$, $V = 1494.12(5)\text{ \AA}^3$, $T = 140.00(10)\text{ K}$, $Z = 4$, $Z' = 1$, $\mu(\text{CuK}\alpha) = 7.447$, 9742 reflections measured, 3033 unique ($R_{int} = 0.0260$) which were used in all calculations. The final wR_2 was 0.0781 (all data) and R_1 was 0.0285 ($I > 2(I)$).

Compound	Complex 5
Formula	$\text{C}_{13}\text{H}_{12}\text{MnN}_3\text{O}_5$
$D_{calc.}/\text{g cm}^{-3}$	1.535
μ/mm^{-1}	7.447
Formula Weight	345.20
Colour	clear pale yellow
Shape	plate
Size/ mm^3	$0.54 \times 0.24 \times 0.16$
T/K	140.00(10)
Crystal System	monoclinic
Space Group	$P2_1/c$
$a/\text{\AA}$	13.2766(3)
$b/\text{\AA}$	6.48128(12)
$c/\text{\AA}$	17.8426(4)
α°	90
β°	103.307(2)
γ°	90
$V/\text{\AA}^3$	1494.12(5)
Z	4
Z'	1
Wavelength/ \AA	1.54184
Radiation type	$\text{CuK}\alpha$
θ_{min}°	5.094
θ_{max}°	75.239
Measured Refl.	9742
Independent Refl.	3033
Reflections with $I > 2(I)$	2936
R_{int}	0.0260
Parameters	224
Restraints	19
Largest Peak/ $e\text{\AA}^{-3}$	0.421
Deepest Hole/ $e\text{\AA}^{-3}$	-0.383
GooF	1.057
wR_2 (all data)	0.0781
wR_2	0.0772
R_1 (all data)	0.0294
R_1	0.0285

Detailed experimental procedure:

A clear pale yellow plate-shaped crystal with dimensions of 0.54×0.24×0.16 mm³ was mounted on a suitable support. Data were collected using a SuperNova, Dual, Cu at zero, Atlas diffractometer operating at $T = 140.00(10)$ K.

Data were measured using ω scans using CuK α radiation. The total number of runs and images was based on the strategy calculation from the program **CrysAlisPro** (Rigaku, V1.171.38.46, 2015). The maximum resolution achieved was $\theta = 75.239^\circ$ (0.83 Å).

The diffraction pattern was indexed. The total number of runs and images was based on the strategy calculation from the program **CrysAlisPro** (Rigaku, V1.171.38.46, 2015) and the unit cell was refined using **CrysAlisPro** (Rigaku, V1.171.38.46, 2015) on 5255 reflections, 54% of the observed reflections.

Data reduction, scaling and absorption corrections were performed using **CrysAlisPro** (Rigaku, V1.171.38.46, 2015). The final completeness is 100.00 % out to 75.239° in θ . A Gaussian absorption correction was performed using **CrysAlisPro** 1.171.38.46 (Rigaku Oxford Diffraction, 2018) Numerical absorption correction based on Gaussian integration over a multifaceted crystal model/Empirical absorption correction using spherical harmonics as implemented in SCALE3 ABSPACK scaling algorithm.. The absorption coefficient μ of this material is 7.447 mm⁻¹ at this wavelength ($\lambda = 1.542\text{Å}$) and the minimum and maximum transmissions are 0.197 and 0.766.

The structure was solved and the space group $P2_1/c$ (# 14) determined by the **ShelXT** ^[6] structure solution program using dual and refined by full matrix least squares on $|F|^2$ using version 2018/3 of **ShelXL** ^[7]. All non-hydrogen atoms were refined anisotropically. Hydrogen atom positions were calculated geometrically and refined using the riding model.

There is a single molecule in the asymmetric unit, which is represented by the reported sum formula. In other words: Z is 4 and Z' is 1.

CCDC- 1958229 contains the supplementary crystallographic data for **5**. These data can be obtained free of charge from *The Cambridge Crystallographic Data Centre* via www.ccdc.cam.ac.uk/data_request/cif.

IR spectrum

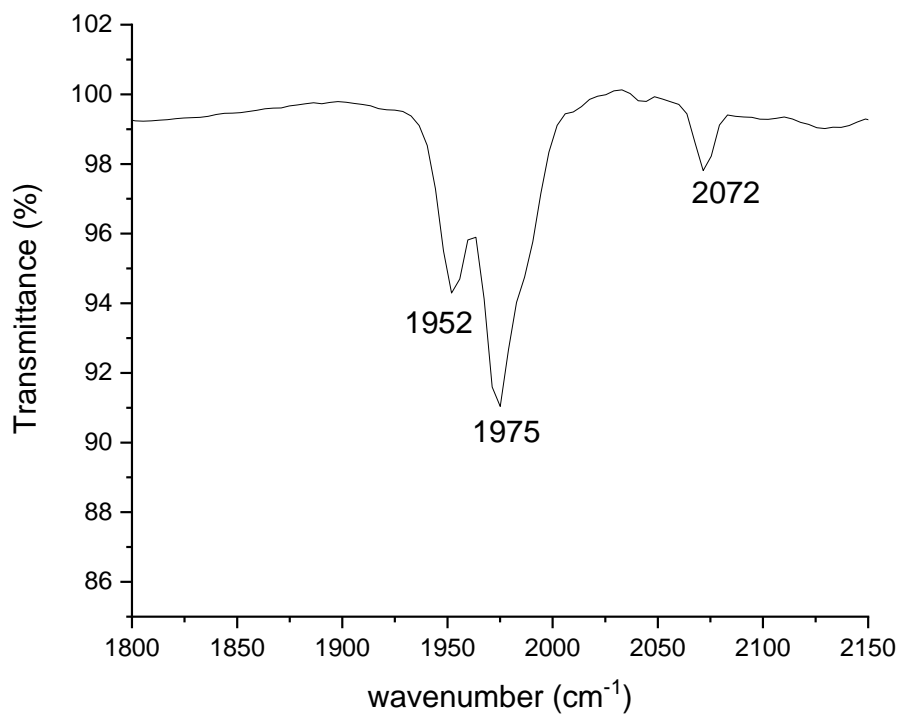
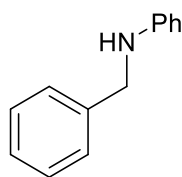


Figure S1 IR spectrum of complex 5 in THF solution

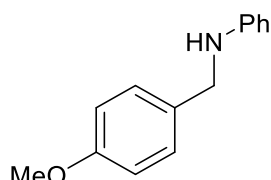
4. Characterization of compounds



8j

$^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 7.45 – 7.28 (m, 5H), 7.25 – 7.17 (m, 2H), 6.81 – 6.73 (m, 1H), 6.71 – 6.63 (m, 2H), 4.36 (s, 2H), 4.08 (s, 1H).

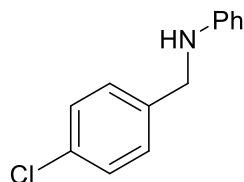
$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 148.34, 139.64, 129.49, 128.85, 127.74, 127.45, 117.81, 113.09, 48.55.



8k

$^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 7.32 (d, $J = 8.7$ Hz, 2H), 7.21 (dd, $J = 8.5, 7.2$ Hz, 2H), 6.92 (d, $J = 8.6$ Hz, 2H), 6.75 (ddd, $J = 8.4, 6.8, 1.1$ Hz, 1H), 6.67 (d, $J = 7.3$ Hz, 2H), 4.28 (s, 2H), 4.01 (s, 1H), 3.83 (s, 3H).

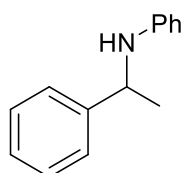
$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 158.98, 148.29, 131.50, 129.36, 128.93, 117.63, 114.14, 112.98, 55.40, 47.92.



8l

$^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 7.36-7.29 (m, 4H), 7.24 – 7.17 (m, 2H), 6.76 (tt, $J = 7.3, 1.1$ Hz, 1H), 6.68 – 6.60 (m, 2H), 4.33 (s, 2H), 4.10 (s, 1H).

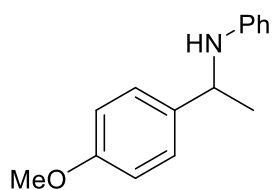
$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 147.91, 138.10, 132.98, 129.42, 128.86, 128.81, 117.94, 113.03, 47.73.



8m

$^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 7.43 – 7.31 (m, 4H), 7.29 – 7.22 (m, 1H), 7.15-7.08 (m, 2H), 6.71 – 6.64 (m, 1H), 6.58-6.50 (m, 2H), 4.52 (q, $J = 6.7$ Hz, 1H), 4.11 (s, 1H), 1.55 (d, $J = 6.7$ Hz, 3H).

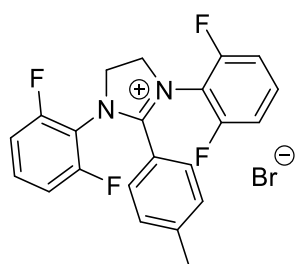
$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 147.36, 145.32, 129.23, 128.76, 127.00, 125.98, 117.40, 113.46, 53.62, 25.13.



8n

^1H NMR (400 MHz, Chloroform-*d*) δ 7.35 – 7.27 (m, 2H), 7.17 – 7.07 (m, 2H), 6.92 – 6.85 (m, 2H), 6.67 (tt, $J = 7.3, 1.1$ Hz, 1H), 6.59 – 6.50 (m, 2H), 4.47 (q, $J = 6.7$ Hz, 1H), 4.06 (s, 1H), 3.80 (s, 3H), 1.52 (d, $J = 6.7$ Hz, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 158.61, 147.42, 137.34, 129.21, 127.02, 117.33, 114.12, 113.47, 55.36, 52.98, 25.09.

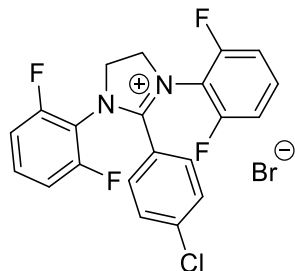


9a

^1H NMR (400 MHz, Acetonitrile-*d*₃) δ 7.64 – 7.48 (m, 2H), 7.24 – 7.05 (m, 8H), 4.68 (s, 4H), 2.27 (s, 3H).

^{13}C NMR (101 MHz, Acetonitrile-*d*₃) δ 170.14, 158.64 (dd, $J = 253.9, 3.3$ Hz), 147.27, 134.06 (t, $J = 10.1$ Hz), 131.02, 129.16, 114.30 (t, $J = 16.1$ Hz), 113.91 (dd, $J = 19.3, 3.6$ Hz), 53.22, 21.67.

^{19}F NMR (376 MHz, CD_3CN) δ -118.50.

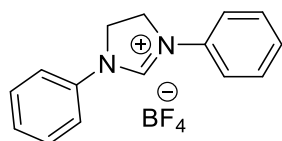


9b

^1H NMR (400 MHz, Acetonitrile-*d*₃) δ 7.56 (tt, $J = 8.6, 6.3$ Hz, 2H), 7.44 – 7.35 (m, 2H), 7.35 – 7.27 (m, 2H), 7.22 – 7.07 (m, 4H), 4.72 (s, 4H).

^{13}C NMR (101 MHz, Acetonitrile-*d*₃) δ 169.04, 158.44 (dd, $J = 254.3, 3.2$ Hz), 141.52, 134.22 (t, $J = 10.1$ Hz), 130.91, 130.85, 119.97, 113.99 (dd, $J = 19.1, 3.6$ Hz), 113.80 (t, $J = 16.0$ Hz), 53.41.

^{19}F NMR (376 MHz, CD_3CN) δ -118.12.

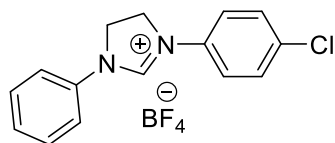


9c

¹H NMR (400 MHz, Acetonitrile-*d*₃) δ 9.08 (s, 1H), 7.62 – 7.52 (m, 4H), 7.51 – 7.37 (m, 6H), 4.58 (s, 4H).

¹³C NMR (101 MHz, CD₃CN) δ 152.42, 136.87, 131.02, 128.85, 119.69, 50.03.

¹⁹F NMR (376 MHz, CD₃CN) δ -151.78.

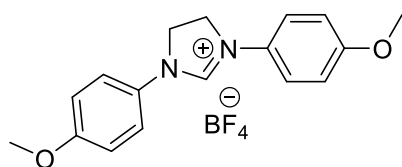


9d

¹H NMR (400 MHz, Acetonitrile-*d*₃) δ 9.07 (s, 1H), 7.60-7.53 (m, 4H), 7.52 – 7.33 (m, 5H), 4.56 (s, 4H).

¹³C NMR (101 MHz, CD₃CN) δ 152.64, 136.75, 135.75, 133.75, 131.06, 130.96, 129.03, 121.35, 119.79, 50.18, 50.12.

¹⁹F NMR (376 MHz, CD₃CN) δ -151.71.

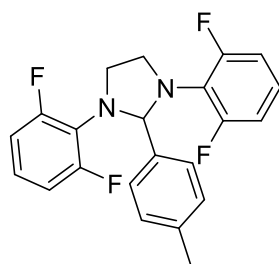


9e

¹H NMR (400 MHz, Acetonitrile-*d*₃) δ 8.81 (s, 1H), 7.38 (d, *J* = 9.1 Hz, 4H), 7.07 (d, *J* = 9.1 Hz, 4H), 4.50 (s, 4H), 3.84 (s, 6H).

¹³C NMR (101 MHz, CD₃CN) δ 160.08, 151.71, 130.03, 121.46, 116.02, 56.43, 50.45.

¹⁹F NMR (376 MHz, CD₃CN) δ -151.79.



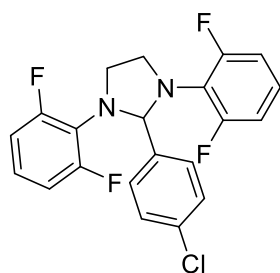
10a

¹H NMR (400 MHz, Acetonitrile-*d*₃) δ 7.23 (d, *J* = 7.6 Hz, 2H), 7.01 – 6.89 (m, 4H), 6.84 (q, *J* = 8.4, 7.5 Hz, 4H), 6.07 (s, 1H), 4.10 – 3.95 (m, 2H), 3.67 – 3.51 (m, 2H), 2.18 (s, 3H).

¹³C NMR (101 MHz, Acetonitrile-*d*₃) δ 159.92 (dd, *J* = 247.2, 7.8 Hz), 139.30, 138.70, 129.52, 128.96, 125.13 (t, *J* = 10.3 Hz), 123.87 (t, *J* = 13.8 Hz), 113.00 (dd), 80.05 (p, *J* = 4.5 Hz), 51.36 (t, *J* = 3.6 Hz), 21.10.

¹⁹F NMR (376 MHz, CD₃CN) δ -119.24.

HRMS (ESI/QTOF) *m/z*: [M + H]⁺ Calcd for C₂₂H₁₉F₄N₂⁺ 387.1479; Found 387.1478



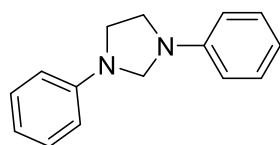
10b

^1H NMR (400 MHz, Acetonitrile- d_3) δ 7.33 (dd, $J = 8.5, 2.1$ Hz, 2H), 7.16 (dd, $J = 8.5, 2.1$ Hz, 2H), 6.98 (qd, $J = 7.8, 7.3, 3.5$ Hz, 2H), 6.86 (q, $J = 8.4, 7.3$ Hz, 4H), 6.03 (s, 1H), 4.07 – 3.94 (m, 2H), 3.71 – 3.54 (m, 2H).

^{13}C NMR (101 MHz, Acetonitrile- d_3) δ 160.00 (dd, $J = 247.4, 7.5$ Hz), 140.71, 134.60, 130.67, 128.92, 125.58 (t, $J = 10.3$ Hz), 123.54 (t, $J = 13.9$ Hz), 113.09 (dd), 79.94 (p, $J = 4.0$ Hz), 51.47.

^{19}F NMR (376 MHz, CD_3CN) δ -119.26.

HRMS (ESI/QTOF) m/z : $[\text{M} + \text{H}]^+$ Calcd for $\text{C}_{21}\text{H}_{16}\text{ClF}_4\text{N}_2^+$ 407.0933; Found 407.0931.

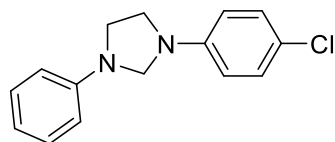


10c

^1H NMR (400 MHz, Chloroform- d) δ 7.33 (t, $J = 7.7$ Hz, 4H), 6.84 (t, $J = 7.4$ Hz, 2H), 6.70 (d, $J = 8.2$ Hz, 4H), 4.69 (s, 2H), 3.67 (s, 4H).

^{13}C NMR (101 MHz, CDCl_3) δ 146.52, 129.48, 117.77, 112.57, 65.98, 46.60.

HRMS (ESI/QTOF) m/z : $[\text{M} + \text{H}]^+$ Calcd for $\text{C}_{15}\text{H}_{15}\text{N}_2^+$ 223.1230; Found 223.1225.

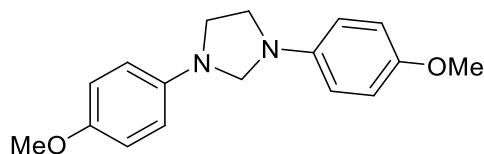


10d

^1H NMR (400 MHz, Chloroform- d) δ 7.30 (t, $J = 7.4$ Hz, 2H), 7.23 (d, $J = 7.9$ Hz, 2H), 6.82 (t, $J = 7.4$ Hz, 1H), 6.66 (d, $J = 8.0$ Hz, 2H), 6.57 (d, $J = 7.5$ Hz, 2H), 4.62 (s, 2H), 3.63 (h, $J = 6.8, 6.4$ Hz, 4H).

^{13}C NMR (101 MHz, CDCl_3) δ 146.39, 145.05, 129.52, 129.30, 122.66, 118.00, 113.57, 112.65, 66.03, 46.72, 46.62.

HRMS (ESI/QTOF) m/z : $[\text{M} + \text{H}]^+$ Calcd for $\text{C}_{15}\text{H}_{14}\text{ClN}_2^+$ 257.0840; Found 257.0846.

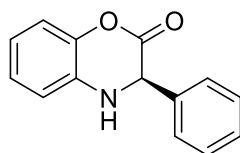


10e

^1H NMR (400 MHz, Methylene Chloride- d_2) δ 6.87 (d, $J = 8.7$ Hz, 4H), 6.63 (d, $J = 8.8$ Hz, 4H), 4.54 (s, 2H), 3.75 (s, 6H), 3.57 (s, 4H).

^{13}C NMR (101 MHz, CD_2Cl_2) δ 152.57, 141.88, 115.22, 113.81, 67.74, 56.04, 47.78.

HRMS (ESI/QTOF) m/z : $[\text{M} + \text{H}]^+$ Calcd for $\text{C}_{17}\text{H}_{21}\text{N}_2\text{O}_2^+$ 285.1598; Found 285.1591.

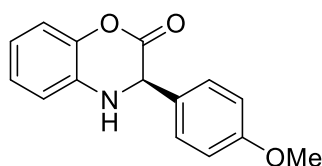


16a

^1H NMR (400 MHz, Chloroform-*d*) δ 7.44-7.33 (m, 5H), 7.07-6.99 (m, 2H), 6.87 (t, $J = 7.8$ Hz, 1H), 6.81 (d, $J = 7.8$ Hz, 1H), 5.06 (s, 1H), 4.22 (br, 1H).

^{13}C NMR (101 MHz, CDCl_3) δ 165.32, 141.03, 136.48, 132.49, 129.11, 127.60, 125.31, 120.51, 117.09, 115.01, 59.39.

HRMS (APPI/LTQ-Orbitrap) m/z : $[\text{M} + \text{H}_1]^+$ Calcd for $\text{C}_{14}\text{H}_{10}\text{NO}_2^+$ 224.0706; Found 224.0697.

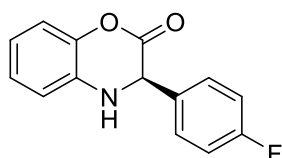


16b

^1H NMR (400 MHz, Chloroform-*d*) δ 7.33 (d, $J = 8.3$ Hz, 2H), 7.09 – 6.98 (m, 2H), 6.89-6.77 (m, 4H), 5.00 (s, 1H), 4.18 (br, 1H), 3.80 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 165.64, 160.22, 141.14, 132.69, 128.93, 128.53, 125.26, 120.53, 117.13, 114.98, 114.53, 58.97, 55.48.

HRMS (APCI/QTOF) m/z : $[\text{M} + \text{H}]^+$ Calcd for $\text{C}_{15}\text{H}_{14}\text{NO}_3^+$ 256.0968; Found 256.0969.



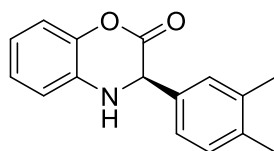
16c

^1H NMR (400 MHz, Chloroform-*d*) δ 7.45-7.36 (m, 2H), 7.12 – 6.98 (m, 4H), 6.89 (td, $J = 7.8, 1.5$ Hz, 1H), 6.83 (dd, $J = 7.8, 1.6$ Hz, 1H), 5.05 (s, 1H), 4.21 (s, 1H).

^{13}C NMR (101 MHz, CDCl_3) δ 165.23, 164.37, 161.91, 141.06, 132.39, 132.24, 132.21, 129.59, 129.50, 125.39, 120.77, 117.17, 116.22, 116.00, 115.08, 58.82.

^{19}F NMR (376 MHz, CDCl_3) δ -112.44.

HRMS (APPI/LTQ-Orbitrap) m/z : $[\text{M} + \text{H}_1]^+$ Calcd for $\text{C}_{14}\text{H}_9\text{FNO}_2^+$ 242.0612; Found 242.0601.

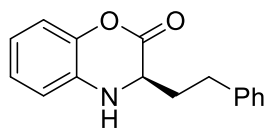


16d

^1H NMR (400 MHz, Chloroform-*d*) δ 7.18 (s, 1H), 7.15-7.09 (m, 2H), 7.07 – 6.97 (m, 2H), 6.90 – 6.83 (m, 1H), 6.83 – 6.76 (m, 1H), 4.99 (s, 1H), 2.25 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3) δ 165.64, 141.14, 137.75, 137.50, 133.85, 132.68, 130.31, 128.88, 125.23, 124.95, 120.46, 117.10, 114.97, 59.26, 19.98, 19.65.

HRMS (APCI/QTOF) m/z : $[\text{M} + \text{H}]^+$ Calcd for $\text{C}_{16}\text{H}_{16}\text{NO}_2^+$ 254.1176; Found 254.1176.

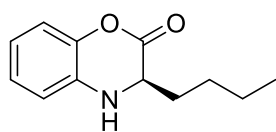


16e

$^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 7.35 – 7.26 (m, 2H), 7.28 – 7.17 (m, 3H), 7.03–6.95 (m, 2H), 6.84 (t, $J = 7.8$ Hz, 1H), 6.66 (d, $J = 7.8$ Hz, 1H), 3.93 (dd, $J = 7.4, 5.2$ Hz, 1H), 3.75 (br, 1H), 2.83 (hept, $J = 7.2$ Hz, 2H), 2.30 (dq, $J = 14.2, 6.0$ Hz, 1H), 2.09 (dq, $J = 15.0, 7.7$ Hz, 1H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 166.55, 141.24, 140.49, 132.32, 128.88, 128.54, 126.60, 125.09, 120.52, 116.91, 115.34, 54.57, 32.79, 31.87.

HRMS (APCI/QTOF) m/z : $[\text{M} + \text{H}]^+$ Calcd for $\text{C}_{16}\text{H}_{16}\text{NO}_2^+$ 254.1176; Found 254.1174.

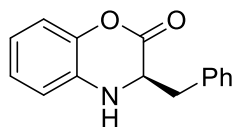


16f

$^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 6.98 (t, $J = 8.2$ Hz, 2H), 6.83 (t, $J = 7.8$ Hz, 1H), 6.77 (d, $J = 7.7$ Hz, 1H), 3.91 (dd, $J = 7.8, 5.2$ Hz, 1H), 3.60–4.25 (br, 1H), 2.00 – 1.84 (m, 1H), 1.77 (h, $J = 8.4, 8.0$ Hz, 1H), 1.51 – 1.27 (m, 4H), 0.92 (t, $J = 7.1$ Hz, 3H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 166.81, 141.17, 132.50, 125.05, 120.28, 116.83, 115.20, 54.93, 31.04, 27.45, 22.46, 13.97.

HRMS (APCI/QTOF) m/z : $[\text{M} + \text{H}]^+$ Calcd for $\text{C}_{12}\text{H}_{16}\text{NO}_2^+$ 206.1176; Found 206.1173.

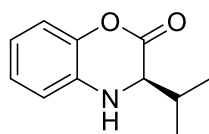


16g

$^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 7.42–7.36 (m, 2H), 7.35 – 7.28 (m, 1H), 7.20 (d, $J = 7.4$ Hz, 2H), 7.04 (dd, $J = 8.0, 1.6$ Hz, 1H), 6.99 (t, $J = 7.6$ Hz, 1H), 6.86 (t, $J = 7.8$ Hz, 1H), 6.65 (d, $J = 7.8$ Hz, 1H), 4.10 (dd, $J = 11.1, 3.3$ Hz, 1H), 3.79 (br, 1H), 3.34 (dd, $J = 13.7, 3.3$ Hz, 1H), 2.92 (dd, $J = 13.7, 11.0$ Hz, 1H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 166.40, 141.27, 136.00, 131.94, 129.42, 129.29, 127.61, 125.26, 120.55, 116.99, 115.48, 56.04, 37.08.

HRMS (APCI/QTOF) m/z : $[\text{M} + \text{H}]^+$ Calcd for $\text{C}_{15}\text{H}_{14}\text{NO}_2^+$ 240.1019; Found 240.1019.

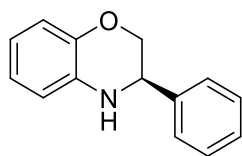


16h

$^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 7.03 – 6.91 (m, 2H), 6.86 – 6.70 (m, 2H), 3.99 (s, 1H), 3.76 (d, $J = 6.1$ Hz, 1H), 2.34 – 2.09 (m, $J = 6.9$ Hz, 1H), 1.06 (d, $J = 7.0$ Hz, 3H), 1.01 (d, $J = 6.7$ Hz, 3H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 165.82, 140.87, 132.23, 125.08, 119.92, 116.72, 114.85, 60.55, 30.06, 18.98, 17.74.

HRMS (APPI/LTQ-Orbitrap) m/z : $[M + H]^+$ Calcd for $C_{11}H_{14}NO_2^+$ 192.1019; Found 192.1011.

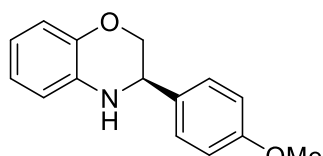


16i

1H NMR (400 MHz, Chloroform-*d*) δ 7.49 – 7.31 (m, 5H), 6.94 – 6.79 (m, 2H), 6.77 – 6.61 (m, 2H), 4.52 (dd, $J = 8.6, 3.0$ Hz, 1H), 4.31 (dd, $J = 10.7, 3.0$ Hz, 1H), 4.02 (dd, $J = 10.7, 8.6$ Hz, 1H), 3.60–4.20 (br, 1H).

^{13}C NMR (101 MHz, $CDCl_3$) δ 143.67, 139.22, 133.93, 128.94, 128.46, 127.32, 121.60, 119.10, 116.72, 115.54, 71.06, 54.34.

HRMS (ESI/QTOF) m/z : $[M + H]^+$ Calcd for $C_{14}H_{14}NO^+$ 212.1070; Found 212.1070.

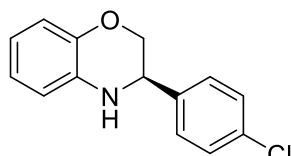


16j

1H NMR (400 MHz, Chloroform-*d*) δ 7.39 – 7.28 (m, 2H), 6.92 (d, $J = 8.0$ Hz, 2H), 6.89 – 6.76 (m, 2H), 6.76 – 6.63 (m, 2H), 4.53 – 4.40 (m, 1H), 4.40 – 4.04 (m, 2H), 3.98 (t, $J = 9.7$ Hz, 1H), 3.82 (s, 3H).

^{13}C NMR (101 MHz, $CDCl_3$) δ 159.79, 143.73, 133.85, 131.13, 128.50, 121.55, 119.20, 116.71, 115.63, 114.35, 71.15, 55.48, 53.76.

HRMS (ESI/QTOF) m/z : $[M + H]^+$ Calcd for $C_{15}H_{16}NO_2^+$ 242.1176; Found 242.1175.

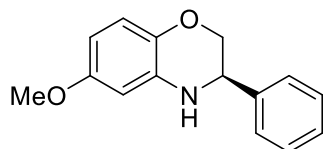


16k

1H NMR (400 MHz, Chloroform-*d*) δ 7.36 (m, 4H), 6.90 – 6.78 (m, 2H), 6.76–6.64 (m, 2H), 4.50 (dd, $J = 8.4, 3.0$ Hz, 1H), 4.00–4.40 (br, 1H), 4.26 (dd, $J = 10.7, 3.0$ Hz, 1H), 3.97 (dd, $J = 10.7, 8.4$ Hz, 1H).

^{13}C NMR (101 MHz, $CDCl_3$) δ 143.68, 137.75, 134.26, 133.48, 129.15, 128.68, 121.74, 119.44, 116.82, 115.70, 70.80, 53.77.

HRMS (ESI/QTOF) m/z : $[M + H]^+$ Calcd for $C_{14}H_{13}ClNO^+$ 246.0680; Found 246.0680.



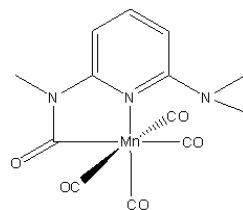
16l

1H NMR (400 MHz, Chloroform-*d*) δ 7.44 – 7.32 (m, 5H), 6.86 (s, 1H), 6.78 (d, $J = 8.4$ Hz, 1H), 6.58 (d, $J = 8.3$ Hz, 1H), 4.48 (d, $J = 8.6$ Hz, 1H), 4.29 (d, $J = 10.7$ Hz, 1H), 3.99 (t, $J = 9.6$ Hz, 1H), 4.55–3.80 (br, 1H)

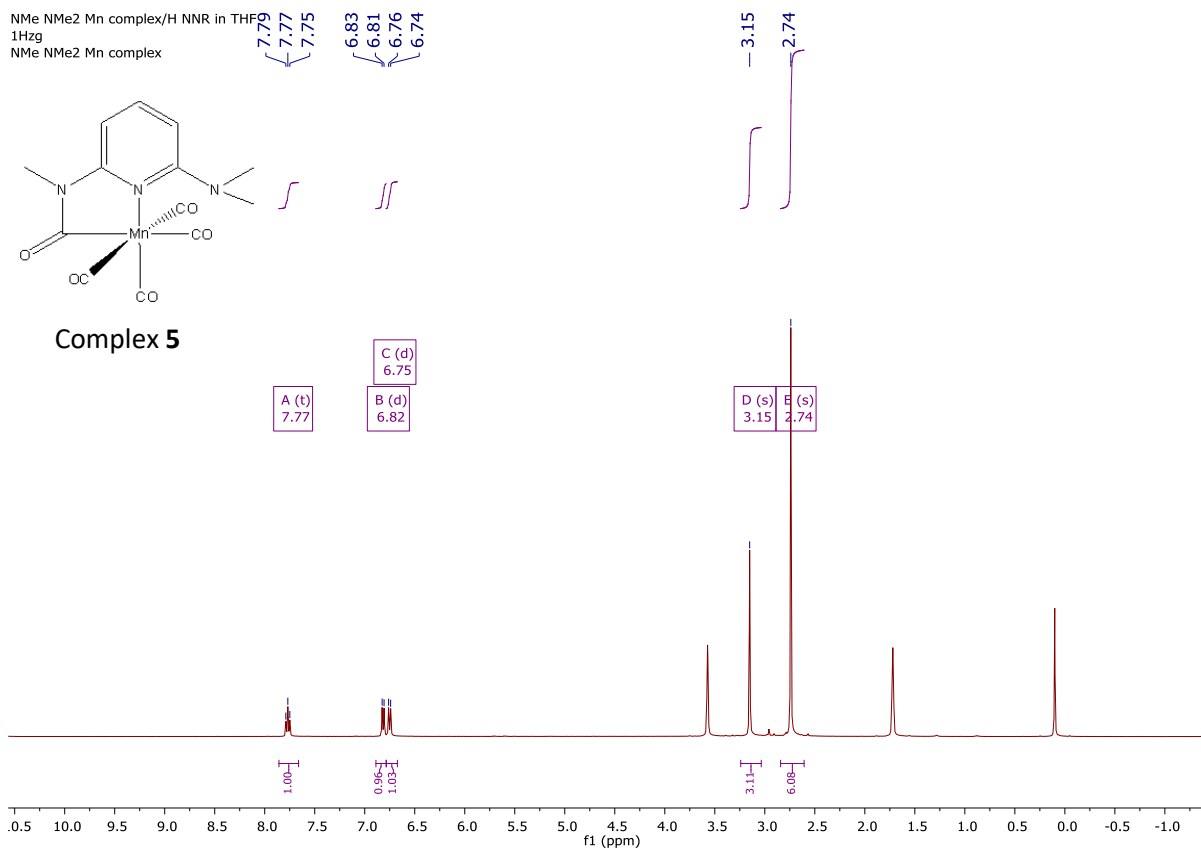
^{13}C NMR (101 MHz, CDCl_3) δ 144.18, 138.71, 132.55, 129.03, 128.65, 127.31, 123.39, 121.42, 116.92, 116.09, 70.98, 54.18.

5. Spectra

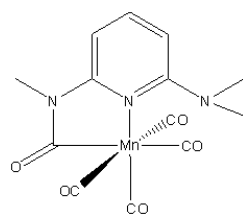
NMe NMe2 Mn complex/H NNR in THF
1Hzg
NMe NMe2 Mn complex



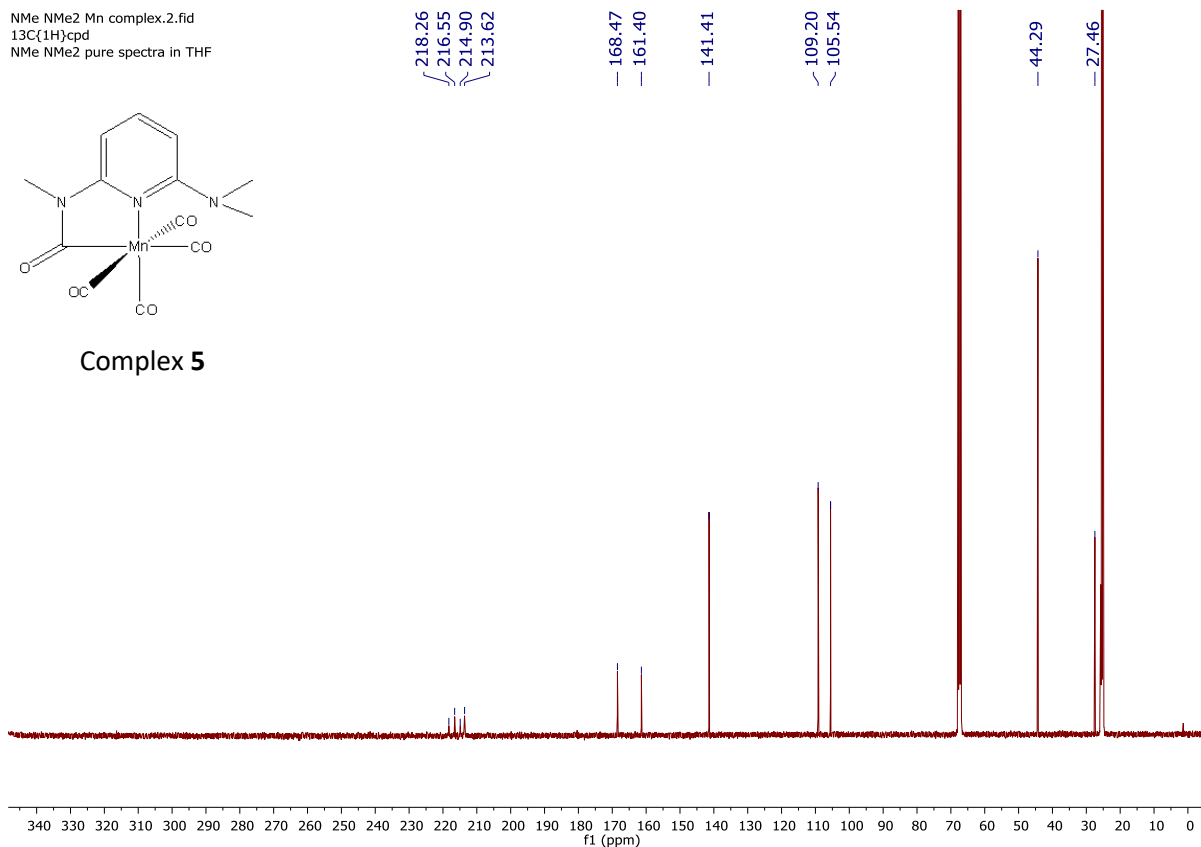
Complex 5



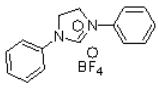
NMe NMe2 Mn complex.2.fid
13C{1H}cpd
NMe NMe2 pure spectra in THF



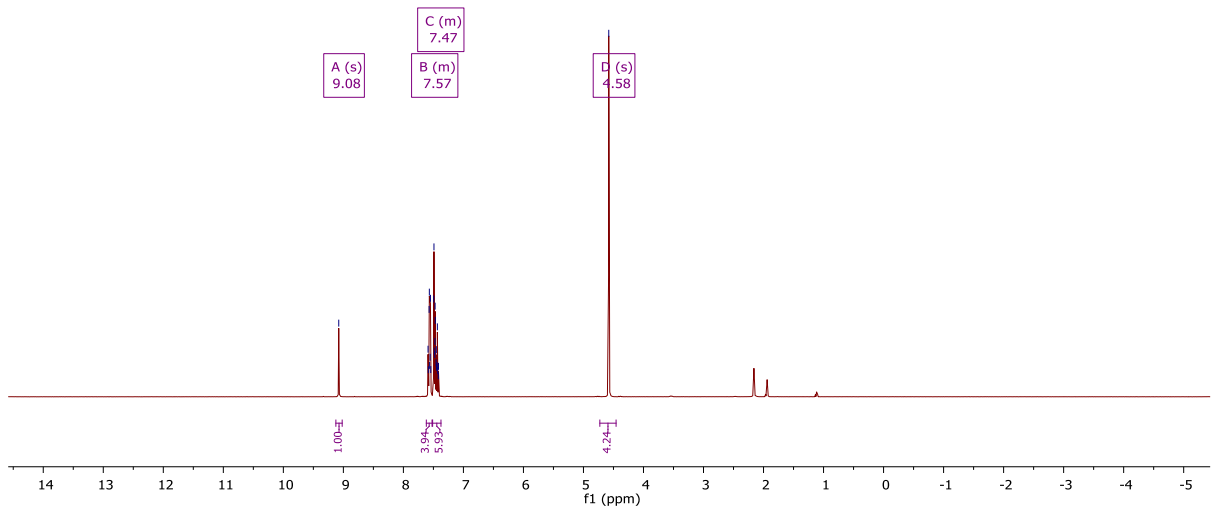
Complex 5



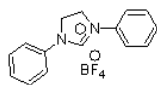
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1Hzg



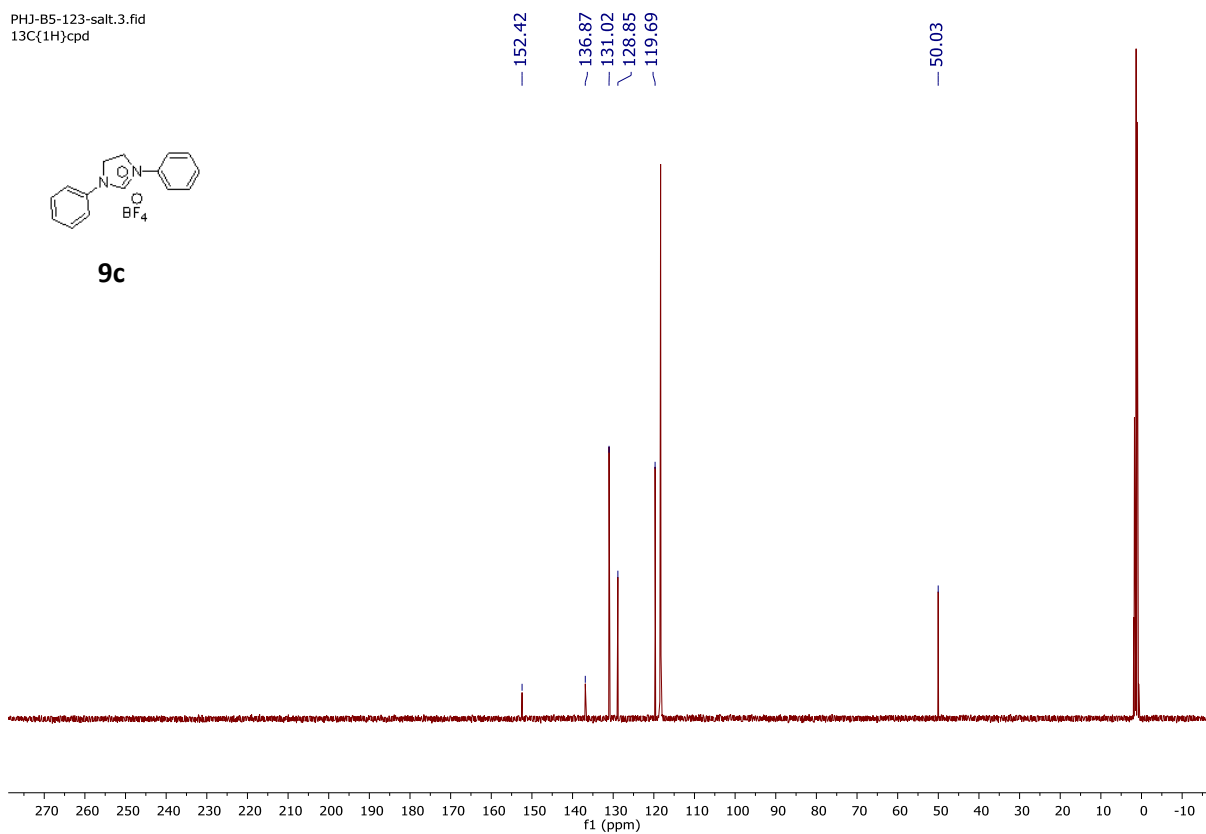
9c



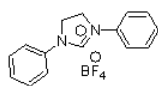
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13C{1H}cpd



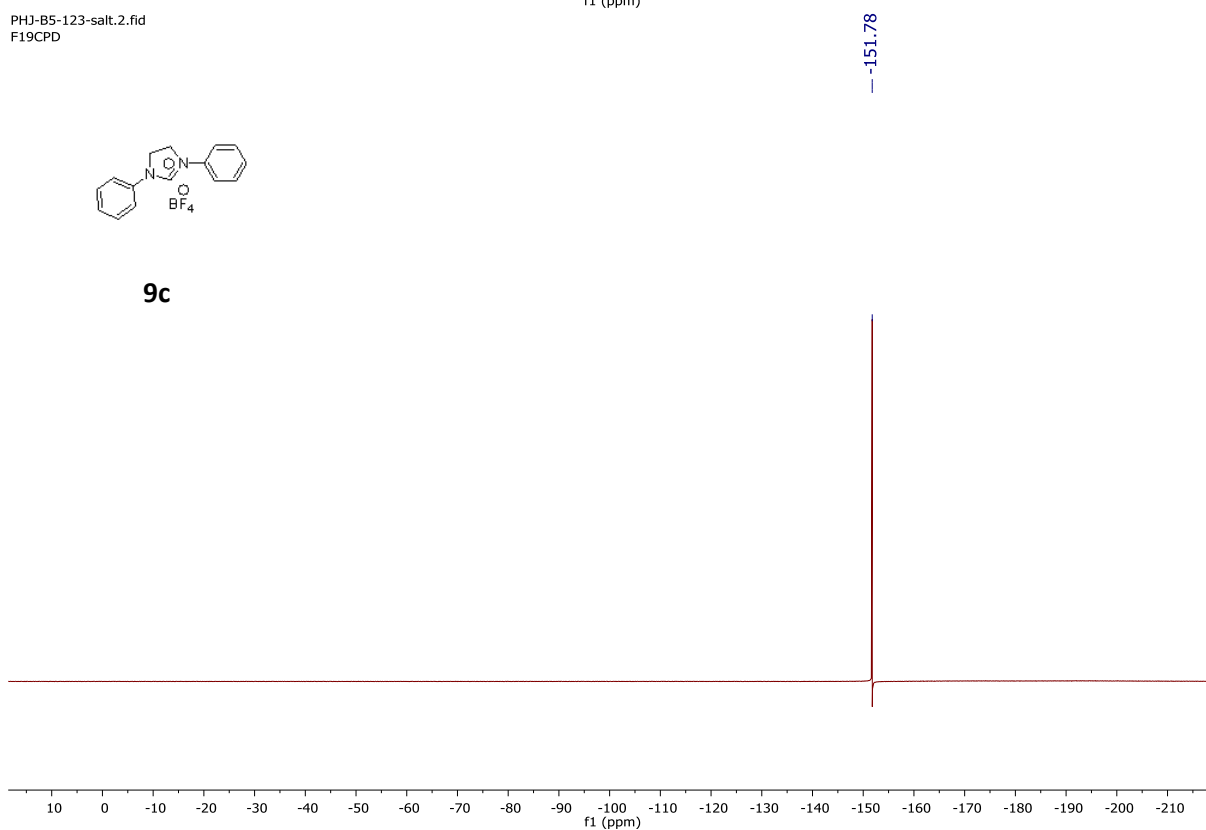
9c

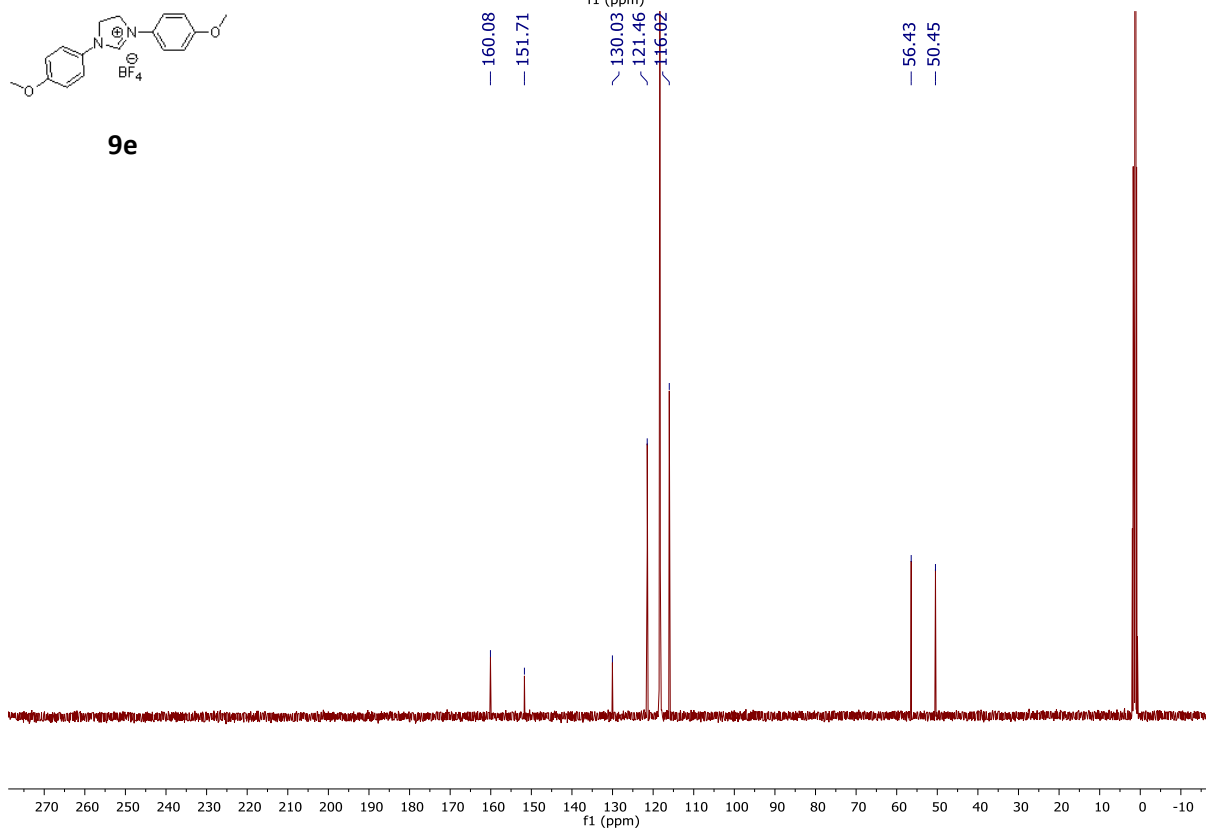
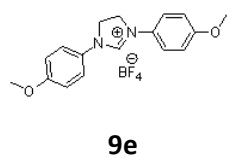
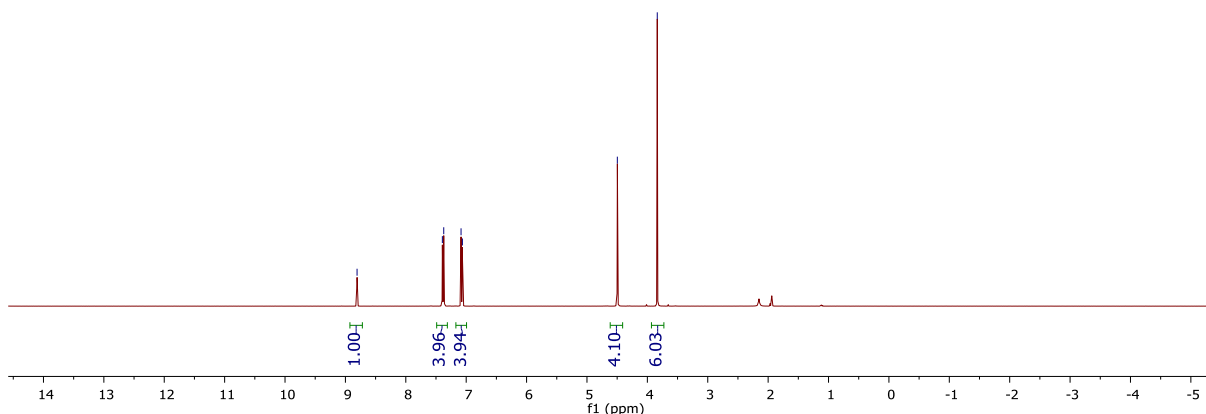
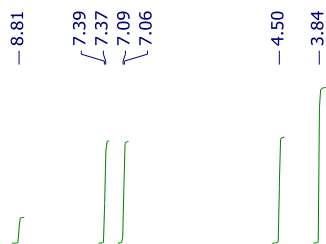
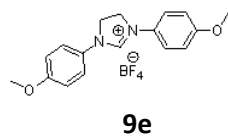


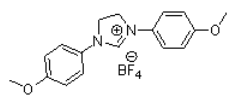
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9c

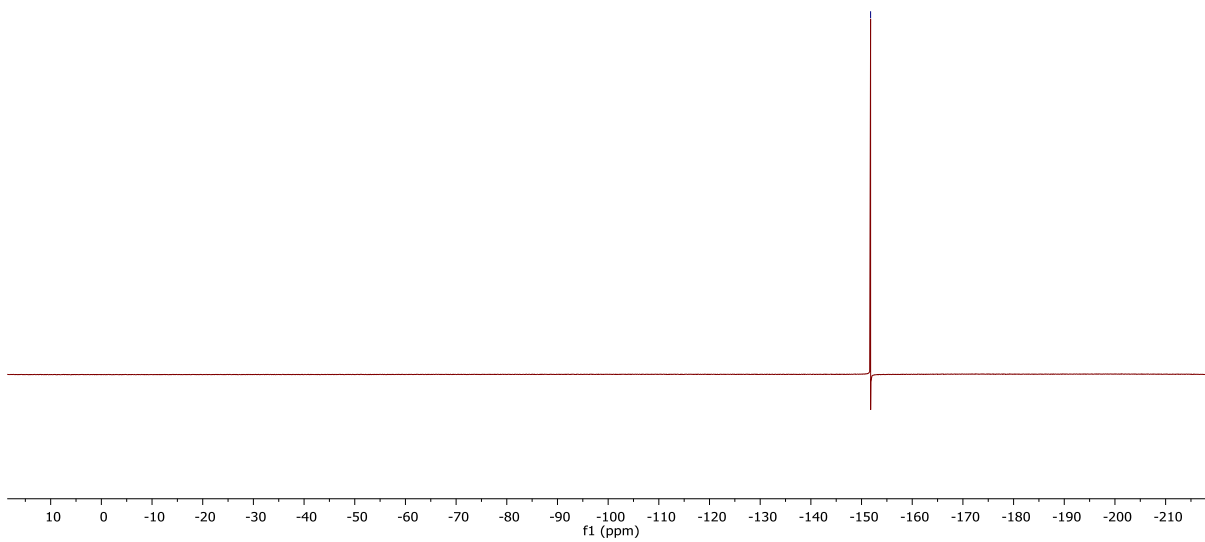


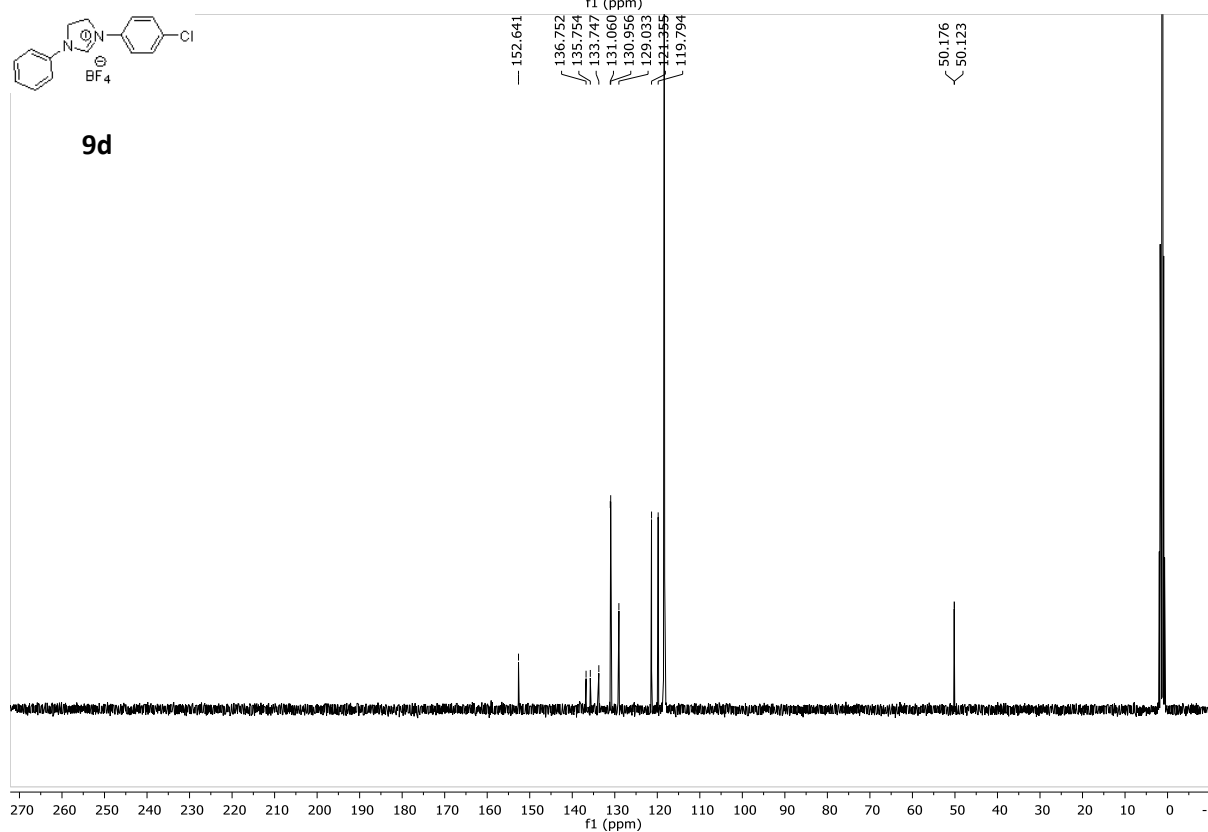
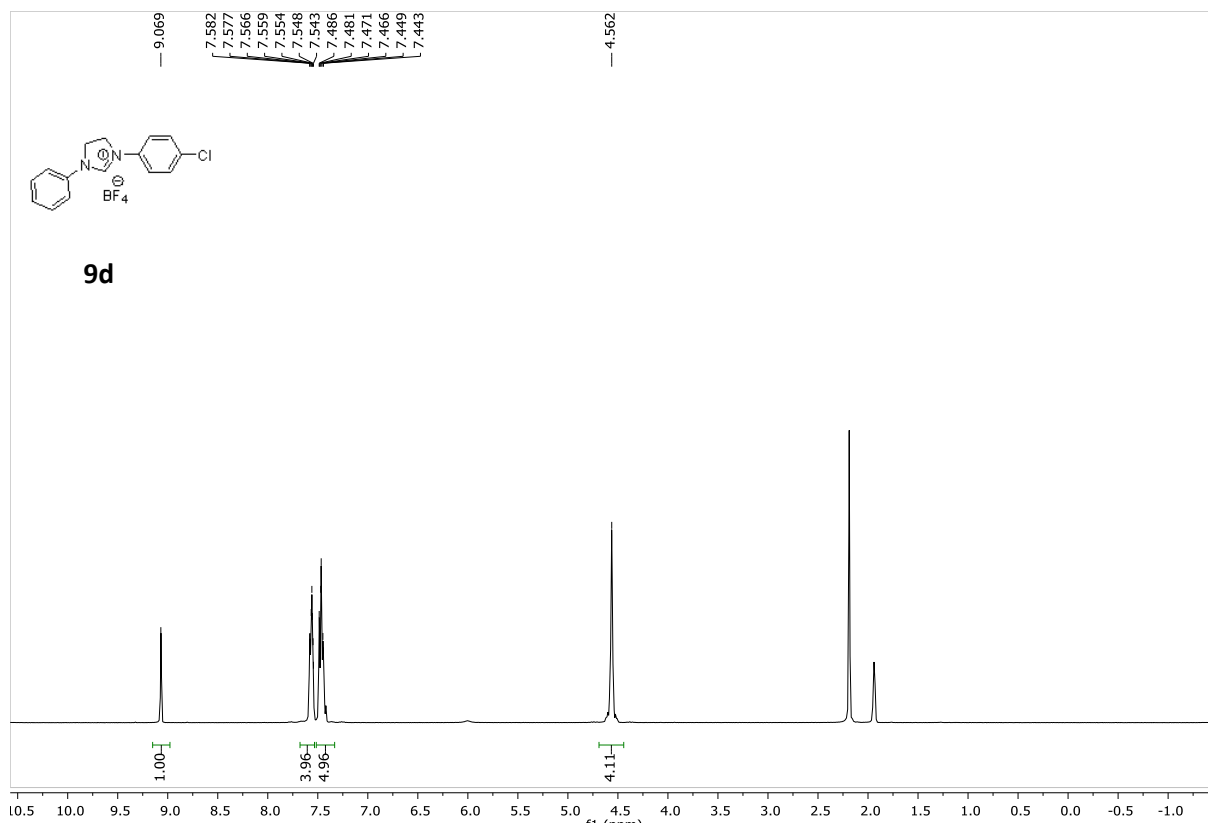


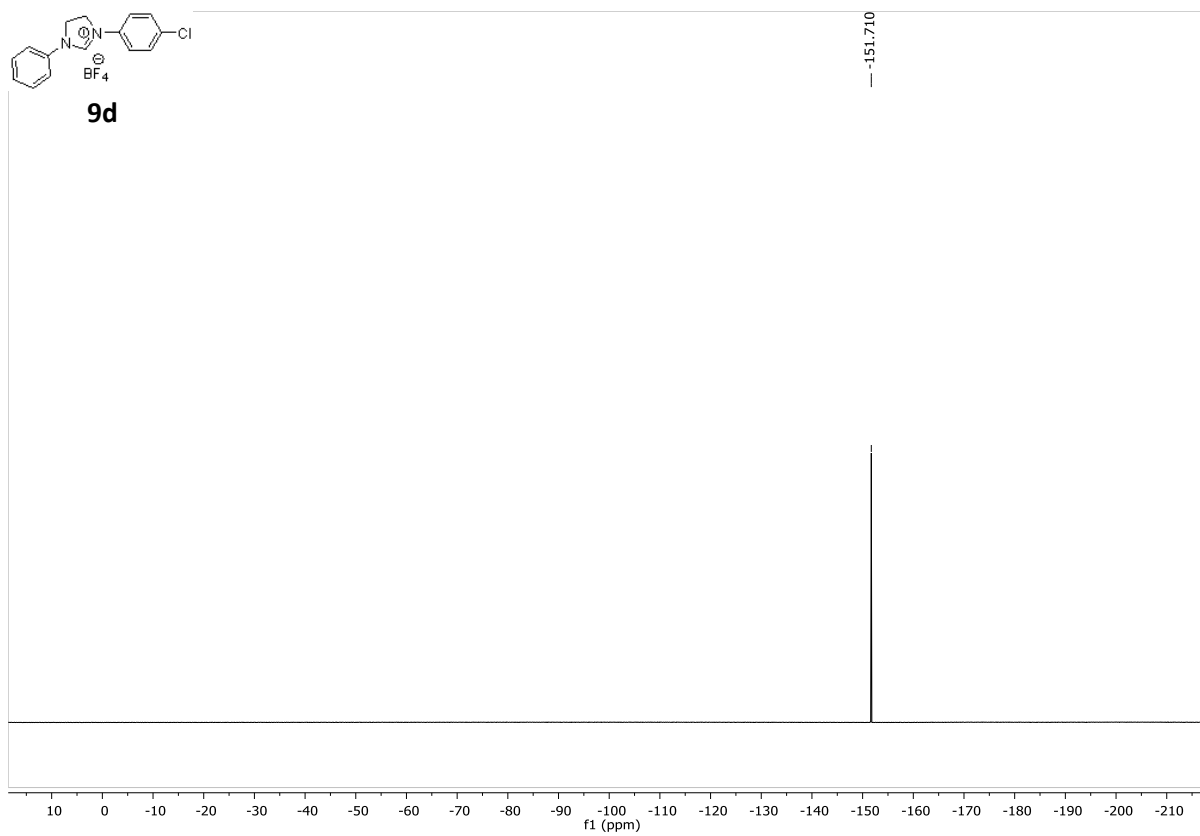


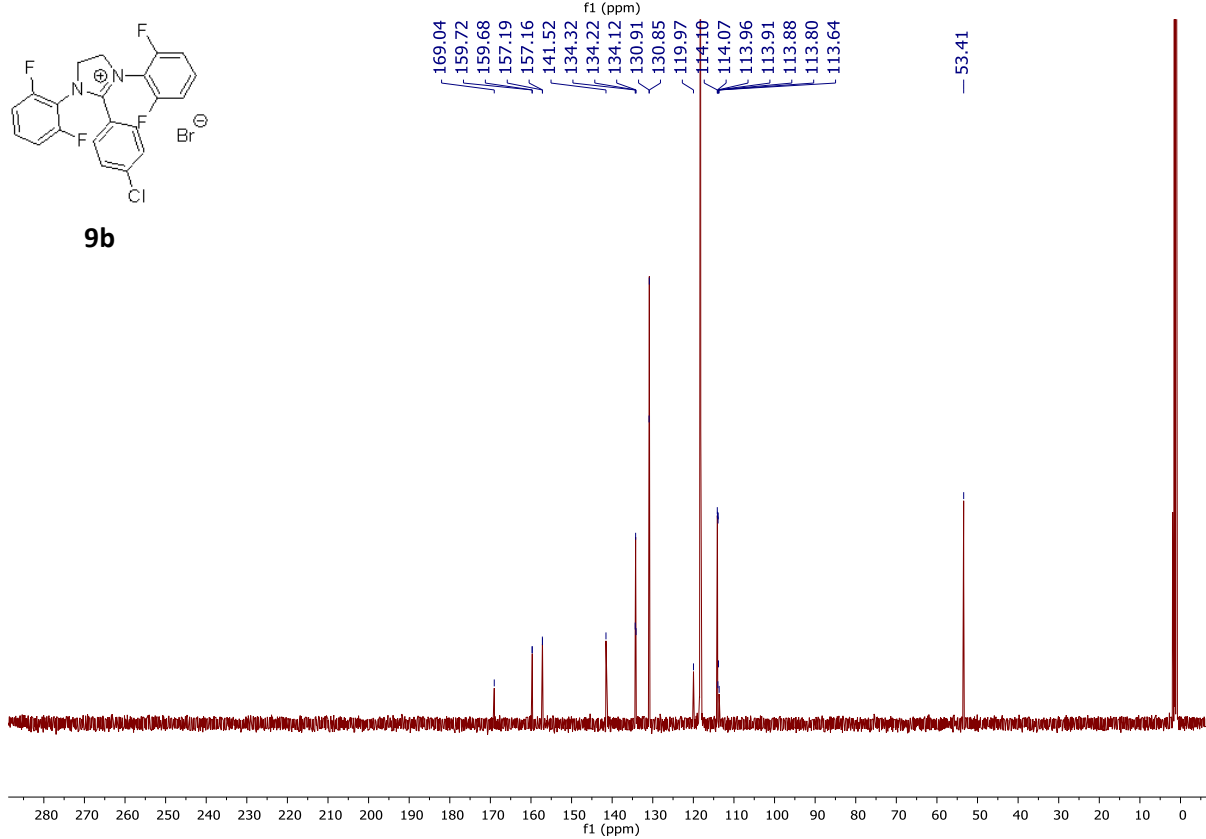
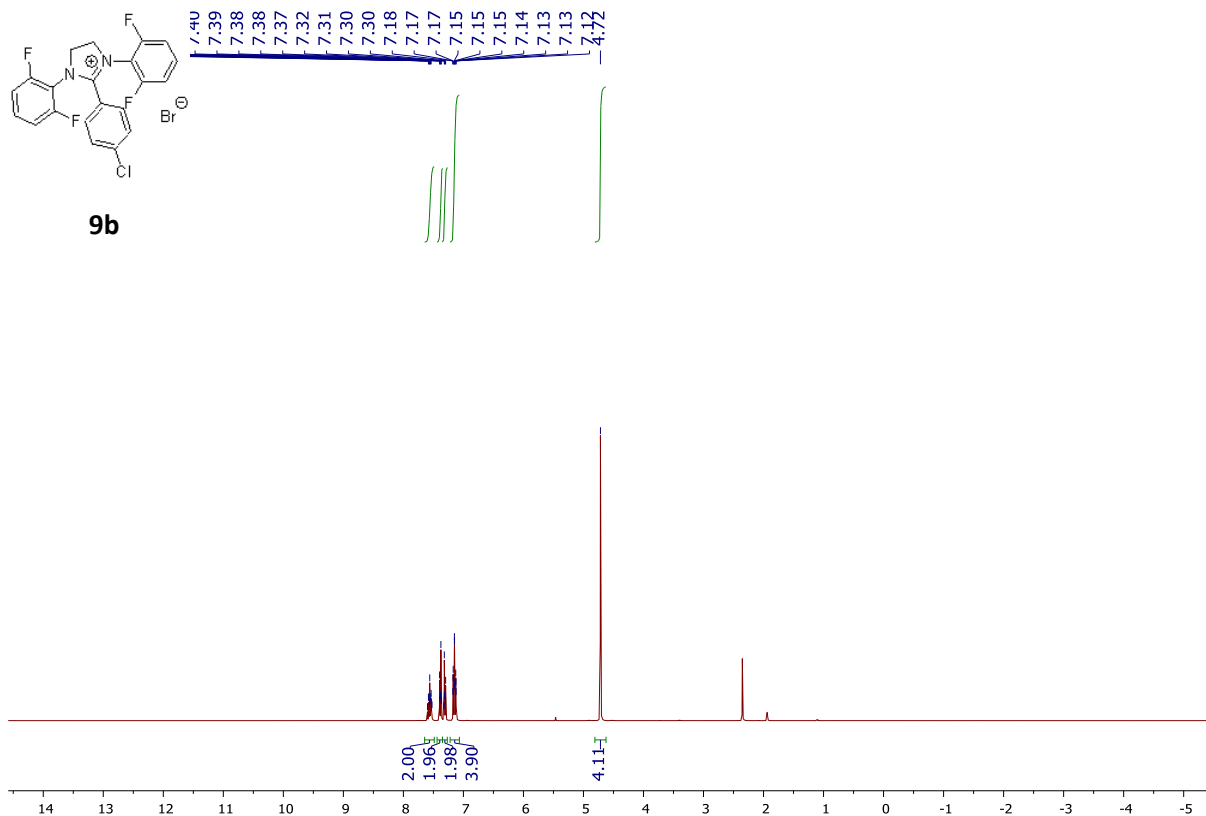
9e

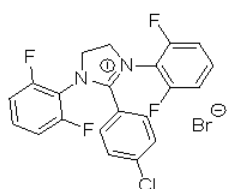
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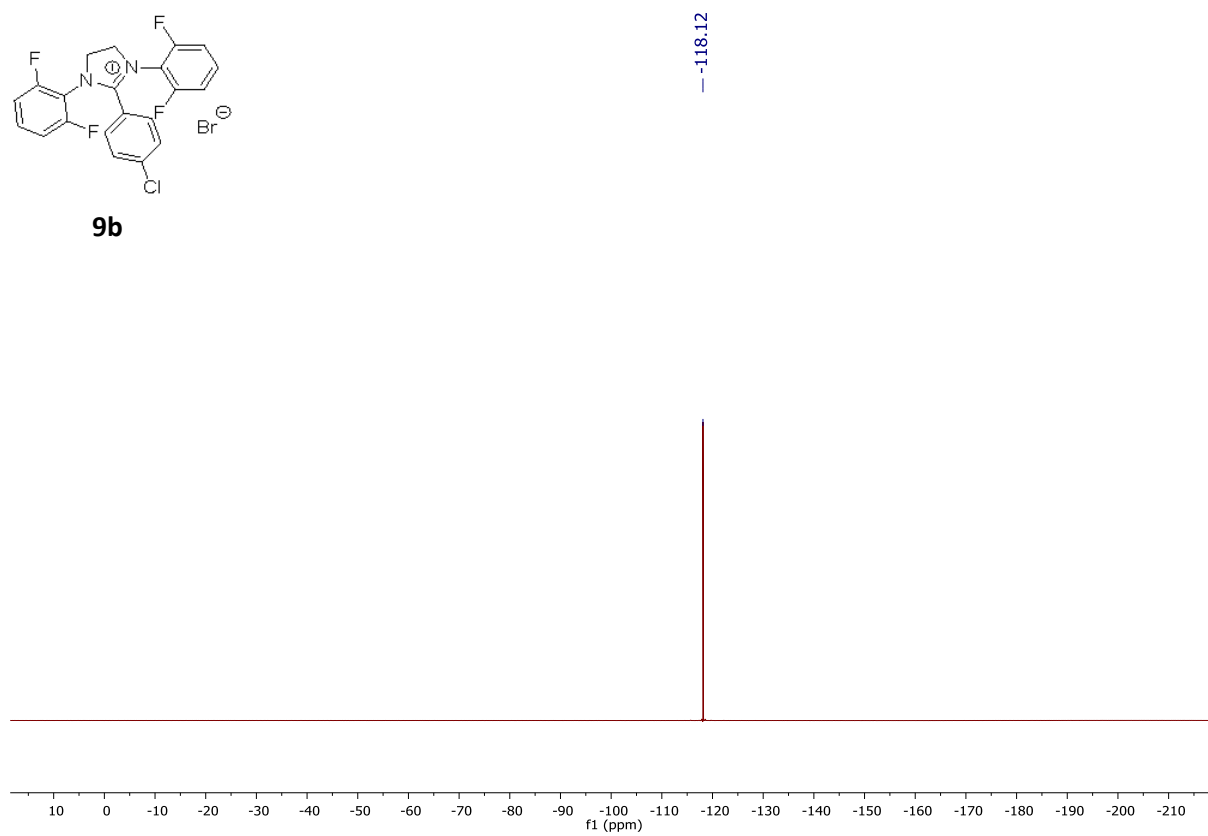


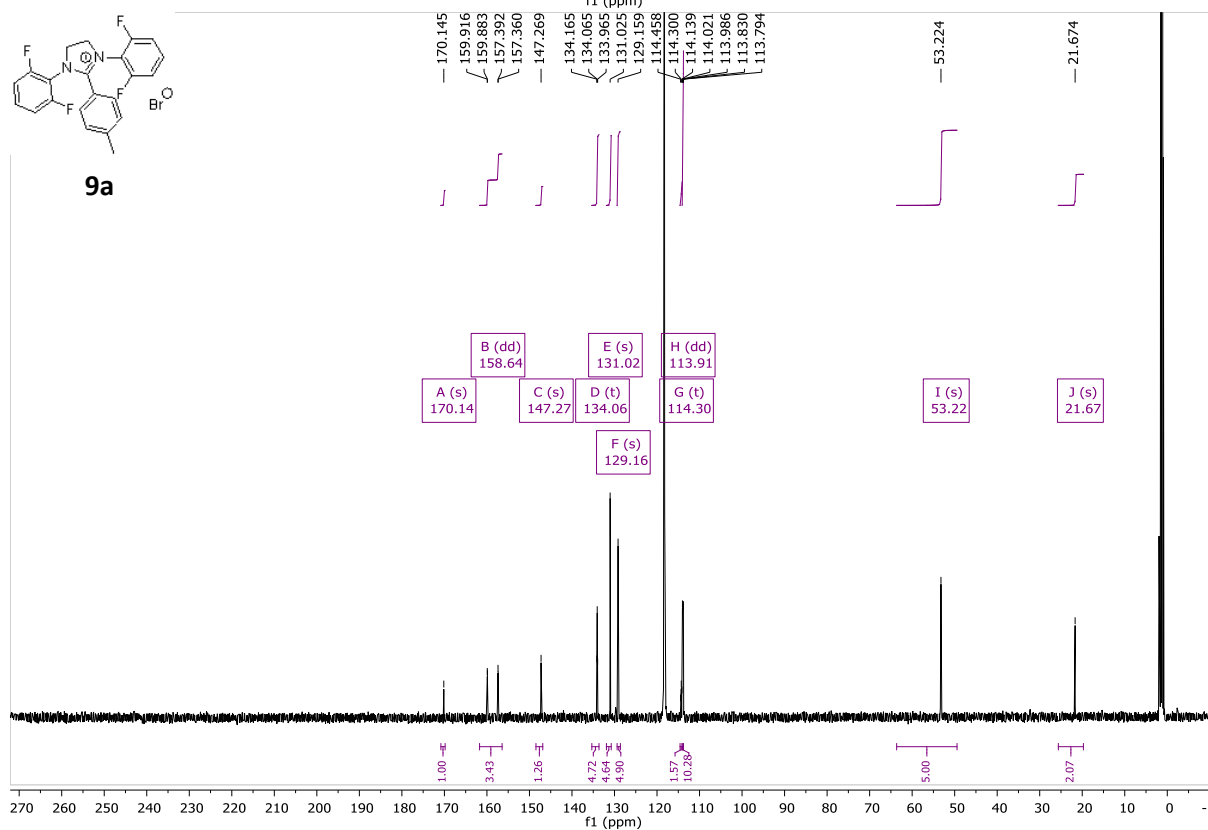
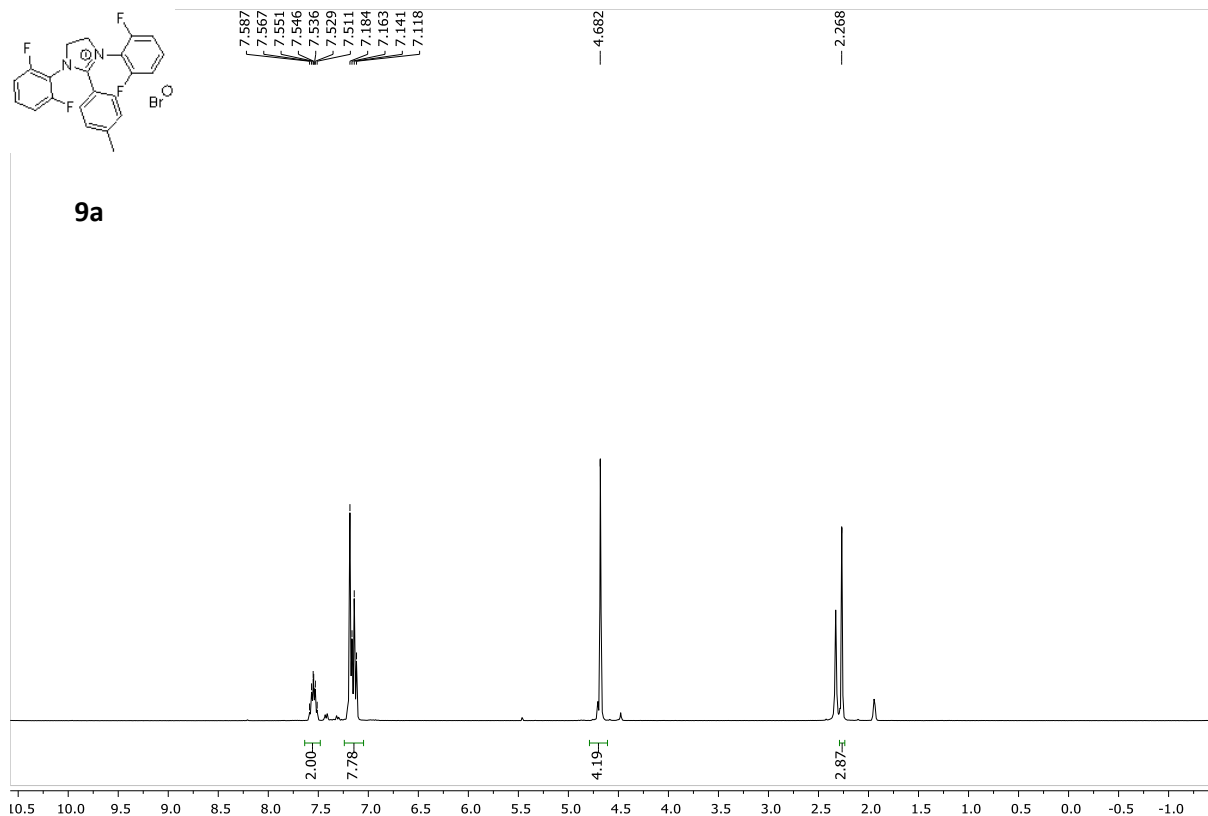


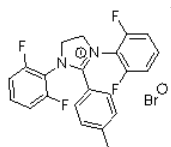




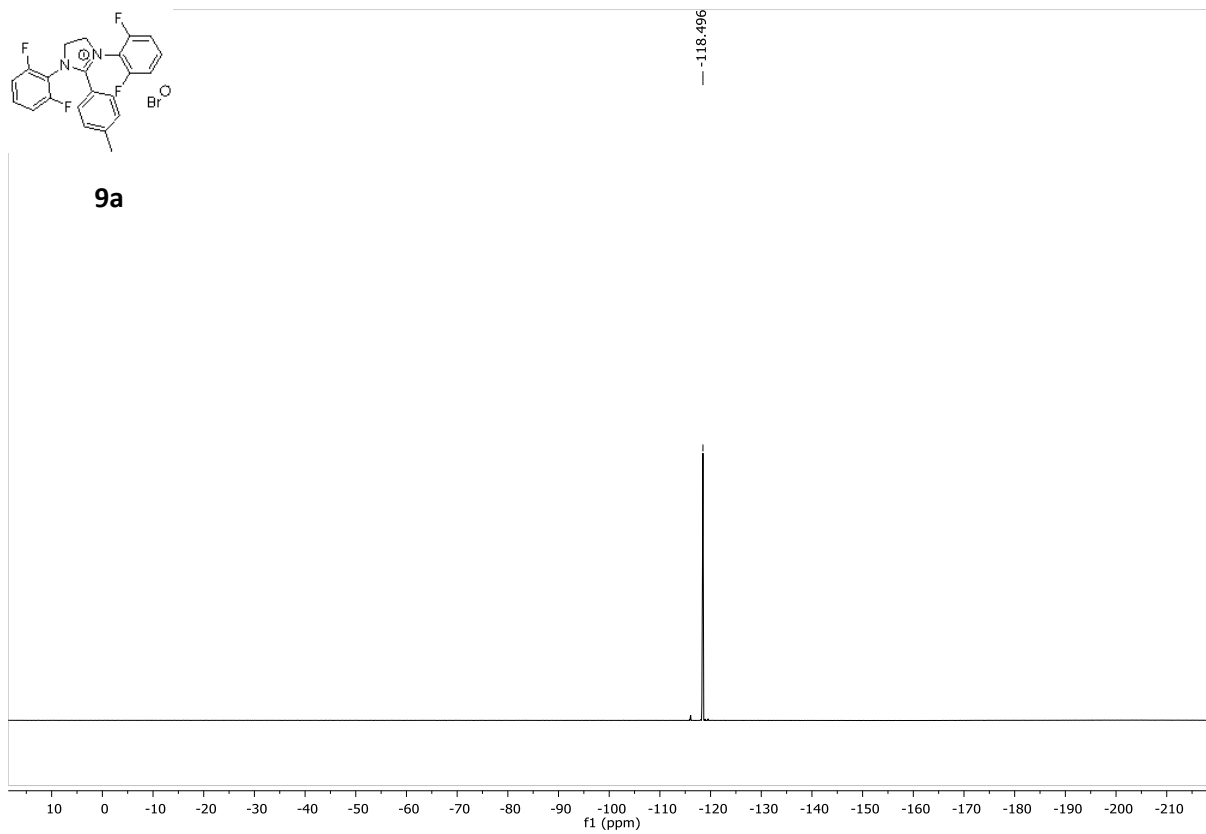
9b

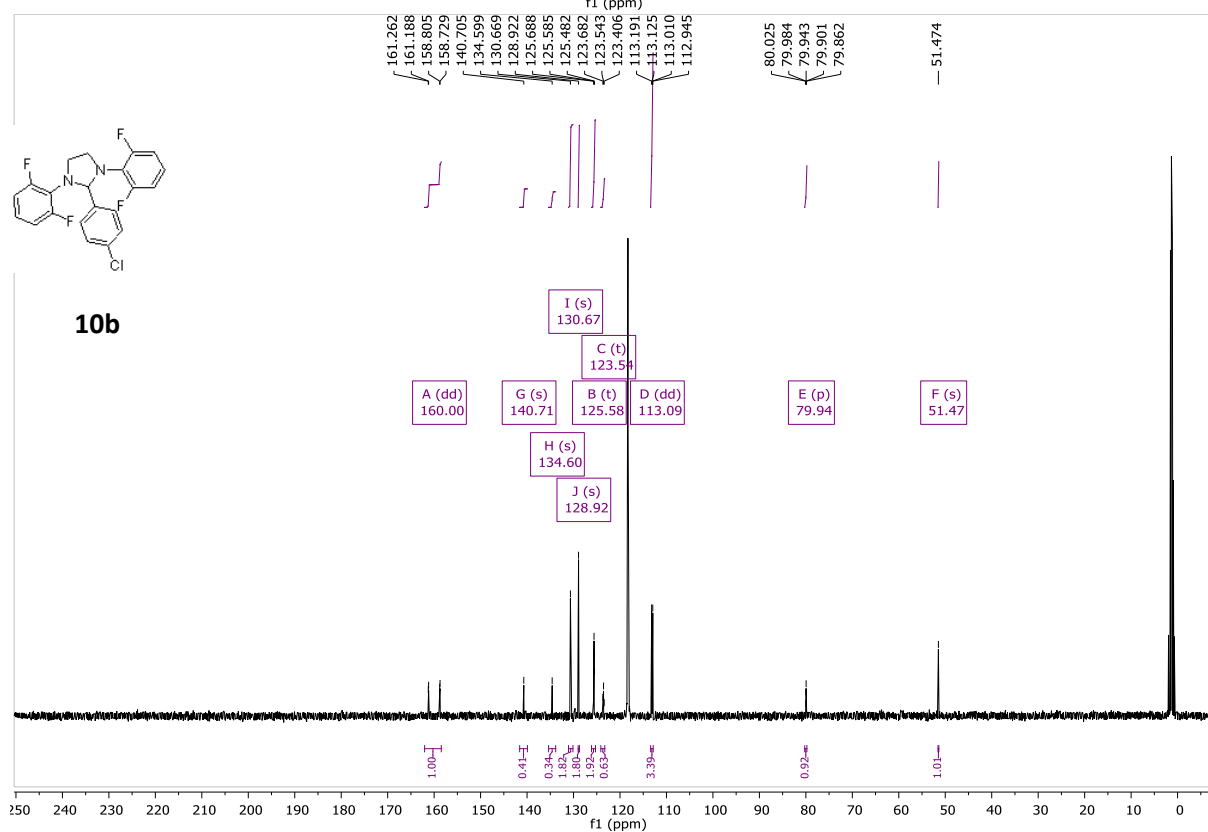
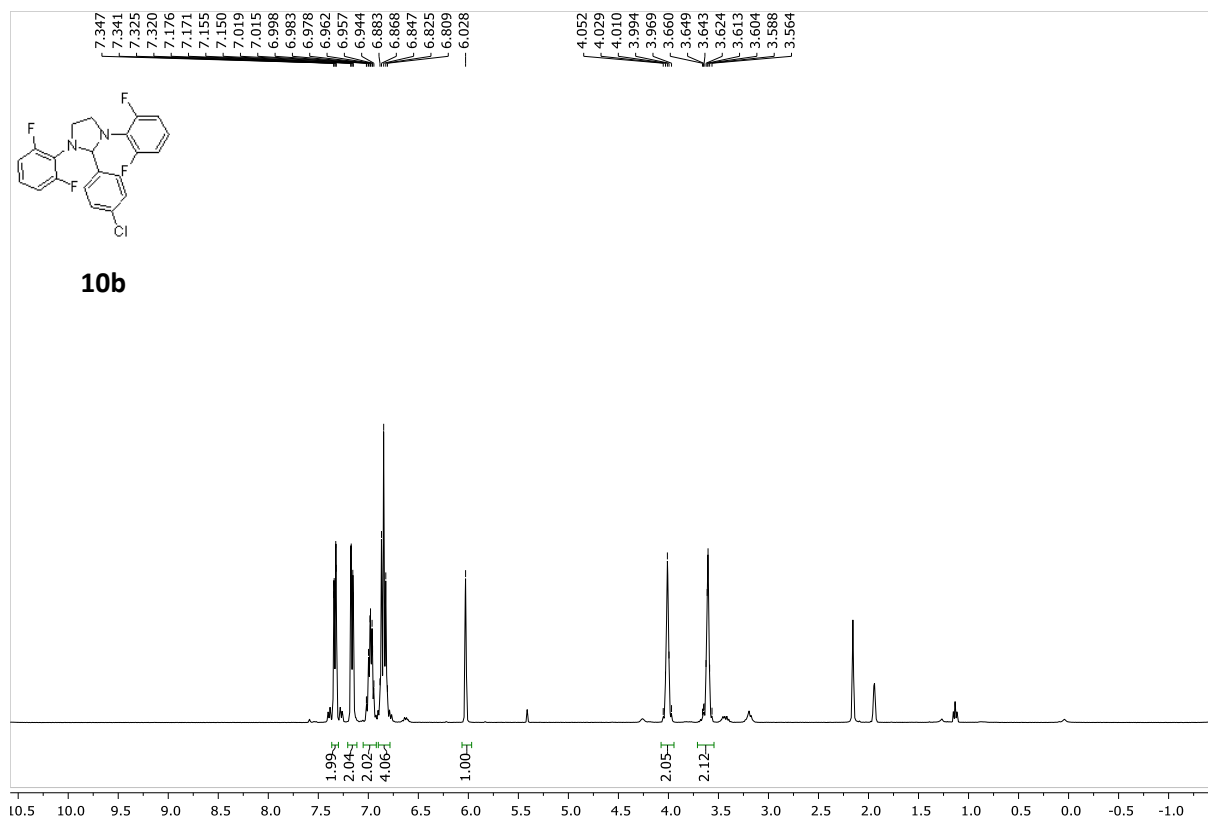


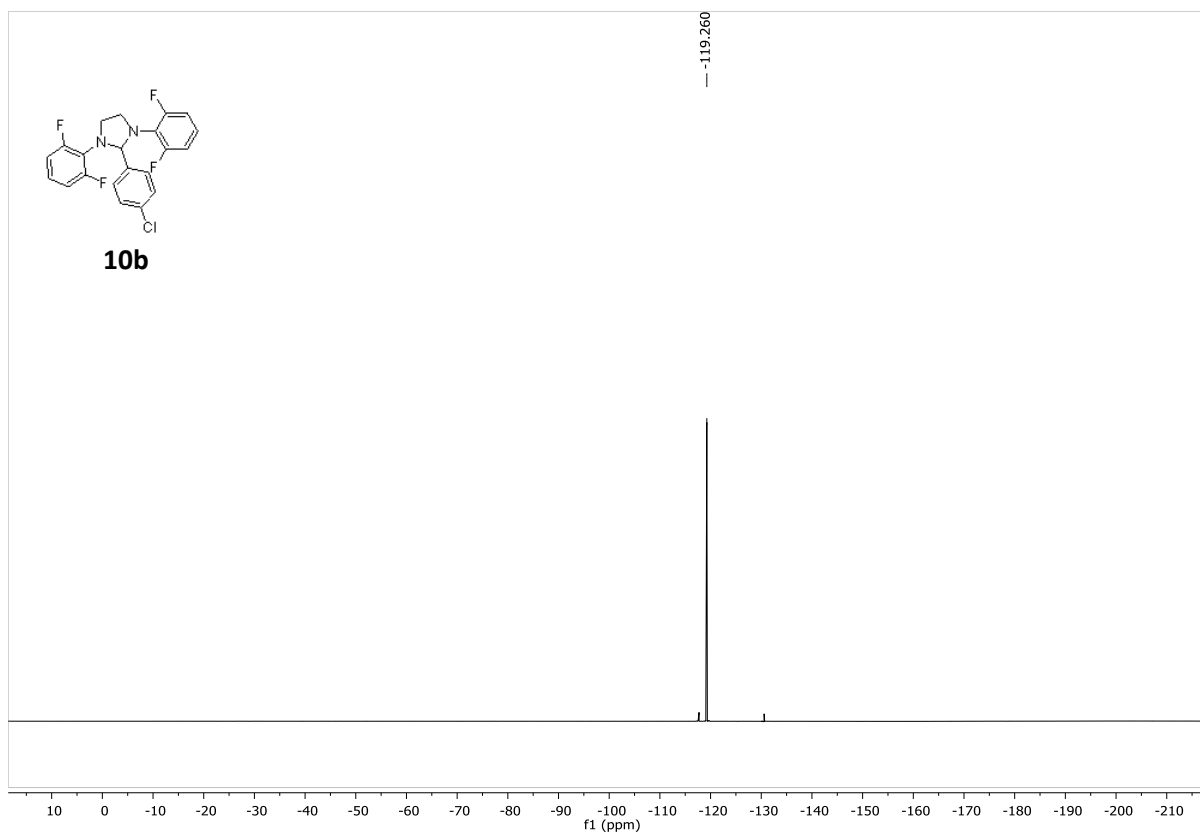


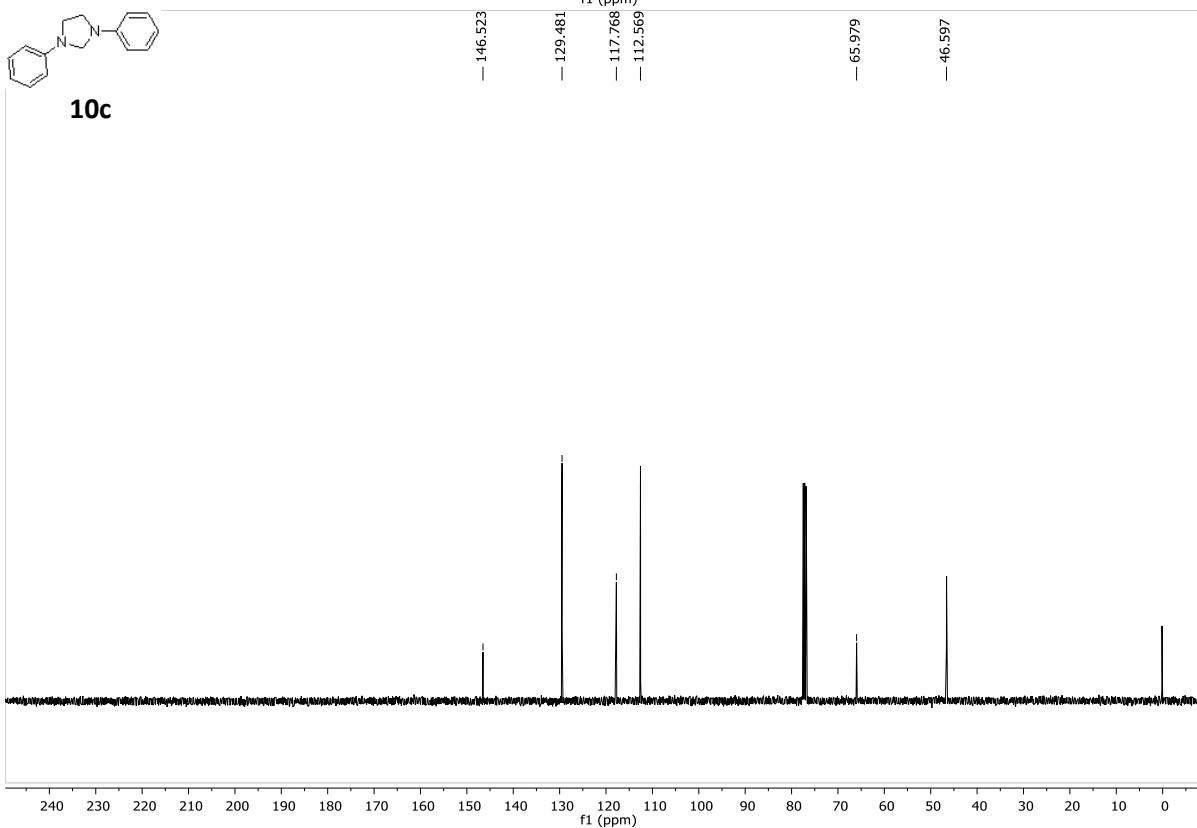
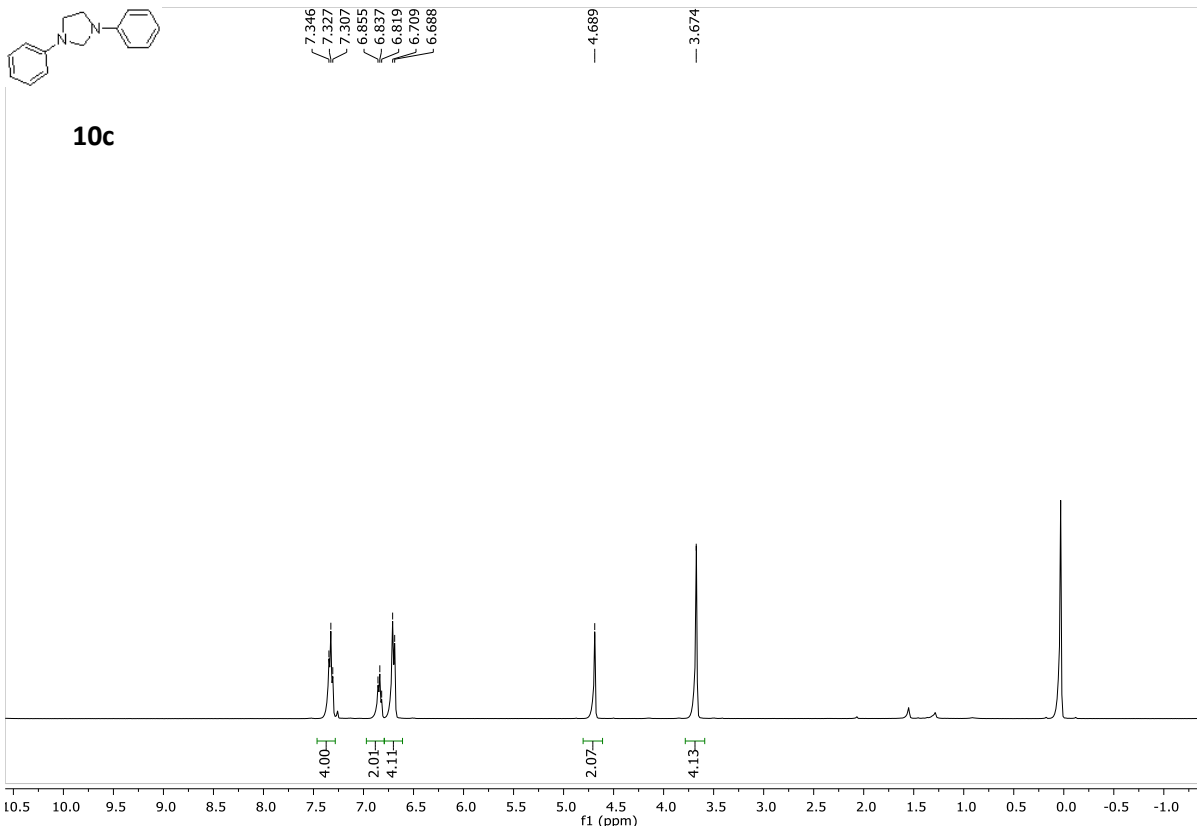


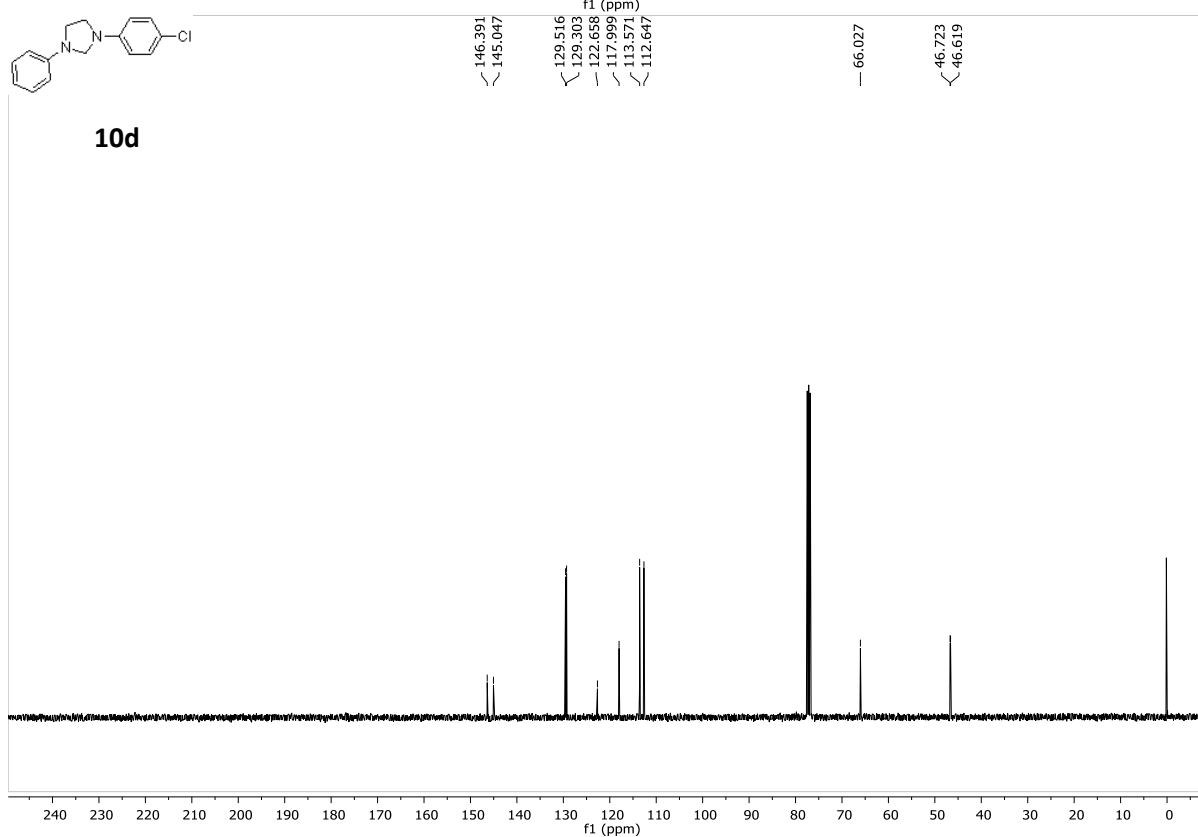
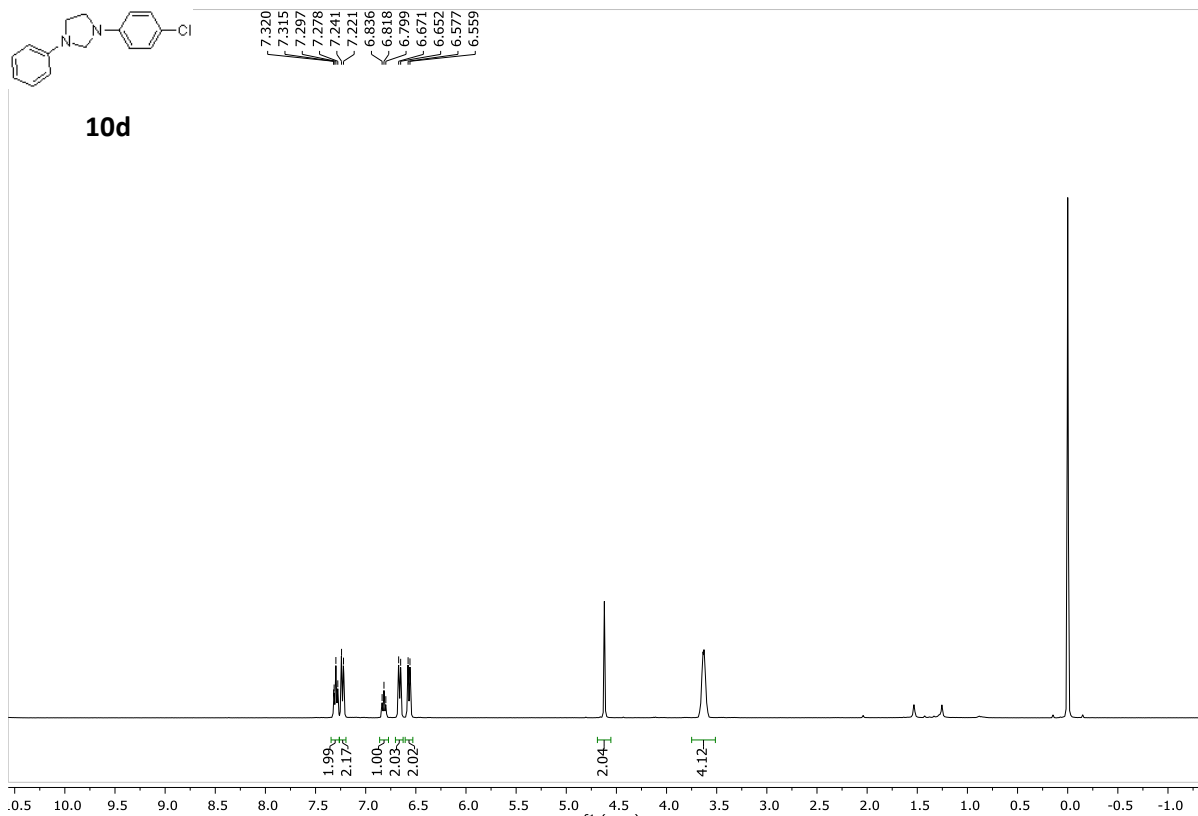
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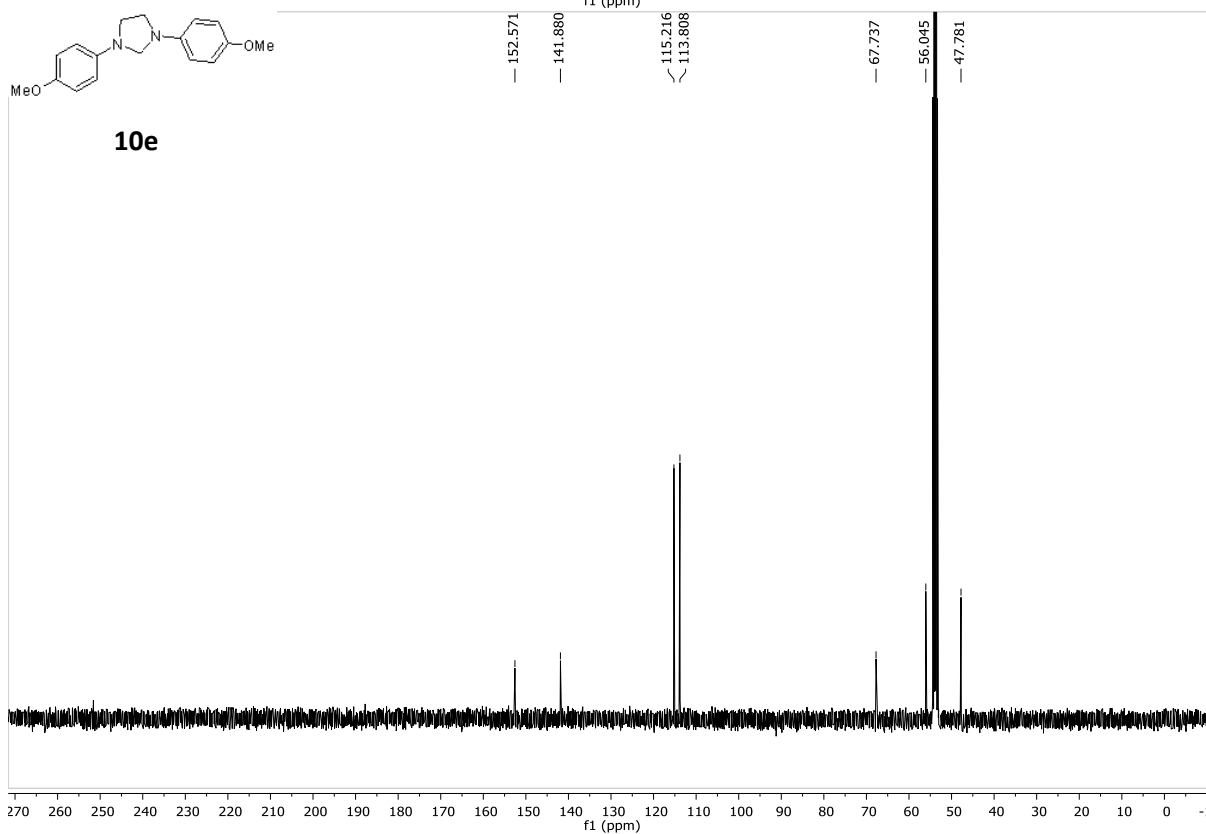
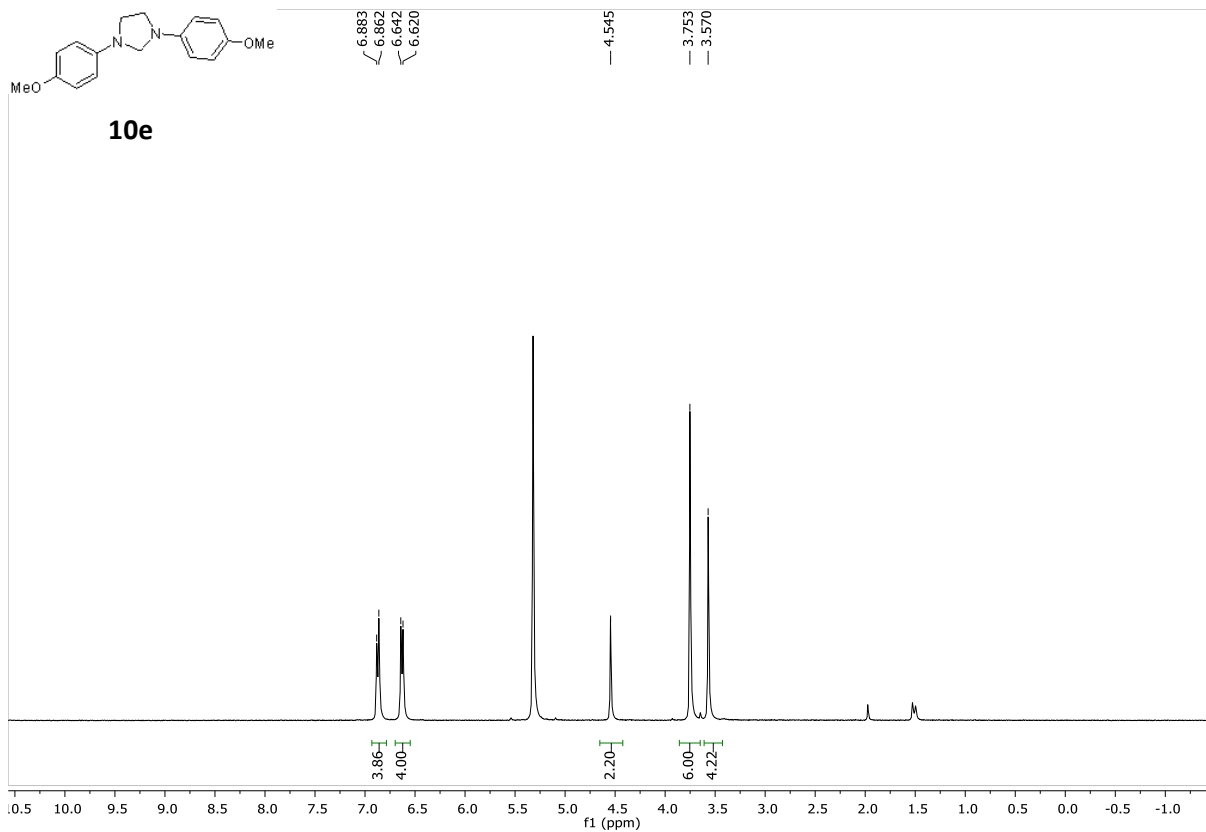


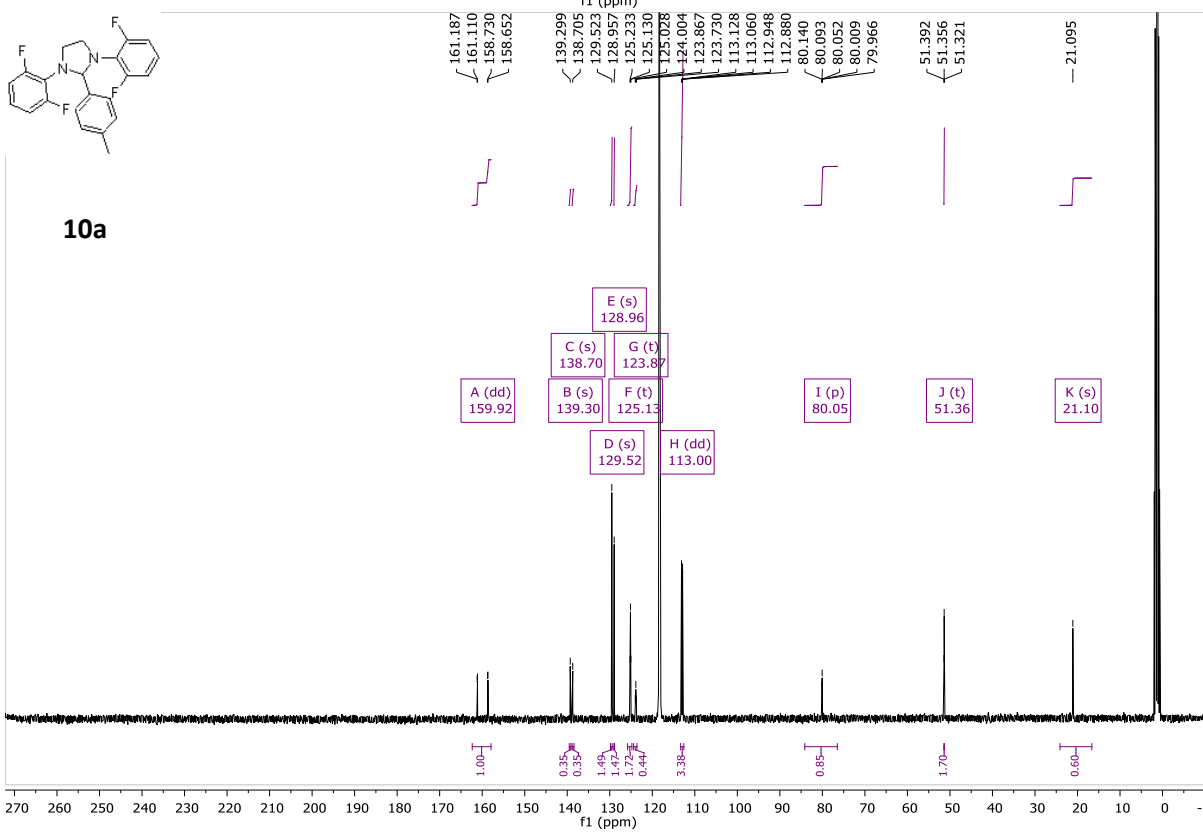
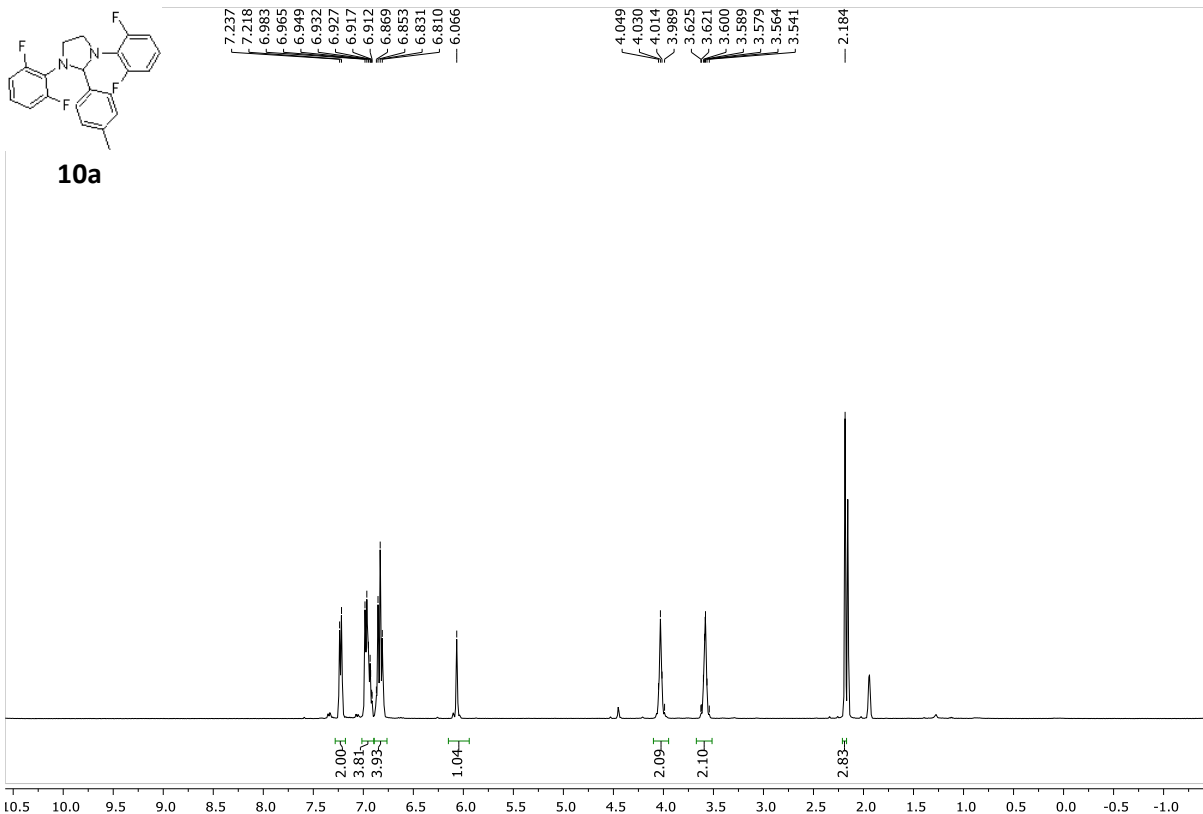


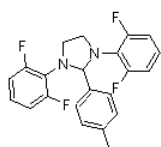




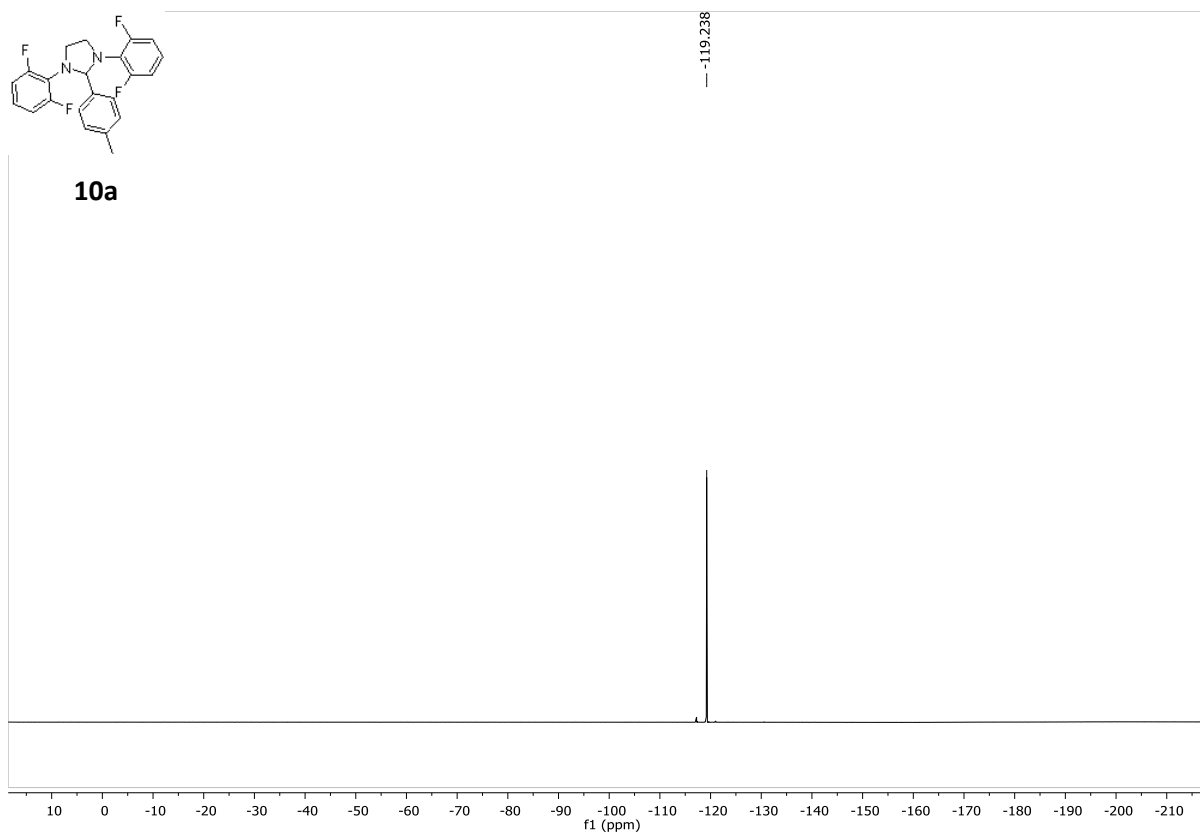


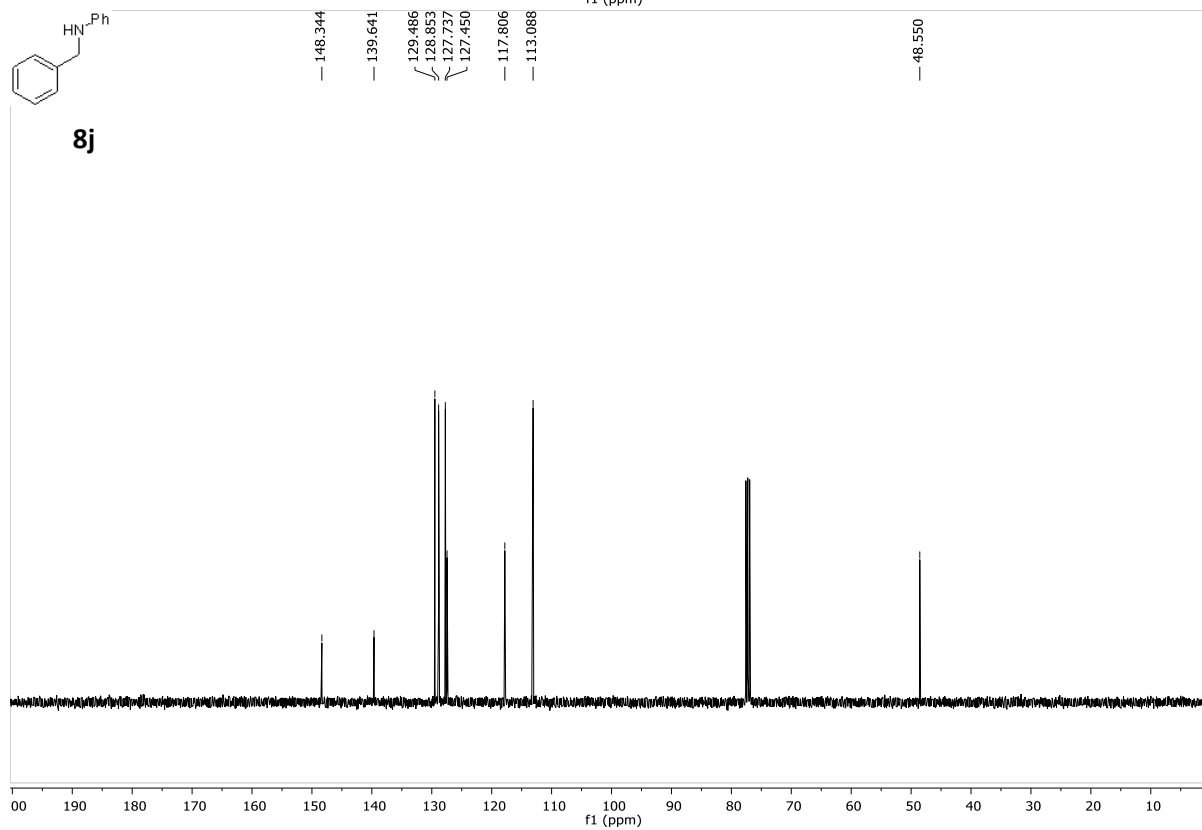
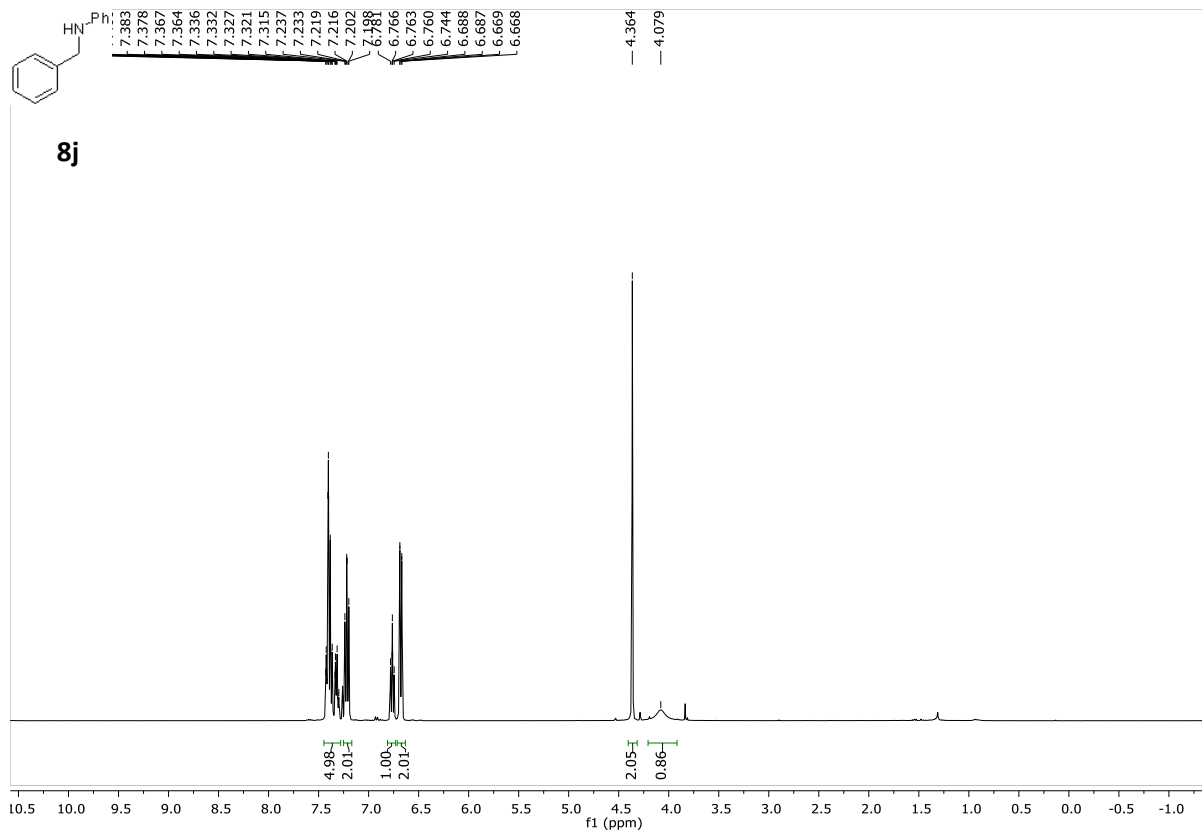


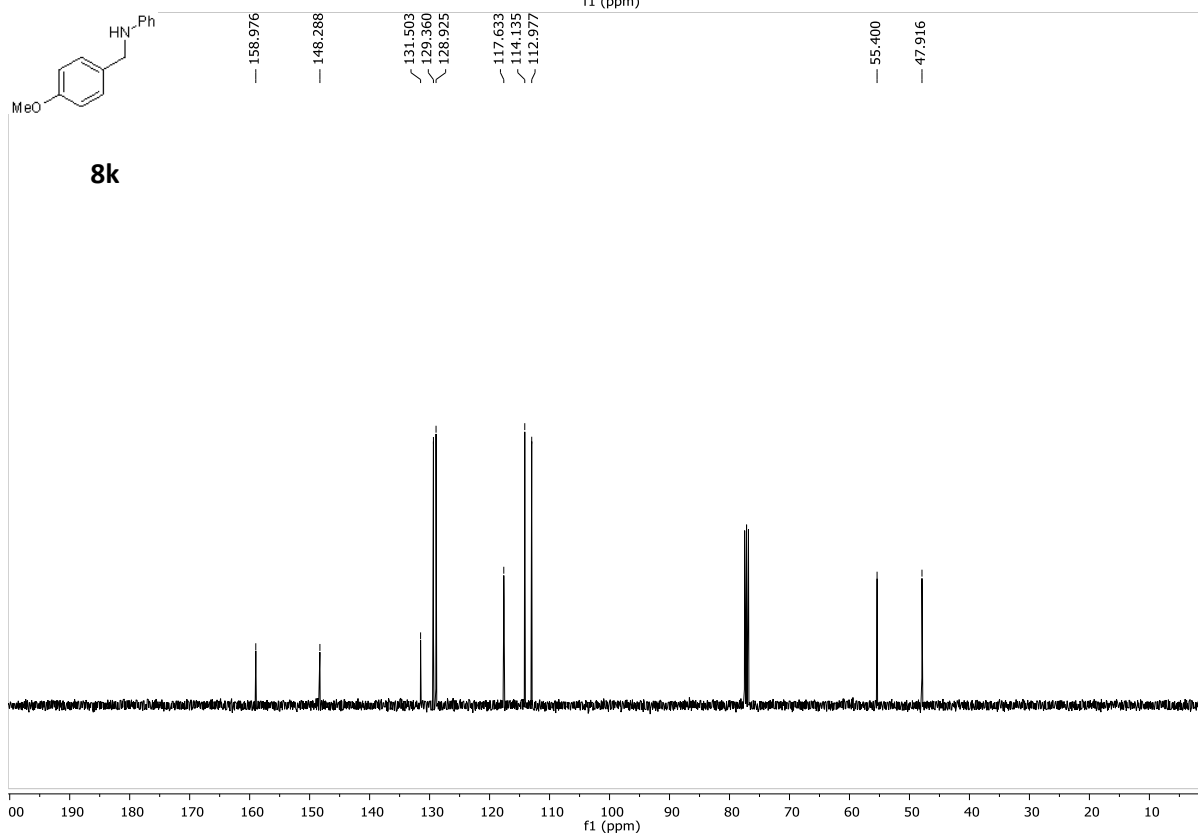
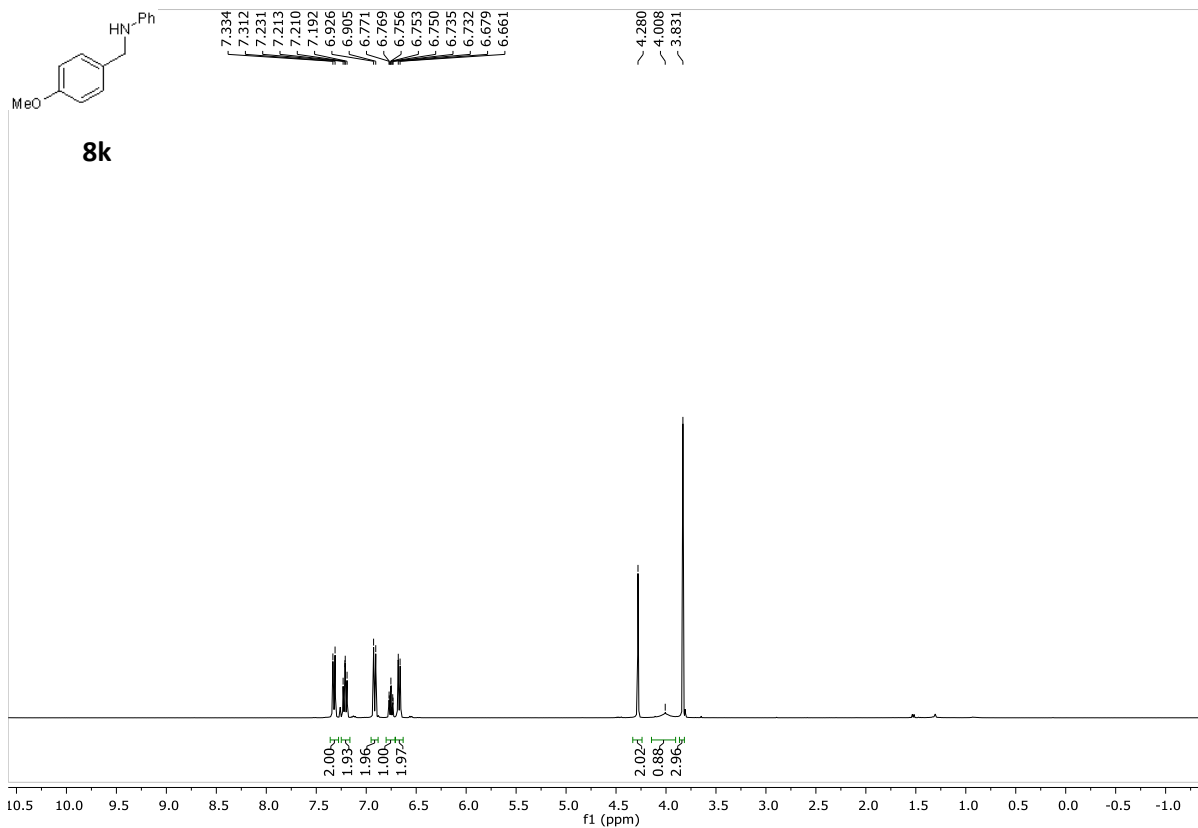


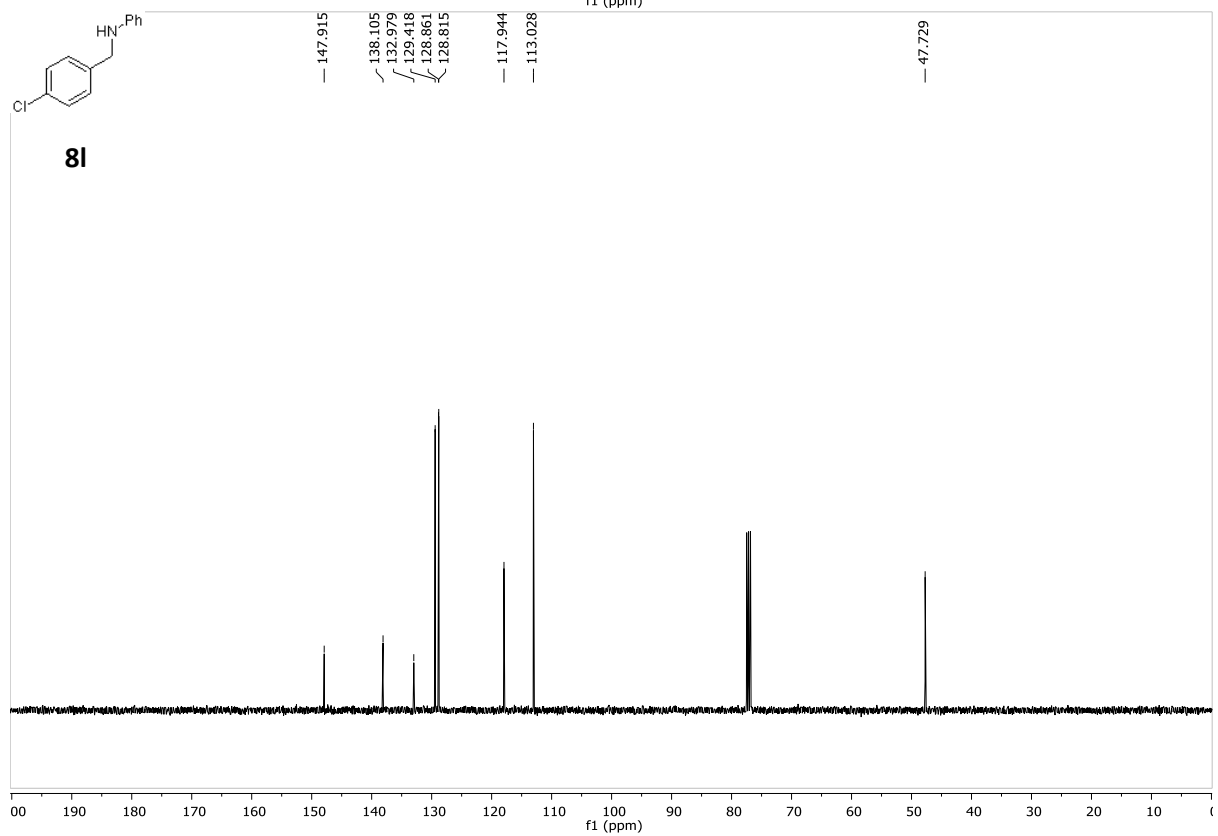
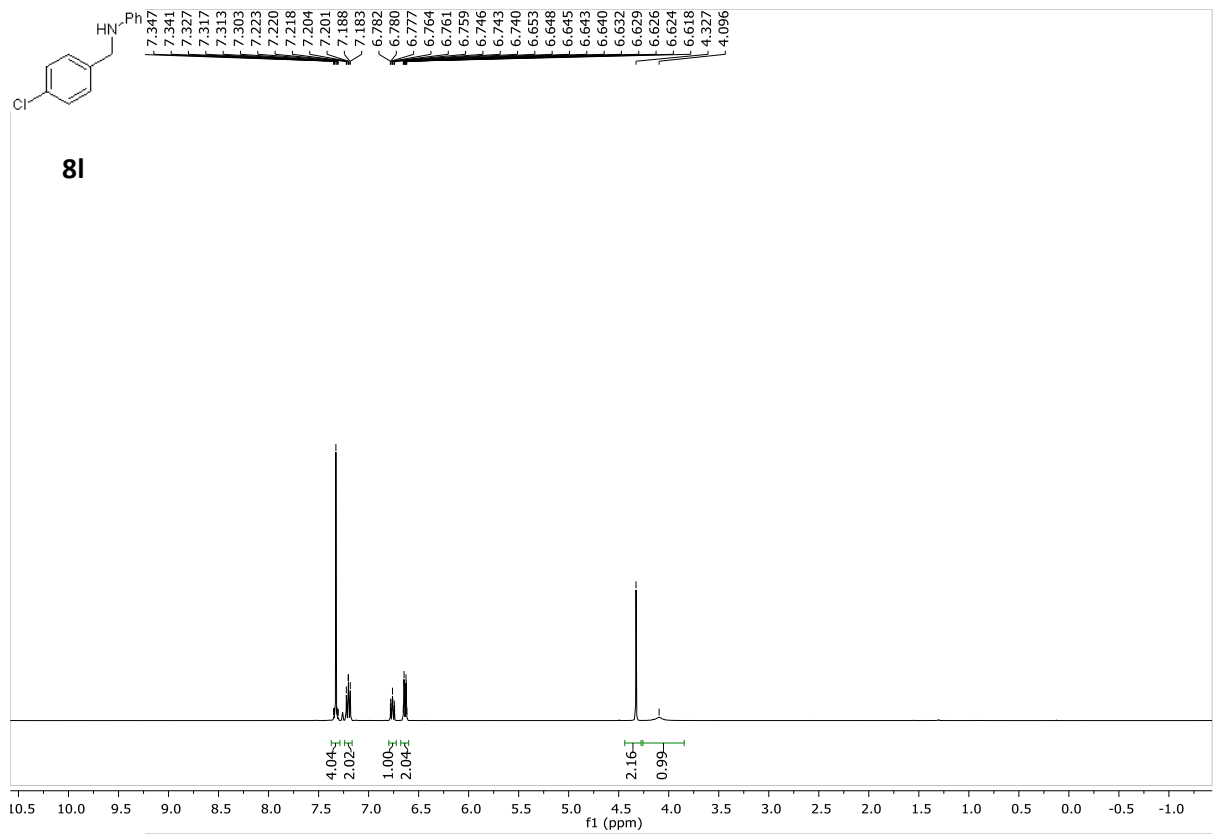


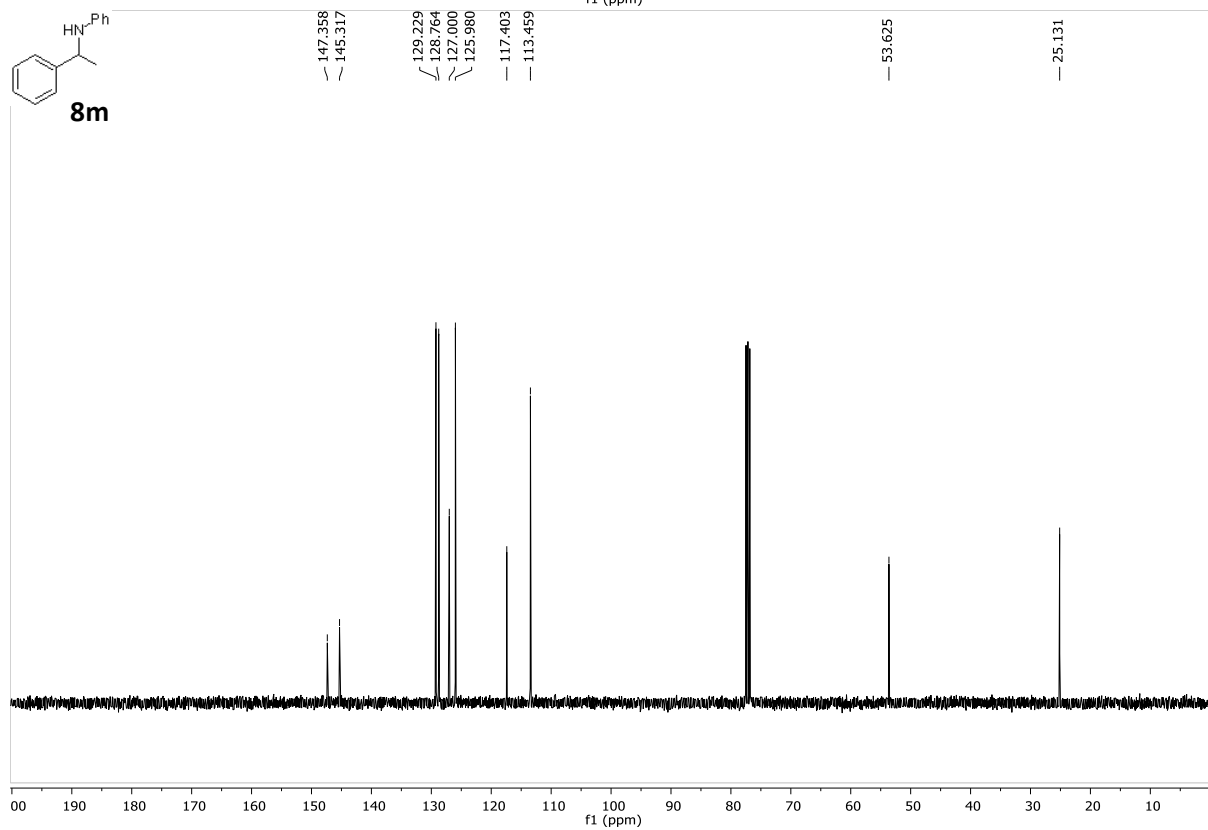
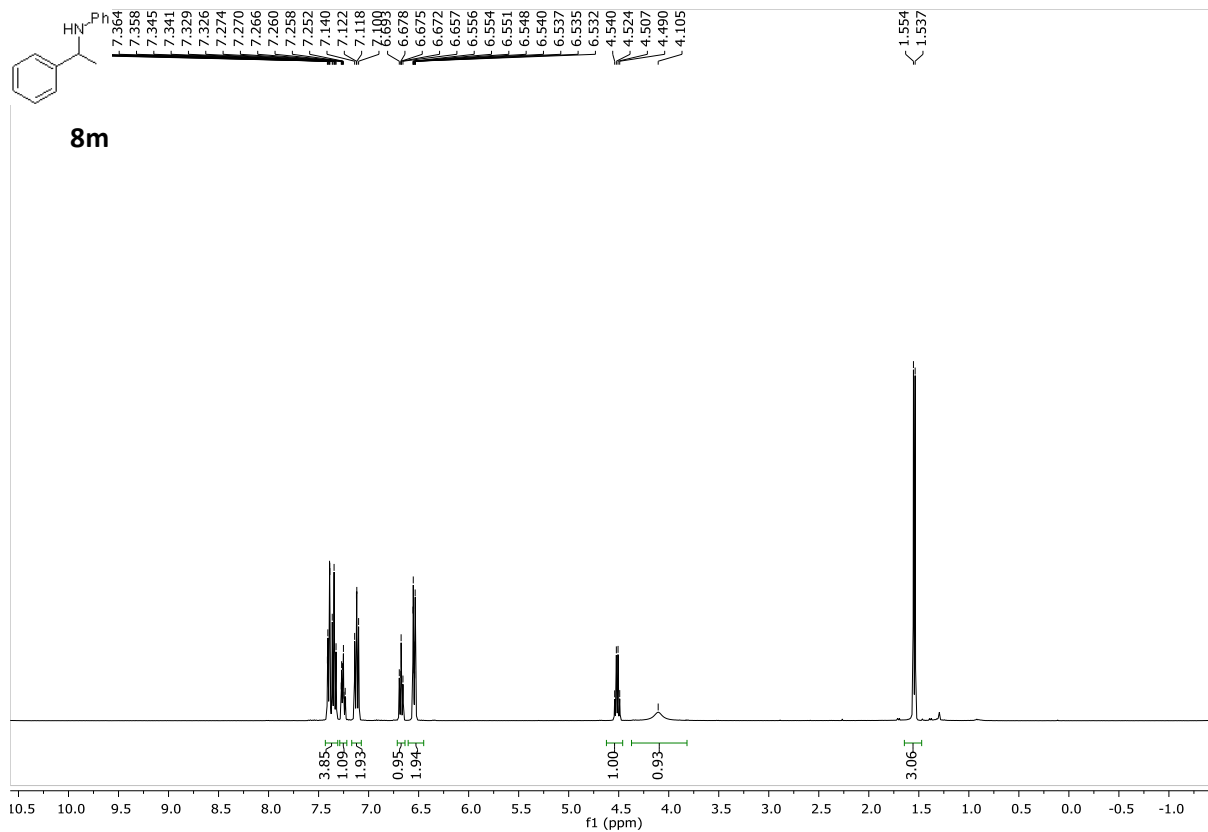
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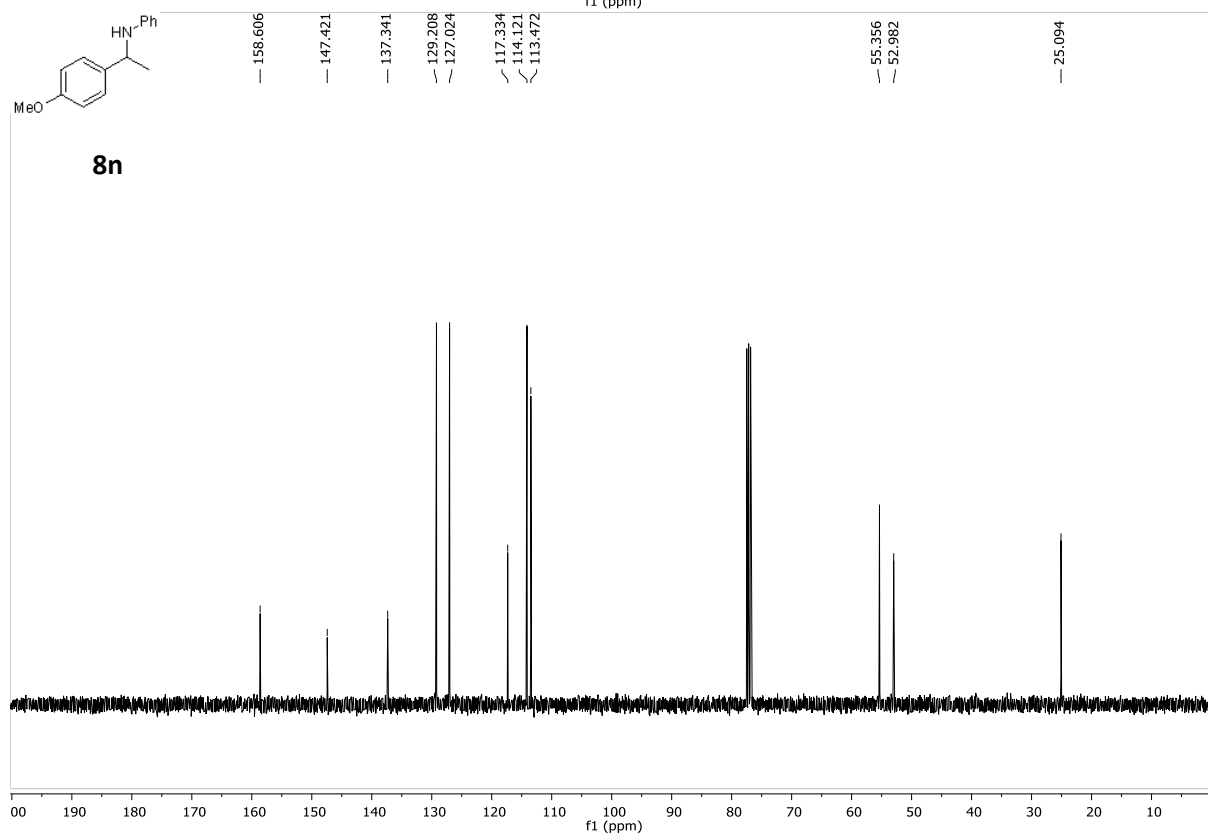
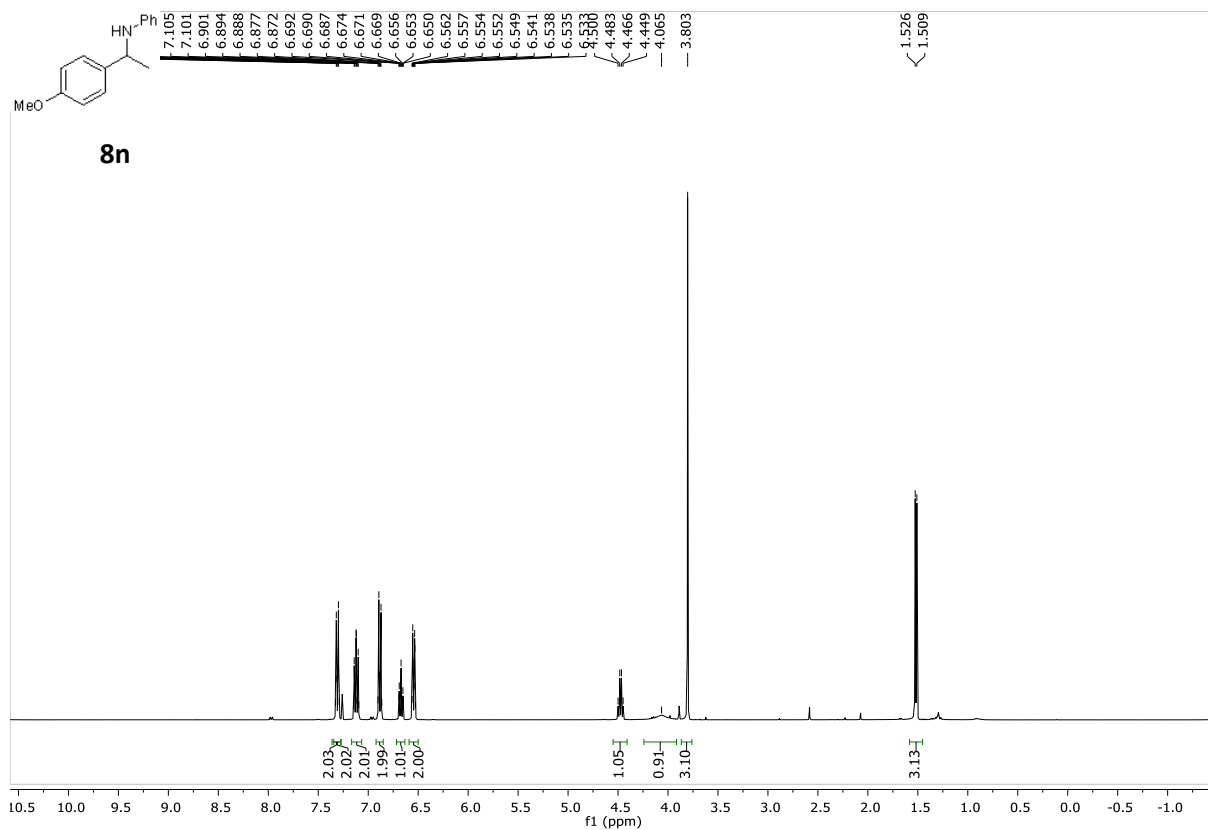


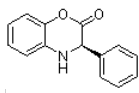




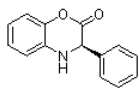
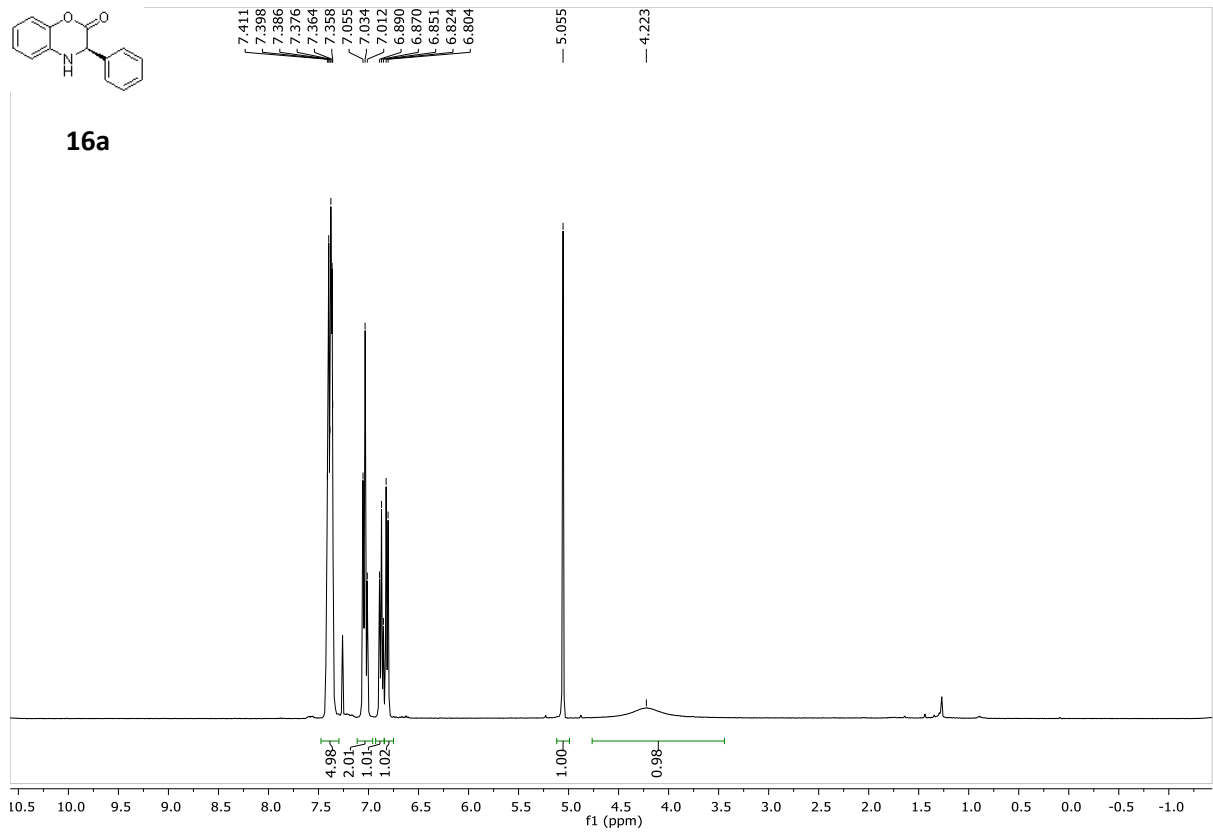




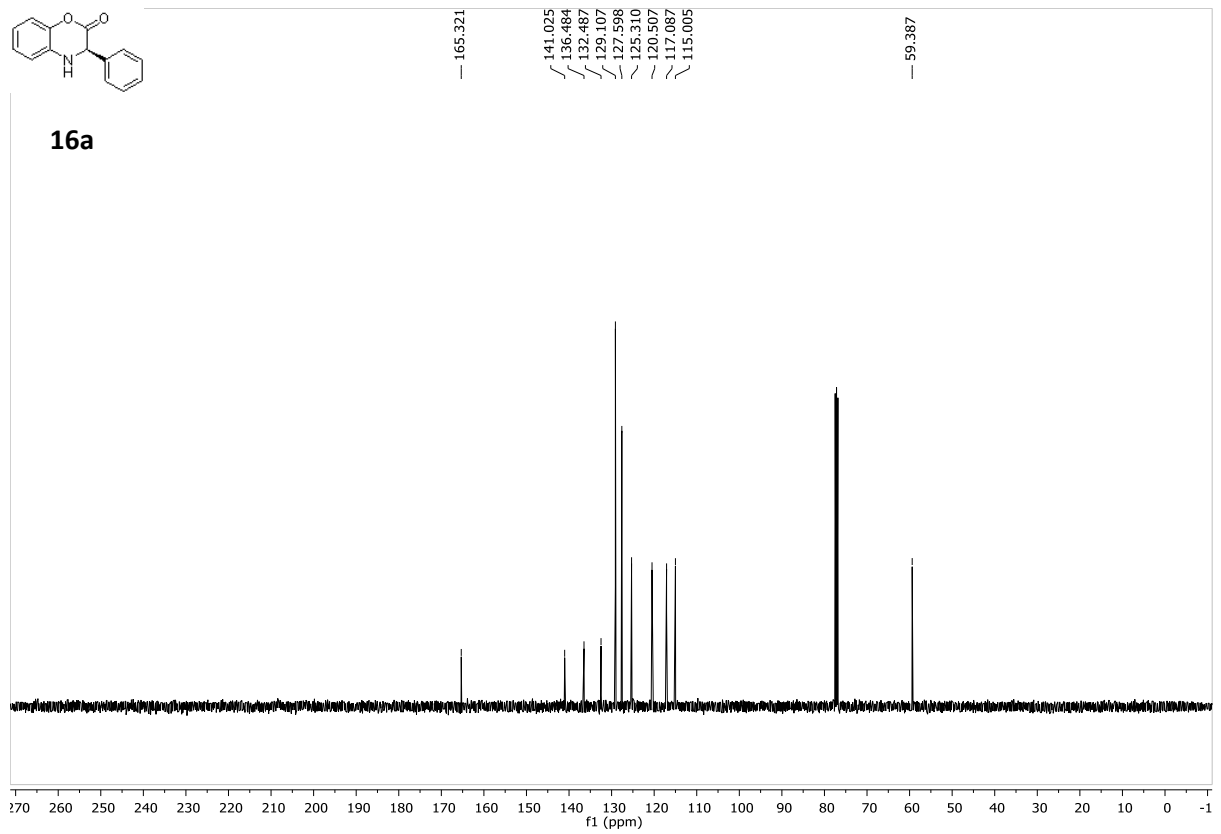


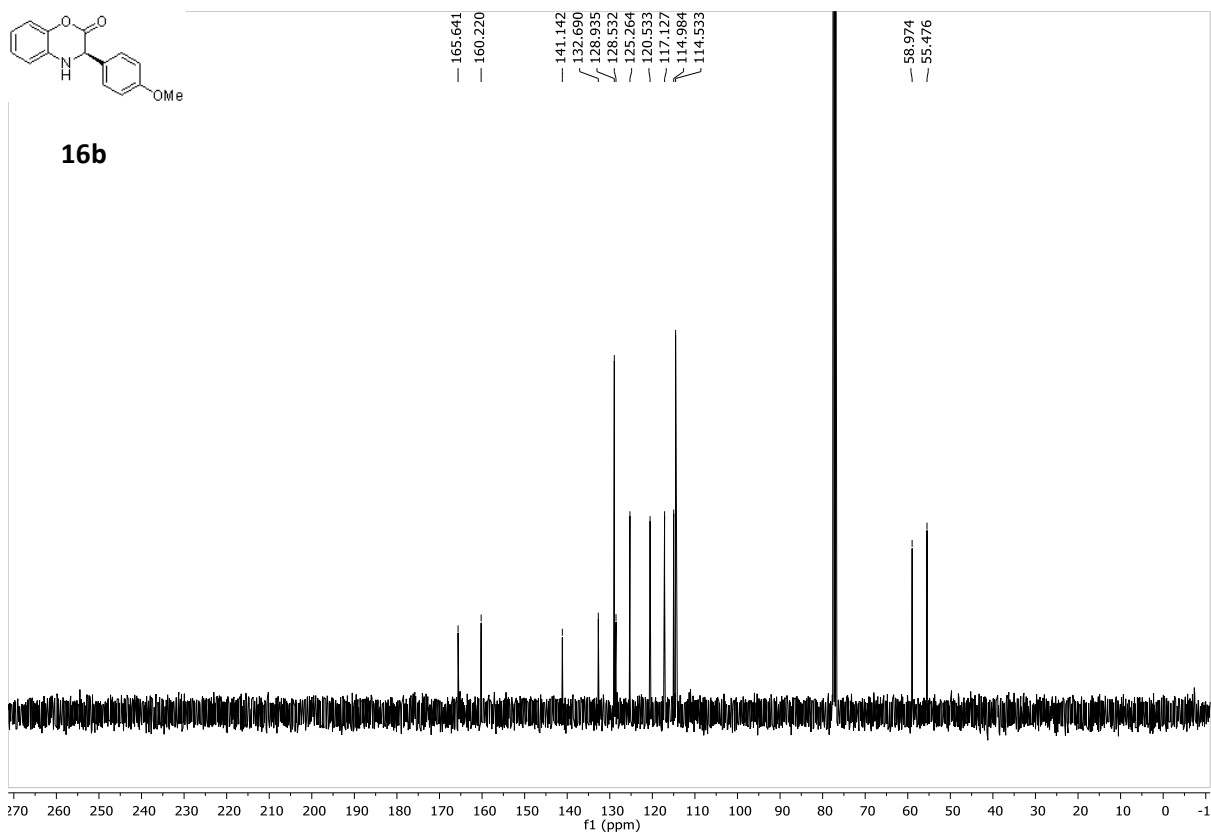
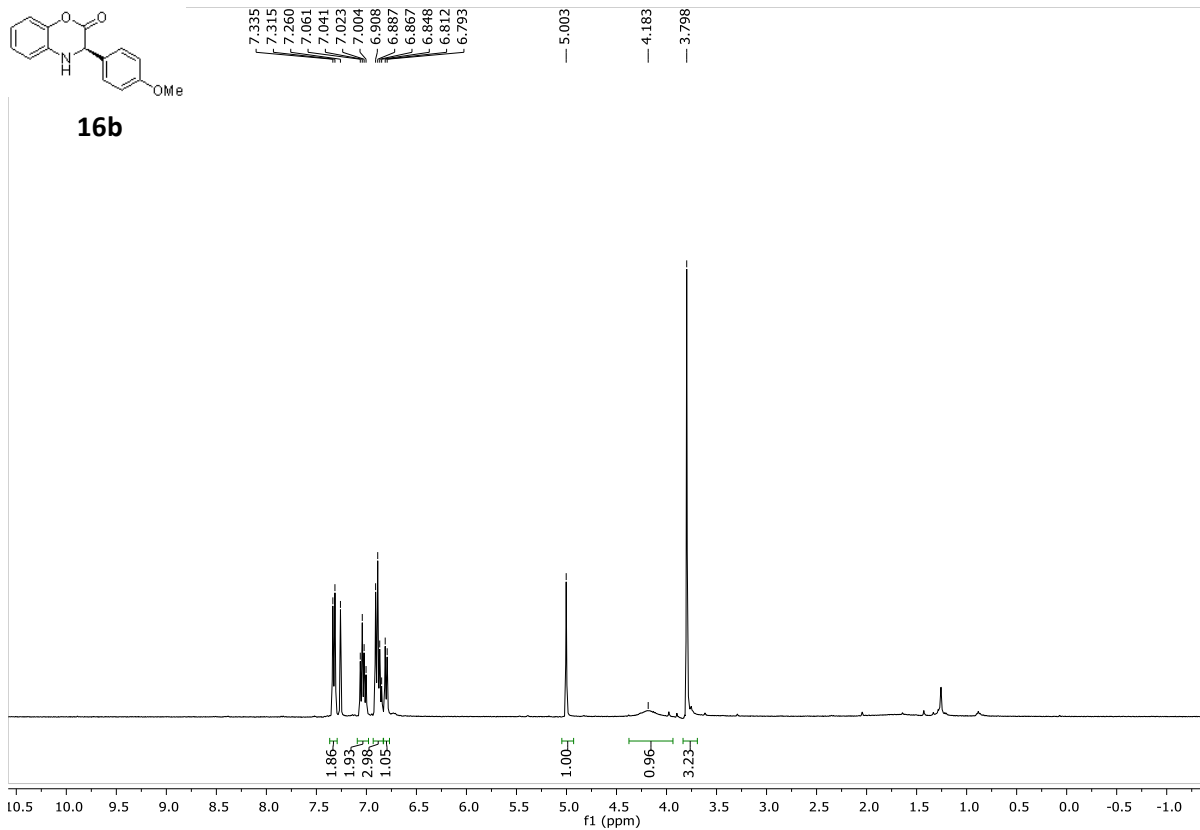


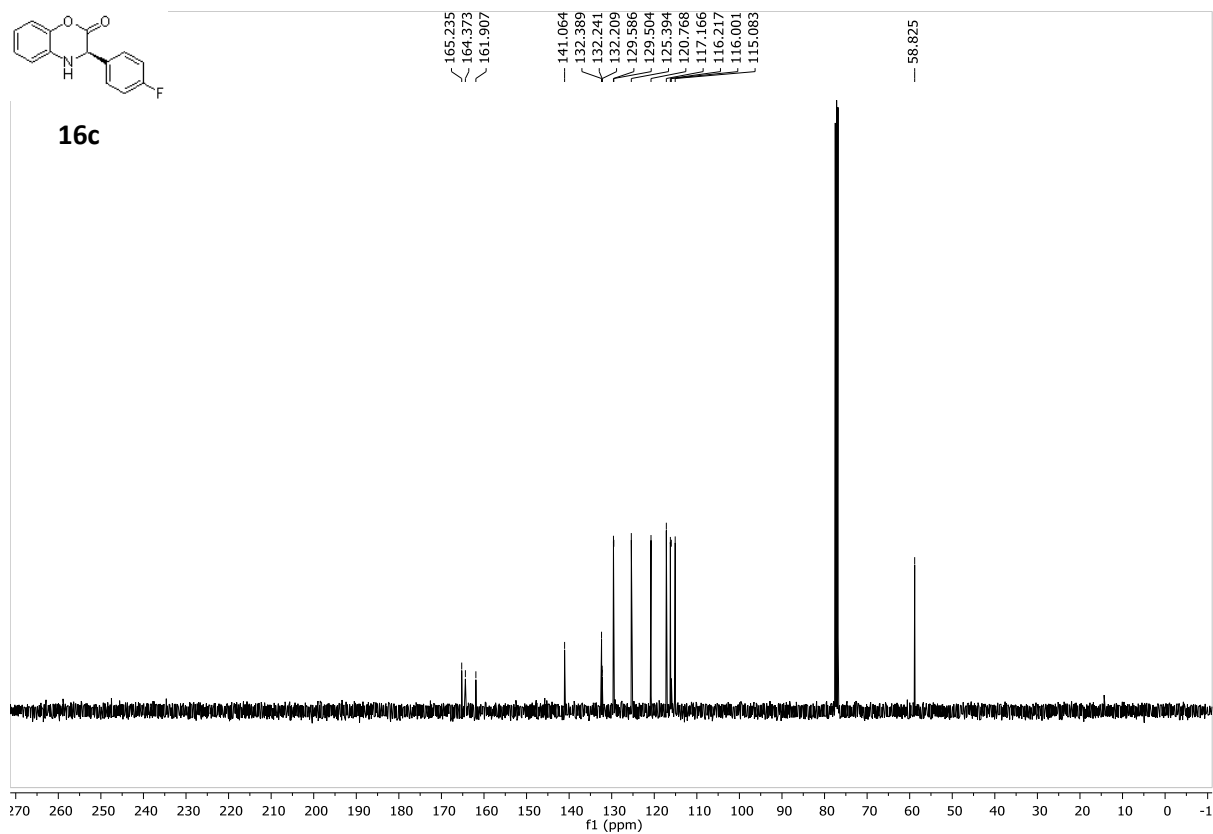
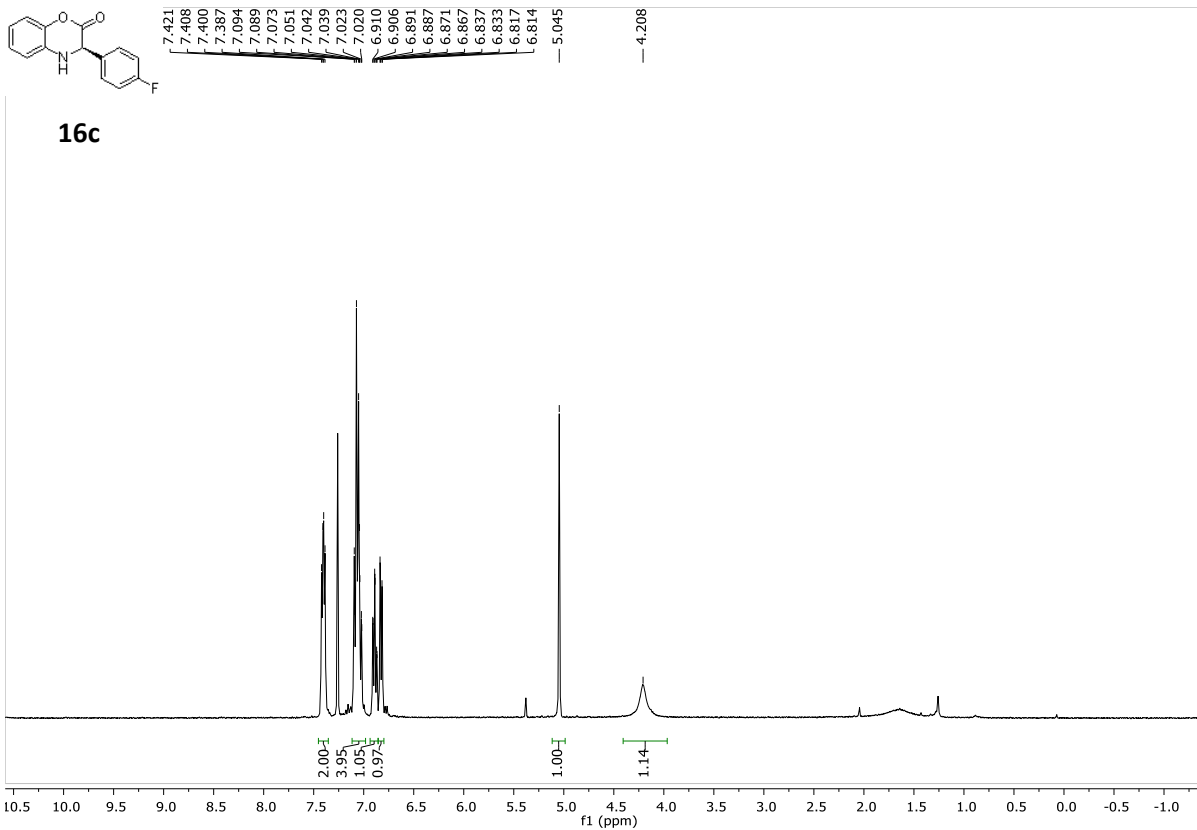
16a

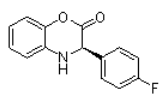


16a

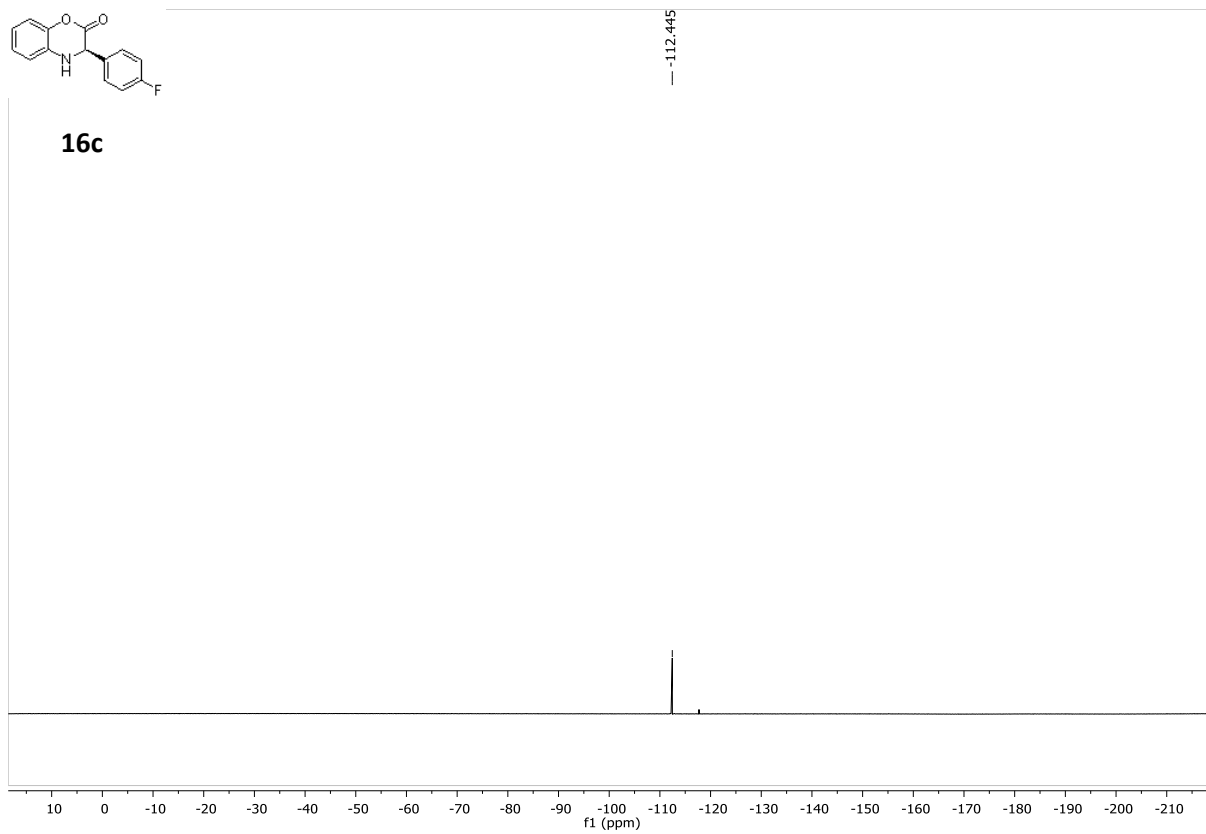


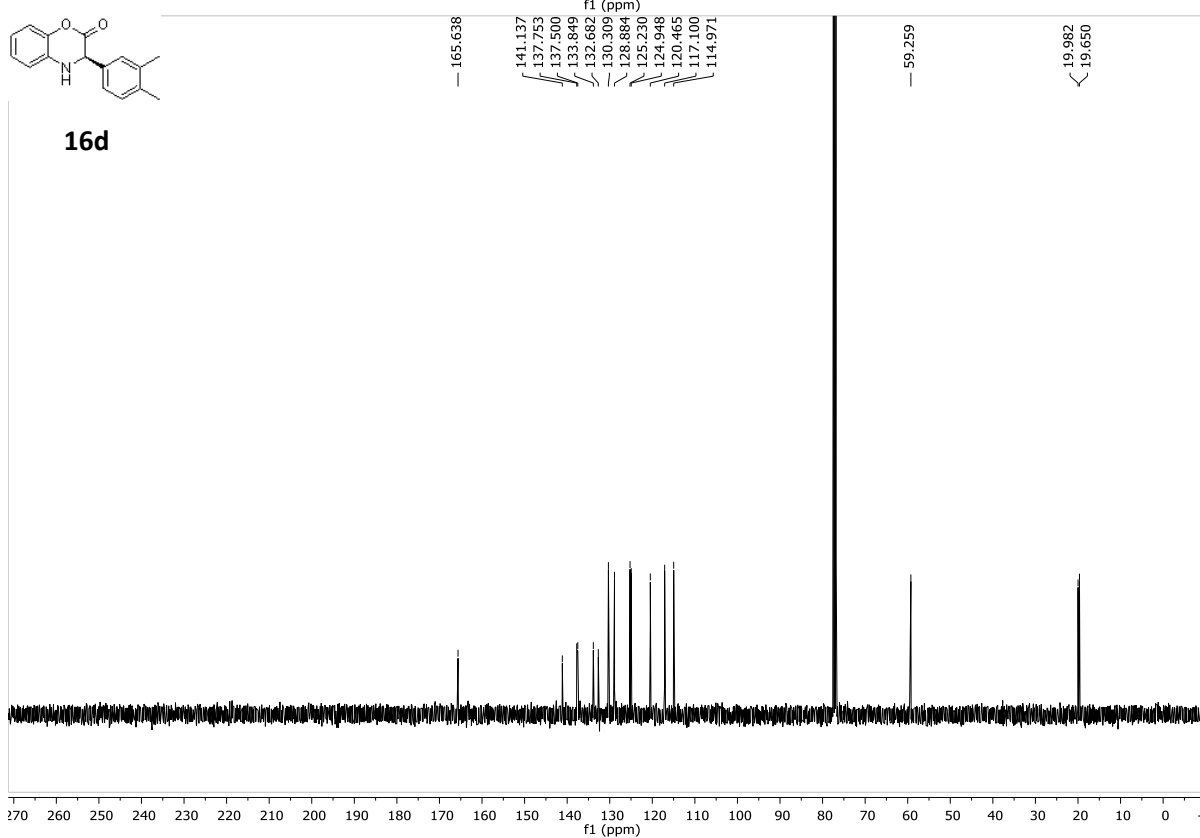
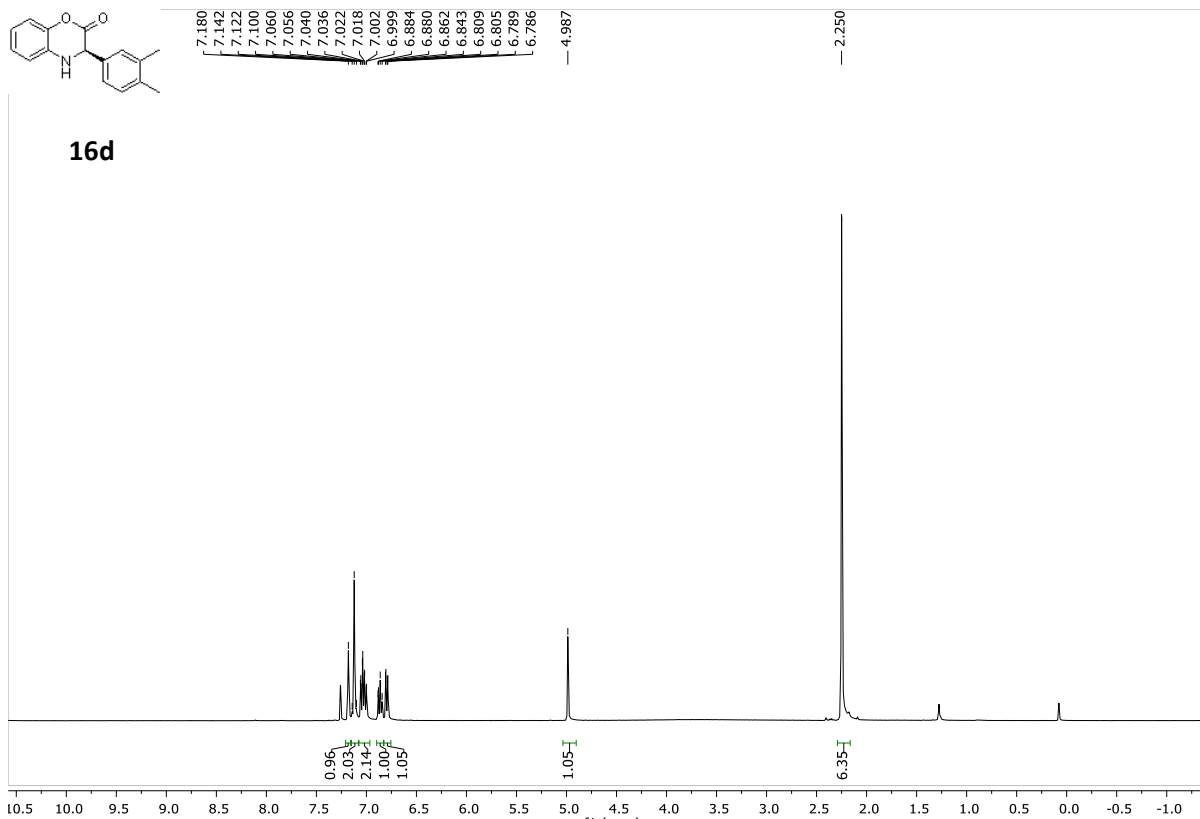


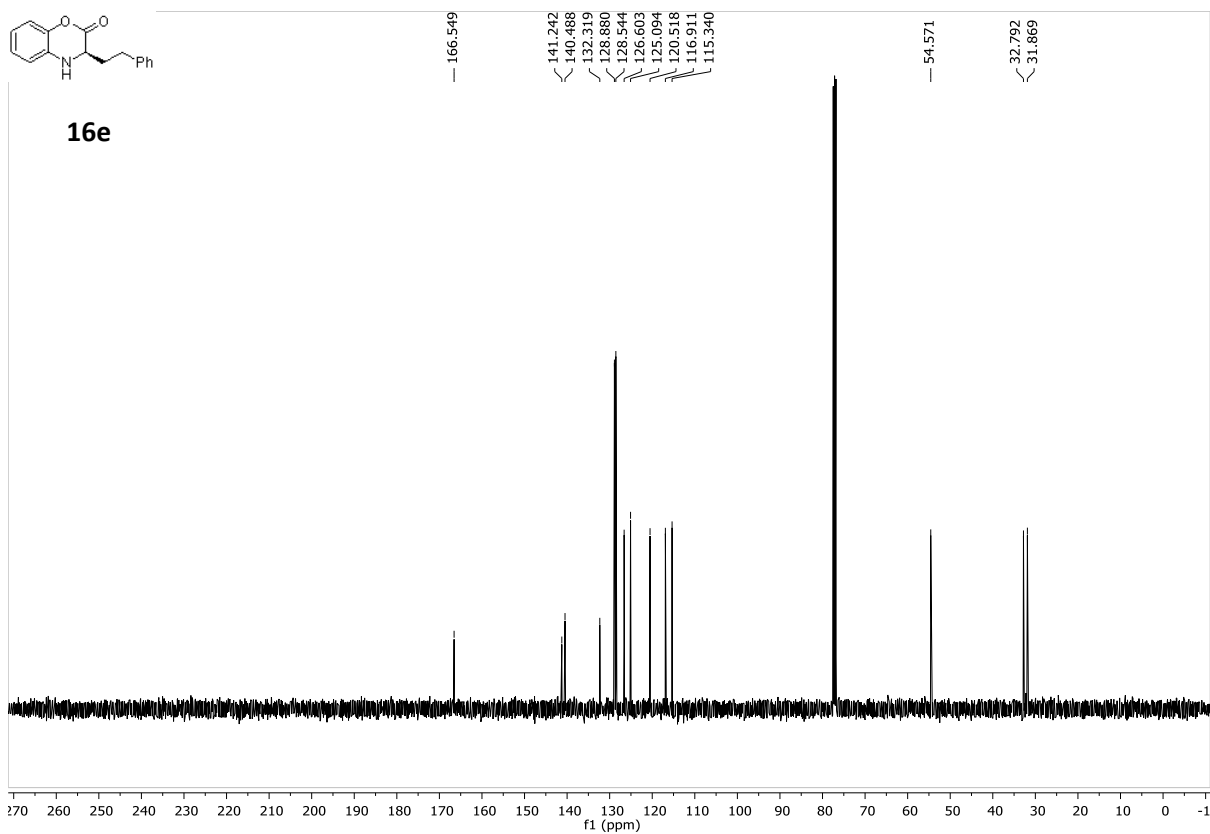
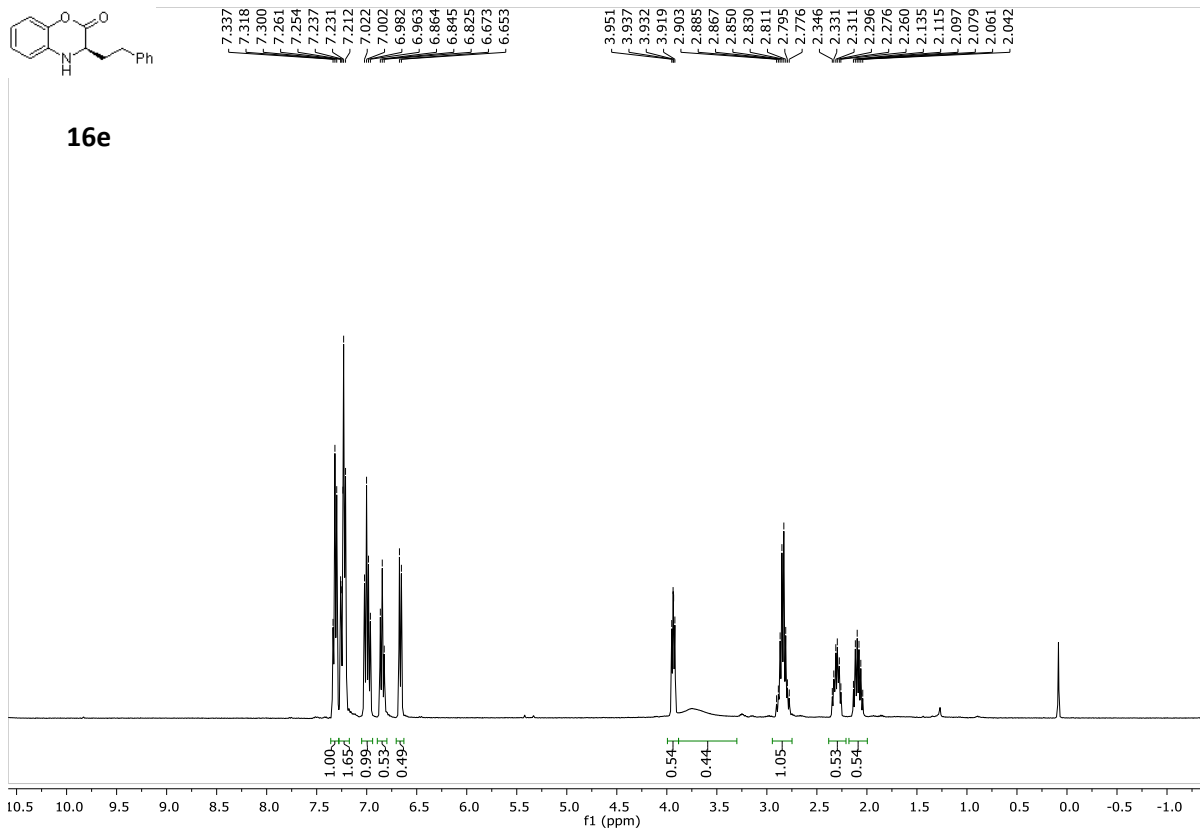


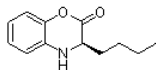


16c

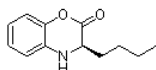
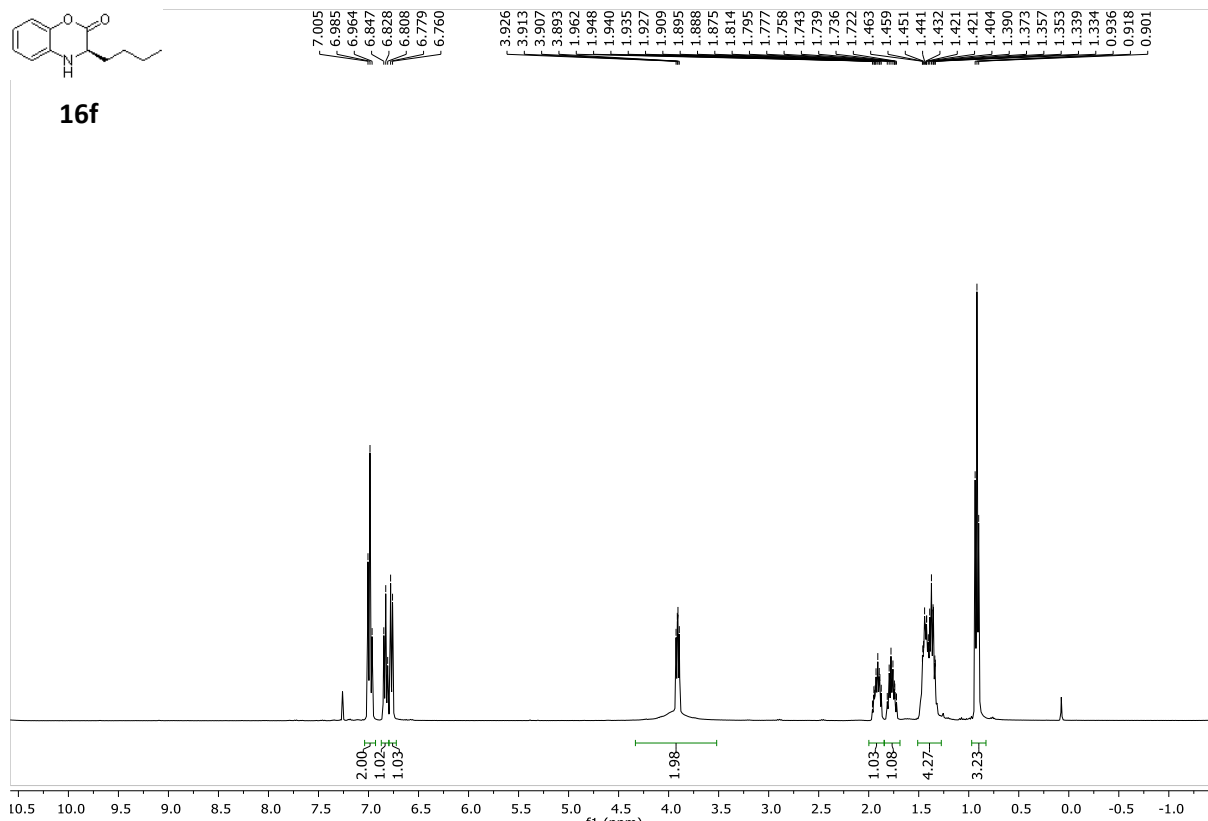




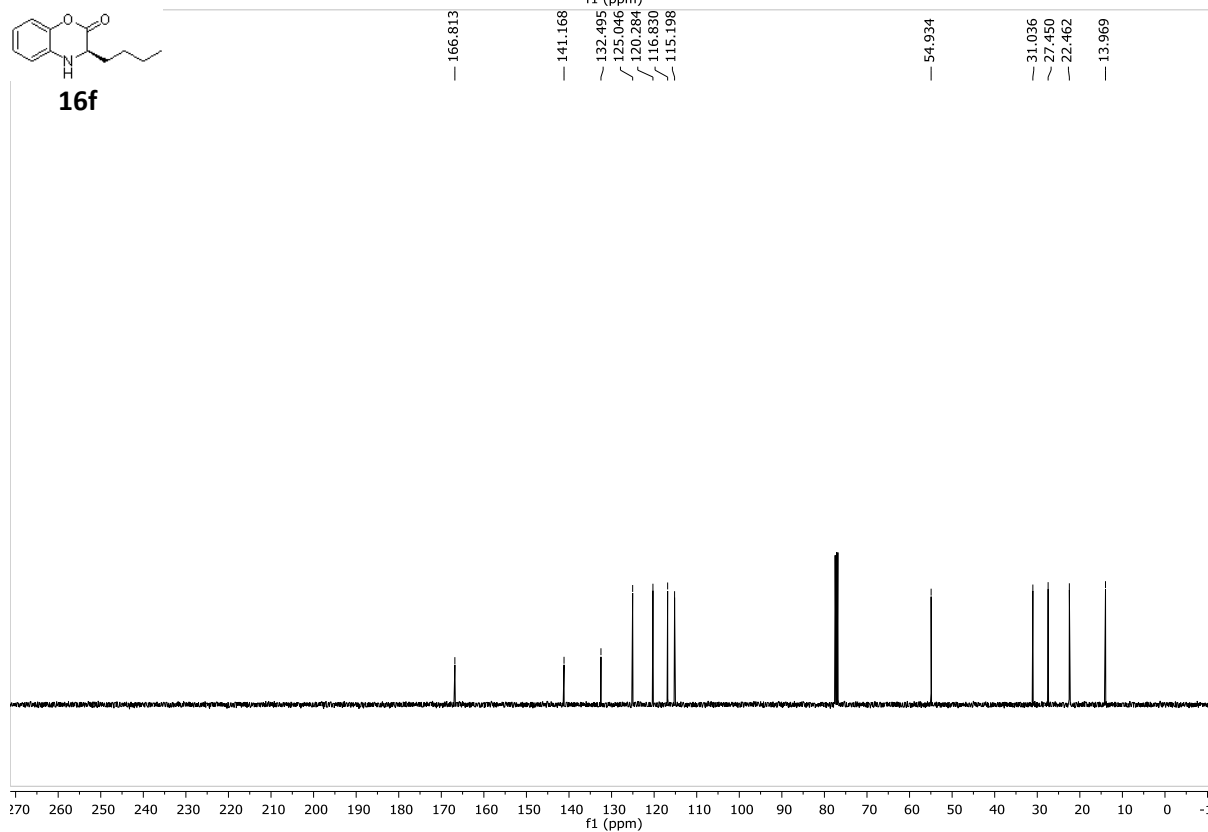


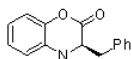


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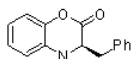
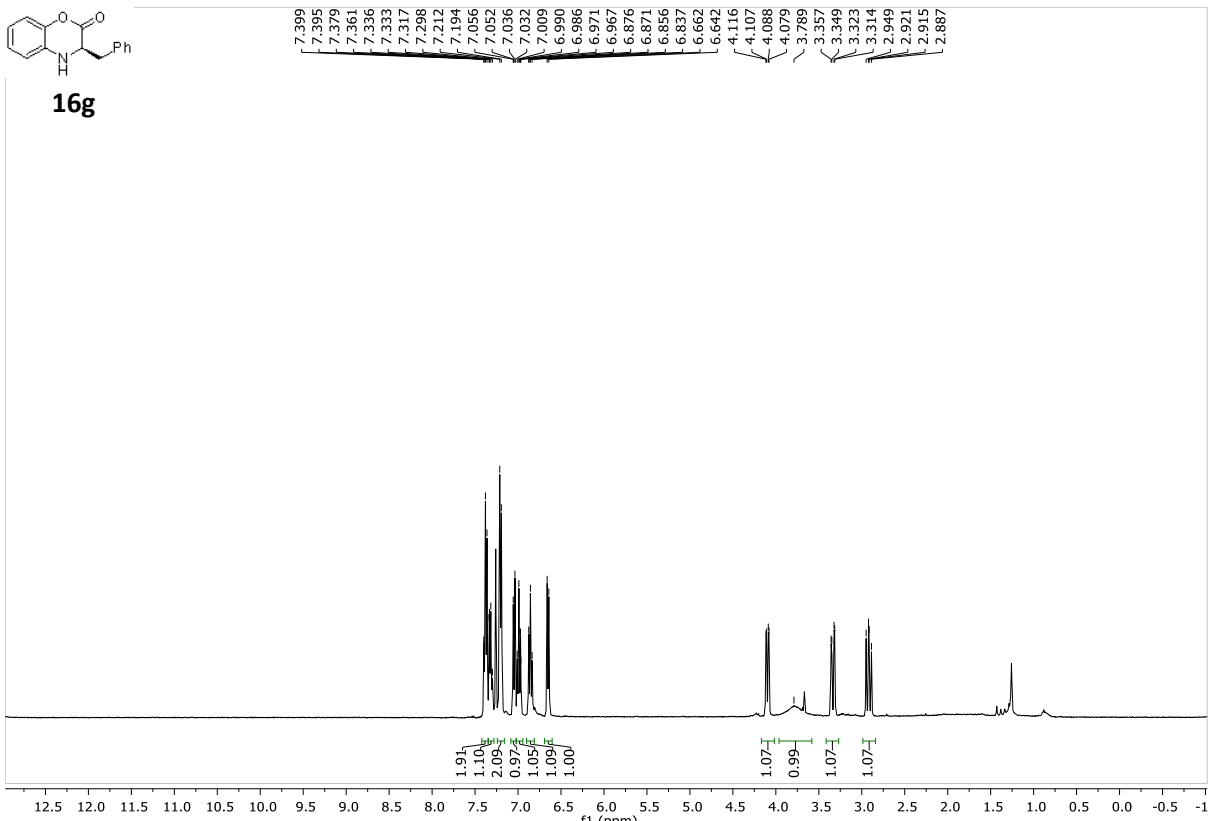


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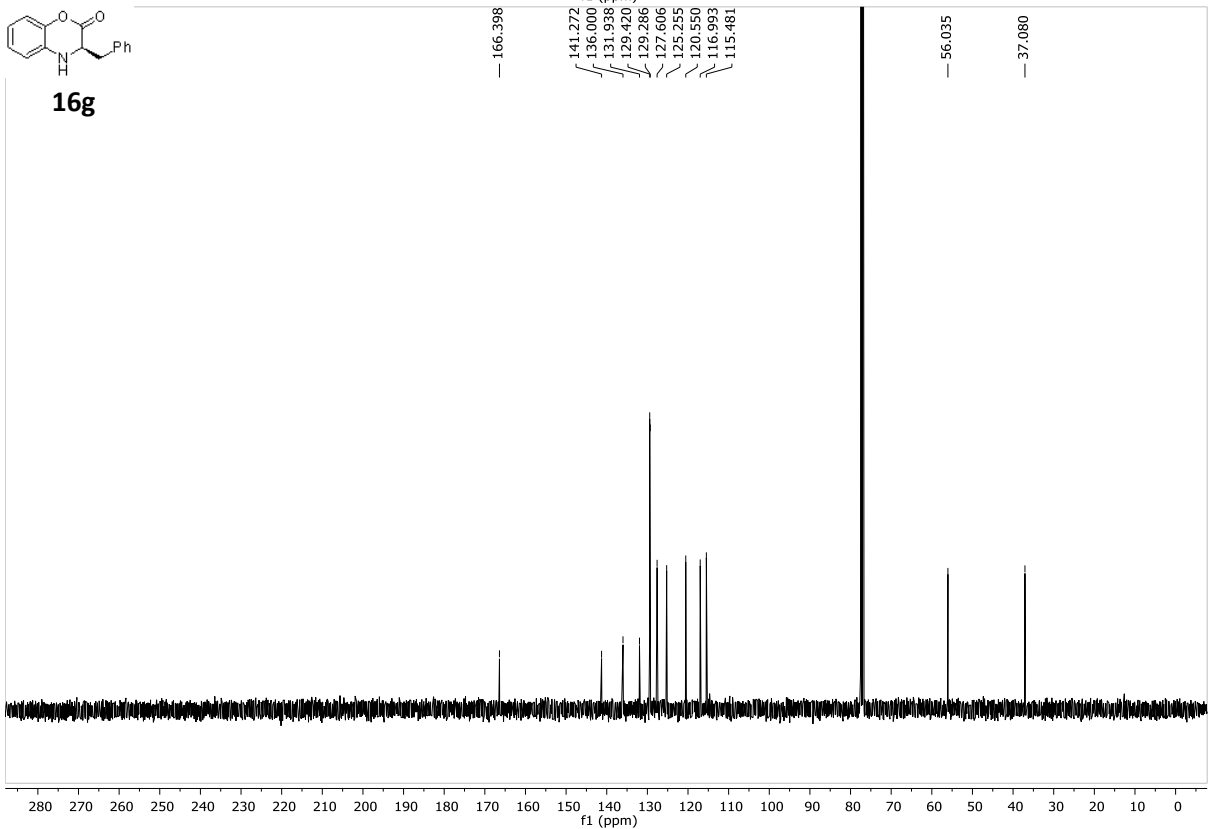


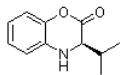


16g

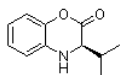
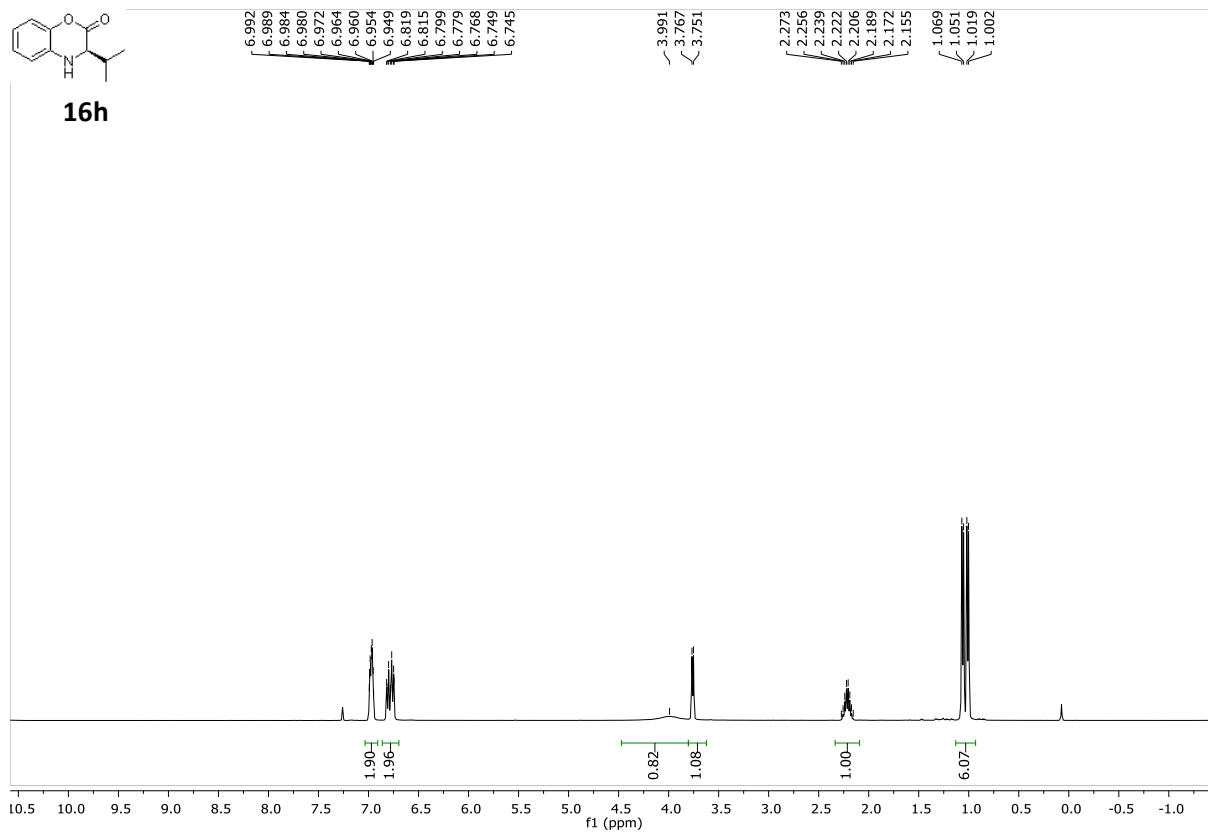


16g

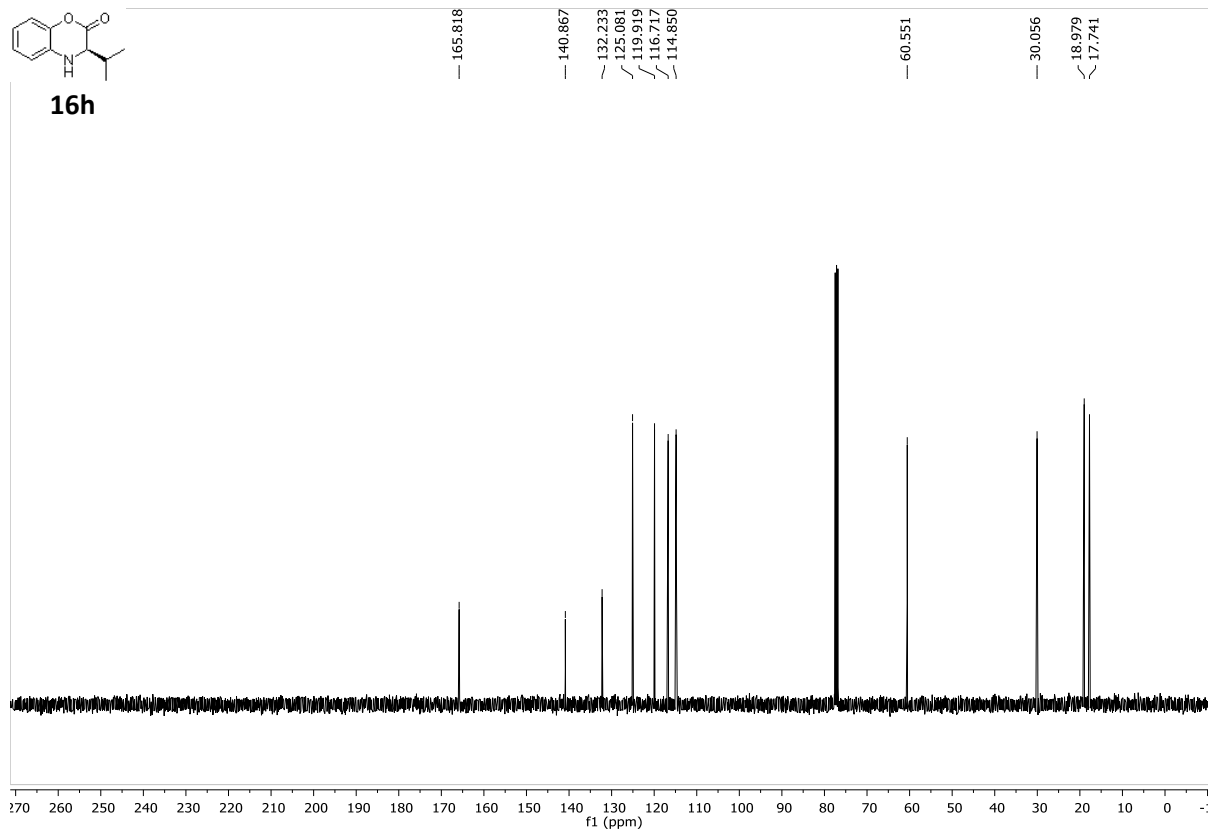


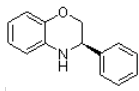


16h

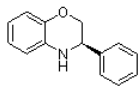
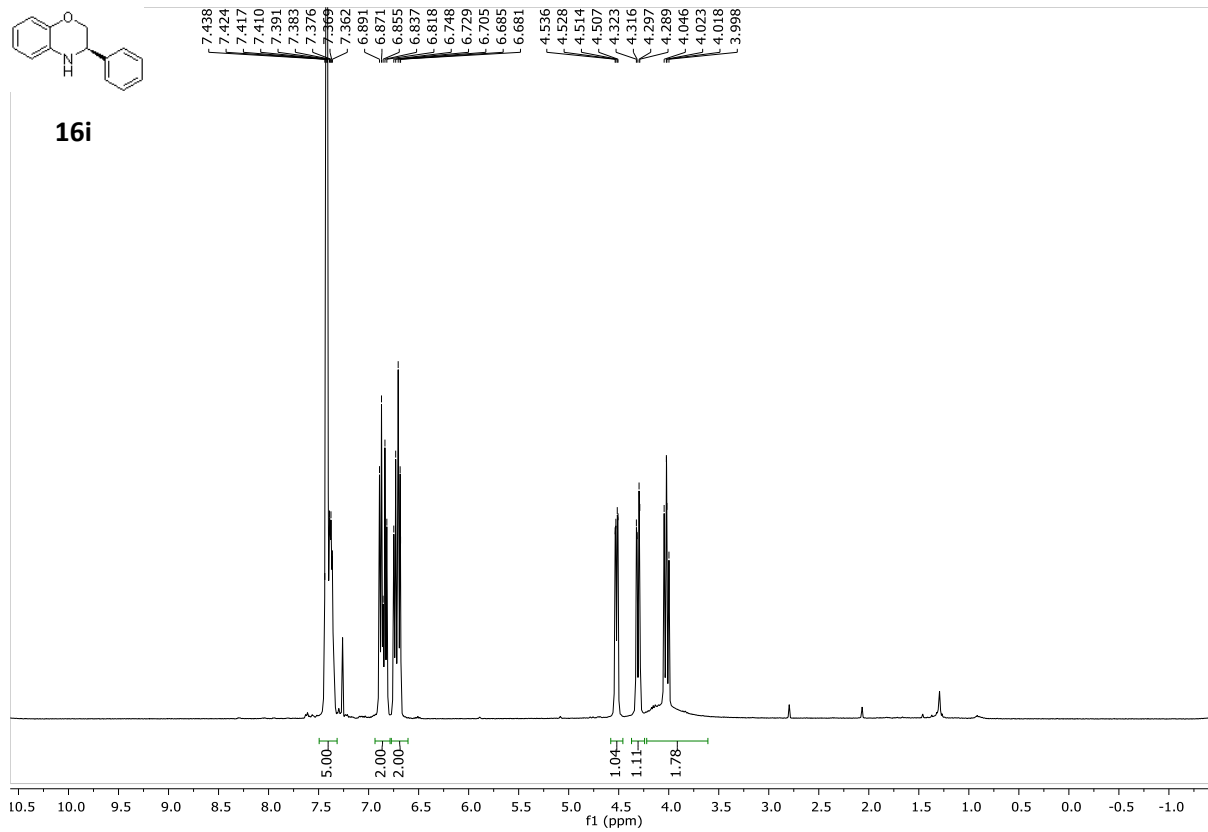


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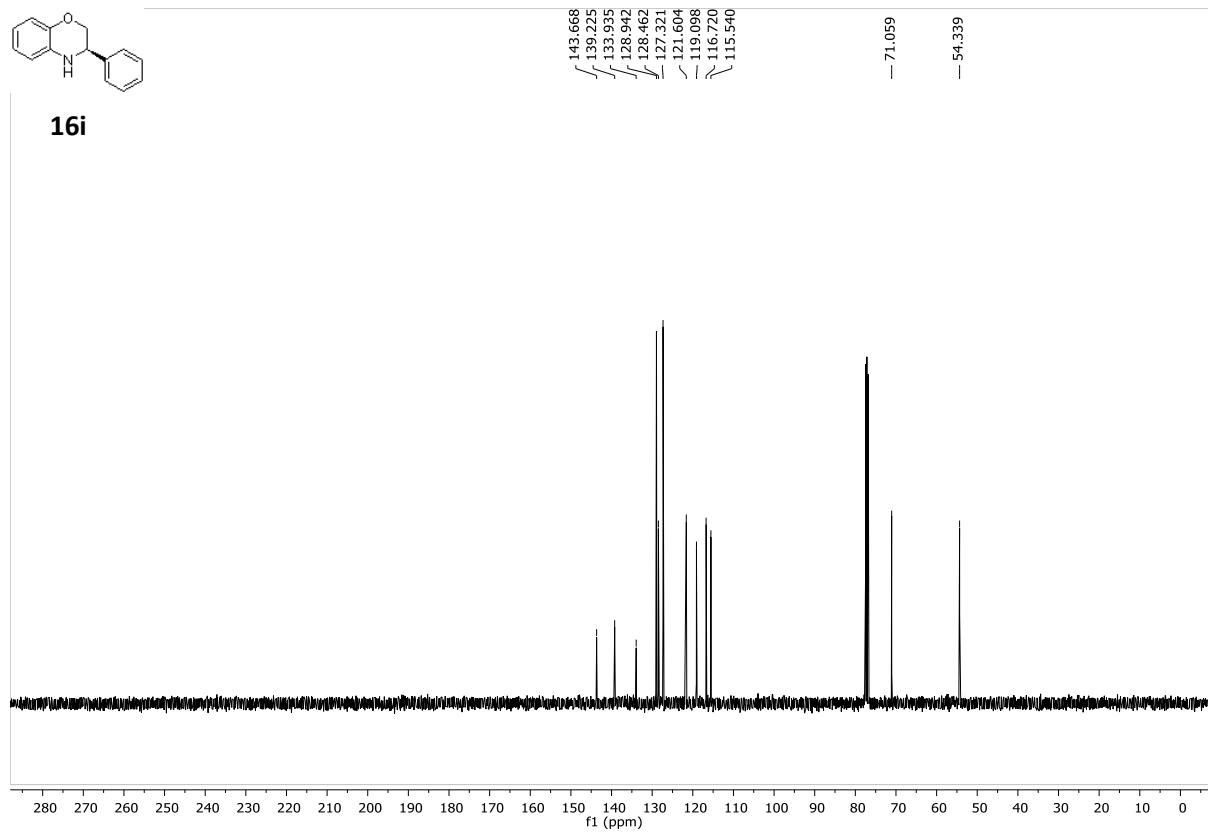


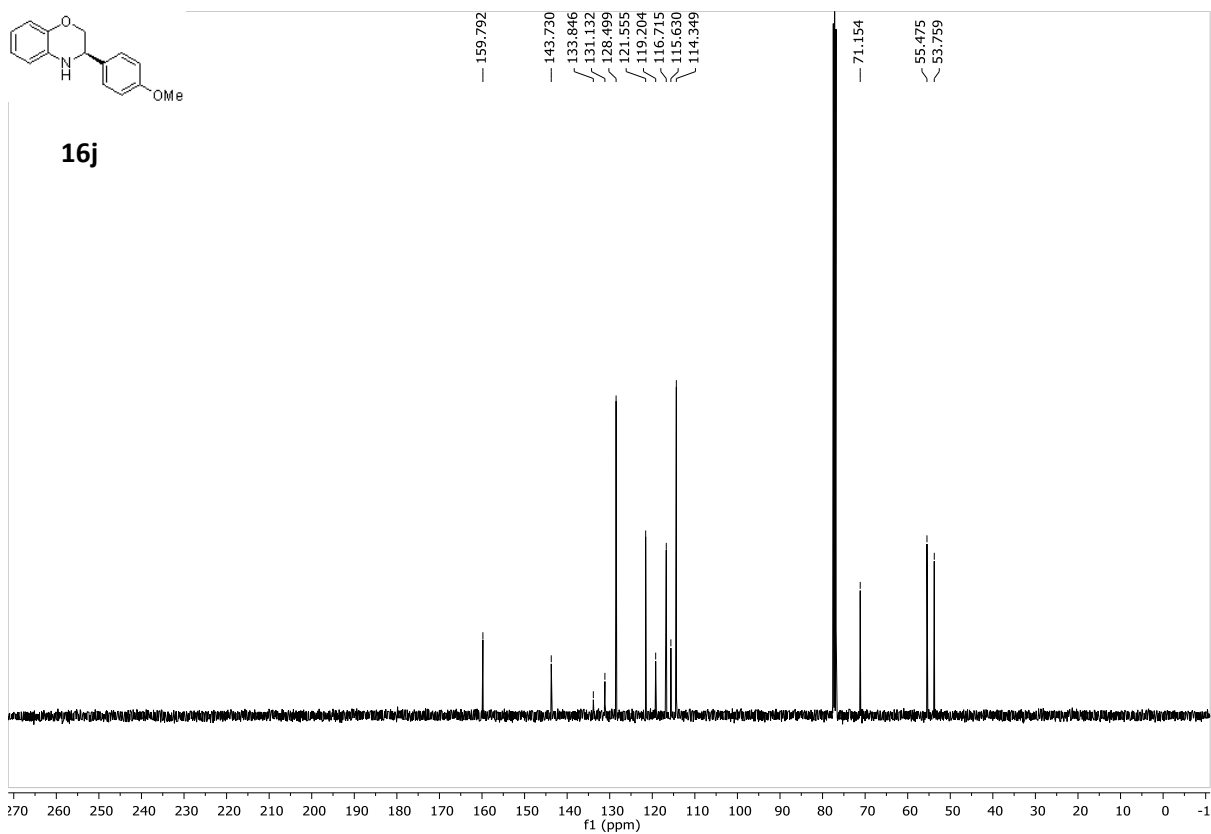
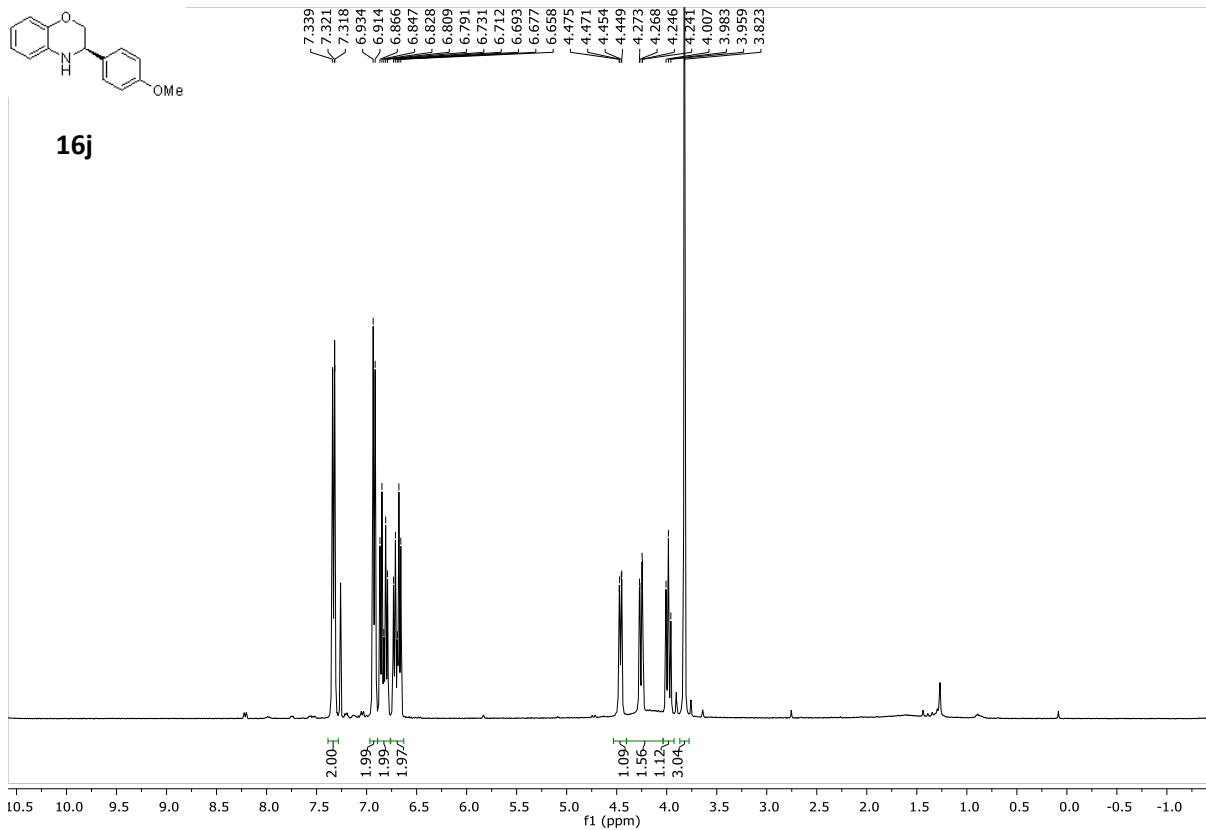


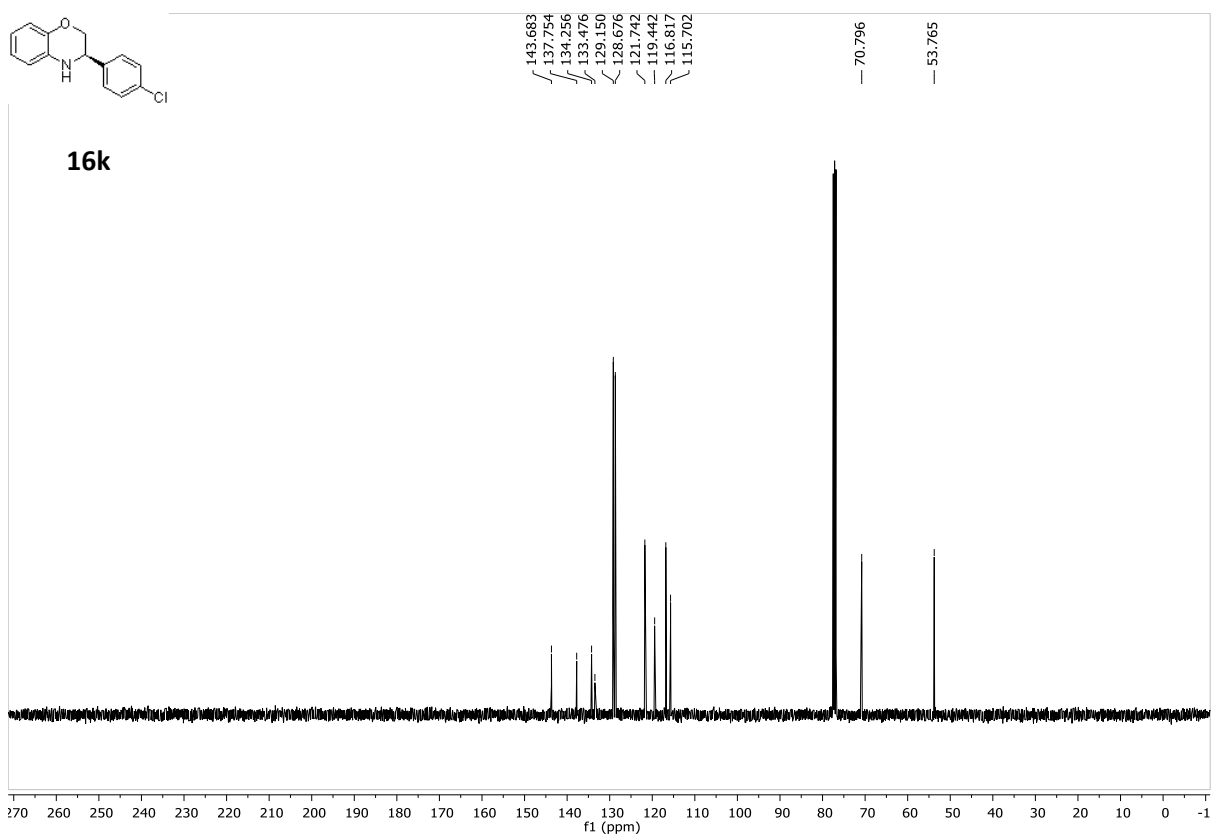
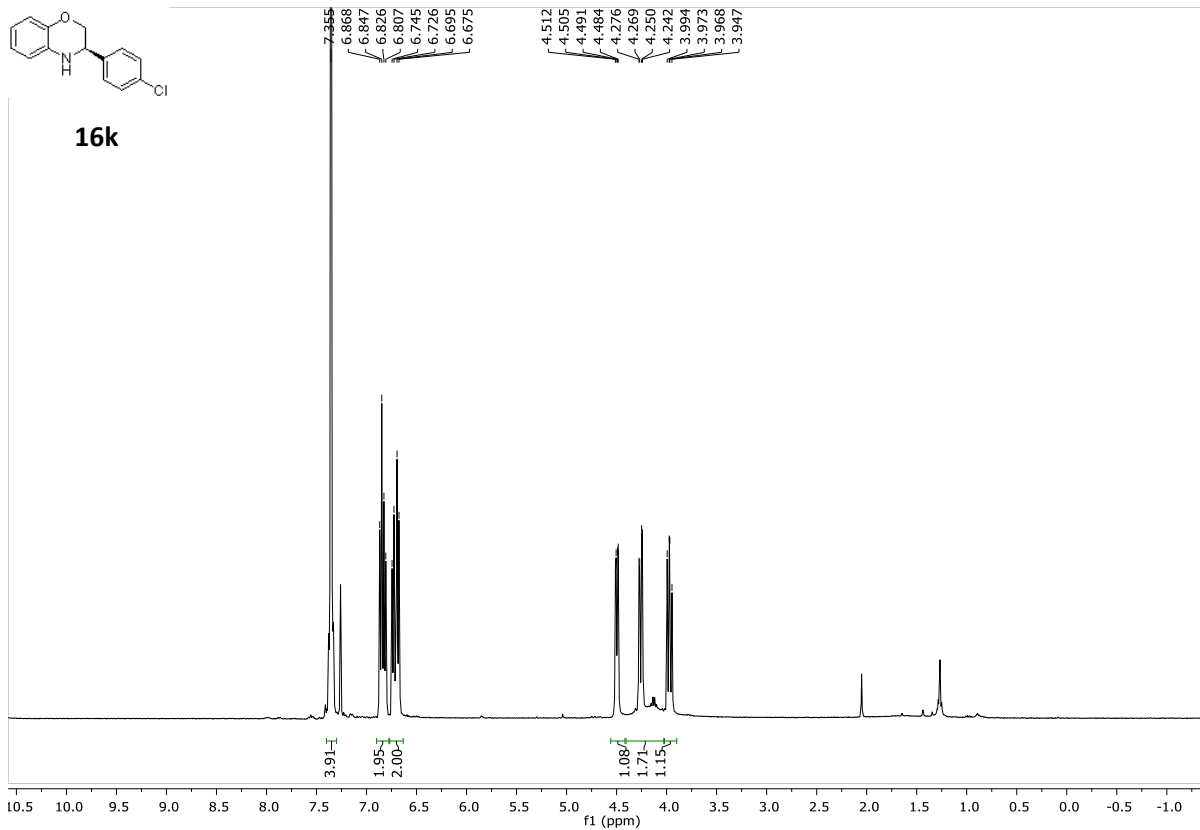
16i

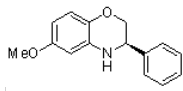


16i

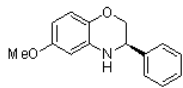
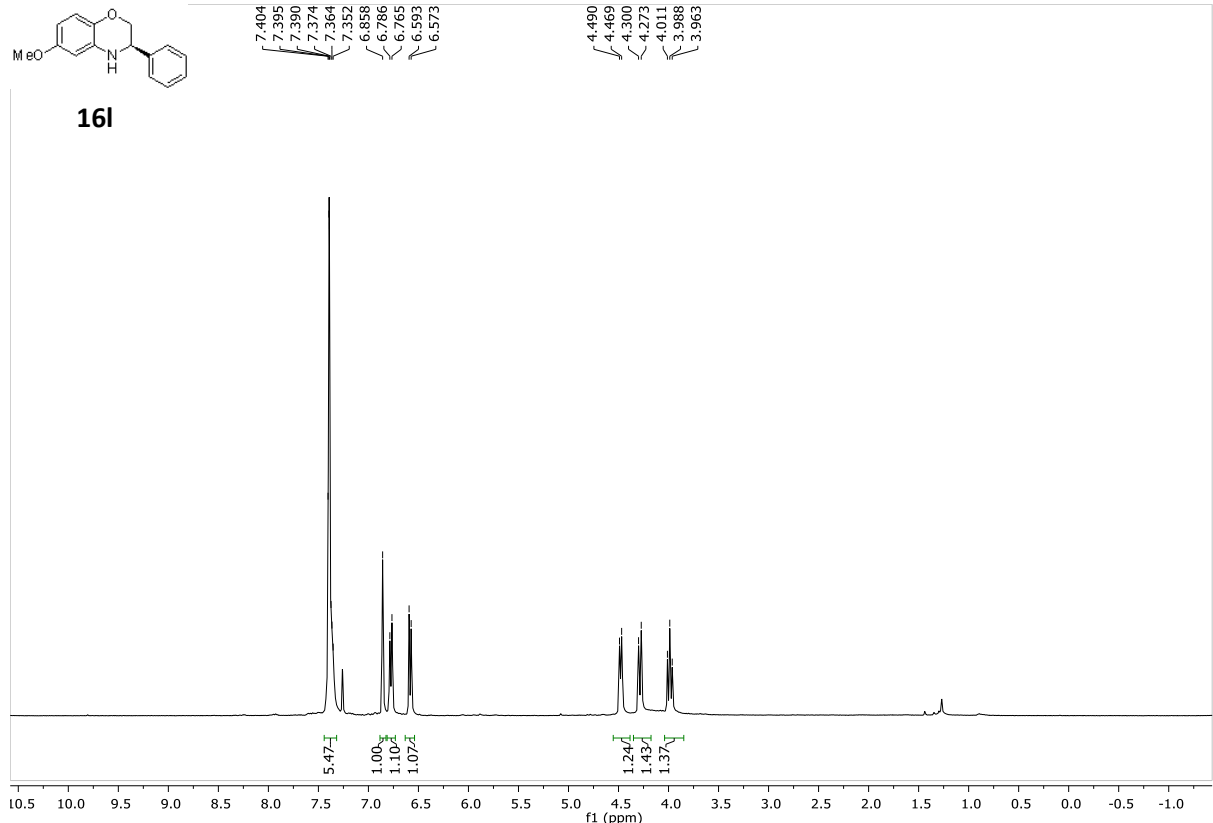




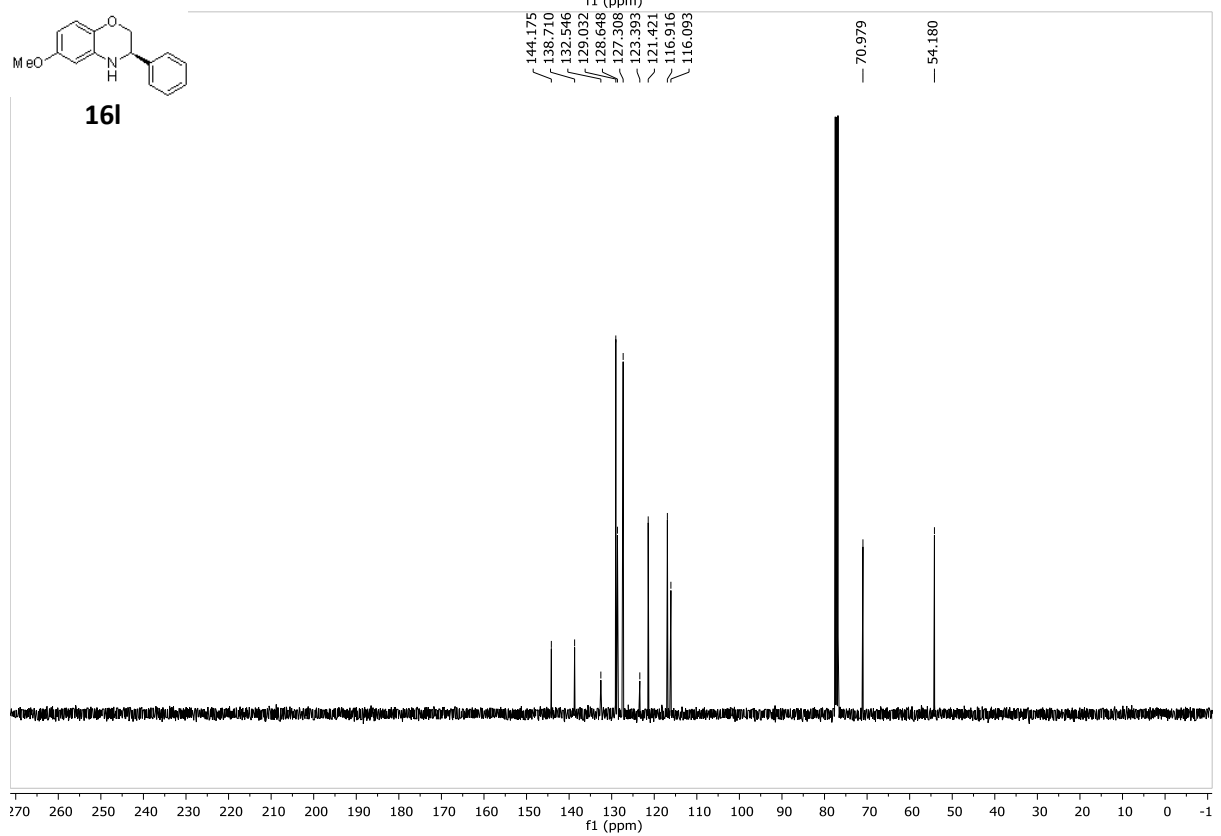




16l



16l



6. HPLC traces

Ee of product **16** was determined by HPLC using chiral column OD-H. Eluents of iPrOH/hexane (30/70 or 10/90) was used. Racemic samples were prepared via hydrogenation using Pd/C catalyst.

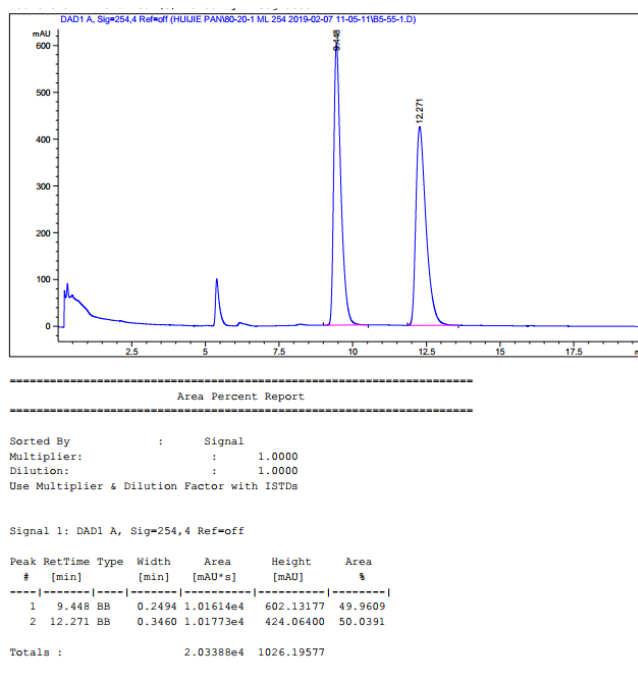
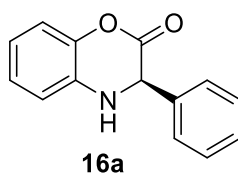


Figure S2 HPLC report of racemic **16a**

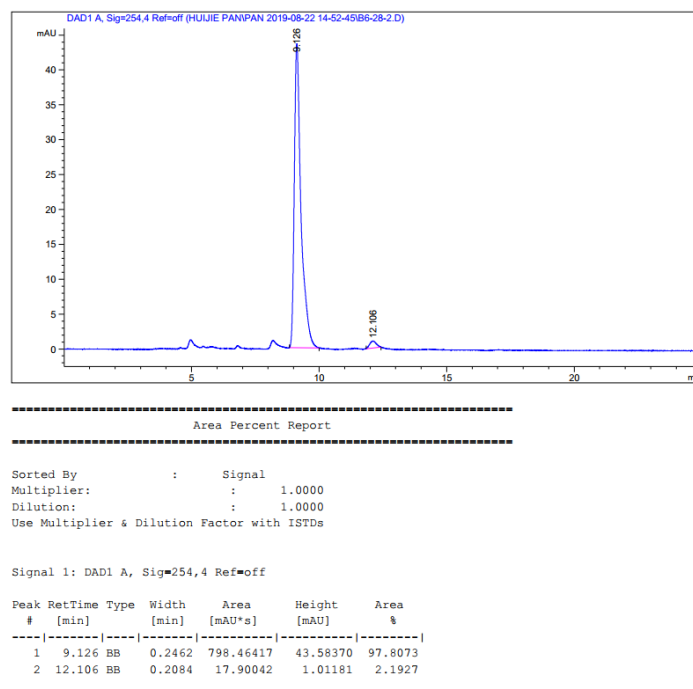
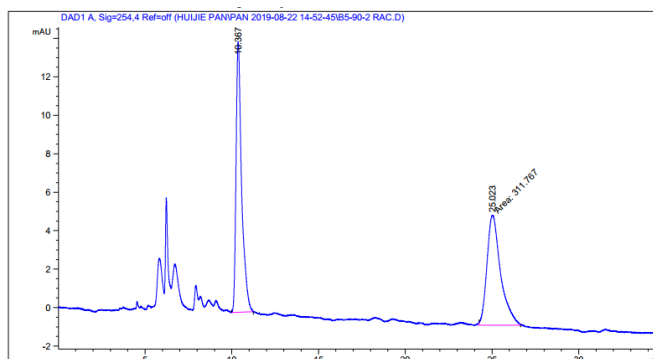
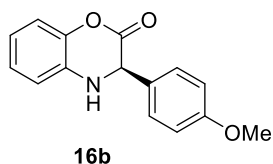


Figure S3 HPLC report of enantioenriched **16a**



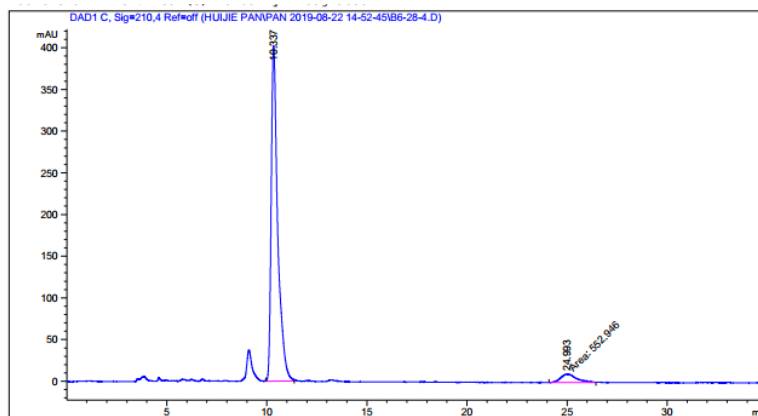
Area Percent Report

Sorted By : Signal
Multiplier: : 1.0000
Dilution: : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.367	BB	0.2652	315.06192	14.07457	50.2628
2	25.023	MM	0.9060	311.76721	5.73494	49.7372

Figure S4 HPLC report of racemic **16b**



Area Percent Report

Sorted By : Signal
Multiplier: : 1.0000
Dilution: : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 C, Sig=210,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.337	VV	0.2790	9112.08203	401.99298	94.2789
2	24.993	MM	0.9087	552.94592	10.14203	5.7211

Figure S5 HPLC report of enantioenriched **16b**

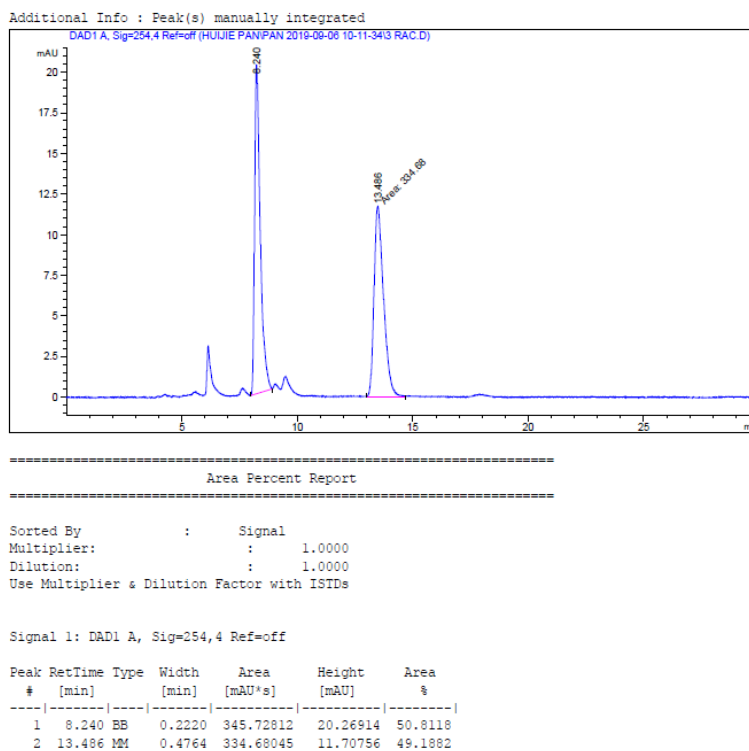
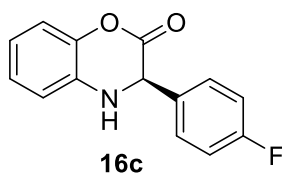


Figure S6 HPLC report of racemic **16c**

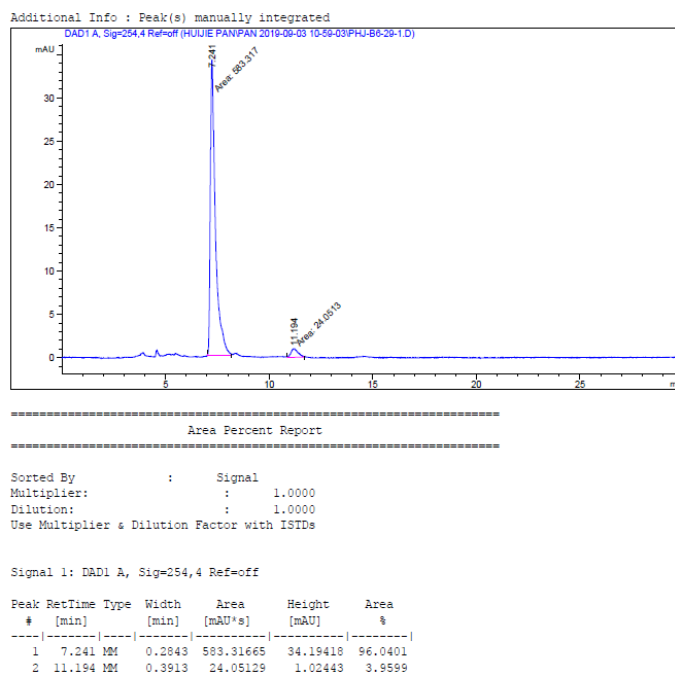
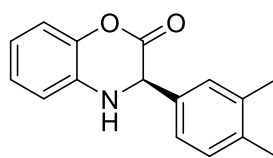
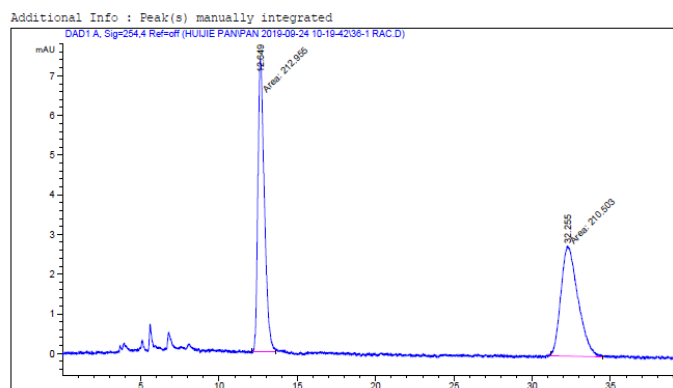


Figure S7 HPLC report of chiral **16c**

HPLC report of enantioenriched **16c**



16d



Area Percent Report

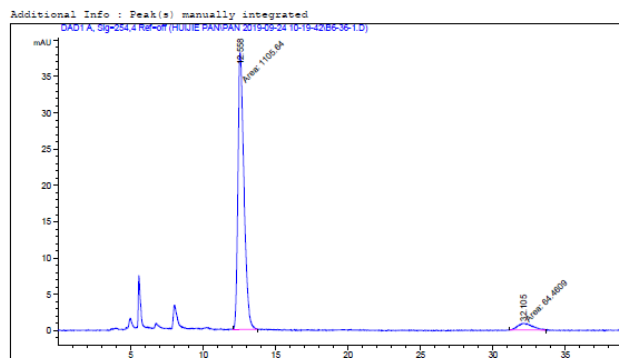
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Sorted By      :      Signal
Multiplier:    :      1.0000
Dilution:      :      1.0000
Use Multiplier & Dilution Factor with ISTDs
    
```

Signal 1: DAD1 A, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.649	MM	0.4809	212.9543	7.35025	50.2896
2	32.255	MM	1.2575	210.50282	2.78990	49.7104

Figure S8 HPLC report of racemic **16d**



Area Percent Report

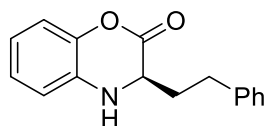
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Sorted By      :      Signal
Multiplier:    :      1.0000
Dilution:      :      1.0000
Use Multiplier & Dilution Factor with ISTDs
    
```

Signal 1: DAD1 A, Sig=254,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.558	MM	0.4821	1105.64209	28.22450	94.4910
2	32.105	MM	1.1639	64.46055	9.23052e-1	5.5090

Figure S9 HPLC report of enantioenriched **16d**



16e

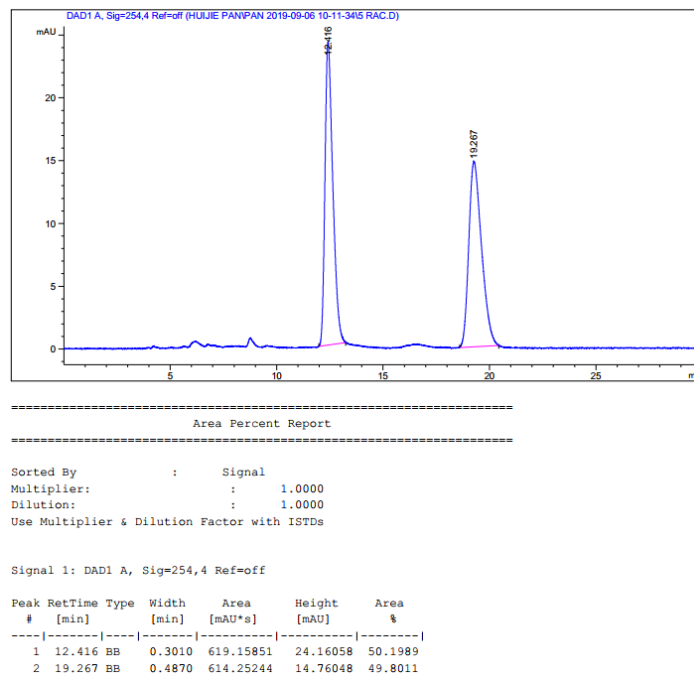


Figure S10 HPLC report of racemic **16e**

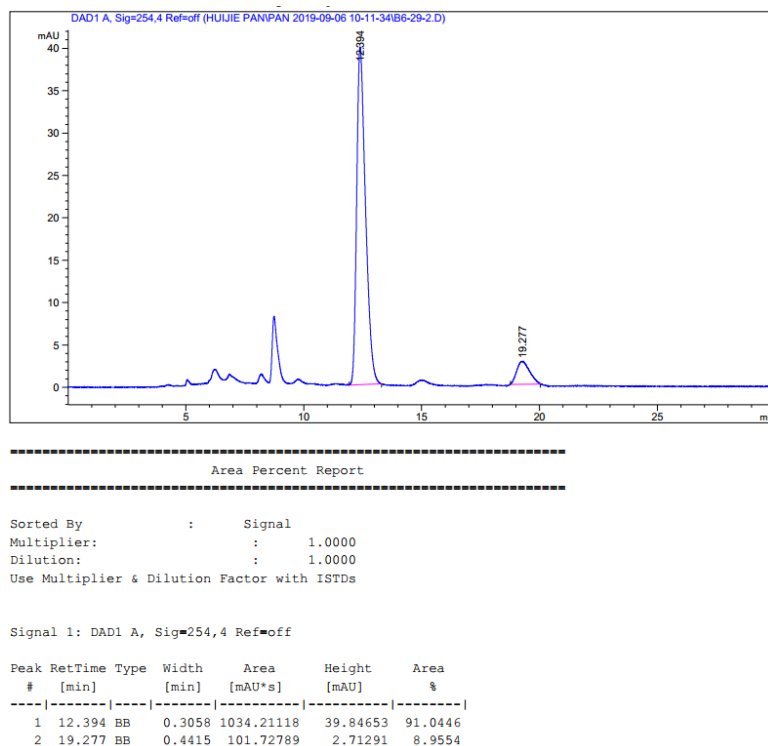
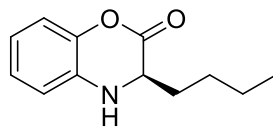
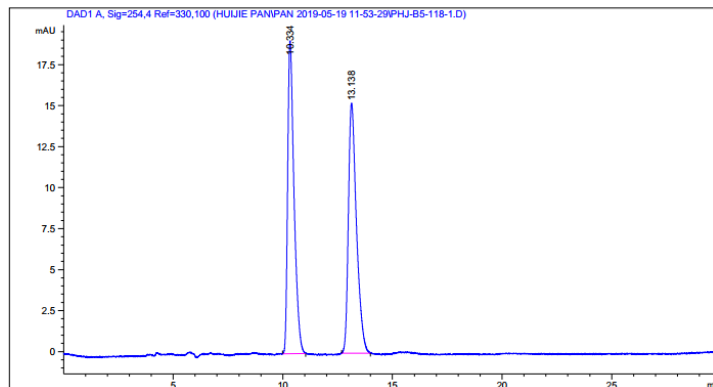


Figure S11 HPLC report of enantioenriched **16e**



16f

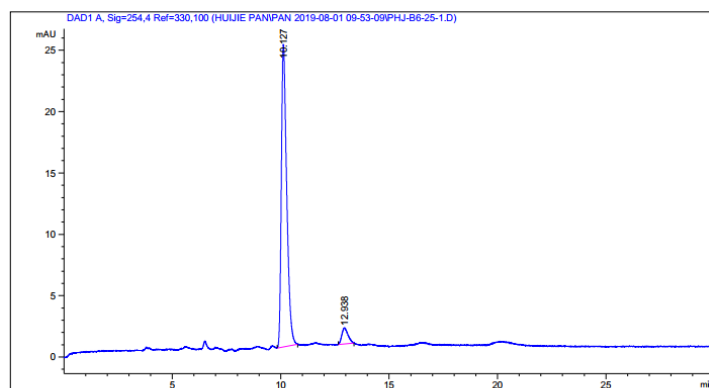


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 Area Percent Report
 =====
 Sorted By : Signal
 Multiplier: : 1.0000
 Dilution: : 1.0000
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=330,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.334	BB	0.2363	383.97266	19.08164	49.5781
2	13.138	BB	0.3009	390.50702	15.24051	50.4219

Figure S12 HPLC report of racemic **16f**



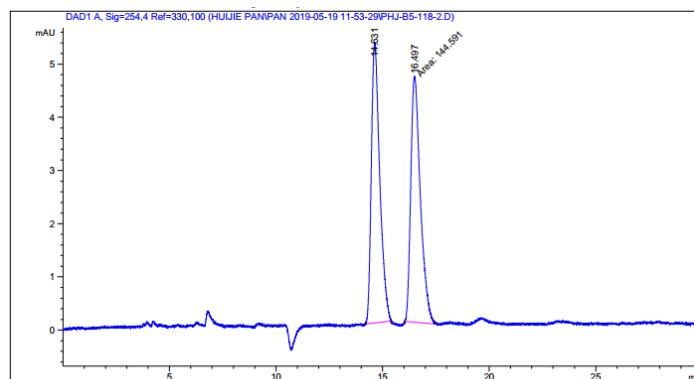
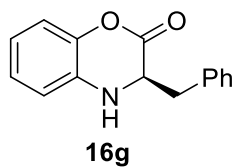
=====
 Area Percent Report
 =====
 Sorted By : Signal
 Multiplier: : 1.0000
 Dilution: : 1.0000
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=330,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.127	BB	0.2184	423.49857	24.65500	94.1730
2	12.938	BB	0.2333	26.20432	1.31865	5.8270

Figure S13 HPLC report of racemic **16f**





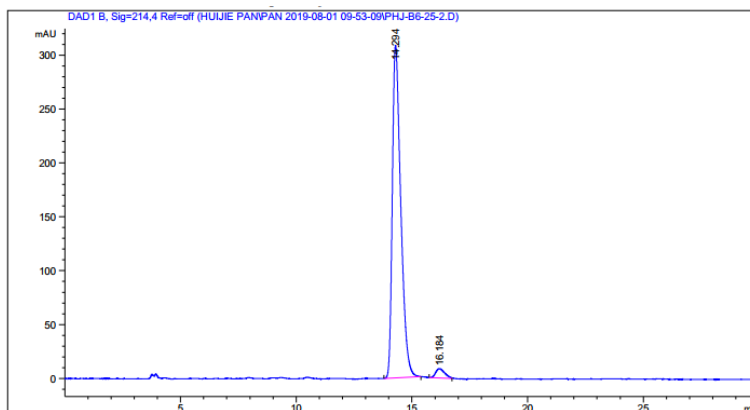
=====
Area Percent Report
=====

Sorted By : Signal
Multiplier: : 1.0000
Dilution: : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=330,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.631	BB	0.3228	144.36594	5.25920	49.9611
2	16.497	MM	0.5209	144.59065	4.62665	50.0389

Figure S14 HPLC report of racemic **16g**



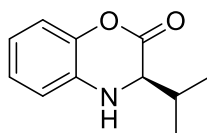
=====
Area Percent Report
=====

Sorted By : Signal
Multiplier: : 1.0000
Dilution: : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 B, Sig=214,4 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.294	BB	0.3547	7799.38379	308.73080	97.0632
2	16.184	BV	0.3156	235.98488	8.79266	2.9368

Figure S15 HPLC report of enantioenriched **16f**



16h

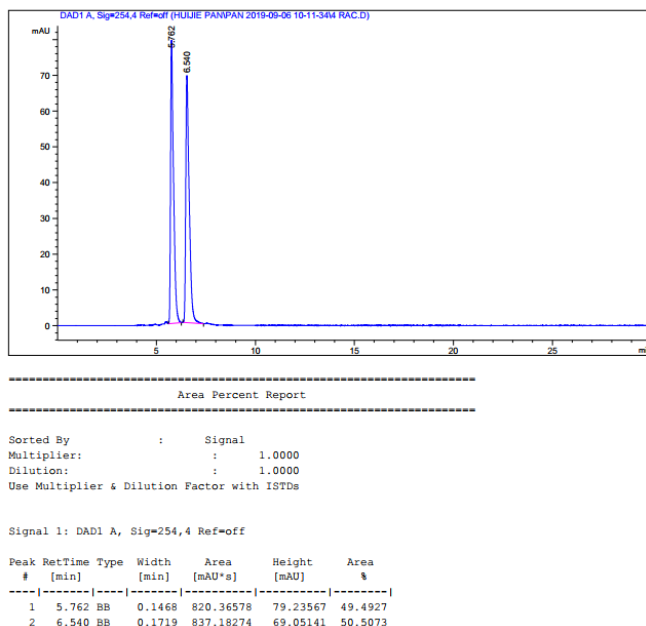


Figure S16 HPLC report of racemic **16h**

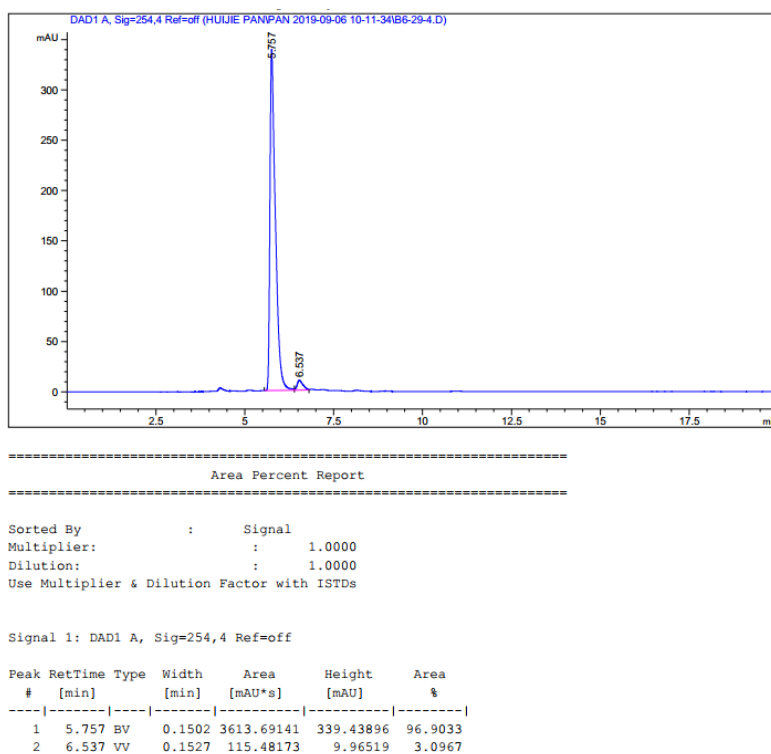
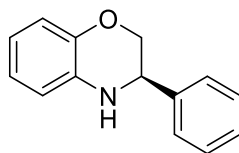
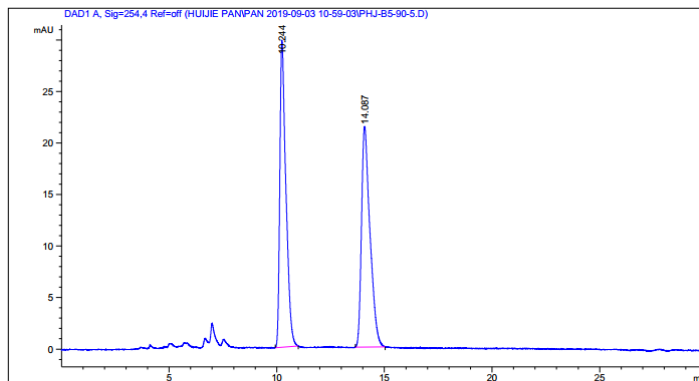


Figure S17 HPLC report of enantioenriched **16h**



16i



```

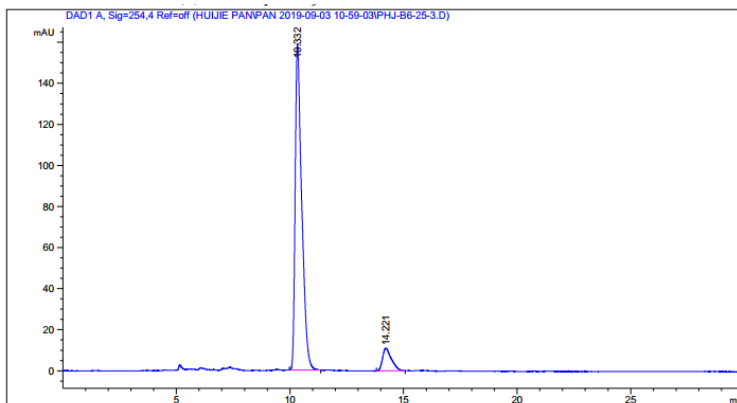
=====
                          Area Percent Report
=====
Sorted By      :      Signal
Multiplier:    :      1.0000
Dilution:      :      1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

```

Signal 1: DAD1 A, Sig=254,4 Ref=off

Peak RetTime Type Width Area Height Area
# [min] [min] [min] [mAU*s] [mAU] %
-----|-----|-----|-----|-----|
1 10.244 BB 0.2608 585.28052 29.77947 50.0409
2 14.087 BB 0.3214 584.32446 21.41022 49.9591
  
```

Figure S18 HPLC report of racemic **16i**



```

=====
                          Area Percent Report
=====
Sorted By      :      Signal
Multiplier:    :      1.0000
Dilution:      :      1.0000
Use Multiplier & Dilution Factor with ISTDs
  
```

```

Signal 1: DAD1 A, Sig=254,4 Ref=off

Peak RetTime Type Width Area Height Area
# [min] [min] [min] [mAU*s] [mAU] %
-----|-----|-----|-----|-----|
1 10.332 BB 0.2839 3180.06323 158.99118 91.3875
2 14.221 BB 0.3192 299.69513 11.07387 8.6125
  
```

Figure S19 HPLC report of enantioenriched **16i**

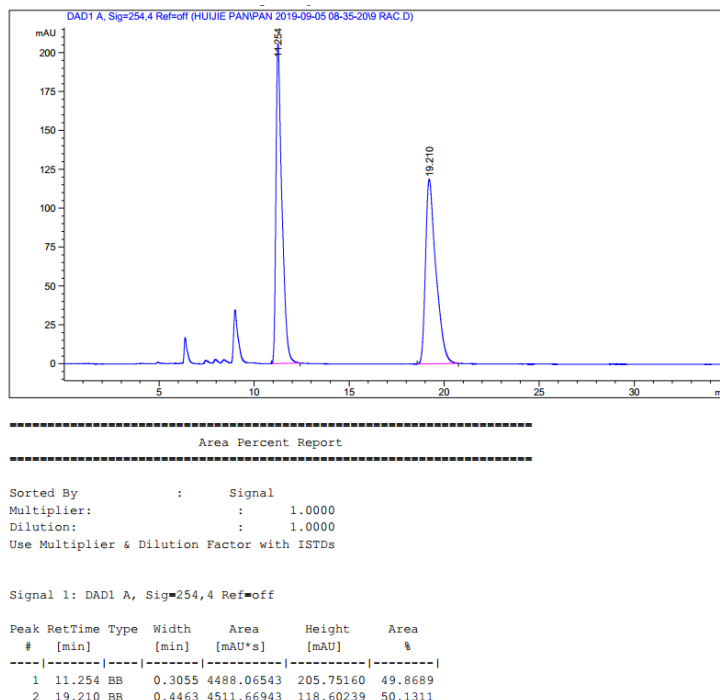
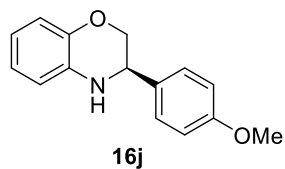


Figure S20 HPLC report of racemic **16j**

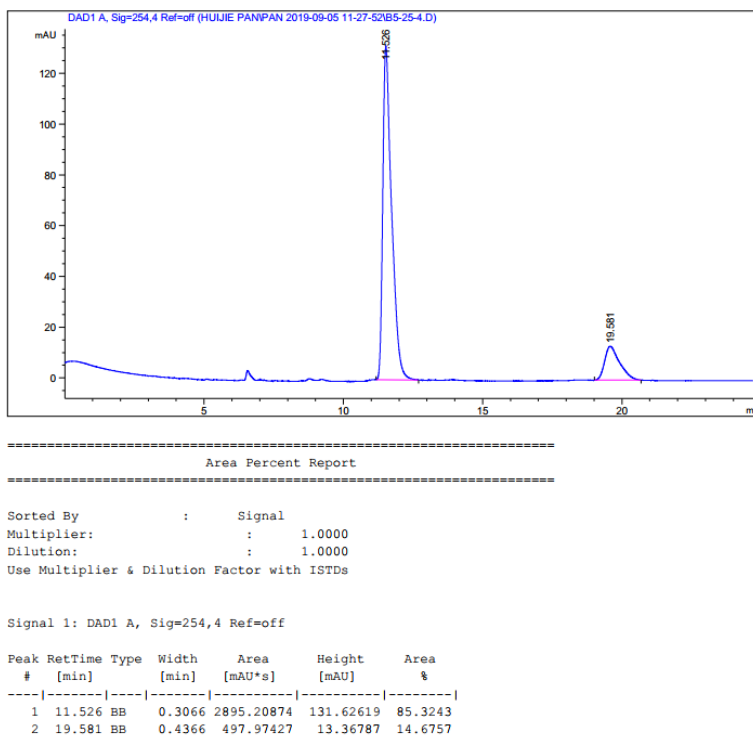


Figure S21 HPLC report of enantioenriched **16j**

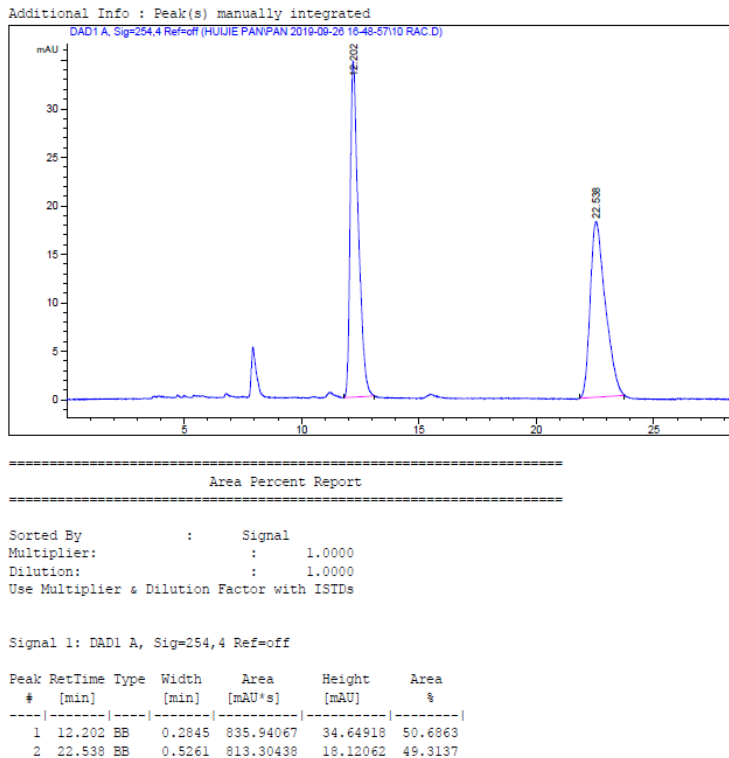
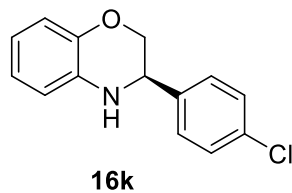


Figure S22 HPLC report of racemic **16k**

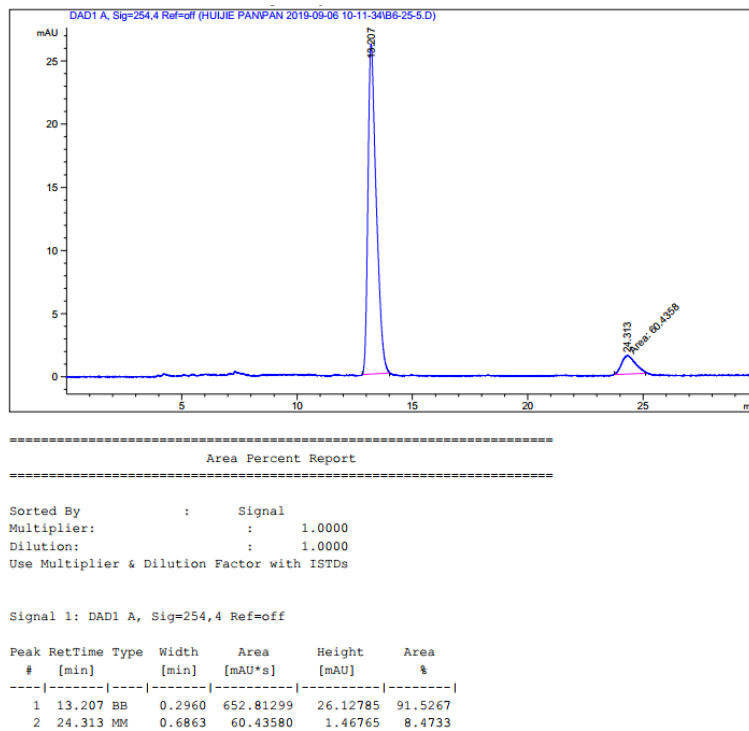


Figure S23 HPLC report of enantioenriched **16k**

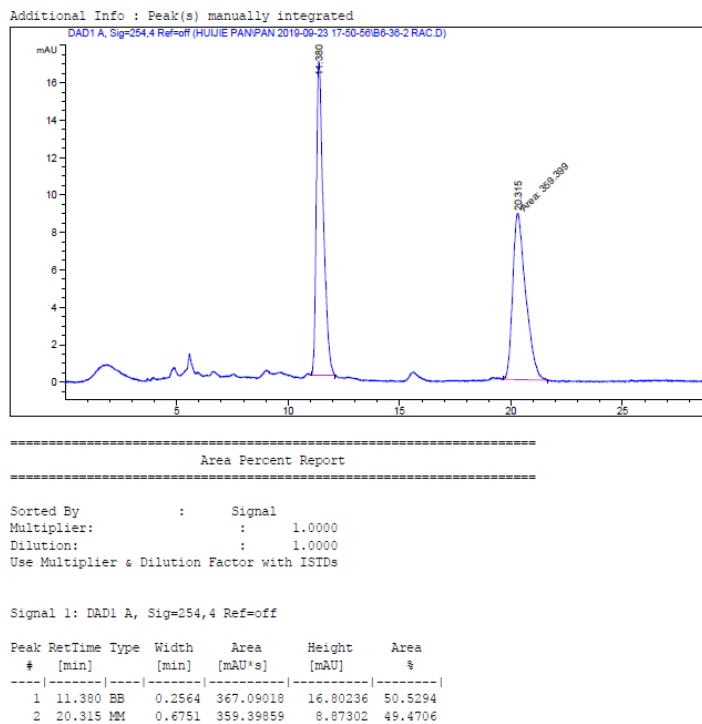
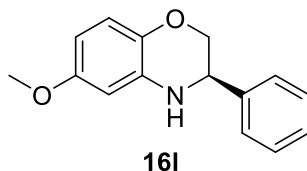


Figure S24 HPLC report of racemic **161**

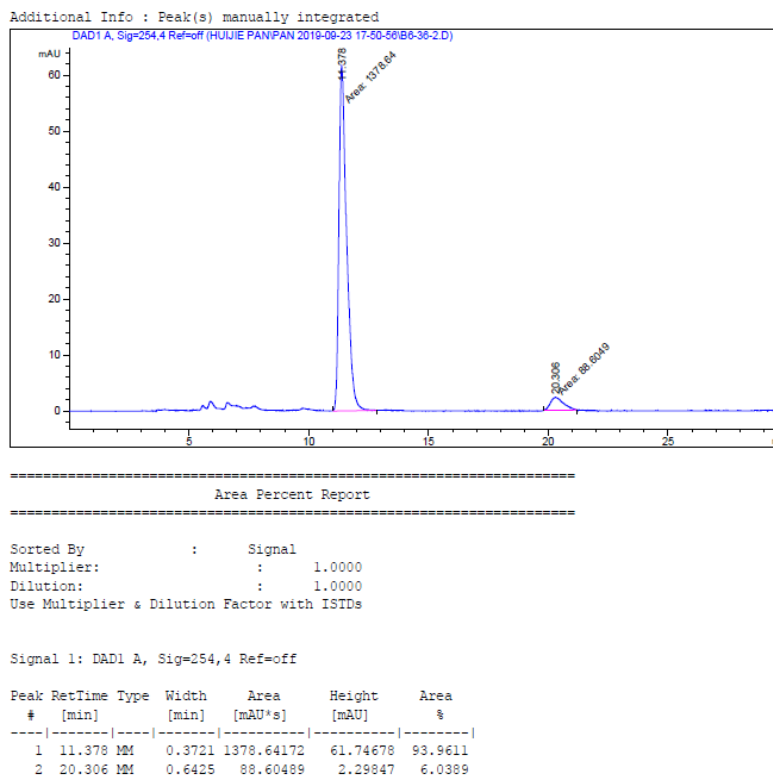


Figure S25 HPLC report of enantioenriched **161**



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