

SUPPORTING INFORMATION

Separation and Identification of Glycan Anomers

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HIGH-RESOLUTION IMS OF GLYCANS

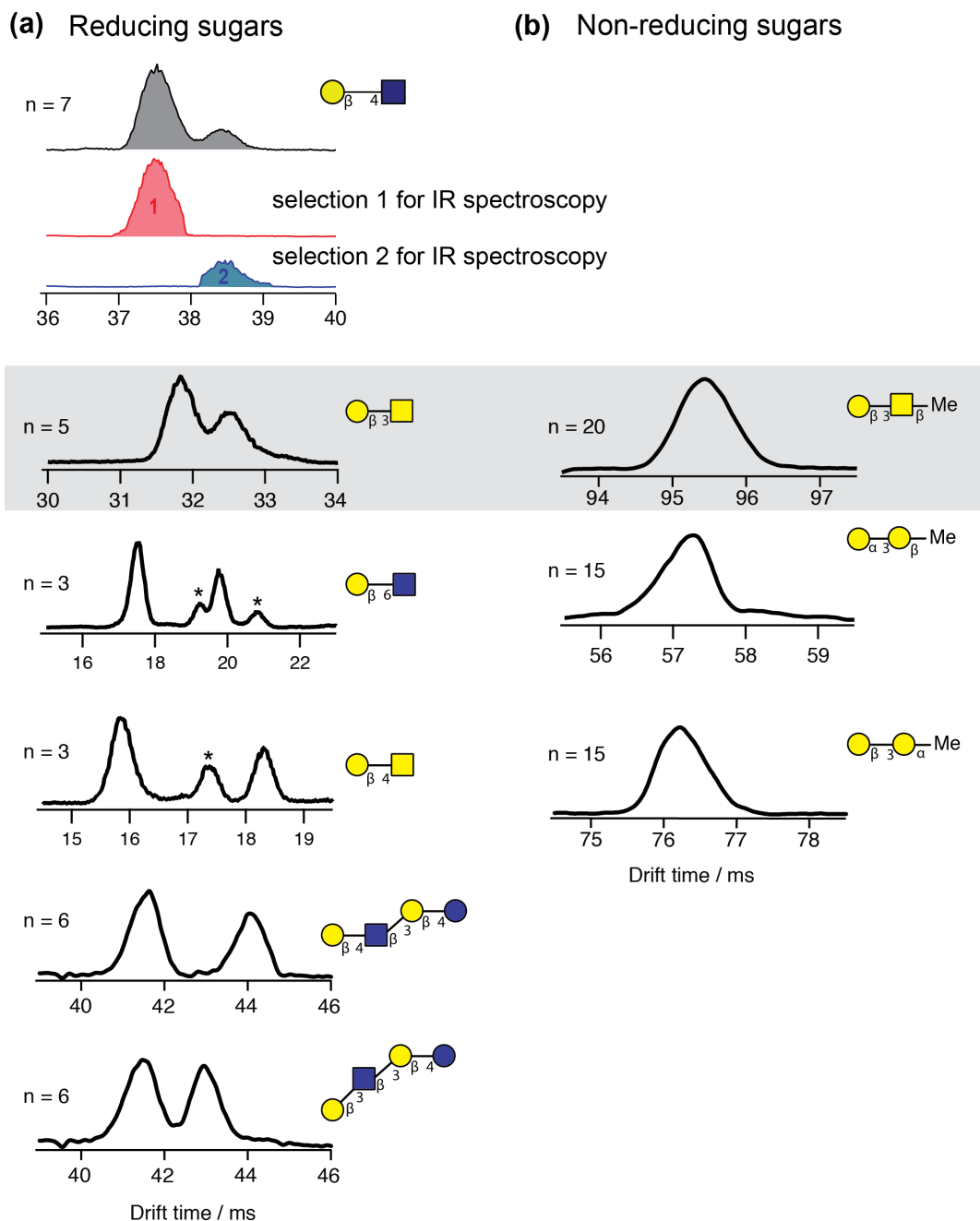


Figure S1: (a) ATDs of glycans with a free reducing-end OH group that are able to form α and β anomers at different number n of IMS cycles on the SLIM board. The type of sugar is annotated in Oxford nomenclature. The top three panels show exemplarily how two different drift peaks are selected into two portions that can then be investigated separately. (b) ATDs of glycans with a methylated reducing-end. As a consequence, the C₁ carbon on the reducing-end of these glycans are locked in their α and β anomeric configuration. An asterisk marks signals that are due to ions of slightly different m/z , which could not sufficiently filtered with the quadrupole MS used.

IR SPECTRA OF MOBILITY-SELECTED DISACCHARIDES

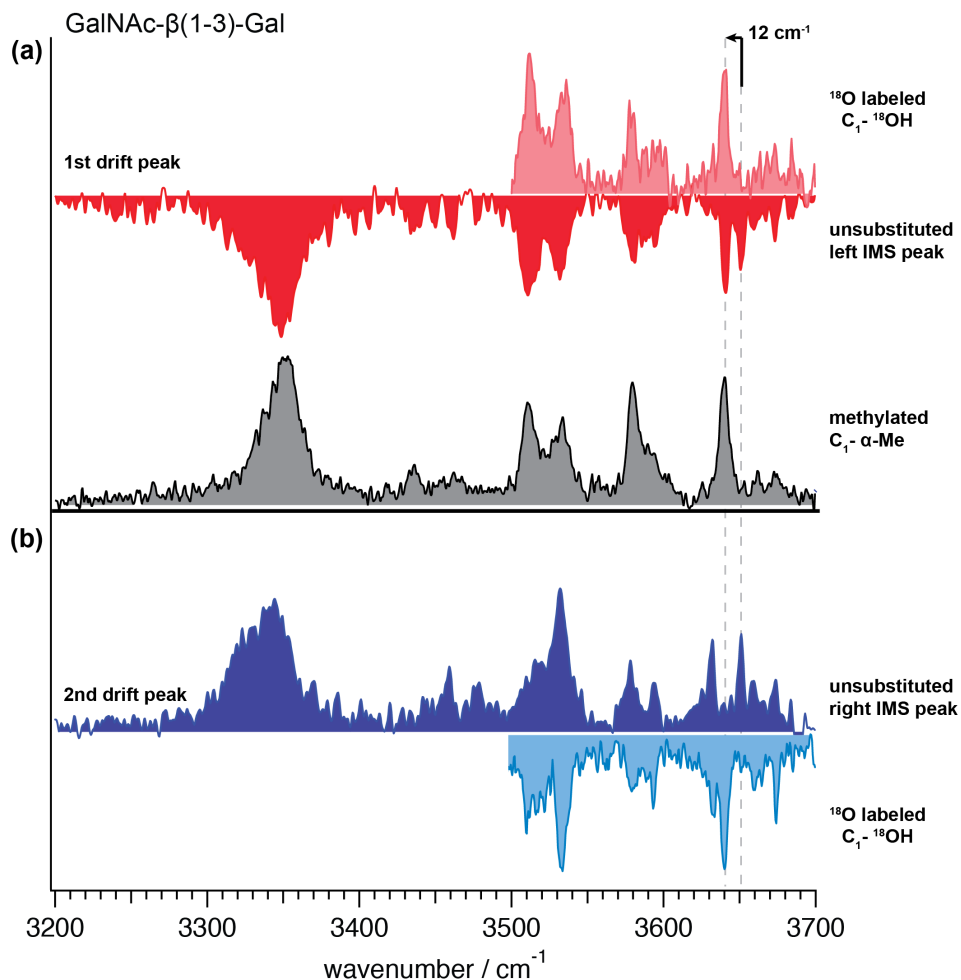


Figure S2: Infrared spectra of the mobility-selected left ((a), dark red) and right ((b), dark blue) drift peak of the disaccharide GalNAc-β(1-3)-Gal and the equivalent species with ¹⁸O-substituted C₁-OH group (light colors), which exhibit a 12 cm⁻¹ redshift in one absorption band. The spectrum of the methylated species GalNAc-β(1-3)-Gal-α-Me (grey) matches the spectrum of the left drift peak in position and intensities of absorption bands, with the exception of the absent C₁-OH oscillator for the methylated species. The left drift peak is thereby identified as the α anomeric form of GalNAc-β(1-3)-Gal and the right peak (by exclusion) as the β anomeric form.

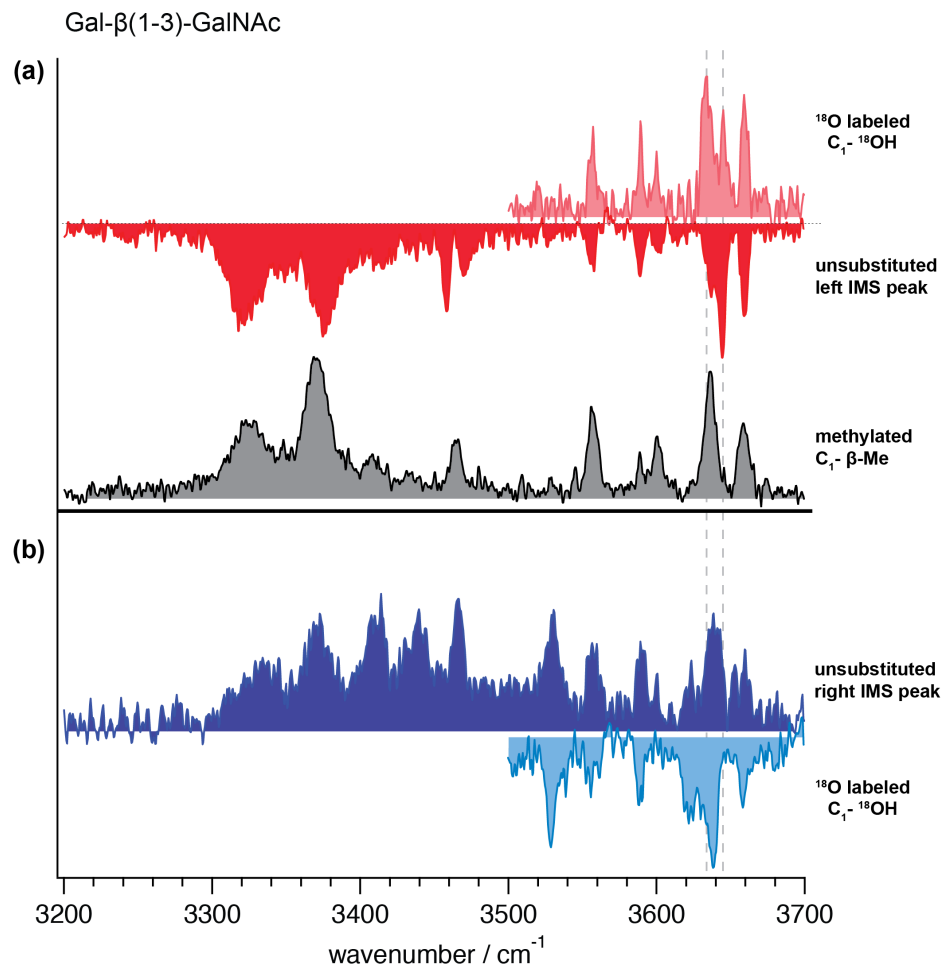


Figure S3: Infrared spectra of the mobility-selected left ((a), dark red) and right ((b), dark blue) drift peak of the disaccharide Gal- β (1-3)-GalNAc and the equivalent species with ^{18}O -substituted $\text{C}_1\text{-OH}$ group (light colors), which exhibit a 12 cm^{-1} redshift in one absorption band. The spectrum of the methylated species Gal- β (1-3)-GalNAc- β -Me (grey) matches the spectrum of the left drift peak in position and intensities of absorption bands, with the exception of the absent $\text{C}_1\text{-OH}$ oscillator for the methylated species. The left drift peak is thereby identified as the β -anomeric form of Gal- β (1-3)-GalNAc and the right peak (by exclusion) as the α anomer.

IR SPECTROSCOPY OF IMS SEPARATED GLUCOSE STRUCTURES

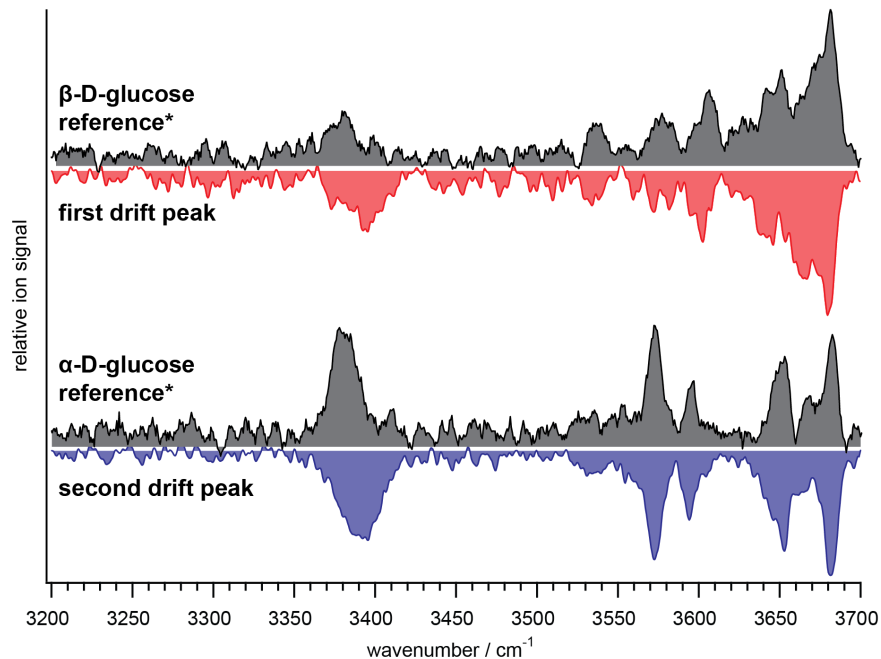


Figure S4: Infrared spectra of the mobility-separated first (red) and second (blue) drift peak of electro sprayed α -D-glucose and previously published reference spectra of the α and β anomers of glucose from a cryogenic, messenger-tagging, double-resonance spectroscopic experiment [1]. The good agreement with the reference spectra leads to the assignment of the α and β anomers to the first and second IMS drift peak, respectively.

GLUCOSE MUTAROTATION

The conversion reaction α -D-glucose $\xrightleftharpoons[k_{\beta}]{k_{\alpha}}$ β -D-glucose can be treated as a first order reaction [2, 3] and the rate equation follows as

$$-\frac{dc_{\alpha}(t)}{dt} = \frac{dc_{\beta}(t)}{dt} = k_{\alpha}c_{\alpha}(t) - k_{\beta}c_{\beta}(t). \quad (S1)$$

Using the substitution for the observable rate $k = k_{\alpha} + k_{\beta}$ and the total concentration c_0 , this can be written as

$$\frac{dc_{\alpha}(t)}{dt} + kc_{\alpha}(t) = k_{\beta}c_0. \quad (S2)$$

Under consideration of the boundary conditions for c_{α} and c_{β}

$$\begin{aligned} c_{\alpha}(t_0) &= c_0 \\ c_{\beta}(t_0) &= 0 \\ c_i(t \rightarrow \infty) &= c_i^{eq} \end{aligned} \quad (S3)$$

the rate differential equation can be solved for $c_{\alpha}(t)$ and $c_{\beta}(t)$, respectively, with

$$\begin{aligned} c_{\alpha}(t) &= c_{\alpha}^{eq} + (c_0 - c_{\alpha}^{eq})e^{-kt} \\ c_{\beta}(t) &= c_{\beta}^{eq} - c_{\beta}^{eq}e^{-kt}. \end{aligned} \tag{S4}$$

The parameters c_{α}^{eq} , c_{β}^{eq} , and k can be obtained from fitting the experimental data describing the abundance of the α and β anomeric forms with these functions. This was done for the data shown in Fig. 5b in the main manuscript by using $c_{\beta}^{eq} = A$ and k as free fit parameters in the fit function $f(t) = A - Ae^{-kt}$. To obtain the specific rate constants we find the relationships

$$\begin{aligned} k_{\beta} &= \frac{c_{\alpha}^{eq}}{c_0}k \\ \text{and } k_{\alpha} &= k - k_{\beta} = \left(1 - \frac{c_{\alpha}^{eq}}{c_0}\right)k. \end{aligned} \tag{S5}$$

REFERENCES

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2. Kendrew, J.C. and Moelwyn-Hughes, E.A.; The Kinetics of Mutarotation in Solution. *Proc R Soc Lond A Math Phys Sci.* **176**, 352-367 (1940).
3. Lin, C.E., Yu, C.J., Chen, C.L., Chou, L.D. and Chou, C.; Kinetics of glucose mutarotation assessed by an equal-amplitude paired polarized heterodyne polarimeter. *J Phys Chem A.* **114**, 1665-1669 (2010).