SUPPORTING INFORMATION:

Cellular Uptake and Intracellular Trafficking of Poly(*N*-(2-Hydroxypropyl) Methacrylamide)

Claudia Battistella, ¹ Romain Guiet, ² Olivier Burri, ² Arne Seitz, ² Stéphane Escrig, ³ Graham W. Knott, ⁴
Anders Meibom^{3,5} and Harm-Anton Klok^{1,*}

¹ École Polytechnique Fédérale de Lausanne (EPFL), Institut des Matériaux et Institut des Sciences et Ingénierie Chimiques, Laboratoire des Polymères, Bâtiment MXD, Station 12, CH-1015 Lausanne, Switzerland.

² École Polytechnique Fédérale de Lausanne (EPFL), Faculté des sciences de la vie, Bioimaging and optics platform, Bâtiment AI, Station 15, CH-1015 Lausanne, Switzerland.

³Laboratory for Biological Geochemistry, School of Architecture, Civil and Environmental Engineering, Ecole Polytechnique Fédérale de Lausanne (EPFL), CH-1015 Lausanne, Switzerland.

⁴ École Polytechnique Fédérale de Lausanne (EPFL), Faculté des sciences de la vie, Bioelectron Microscopy Core Facility, Bâtiment AI, Station 19, CH-1015 Lausanne, Switzerland.

⁵Center for Advanced Surface Analysis, Institute of Earth Sciences, University of Lausanne, CH-1015

Lausanne, Switzerland.

CORRESPONDING AUTHOR: harm-anton.klok@epfl.ch; Tel: + 41 21 693 4866

Equation 1 and 2 were used for the determination of Mander's coefficients M1 and M2 for Ch1 (GFP, EE or L) and for Ch2 (Rhodamine, ¹⁹F-PHPMA-Rhodamine), respectively.¹

$$M_1 = \frac{\sum_{i} GFP_{i,coloc}}{\sum_{i} GFP_i} \tag{1}$$

$$M_2 = \frac{\sum_i Rho_{i,coloc}}{\sum_i Rho_i} \tag{2}$$

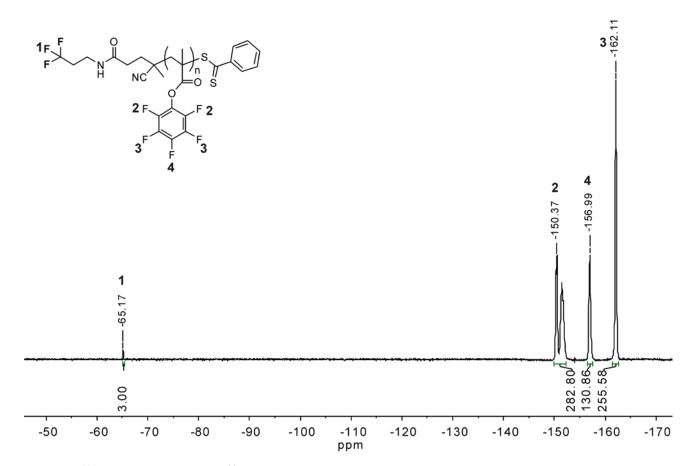


Figure S1. ¹⁹F-NMR spectrum of ¹⁹F-PPFMA in CDCl₃.

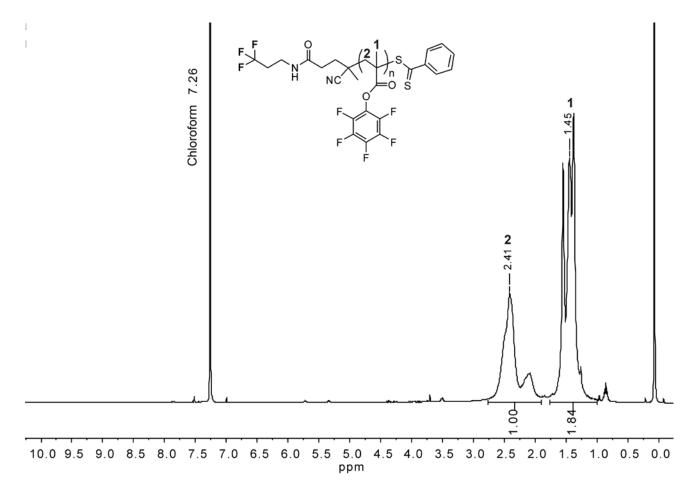


Figure S2. ¹H-NMR spectrum of ¹⁹F-PPFMA in CDCl₃.

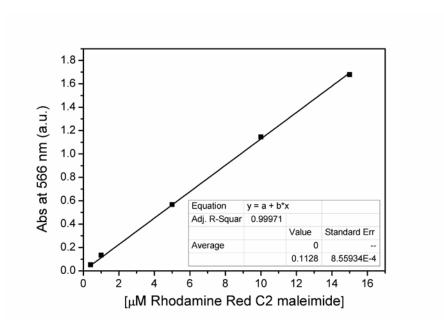
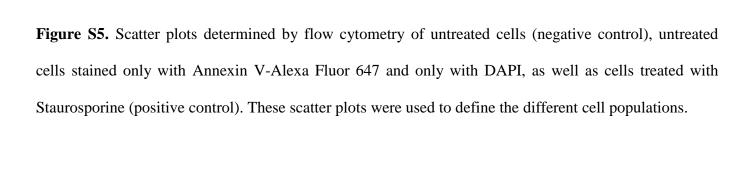


Figure S3. Calibration curve of Rhodamine Red C2 maleimide (ϵ : 0.112890 mM⁻¹ cm⁻¹; Solvent DMSO, $\lambda = 566$ nm).

Figure S4. ¹H-NMR spectrum of ¹⁹F-PHPMA in CD₃OD.



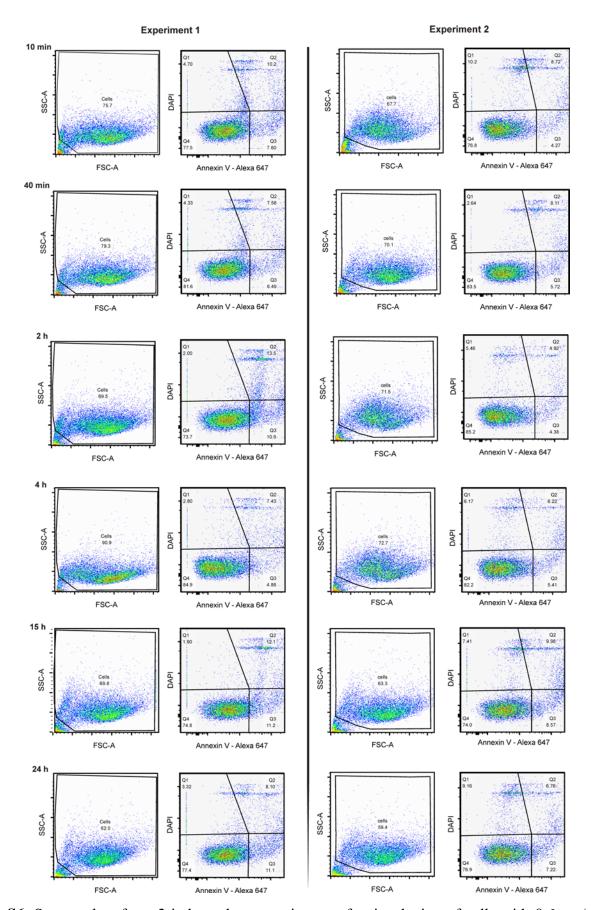


Figure S6. Scatter plots from 2 independent experiments after incubation of cells with 0.6 mg/mL ¹⁹F-PHPMA-Rhodamine for different times as determined by flow cytometry using Annexin V-Alexa Fluor

647/DAPI assay. Q2,3,4 represent the geometric mean fluorescence determined for the three different cell populations (live cells, early apoptotic cells and late apoptotic/necrotic cells).

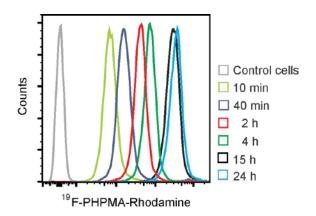


Figure S7. Flow cytometry histogram of Hela cells upon incubation with ¹⁹F-PHPMA-Rhodamine at different incubation times (10 min, 40 min, 2 h, 4 h, 15 h, 24 h) as determined by flow cytometry.

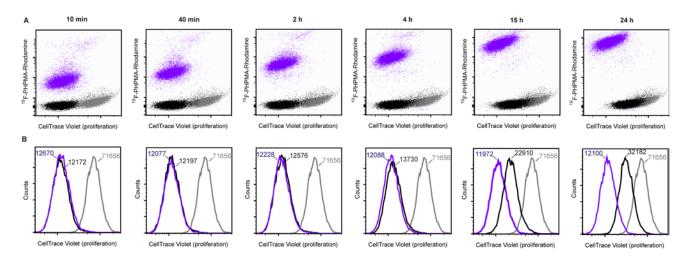
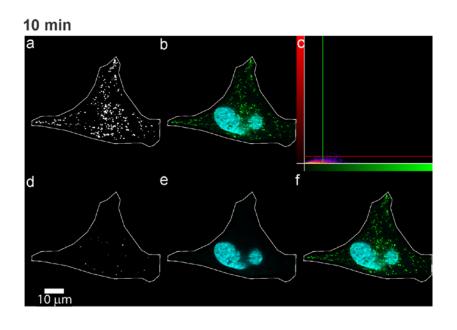
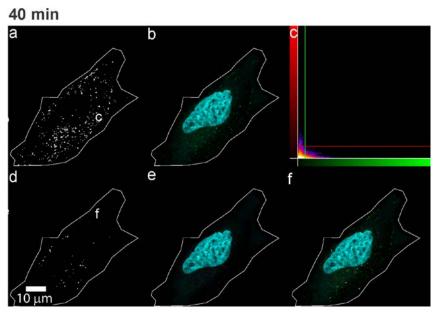
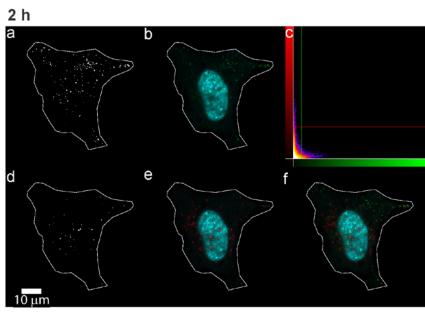
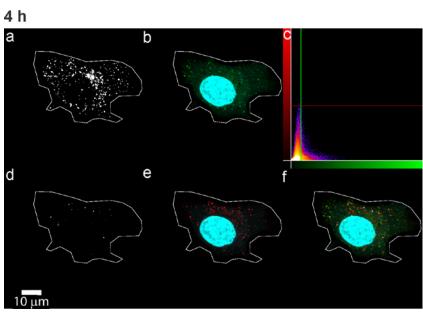


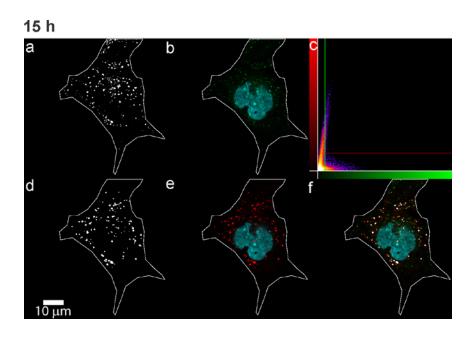
Figure S8. Shift of CellTrace Violet cell fluorescence from the initial (Seeding time, in grey) and the final (End time, in violet) cell fluorescence deriving from cell division during the time frame of the experiment as determined by flow cytometry. Results are represented as both (A) scatter dot plots, which highlight the enhanced ¹⁹F-PHPMA-Rhodamine accumulation in cells over time and as (B) histogram plots, which also report the geometric mean fluorescence values.











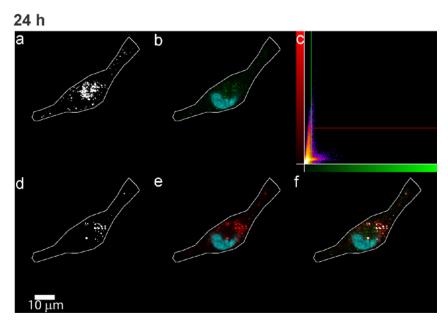
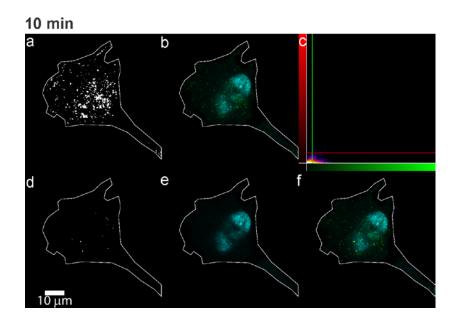
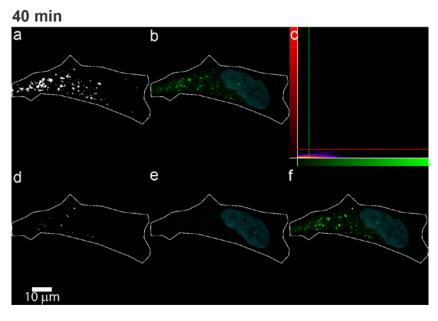
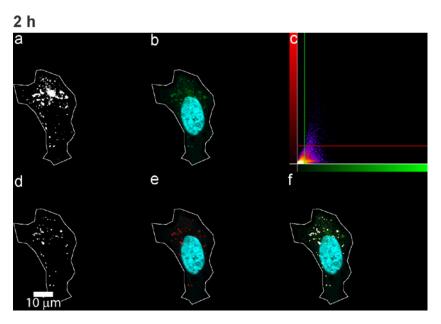
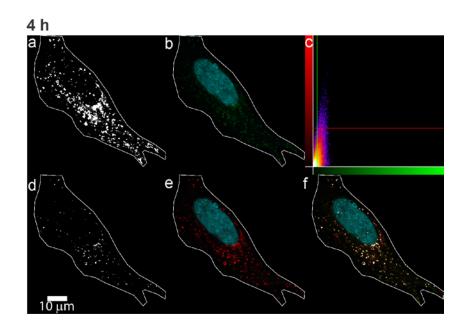


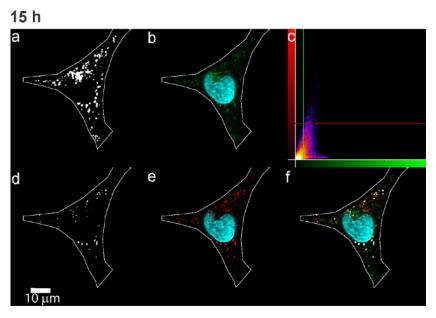
Figure S9. Co-localization studies of ¹⁹F-PHPMA-Rhodamine (magenta) with GFP-EE (green) for 10 min, 40 min, 2 h, 4 h, 15 h and 24 h ¹⁹F-PHPMA-Rhodamine incubation (0.6 mg/mL) for one single experiment. Representative observations show the threshold images (a and d), corresponding to endosomal vesicles (b, in green) and polymer (e, in red), represented together with the DAPI channel (b, e and f, in Cyan). The merged channels with the "co-localized" pixel (f) as well as the fluorogram (c) are also reported. The contours of the manually drawn ROIs are shown as a white line.











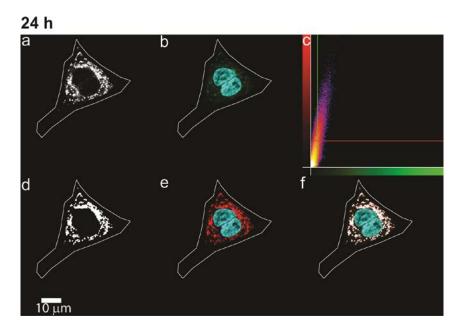


Figure S10. Co-localization studies of ¹⁹F-PHPMA-Rhodamine (magenta) with GFP-L (green) for 10 min, 40 min, 2 h, 4 h and 15 h ¹⁹F-PHPMA-Rhodamine incubation (0.6 mg/mL) for one single experiment. Representative observations show the threshold images (a and d), corresponding to lysosomal vesicles (b, in green) and polymer (e, in red), represented together with the DAPI channel (b, e and f, in Cyan). The merged channels with the "co-localized" pixel (f) as well as the fluorogram (c) are also reported. The contours of the manually drawn ROIs are shown as a white line.

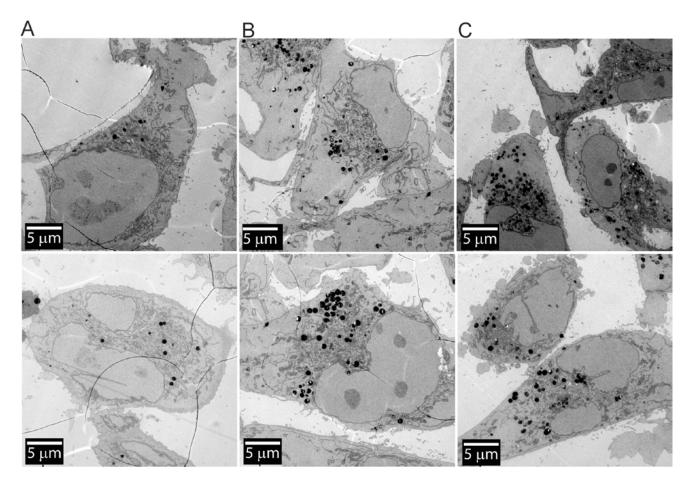


Figure S11. TEM images of (A) control cells, as well as cells incubated for (B) 4 h and (C) 15 h with ¹⁹F-PHPMA-Rhodamine.

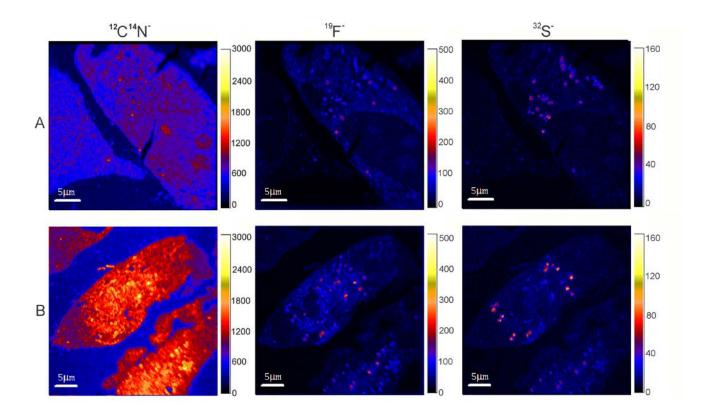


Figure S12. Secondary ion maps of ¹⁴N¹²C⁻, ¹⁹F⁻ and ³²S⁻ in Hela cells treated with 0.6 mg/mL ¹⁹F-PHPMA-Rhodamine for (A) 4 hours and (B) 15 hours.

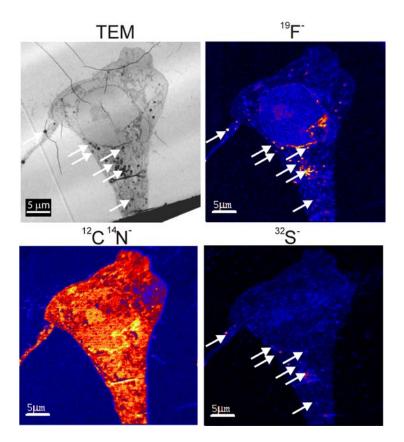


Figure S13. TEM image and the corresponding ¹⁹F, ³²S and ¹²C¹⁴N secondary ion maps obtained after 15 h incubation of Hela cells with 0.6 mg/mL ¹⁹F-PHPMA-Rhodamine. White arrows indicated some of the observed correlations between cell vesicles, ¹⁹F- and ³²S- signals, as an example.

REFERENCES

1. Manders, E. M. M.; Verbeek, F. J.; Aten, J. A., Measurement of Colocalization of Objects in Dual-Color Confocal Images. *Journal of Microscopy-Oxford* **1993**, 169, 375-382.