SCIENTIFIC REPORTS

OPEN

SUBJECT AREAS: ECOLOGY BIOGEOCHEMISTRY

Received 18 February 2014

> Accepted 28 April 2014

Published 15 May 2014

Correspondence and requests for materials should be addressed to C.F.

(christinafasching@ gmx.net) or T.J.B. (tom. battin@univie.ac.at)

* Current address: Department of Ecohydrology, Leibniz-Institute of Freshwater Ecology and Inland Fisheries (IGB), Müggelseedamm 310, 12587 Berlin, Germany.

Microbial degradation of terrigenous dissolved organic matter and potential consequences for carbon cycling in brown-water streams

Christina Fasching¹, Barbara Behounek¹, Gabriel A. Singer^{2*} & Tom J. Battin^{1,2}

¹Department of Limnology and Oceanography, University of Vienna, Althanstrasse 14, A-1090 Vienna, Austria, ²WasserCluster Lunz GmbH, Dr. Carl Kupelwieser Promenade 5, 3293 Lunz am See, Austria.

Streams receive substantial terrestrial deliveries of dissolved organic matter (DOM). The chromophoric (CDOM) fraction of terrestrial deliveries confers the brown colour to streamwater, often understood as browning, and plays a central role in aquatic photochemistry and is generally considered resistant to microbial metabolism. To assess the relevance of terrigenous DOM for carbon fluxes mediated by stream microorganisms, we determined the bioavailable fraction of DOM and microbial carbon use efficiency (CUE), and related these measures to partial pressure of CO_2 in headwater streams spanning across a browning gradient. Fluorescence and absorbance analyses revealed high molecular weight and aromaticity, and elevated contributions from humic-like components to characterize terrestrial CDOM. We found that microorganisms metabolized this material at the cost of low CUE and shifted its composition (from fluorescence and absorbance) towards less aromatic and low-molecular weight compounds. Respiration (from CUE) was related to CO_2 supersaturation in streams and this relationship was modulated by DOM composition. Our findings imply that terrigenous DOM is respired by microorganisms rather than incorporated into their biomass, and that this channelizes terrigenous carbon to the pool of CO_2 potentially outgassing from streams into the atmosphere. This finding may gain relevance as major terrigenous carbon stores become mobilized and browning progresses.

significant amount of the terrestrial primary production is transported laterally along the fluvial continuum from soils to the oceans¹. Evidence suggests that this lateral flux as DOM into inland waters is increasing in North America, North and Central Europe, and in Southeast Asia¹⁻⁵ causing visible browning of these waters⁶. Especially the CDOM fraction of these terrestrial deliveries impacts on freshwater ecosystems as CDOM influences the light regime, primary production and related trophic processes^{4,7}. At high concentration, CDOM can even have implications for drinking water supply⁸. The photochemical degradation of CDOM can also contribute to CO₂ outgassing fluxes from streams, rivers and lakes⁹.

It is therefore of paramount relevance to improve our understanding of the carbon cycling at the terrestrialaquatic interface¹⁰⁻¹². This is especially true for headwater streams, which are most abundant in fluvial networks and tightly connected with the terrestrial milieu. Small streams are also major sources of CO₂ to the atmosphere^{10,13-14} but the contributions of DOM metabolism to CO₂ outgassing from these streams remain poorly understood. The common wisdom that terrestrial CDOM (that is, mostly humics) is resistant¹⁵ to microbial metabolism has shaped our thinking over the last decades¹². This perception is now being re-evaluated based on evidence that chemical recalcitrance and high Δ^{14} C age as commonly assumed properties of terrigenous DOM do not necessarily predict its bioavailability^{12,16-17}. It was shown for instance that terrestrial DOM appears to drive respiration and CO₂ outgassing from the Amazon River^{17,18}.

The aim of this study was to investigate the metabolic fate of terrestrial DOM in headwater streams and to assess its potential contribution to carbon cycling in these streams. Study streams encompassed a brown-colour gradient and we used a space-for-time substitution approach to evaluate the effect of browning on carbon dynamics. We hypothesize that heterotrophic metabolism in stream ecosystems, as depicted by pCO_2 , results from dissolved organic carbon (DOC) concentration, its bioavailable fraction (%BDOC) and the catabolic response of the microorganisms as carbon use efficiency (CUE). We assume that %BDOC and CUE both depend

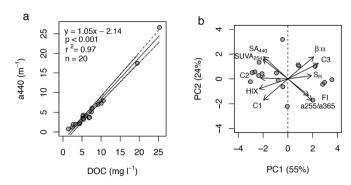


Figure 1 | Quantitative and qualitative aspects of DOM. (a) Colour (as a_{440}) increases with DOC concentration. Dashed lines represent 95% confidence intervals. (b) Principal component analysis (PCA) based on the optical properties of the 20 streams distinguishes terrigenous from autochthonous DOM (PC1). Terrigenous DOM was characterized by high specific absorption at 254 nm (SUVA₂₅₄) and 440 nm (SA₄₄₀), and a high humification index (HIX). Fluorometry revealed two humic-like compounds of terrigenous origin (C1 and C2), while the third component (C3) was putatively associated with new biological production (*Supplementary Information*). High values of the freshness index (β/α) and the fluorescence index (FI) but also lower apparent molecular weight (indicated by a255/a365 and the slope ratio S_R) describe DOM with a more autochthonous character. Arrows are based on PCA structural coefficients.

on DOM composition^{19,20}. To test this hypothesis, we related DOC concentration and DOM optical properties (from fluorescence spectrometry and absorbance) to %BDOC, CUE and to the potential evasion of CO_2 in brown-water streams.

Results

DOM quantity and composition. We investigated attributes of DOM from twenty 1st-order streams in Austria (Table S1 and Table S2) that drain catchments spanning over a gradient in brown colour as a response of varying in peat cover. In fact, the relative contributions of peat and wetlands to land cover are known to imprint on CDOM and its colour as a commonly used indicator of terrestrial DOM deliveries to freshwater ecosystems²¹⁻²⁴. We found streamwater DOC concentration ranging from 1.41 to 24.31 mg C L⁻¹ and CDOM colour (*a*₄₄₀) ranging from 0.78 to 26.64 m⁻¹ (Table S1) across all streams; concentration and colour correlated significantly (Figure 1a).

We analysed the composition of streamwater DOM as inferred from absorbance and fluorescence measures including fluorescent components modelled from excitation emission matrices (EEMs) by parallel factor analysis (PARAFAC)²⁵. A principal component analysis (PCA) based on these measures revealed a gradient of streams where terrigenous DOM mixes in varying proportions with autochthonous DOM. Elevated aromaticity (SUVA₂₅₄), colour (a_{440}), higher degree of humification (HIX) and apparent molecular-weight compounds (S_R), and the humic-like fluorescent components C1 and C2 (Figure 2) are all indicative of terrigenous DOM deliveries to these streams. On the other hand, the freshness index (β/α), low apparent molecular weight and aromaticity, but also the humic-like component C3 (Figure 2) assumedly associated with biological production²⁶, indicate in-stream production of DOM.

Relating the same set of concentration-independent optical measures that describe DOM composition to a_{440} , we found that the terrigenous imprint on DOM increases with brown colour across all streams (Figure 3a). To rule out the possibility of a spurious correlation driven by DOC concentration, we excluded SA₄₄₀ and SUVA₂₅₄ from the analysis and were able to confirm the observed composition shift towards terrigenous DOM paralleled the gradient of brown colour (Figure S1).

DOM biodegradation and CUE. We used classical batch biodegradation assays (20 days) to monitor changes in DOM composition upon microbial degradation and calculated BDOC (as percent of the initial DOC concentration)²⁷ and CUE (as biomass production *versus* biomass production plus respiration). Based on canonical correlations, we did not find any obvious relationship between %BDOC and DOM composition as inferred from its optical properties (Figure 3b and Figure S2). To back up our BDOC measurements we also computed microbial carbon demand from respiration and biomass build-up²⁰, which should essentially equal BDOC. The linear relationship between both measures ($r^2 = 0.80$, p < 0.001, n = 20) (see *Supplementary Information*) supports our BDOC measurements.

In contrast to %BDOC, CUE varied more broadly (4.57% to 37.10%) over all twenty study streams and averaged 12.73 \pm 9.58%; CUE was significantly related to DOM composition (canonical correlation, r = 0.88, p < 0.05, n = 20). CUE increased with putatively fresh DOM of apparently low molecular weight and with a microbial imprint; it declined with increasing contributions from terrigenous DOM (Figure 3c and Figure S3). To test whether coloured DOM affects CUE, we also related the gradient of DOM quality associated with CUE to the gradient of DOM quality associated with

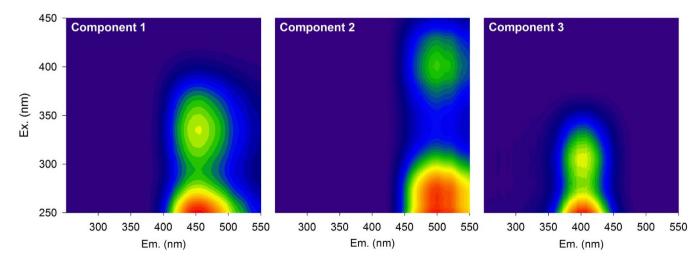


Figure 2 | Three fluorescent components were modeled by parallel factor analysis (PARAFAC) from excitation emission matrices. All components were assigned as humic-like.

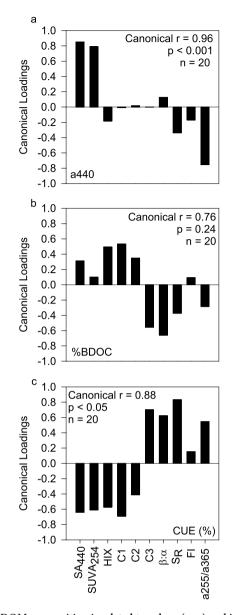


Figure 3 | DOM composition is related to colour (a_{440}) and impacts microbial metabolism. Canonical correlation analyses reveal DOM optical properties most strongly related to (a) colour (a_{440}) , (b) the bioavailable fraction of DOC (%BDOC) and (c) carbon use efficiency (CUE). Canonical loadings (correlations between the one canonical variate of our analysis and the respective variables) indicate the strength and direction of the relationship between the respective constraint and individual optical

brown colour and found a significant correlation (Spearman correlation, r = -0.50, p < 0.05).

measures (see Methods).

DOM optical properties shifted upon microbial degradation as indicated from a decrease of humification (as HIX) (t-test, p < 0.001, n = 20; Figure 4a) and a concurrent increase of the apparent freshness (as β/α) of DOM (t-test, p < 0.001, n = 20; Figure 4b). Especially when the initial terrigenous imprint was strong, apparent freshness increased disproportionally upon degradation. We depicted these shifts in DOM composition occurring during the degradation assays by using the eigenvectors of the PCA describing DOM composition (based on the optical information across the 20 streams before running biodegradation assays) to predict PCA-scores at the end of the experiments (Figure 4c and Figure S4). This revealed that the initial terrigenous imprint on DOM decreased

upon microbial degradation, which resulted in an enhanced autochthonous DOM signature. Relating the length of these shifts in carbon composition to CUE, we found that that DOM with a stronger terrestrial imprint induced lower CUE (Figure 5).

Relationship between BDOC, CUE and streamwater CO₂. To evaluate carbon cycling in our study streams we first determined CO₂ partial pressure (*p*CO₂) in the streamwater and found an average *p*CO₂ of 1,611 ± 1,055 µatm. We also calculated the degree of CO₂ supersaturation (expressed as Δ CO₂, which is the concentration of streamwater CO₂ minus the CO₂ concentration expected at complete atmospheric equilibrium) as a proxy for stream ecosystem metabolism. Positive Δ CO₂ values (73.00 ± 64.48 µmol l⁻¹) indicate that all streams were supersaturated in CO₂ and hence a potential source of CO₂ to the atmosphere. Next we related respiration rates measured in the biodegradation assays and found them closely related to Δ CO₂ in the streams (r² = 0.66, p < 0.001, n = 20). To further illuminate the processes driving this relationship, we propose the following linear model

$$\begin{split} \log &\Delta \text{CO}_2 = b_0 * \log DOC + b_1 * \log \% BDOC \\ &+ b_2 * \log(1 - CUE), \end{split}$$

which is derived from our understanding that stream metabolism is the product of the total amount of DOC *times* its bioavailable fraction (%BDOC) *times* the catabolic response of the microorganisms, as the fraction respired carbon (1-CUE). Regression analysis revealed that DOC concentration and CUE explained 51% (p < 0.01) of the variance in ΔCO_2 across all study sites (Figure S5). Although DOC concentration (beta = 0.45, p < 0.05, sum of squares = 3.91) was the strongest predictor for ΔCO_2 followed by CUE as suggested by its sums of squares (beta = 0.53, p < 0.05, sum of squares = 1.68). BDOC was not a significant predictor (beta = -0.17, p = 0.43, sum of squares = 0.2).

Discussion

DOM quantity and composition. Inland waters receive large amounts of DOM from the terrestrial environment¹. The traditional perception that the chromophoric and humic fraction of this delivery is largely resistant to the microbial metabolism is now changing^{12,16-17}. Our findings expand this changing view by showing the link between the microbial degradation of terrestrial CDOM and potential CO₂ outgassing from small headwater streams.

Our results reveal that terrestrial deliveries not only increase the DOC concentration in streams but also shift the composition of DOM. The DOC concentrations we measured are closely bracketed by values from headwaters in the UK² and North America³⁰, for instance, and colour (a_{440}) is in the range of values (0.69–29.94 m⁻¹) from boreal streams and rivers⁹. PARAFAC analyses revealed that humic-like compounds and elevated aromaticity (as SUVA₂₅₄) characterised terrigenous DOM. Values of SUVA₂₅₄ from our streams are in the upper range of values reported from streams and lakes in North America²⁸ (range: 0.3–8.7 mg L⁻¹ m⁻¹) and closely bracketed by values (3.5 to 4.9 mg L⁻¹ m⁻¹) reported from channels draining tropical peat swamp forests⁵. The browning gradient that we captured with our study streams may thus be representative for other systems influenced by increased terrestrial DOM deliveries.

The relationship between DOC concentration and colour indicates that the variability in DOC concentration across the streams is driven to a large extent by terrigenous DOM deliveries^{9,21,29}. Clearly, this relationship supports the quantitative effect of "brown" and terrigenous DOM deliveries on stream DOC concentration as reported previously^{2,31}. The canonical correlation analysis furthermore revealed that an increase in colour was accompanied by high aromaticity, high apparent molecular weight and high humification



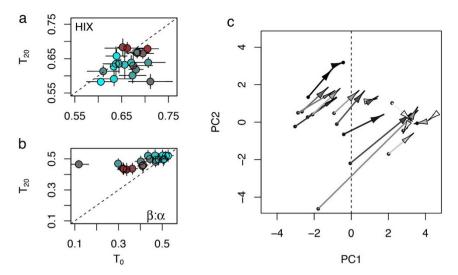


Figure 4 | DOM composition during microbial degradation. (a) The humification index (HIX) and (b) the freshness index (β/α) at start of the degradation assays (T_0) and after the 20-day incubation period (T_{20}). Given is the mean \pm SD (n = 3 replicates) and the dashed line indicates a 1:1 relationship (i.e., no change during degradation) Original DOM composition as derived from the PCA is indicated by colour. Brown colours denote streams with predominantly terrigenous DOM and blue colours represent streams with relatively more autochthonous DOM. (c) Shifts in DOM composition over the 20-day experiments. Starting coordinates of arrows (indicated by black circles) are identical with scores of the PCA based on optical measures (Figure 1b). End coordinates are predicted scores computed with the PCA eigenvectors and the optical measures at the end of the experiments. Lengths of arrows denote the intensity of the shift in DOM composition. The greyscale indicates CUE; ranging from black (lowest CUE) to white (highest CUE).

collectively pointing at the increasing terrigenous character of DOM. This is also supported by lower FI values indicative of allochthonous DOM sources³² and by elevated contributions of the humic-like fluorescence components C1 and C2. Both fluorescence components are of terrestrial origin³³ and often encountered in freshwater eco-systems³⁴, and thus appropriate to track DOM sources³⁵. Our findings corroborate previous studies relating DOM colour to chemical composition, including apparent molecular weight³⁶ or lignin phenols³⁷.

The PCA based on DOM absorbance and fluorescence also suggests that autochthonous compounds imprint to varying degrees on the optical properties of the DOM pool. Such autochthonous DOM compounds may be of recent microbial origin as indicated by ele-

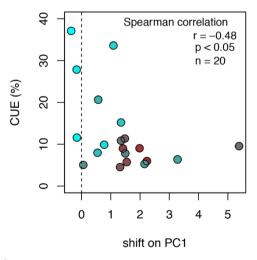


Figure 5 | CUE decreases with the absolute shift in DOM composition along PC1 (i.e., each stream's PC1 score *after* minus the respective score *before* degradation). Original DOM composition as derived from the PCA (PC1) is indicated by colour. Brown colour denotes streams with predominantly terrigenous DOM and blue colour represents streams with relatively more autochthonous DOM.

vated values of the freshness index $(\beta/\alpha)^{38,39}$. High values of the fluorescence index (FI) commonly used as an indicator of DOM source (i.e., terrigenous *versus* microbially derived)³² along with lower apparent molecular weight and aromaticity further support our notion of microbial contributions to the DOM pool.

The relative mix of allochthonous (that is, terrestrial) and autochthonous DOM across the browning gradient studied here possibly affects the susceptibility of the DOM pool to the microbial metabolism in the streams. We could not find any significant effect of DOM composition on its bioavailability (as %BDOC). This seems to run counter to findings from coastal temperate streams, for instance, where protein-like fluorescence predicted %BDOC ranging on average from 12.6% and 30.1%⁴⁰. We attribute this discrepancy to the comparatively low %BDOC values, but close to those reported from other streams draining peat⁴¹, and to their low variability (average \pm SD: 4.55 \pm 2.14%) across our study streams. CUE, however, varied broadly across the streams and was related to DOM composition. This agrees with previous findings¹⁹, showing, for instance, a relationship between CUE and apparent molecular weight (as a254/ a365)⁴². In line with this, CUE increased with putatively fresh DOM of apparently low molecular weight and with a microbial imprint; it declined with increasing contributions from terrigenous DOM. CUE was lower than values reported from various aquatic ecosystems¹⁹, which may be attributable to the largely terrigenous character of DOM as the consumption of terrigenous carbon may support a low but continuous level of metabolic activity⁴³. Our data on nutrient concentration in the biodegradation assays (Table S2) preclude that nutrient limitation affected CUE. This is important to note as nutrient limitation can shift the balance between respiration and biomass production when excess carbon is released by respiration⁴⁴. CUE may also be low in the absence of nutrient limitation. For instance, the relative energy content per unit carbon may decrease with increasing terrestrial imprint due to more oxidized compounds^{19,45}. The degradation of such compounds requires the production of exoenzymes (e.g., phenoloxidases), which increases carbon demand but which may also enhance respiration⁴⁵.

The fact that the gradient of DOM composition associated with CUE paralleled the gradient of DOM composition (that is, CDOM associated with the brown colour) indicates that changes in this CDOM lead to a decrease in CUE. This suggests an impact of brown DOM on carbon processing in streams. In streams with elevated brown colour, a smaller fraction of DOC is incorporated into microbial biomass, while a larger fraction is used for the maintenance of microbial metabolism.

Biodegradation assays revealed that microbial activity apparently imparted an autochthonous signature on an otherwise allochthonous DOM pool characterised by chromophoric and humic-like terrigenous compounds. Our findings suggest that microorganisms removed humic-like compounds while simultaneously producing novel compounds. This is consistent with the perception of the dual role of microorganisms as consumers and producers of DOM⁴⁶. Uptake of humic substances or polyphenolic compounds by microorganisms in streams has been reported previously^{43,45,47}. DOM with a strong initial terrigenous imprint was extensively reworked as indicated by the pronounced shifts towards autochthonous DOM compared to DOM that initially has a more autochthonous signature. The apparent freshness for instance increased disproportionally when the initial terrigenous imprint on DOM was strong, which is reflected in the overall shifts in DOM composition upon degradation. This agrees with the observation of lignin and related phenolic compounds being extensively reworked by microorganisms and possibly causing a complete overturn of the DOM pool within weeks in the Amazon River¹⁷. Moreover, low-molecular-weight DOM of terrigenous origin was readily available and also quantitatively important for microbial metabolism in boreal freshwaters²⁰. However, optical properties of relatively more autochthonous DOM changed comparatively less upon microbial degradation, which may be due to the simultaneous production of novel compounds which may resemble the optical properties of the initial material.

DOM with a strong terrigenous imprint induced lower CUE upon degradation, supporting the results from the canonical correlation analysis. This confirms that this carbon pool fuelled microbial respiration rather than sustaining microbial growth. This unbalance between microbial respiration and growth may impart the microbial imprint on DOM observed during biodegradation. As evoked by the shifts in DOM composition this may occur especially when the initial terrestrial imprint was strong. Our results suggest that elevated catabolism was likely required to maintain microbial metabolism of terrigenous DOM¹⁹.

It is now well established that headwater streams are supersaturated in $CO_2^{9,14,13,48-51}$, yet it remains largely unclear to what extent this CO_2 originates from catchment or from *in situ* respiration¹². While there is evidence that groundwater delivers CO_2 from terrestrial respiration into small headwater streams^{49,50,52}, it is also recognized that these streams are net heterotrophic (that is, they metabolize terrigenous organic carbon)¹⁰. That inland waters are to a large extent net heterotrophic is also supported by surveys showing that lake and stream pCO_2 is linked to *in situ* metabolism of $DOC^{9,48,51}$. In line with these observations, the respiration measured in our biodegradation assays was closely related to streamwater ΔCO_2 , which indicates indeed that metabolism contributes to the build-up of CO_2 in headwater streams. This result is remarkable because it links DOC metabolism to CO_2 supersaturation in the streams despite the fact that these are independent measurements.

To further explore the relationship between respiration and CO₂ supersaturation in the streams we proposed a simple model based on the fact that the fraction of respired DOC can be inferred from BDOC and CUE. Our model revealed DOC concentration and CUE as significant predictors for streamwater ΔCO_2 . DOC concentration may affect streamwater ΔCO_2 on a mass basis, which complies with large-scale surveys from lakes and streams^{48,51}. We recognise that a common terrestrial origin of both DOC and CO₂ may drive to some extent at least the observed relationship between DOC and streamwater CO₂ as reported from Amazonian headwater streams⁵². It is

clear that quantitative contributions of DOC and CUE to CO2 supersaturation cannot be inferred from our model; however, it points to the microbial metabolism of terrigenous DOM as an important driver of CO₂ dynamics in brown-water streams. While DOC represents the potential energy basis for the heterotrophic metabolism, BDOC better reflects the amount of organic carbon that is actually available for microbial metabolism. That %BDOC was not a significant term points to bioavailable DOM as a relatively constrained fraction of the carbon pool, which in fact agrees with the missing relationship between %BDOC and DOM composition. In contrast, model results stress that the coupling of CUE with DOM composition constitutes a control on ΔCO_2 besides DOC quantity. Our findings thus provide evidence that DOM quantity and composition affect CO₂ build-up in headwater streams. The quantity of terrigenous DOM deliveries to streams may increase the pool of carbon that is potentially available for microorganisms, while its composition affects its metabolic fate in the microbial compartment. Coloured DOM was preferentially respired with little trophic transfer to the microbial food web, which may contribute to CO₂ supersaturation and evasion to the atmosphere.

Our findings highlight terrestrial deliveries of chromophoric and humic DOM as a relevant component of the carbon cycle in headwater streams and thus complement recent findings from boreal^{16,20} and tropical¹⁷ systems. Upon entrance into streams, this DOM pool becomes subject to microbial degradation at the cost of low carbon use efficiency and potentially contributes to CO₂ evasion to the atmosphere even without photochemical facilitation. Collectively, these findings contradict the common wisdom that "brown" DOM is resistant to microbial degradation and therefore largely exempt from metabolism en route from terrestrial sources to marine sinks⁹. Our findings may have important implications as terrigenous DOM deliveries into inland waters are predicted to increase and to cause browning³, especially where carbon loss from major stores such as peat and permafrost can be massive as climate change progresses.

Methods

Site description and sampling. We sampled twenty streams (1st-order, Austria) draining catchments with differing coverage of coniferous forest and peat (Table S1 and S2). Streamwater samples for DOC and optical analyses were filtered (Whatman GF/F filters) and stored in clean borosilicate vials. Triplicate streamwater samples were collected for the determination of pCO_2 into 50-ml glass vials pre-conditioned with NaN₃ (0.02% final concentration); at each site we also collected samples for atmospheric CO₂. Streamwater for degradation assays was filtered as DOC samples and collected in clean polypropylene copolymer containers.

Biodegradation assays. We conducted biodegradation assays to determine BDOC and CUE, and to relate them to DOM composition and to the observed shifts in DOM composition during microbial degradation. For each stream we conducted triplicate assays over 20 days (in the dark, 18°C). Sterile-filtered streamwater was inoculated with the native microbial community from the respective stream in 250 ml Schottbottles with a headspace (38%). Samples for DOC concentration, DOM fluorescence and absorbance and CO_2 were obtained at the start of the incubation and at day 3, 6, 10, 15 and 20, respectively. DOM samples were filtered and stored as described above. CO2 samples (10 ml) were collected from the headspace and injected into preevacuated exetainers. Abundance of microbial cells at each time point was determined using epifluorescence microscopy (Zeiss AxioImager) and microbial biomass was estimated from cell size and conversion factors. BDOC was calculated as the loss in DOC concentration over the 20-d incubation period. CUE was calculated as the increase in microbial biomass versus the increase in microbial biomass plus respiration (as dissolved inorganic carbon increase calculated from CO2 measurements over the same time period). Further details are provided in the Supplementary Information.

DOM analyses. DOC concentration was measured using a total organic carbon analyzer (Sievers 5310C, GE, USA). DOM fluorescence and absorbance were determined using an Aqualog (Horiba, USA). Parallel factor analysis (PARAFAC) on excitation emission matrices²⁵ resulted in three humic-like components but no protein-like component; this is plausible given the humic character of the streamwater (Figure 2). Components (C1, C2, C3) are expressed in percent relative to their total fluorescence. From fluorescence we further derived the humification index (HIX) indicative of the extent of humification⁵³, the β/α index indicative of fresh microbially produced DOM⁴¹, and the fluorescence index (FI) as a proxy for DOM source (i.e., terrigenous *versus* microbially derived DOM)⁴². From absorption coefficients we derived the brown colour of DOM $(a_{440})^{54}$, the specific UV absorption (SUVA₂₅₄) as a proxy for aromaticity⁵⁵, and the slope ratio $(S_R)^{36}$ and the ratio of absorption coefficients a_{254} : a_{365}^{42} , which are both related to apparent DOM molecular weight. Further details are provided in the *Supplementary Information*.

Streamwater ΔCO_2 and respiration. CO_2 concentration was measured using gas chromatography (Agilent) directly from the exetainers containing headspace samples from the degradation assays or after equilibration from the headspace (N₂) in the streamwater samples. Anticipating shifts in carbonate fractions during sample equilibration we recalculated dissolved inorganic carbon (DIC) concentration using Henry's law and considering alkalinity and the respective equilibrium constants for the carbonate fractions (adjusted for temperature and ionic strength)^{45,56}. We derived streamwater CO_2 concentration (mol 1⁻¹) from DIC, pH and the equilibrium constants for the carbonate fractions⁵⁶. The degree of CO_2 supersaturation (expressed as ΔCO_2 in µmol 1⁻¹) in the streams is the concentration of streamwater CO_2 minus the CO_2 concentration expected at complete atmospheric equilibrium; it indicates the CO_2 evasion potential from the streams and serves as a proxy for heterotrophic metabolism. Further details are provided in the *Supplementary Information*.

Statistical analyses. Principal component analysis (PCA) using optical measures normalized for DOC concentration served to explore DOM composition and to differentiate bulk DOM characterised by terrigenous or by autochthonous DOM contributions. Three separate canonical correlation analyses identified three linear sets of optical measures (as the respective first canonical axis) most strongly related to a_{440} (brown water colour), %BDOC and CUE as the constraints. Canonical loadings (i.e., correlations between the underlying variables and the one canonical axis) indicate the strength and the direction of the relationship of the individual DOM optical measure to the constraint. The strength of the relationship between the canonical axis and the constraint was computed as the canonical correlation coefficient, significance of each canonical correlation was computed by permutation.

- 1. Regnier, P. *et al.* Anthropogenic perturbation of the carbon fluxes from land to ocean. *Nature Geosci.* **6**, 597–607 (2013).
- Evans, C. D., Monteith, D. T. & Cooper, D. M. Long-term increases in surface water dissolved organic carbon: Observations, possible causes and environmental impacts. *Environ. Pollut.* 137, 55–71 (2005).
- Monteith, D. T. *et al.* Dissolved organic carbon trends resulting from changes in atmospheric deposition chemistry. *Nature* 450, 537–541 (2007).
- 4. Hansson, L. A. *et al.* Food-chain length alters community responses to global change in aquatic systems. *Nat. Clim. Change* **3**, 228–233 (2012).
- Moore, S. et al. Deep instability of deforested tropical peatlands revealed by fluvial organic carbon fluxes. *Nature* 493, 660–664 (2013).
- Roulet, N. & Moore, T. R. Environmental chemistry browning the waters. *Nature* 444, 283–284 (2006).
- Karlsson, J. et al. Light limitation of nutrient-poor lake ecosystems. Nature 460, 506–510 (2009).
- Tuvendal, M. & Elmqvist, T. Ecosystem services linking social and ecological systems: river brownification and the response of downstream stakeholders. *Ecol. Soc.* 16, doi:10.5751/ES-04456-160421 (2011).
- Lapierre, J.-F., Guillemette, F., Berggren, M. & del Giorgio, P. A. Increases in terrestrially derived carbon stimulate organic carbon processing and CO₂ emissions in boreal aquatic ecosystems. *Nat. Commun.* 4, doi:10.1038/ ncomms3972 (2013).
- Battin, T. J. et al. Biophysical controls on organic carbon fluxes in fluvial networks. Nat. Geosci. 1, 95–100 (2008).
- 11. Battin, T. J. et al. The boundless carbon cycle. Nat. Geosci. 2, 598-600 (2009).
- Marín-Spiotta, E. et al. Paradigm shifts in soil organic matter research affect interpretations of aquatic carbon cycling: transcending disciplinary and ecosystem boundaries. Biogeochemistry doi:10.1007/s10533-013-9949-7 (2014).
- 13. Butman, D. & Raymond, P. A. Significant efflux of carbon dioxide from streams and rivers in the United States. *Nature Geosci.* **4**, 839–842 (2011).
- 14. Raymond, P. A. *et al.* Global carbon dioxide emissions from inland waters. *Nature* **503**, 355–359 (2013).
- Schmidt, M. W. *et al.* Persistence of soil organic matter as an ecosystem property. *Nature* 478, 49–56 (2011).
- McCallister, S. L. & del Giorgio, P. A. Evidence for the respiration of ancient terrestrial organic C in northern temperate lakes and streams. *PNAS* 109, 16963–16968 (2012).
- 17. Ward, N. D. *et al.* Degradation of terrestrially derived macromolecules in the Amazon River. *Nat. Geosci.* **6**, 530–533 (2013).
- Mayorga, E. *et al.* Young organic matter as a source of carbon dioxide outgassing from Amazonian rivers. *Nature* 436, 538–541 (2005).
- del Giorgio, P. A. & Cole, J. J. Bacterial growth efficiency in natural aquatic systems. Ann. Rev. Ecol. Syst. 29, 503–541 (1998).
- Berggren, M., Laudon, H., Haei, M., Ström, L. & Jansson, M. Efficient aquatic bacterial metabolism of dissolved low-molecular-weight compounds from terrestrial sources. *ISME J.* 4, 408–416 (2010).
- 21. Stedmon, C. A. *et al.* Dissolved organic matter (DOM) Export to a temperate estuary: seasonal variations and implications of land use. *Estuar. Coast.* **29**, 338–400 (2006).

- Osburn, C. L. & Stedmon, C. A. Linking the chemical and optical properties of dissolved organic matter in the Baltic–North Sea transition zone to differentiate three allochthonous inputs. *Mar. Chem.* 126, 281–294 (2011).
- Fichot, C. G. & Benner, B. The spectral slope coefficient of chromophoric dissolved organic matter (S275–295) as atracer of terrigenous dissolved organic carbon in river-influenced ocean margins. *Limnol. Oceanogr.* 57, 1453–1466 (2012).
- Spencer, R. G. M. et al. Chromophoric dissolved organic matter export from U.S. rivers. Geophys. Res. Lett. 40, 1575–1579 (2013).
- Stedmon, C. A. & Bro, R. Characterizing dissolved organic matter fluorescence with parallel factor analysis: a tutorial. *Limnol. Oceanogr. Meth.* 6, 572–579 (2008).
- Cory, R. M. & Kaplan, L. A. Biological lability of streamwater fluorescent dissolved organic matter. *Limnol. Oceanogr.* 57, 1347–1360 (2012).
- Servais, P., Billen, G. & Hascoet, M. C. Determination of the biodegradable fraction of dissolved organic matter in waters. *Water. Res.* 21, 445–450 (1987).
- Jaffé, R. *et al.* Spatial and temporal variations in DOM composition in ecosystems: The importance of long-term monitoring of optical properties. *J. Geophys. Res.* 113, doi:10.1029/2008JG000683 (2008).
- Spencer, R. G. M. *et al.* The estuarine mixing behaviour of peatland derived dissolved organic carbon and its relationship to chromophoric dissolved organic matter in two North Sea estuaries (U.K.). *Estuar. Coast. Shelf S.* **74**, 131–144 (2007).
- Mullholland, J. M. Dissolved organic matter concentration and flux in streams. J. N. Am. Benthol. Soc. 16, 131–141 (1997).
- Weyhenmeyer, G. *et al.* Selective decay of terrestrial organic carbon during transport from land to sea. *Glob. Change Biol.* 18, 349–355 (2012).
- McKnight, D. M. et al. Spectrofluorometric characterization of dissolved organic matter for indication of precursor organic material and aromaticity. *Limnol.* Oceanogr. 46, 38–48 (2001).
- Coble, P. G., Del Castillo, C. E. & Avril, B. Distribution and optical properties of CDOM in the Arabian Sea during the 1995 Southwest Monsoon. *Deep-Sea Res.* 45, 2195–2223 (1998).
- Stedmon, C. A., Markager, S. & Bro, R. Tracing dissolved organic matter in aquatic environments using a new approach to fluorescence spectroscopy. *Mar. Chem.* 82, 239–254 (2003).
- Stedmon, C. A. *et al.* Characteristics of Dissolved Organic Matter in Baltic Coastal Sea Ice: Allochthonous or Autochthonous Origins? *Environ. Sci. Technol.* 41, 7273–7279 (2007).
- Helms, J. R. et al. Absorption spectral slopes and slope ratios as indicators of molecular weight, source, and photobleaching of chromophoric dissolved organic matter. *Limnol. Oceanogr.* 53, 955–969 (2008).
- 37. Spencer, R. G. M., Aiken, G. R., Wickland, K. P., Striegl, R. G. & Hernes, P. J. Seasonal and spatial variability in dissolved organic matter quantity and composition from the Yukon River basin, Alaska, *Global Biogeochem. Cycles* 22, doi:10.1029/2008GB003231 (2008).
- Parlanti, E., Wortz, K., Geoffroy, L. & Lamotte, M. Dissolved organic matter fluorescence spectroscopy as a tool to estimate biological activity in a coastal zone submitted to anthropogenic inputs. *Org. Geochem.* 31, 1765–1781 (2000).
- Wilson, H. F. & Xenopoulos, M. A. Effects of agricultural land use on the composition of fluvial dissolved organic matter. *Nat. Geosci.* 2, 37–41 (2009).
- Fellman, J. B., Hood, E., Edwards, R. T. & D'Amore, D. V. Changes in the concentration, biodegradability, and fluorescent properties of dissolved organic matter during stormflows in coastal temperate watersheds. *J. Geophys. Res.* 114, doi:10.1029/2008JG000790 (2009).
- Stutter, M. I., Richards, S. & Dawson, J. J. C. Biodegradability of natural dissolved organic matter collected from a UK moorland stream. *Wat. Res.* 47, 1169–1180 (2013).
- Berggren, M., Laudon, H. & Jansson, M. Hydrological Control of Organic Carbon Support for Bacterial Growth in Boreal Headwater Streams. *Microbial Ecol.* 57, 170–178 (2009).
- Guillemette, F. & del Giorgio, P. A. Reconstructing the various facets of dissolved organic carbon bioavailability in freshwater ecosystems. *Limnol. Oceanogr.* 56, 734–748 (2011).
- Hessen, D. O. & Anderson, T. R. Excess carbon in aquatic organisms and ecosystems: physiological, ecological, and evolutionary implications. *Limnol. Oceanogr.* 53, 1685–1696 (2008).
- Berggren, M., Lapierre, J.-F. & del Giorgio, P. A. Magnitude and regulation of bacterioplankton respiratory quotient across freshwater environmental gradients. *ISME J.* 6, 984–993 (2012).
- Guillemette, F. & del Giorgio, P. A. Simultaneous consumption and production of fluorescent dissolved organic matter by lake bacterioplankton. *Environ. Microbiol.* 14, 1432–1443 (2012).
- Volk, C. J., Volk, C. B. & Kaplan, L. A. Chemical composition of biodegradable dissolved organic matter in streamwater. *Limnol. Oceanogr.* 42, 39–44 (1997).
- Sobek, S., Algesten, G., Bergström, A.-K., Jannson, M. & Tranvik, L. J. The catchment and climate regulation of pCO₂ in boreal lakes. *Glob. Change Biol.* 9, 630–64 (2003).
- Teodoru, C. R., del Giorgio, P. A., Prairie, Y. T. & Camire, M. Patterns in pCO2 in boreal streams and rivers of northern Quebec, Canada. *Global Biogeochem. Cyc.* 23, doi:10.1029/2008GB003404 (2009).

- Wallin, M., Buffam, I., Öquist, M., Laudon, H. & Bishop, K. Temporal and spatial variability of dissolved inorganic carbon in a boreal stream network: concentrations and downstream fluxes. *J. Geophys. Res.* 115, doi:10.1029/ 2009JG001100 (2010).
- Lapierre, J. F. & del Giorgio, P. A. Geographical and environmental drivers of regional differences in the lake pCO₂ versus DOC relationship across northern landscapes. J. Geophys. Res. – Biogeosci. 117, 15–24 (2012).
- Johnson, M. S. et al. CO₂ efflux from Amazonian headwater streams represents a significant fate for deep soil respiration. *Geophys. Res. Lett.*, 35, doi:10.1029/ 2008GL034619 (2008).
- 53. Zsolnay, A., Baigar, E., Jimenez, M., Steinweg, B. & Saccomandi, F. Differentiating with fluorescence spectroscopy the sources of dissolved organic matter in soils subjected to drying. *Chemosphere* 38, 45–50 (1999).
- Cuthbert, I. D. & del Giorgio, P. A. Toward a standard method of measuring color in freshwater. *Limnol. Oceanogr.* 37, 1319–1326 (1992).
- Weishaar, J. L. *et al.* Evaluation of specific ultraviolet absorbance as an indicator of the chemical composition and reactivity of dissolved organic carbon. *Environ. Sci. Technol.* 37, 4702–4708 (2003).
- Dickson, A. G., Sabine, C. L. & Christian, J. R. (Eds.) Guide to best practices for ocean CO2 measurements. PICES Special Publication 3, 191 pp. (2007).

Acknowledgments

We are grateful to Hubert Kraill, Markus Dengg and Thomas Fuss for assisting in the lab. We thank Linda Wilhelm for discussion and advice. Financial support came from the Austrian Science Fund (START Y420-B17) to TJB.

Author contributions

C.F., G.A.S. and T.J.B. planned the research. C.F. and B.B. accomplished the fieldwork and the biodegradation experiments. C.F. conducted the CO₂ measurements and all microbial work, B.B conducted the fluorescence spectrometry. C.F. performed the statistical analyses aided by G.A.S. T.J.B. wrote the manuscript with assistance from G.A.S., B.B. and C.F.

Additional information

Supplementary information accompanies this paper at http://www.nature.com/ scientificreports

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Fasching, C., Behounek, B., Singer, G.A. & Battin, T.J. Microbial degradation of terrigenous dissolved organic matter and potential consequences for carbon cycling in brown-water streams. *Sci. Rep.* **4**, 4981; DOI:10.1038/srep04981 (2014).



This work is licensed under a Creative Commons Attribution 3.0 Unported License. The images in this article are included in the article's Creative Commons license, unless indicated otherwise in the image credit; if the image is not included under the Creative Commons license, users will need to obtain permission from the license holder in order to reproduce the image. To view a copy of this license, visit http://creativecommons.org/licenses/by/3.0/