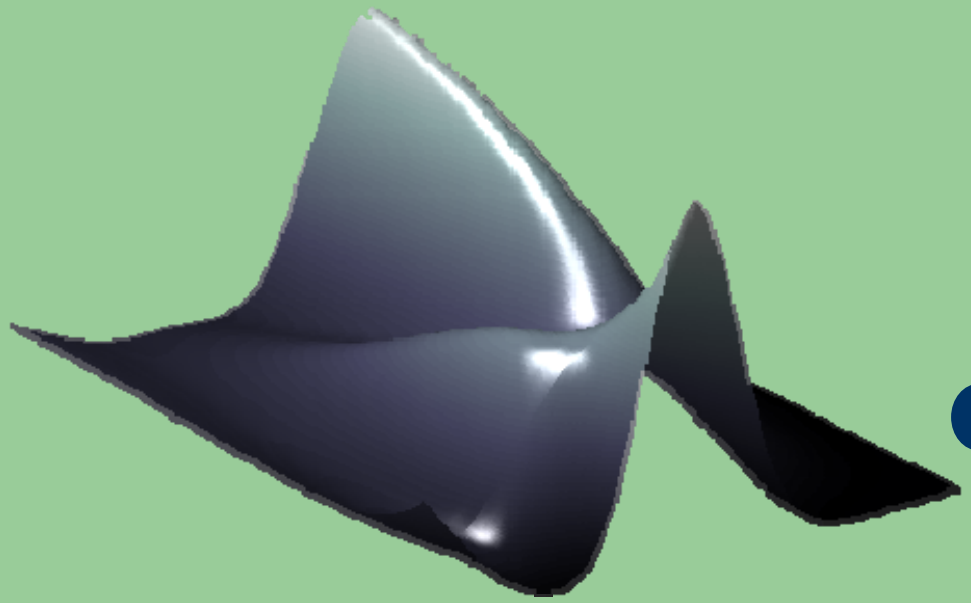
Soft- & Hard Modelling of Kinetic Data



An introduction to the basic principles for the analysis of calorimetric & spectroscopic data

- Julien Billeter & Bobby Neuhold

ETH
Eidgenössische Technische Hochschule Zürich
Swiss Federal Institute of Technology Zurich

Safety and Environmental Technology Group

Kinetic Data Structures

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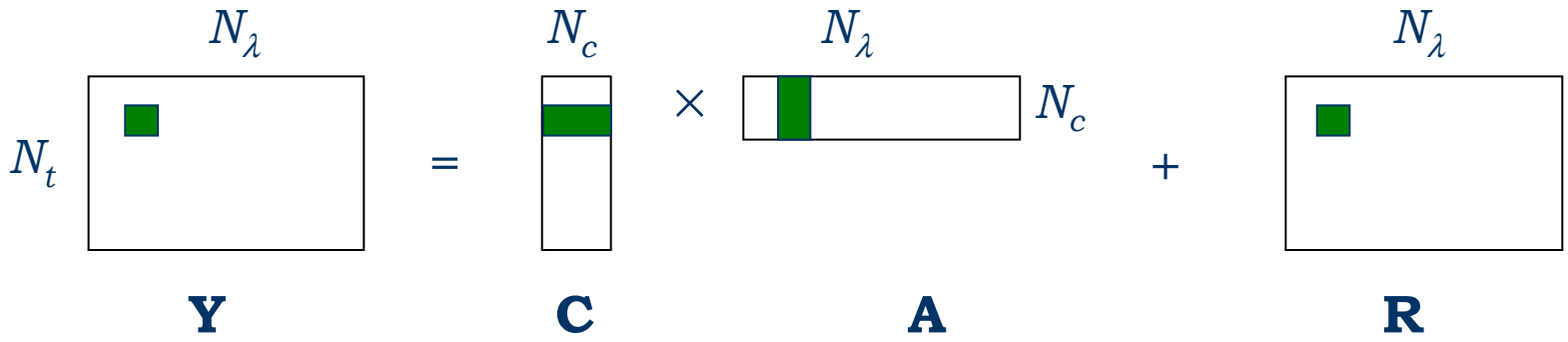

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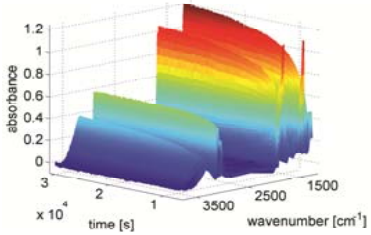
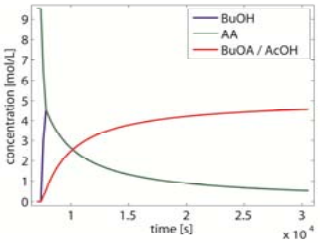
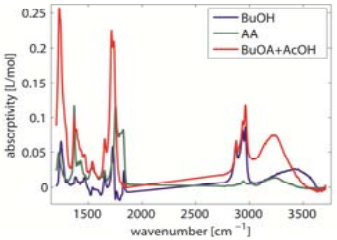


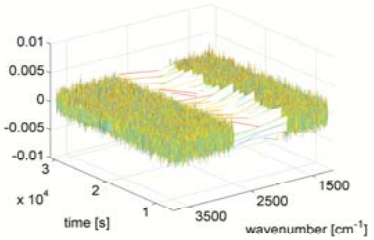
Y



C



A



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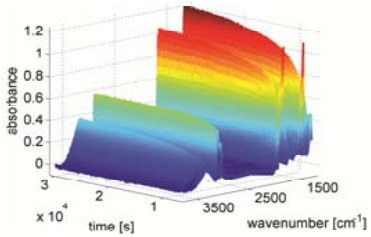
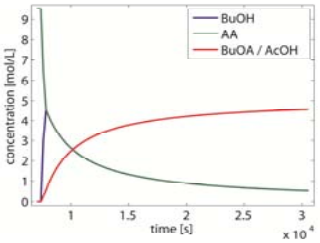
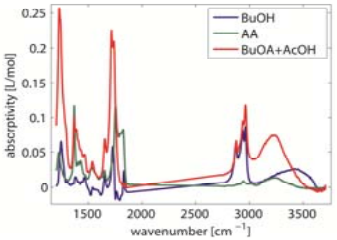
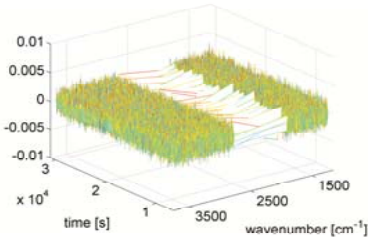


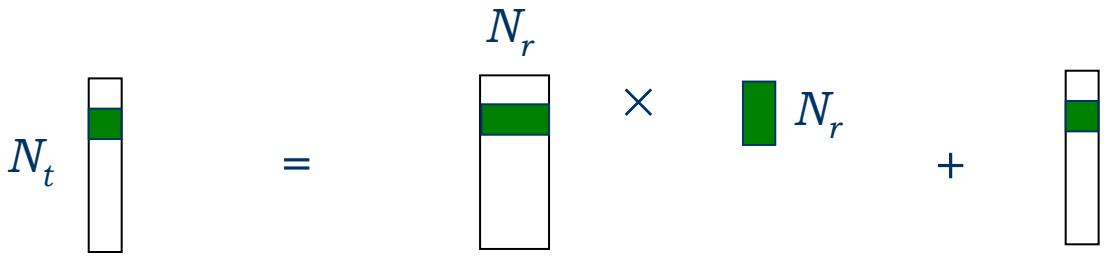






Y **C** **A** **R**

• **Calorimetry**: heat release (**q**) is proportional to the change in the reaction extent ($d\xi/d\mathbf{t}$)



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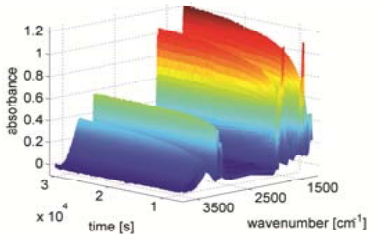

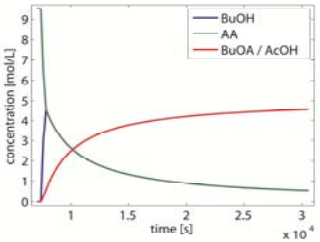

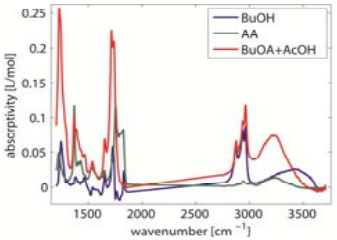

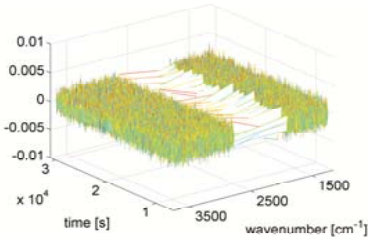

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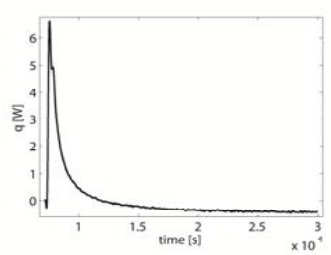
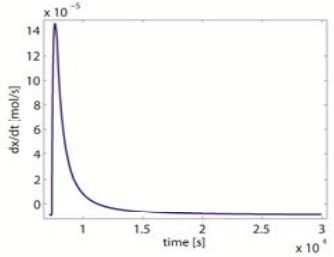
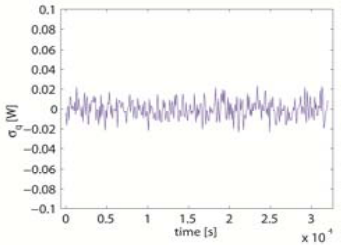




43 kJ/mol



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Soft- & Hard Modelling of Kinetic Data

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Soft Modelling (Part 1)

- Modelling experimental kinetic data based only on simple ‘a priory’ knowledge on the data structure and the results
  - Multivariate (multi wavelength spectroscopic) data and linear dependence of concentrations and data signal (Beer’s law)
    - non-negativity of concentrations and species spectra
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Hard Modelling (Part 2)

- Modelling experimental kinetic data based on a parameterised physical-chemical ‘hard’ model
- rate law defines the concentration profiles of the contributing species as a function of the rate constants
- applicable to both, calorimetry & spectroscopy (univariate & multivariate)

Soft- & Hard Modelling of Kinetic Data

Part 1: Soft Modelling

Soft Modelling of Spectroscopic Kinetic Data

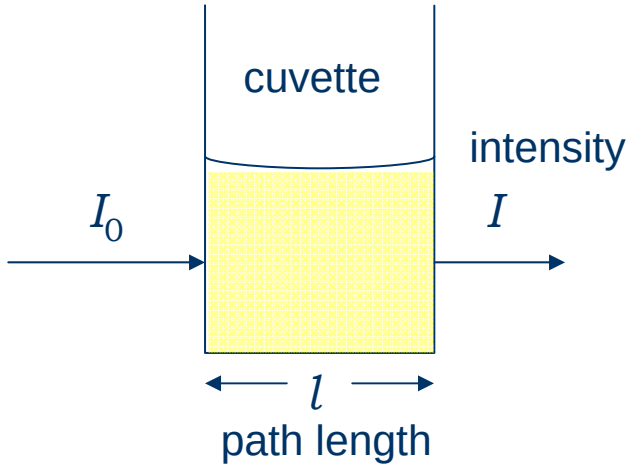
Topics

- Absorption spectroscopy
  - Beer's law in elegant matrix notation (**Y** = **C** $\times$ **A**)
  - Non-unique factorisation of **Y** / rotational ambiguity
- Principal Component Analysis (**PCA**)
  - Abstract Factor analysis (**AFA**) by Singular Value Decomposition (**SVD**)
  - Chemical rank of the measurement matrix
  - The number of absorbing species
- Evolving Factor Analysis (**EFA**)
  - Evolutionary rank analysis by repeated SVD of sub matrices of **Y**
  - The 'Appearance' & 'Disappearance' of absorbing species
- Multivariate Curve Resolution by Alternating Least-Squares (**MCR-ALS**)
  - Model-free iterative decomposition of **Y** = **C** $\times$ **A** + **R**
  - Ideas, principles, limitations

Absorption Spectroscopy – Beer's Law

Absorbance *y* at wavelength λ:

$$y_\lambda = -\log\left(I/I_0\right)_\lambda$$



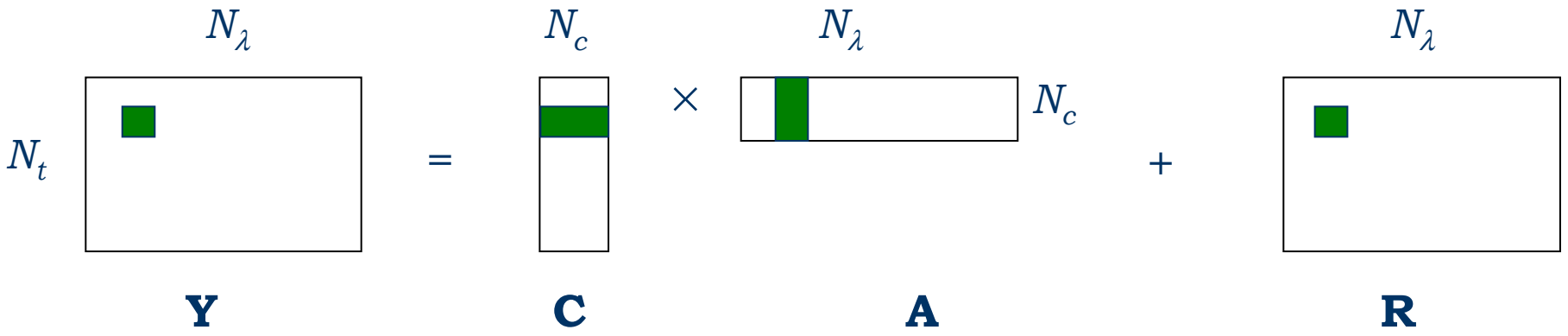
- Absorbance signal y_λ is linearly dependent on contributing species concentrations c_k, the corresponding coefficients are the molar absorptivities $a_{k,\lambda}$ that form the pure species spectra

Beer's Law:

$$y_\lambda = \sum_{k=1}^{N_c} c_k a_{k,\lambda} \times l$$

For simplicity:
path length l =1

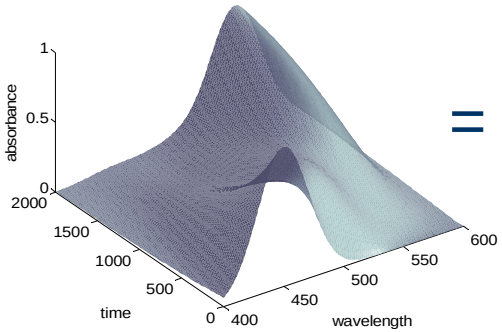
Beer's law in elegant matrix notation



$$y_{t,\lambda} = \sum_{k=1}^{N_c} c_{t,k} a_{k,\lambda} + r_{t,\lambda}$$

Rotational Ambiguity

e.g. A→B→C



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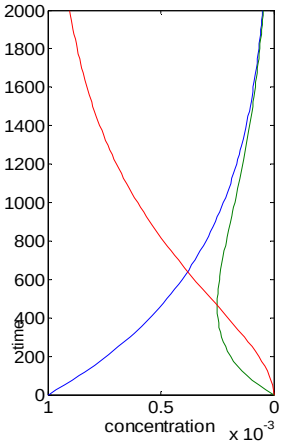

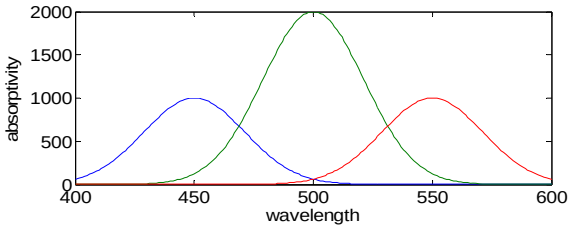

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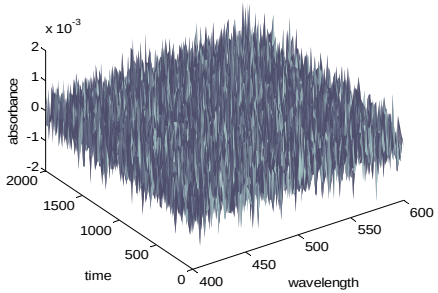

$$\mathbf{Y} = \mathbf{C} \times \mathbf{A} + \mathbf{R}$$

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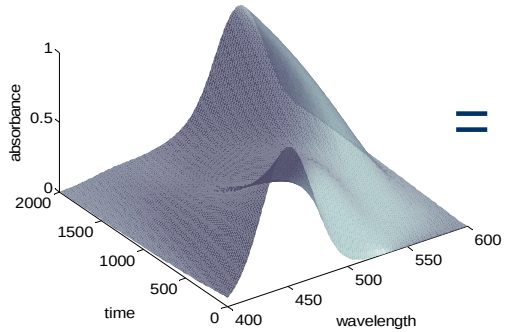

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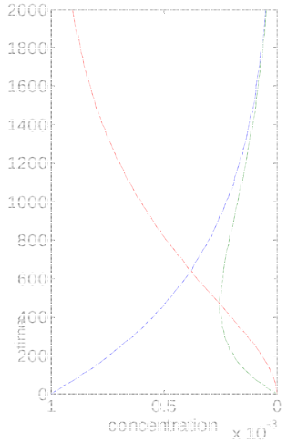

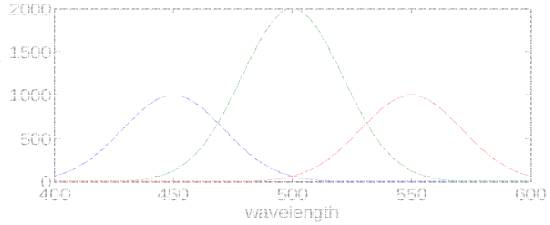

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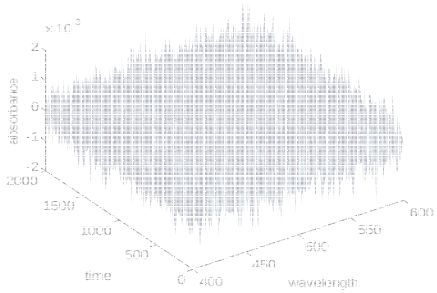

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Goal: Find concentration profiles **C** and species spectra **A** such that the residuals **R**=**Y**-**CA** become small only using a 'soft model', i.e. by linear factorisation

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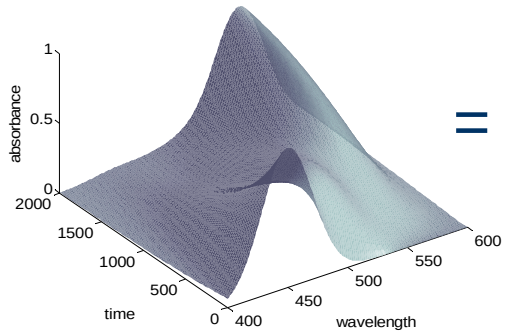

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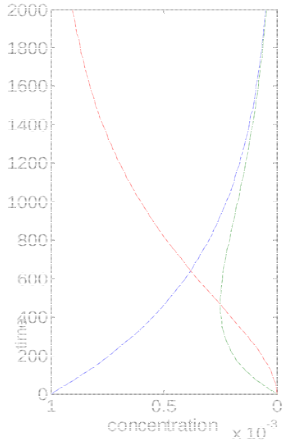

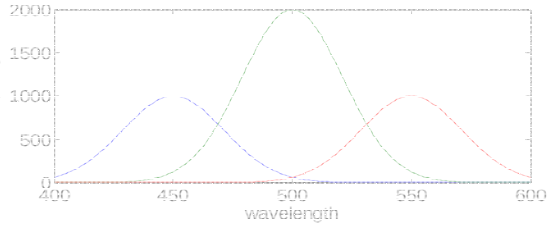

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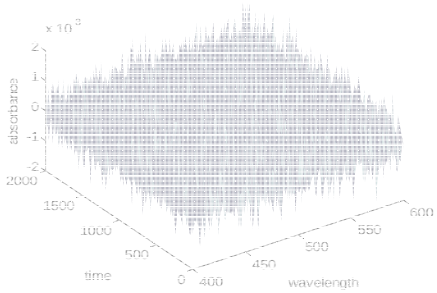

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Problem: Factorisation is not unique (rotational ambiguity)

Major Soft-Modelling Classes

- By using appropriate ‘soft’ restrictions on **C** and **A**, e.g. non-negativity, windows of existence, closure, unimodality, known spectra, the number of possible solutions can be reduced, sometimes can even lead to a unique solution for **C** & **A**

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- There are 2 major classes

Factor Analysis (AFA) based

$$\mathbf{Y} = \bar{\mathbf{U}}\bar{\mathbf{S}}\bar{\mathbf{V}} = \bar{\mathbf{U}}\mathbf{T}\mathbf{T}^{-1}\bar{\mathbf{S}}\bar{\mathbf{V}} = \mathbf{C}\mathbf{A}$$

Find $\mathbf{T}$ such that

$$\mathbf{C} = \bar{\mathbf{U}}\mathbf{T}, \text{ and } \mathbf{A} = \mathbf{T}^{-1}\bar{\mathbf{S}}\bar{\mathbf{V}}$$

Alternating Least Squares (ALS) based

Start from some guessed **C**,

then recalculate **A** and **C** until satisfied:

$$\mathbf{A} = (\mathbf{C}^t\mathbf{C})^{-1}\mathbf{C}^t\mathbf{Y} = \mathbf{C}^+\mathbf{Y}$$

$$\mathbf{C} = \mathbf{Y}\mathbf{A}^t(\mathbf{A}\mathbf{A}^t)^{-1} = \mathbf{Y}\mathbf{A}^+$$

Principal Component Analysis (PCA)

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- One very well defined solution is the one received from Abstract Factor Analysis (AFA) using Singular Value Decomposition (SVD)

$$\underset{N_t}{\boxed{\mathbf{Y}}}\ \overset{N_\lambda}{=}\ \underset{}{\boxed{\mathbf{U}}}\ \underset{}{\boxed{\mathbf{S}}}\ \underset{N_\lambda}{\boxed{\mathbf{V}}} = \underset{}{\boxed{\mathbf{US}}}\ \underset{N_\lambda}{\boxed{\mathbf{V}}} = \underset{N_c}{\boxed{\mathbf{C}}}\ \underset{N_\lambda}{\boxed{\mathbf{A}}}$$

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in Matlab:

```
[U,S,Vt]=svd(Y,0);
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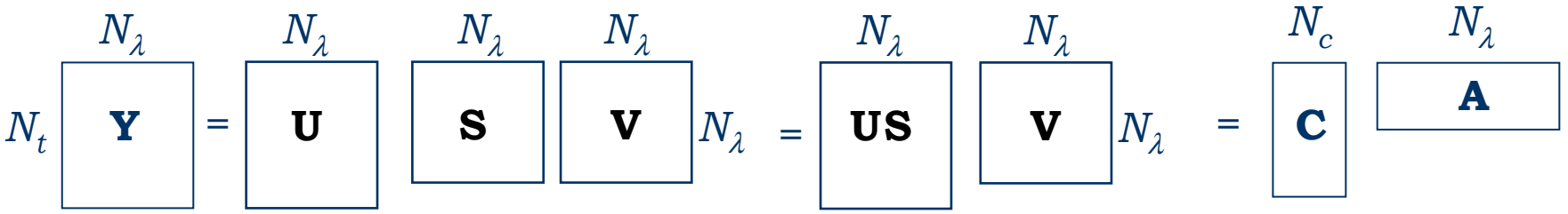

- Some properties of **U**, **S** and **V**

in Matlab:

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$$\mathbf{Y}\mathbf{Y}^t\mathbf{U} = \mathbf{U}\boldsymbol{\Lambda}$$

$$\mathbf{Y}^t\mathbf{Y}\mathbf{V}^t = \mathbf{V}^t\boldsymbol{\Lambda}$$

$$\boldsymbol{\Lambda} = \mathbf{S}^2$$

$$\mathbf{U}^t\mathbf{U} = \mathbf{V}\mathbf{V}^t = \mathbf{I}$$

columns of **U** (rows of **V**) are eigenvectors of $\mathbf{Y}\mathbf{Y}^t$ ($\mathbf{Y}^t\mathbf{Y}$)

S is a diagonal matrix with the square root of their eigenvalues

U and $\mathbf{V}^t$ are orthonormal

Chemical rank – number of absorbing species

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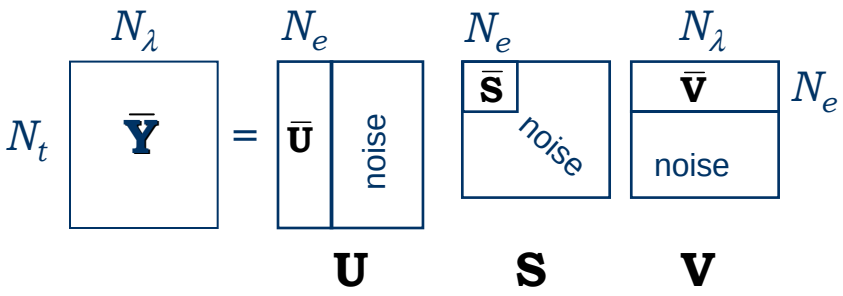
- Many of them just represent ‘noise’ and can be neglected; the significant ‘factors’, the Principal Components, are retained in $\bar{\mathbf{U}}$ and $\bar{\mathbf{V}}$ and form ‘abstract’ concentration profiles and spectra



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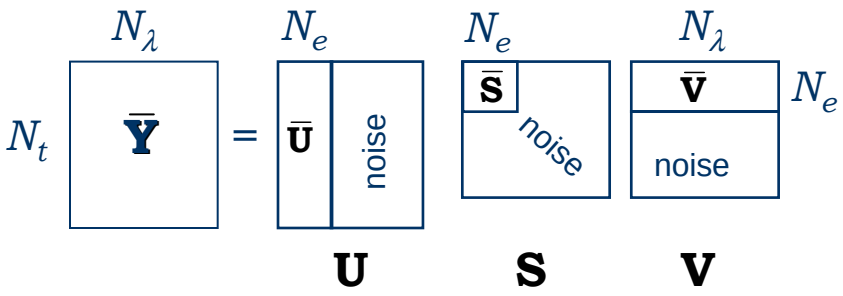

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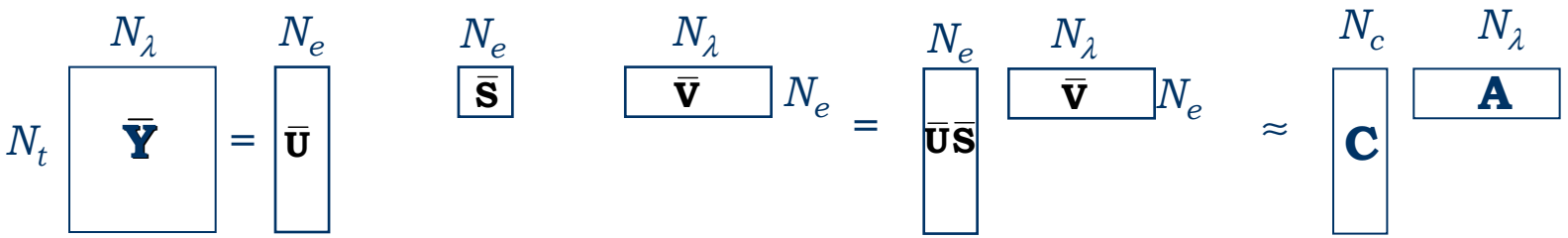
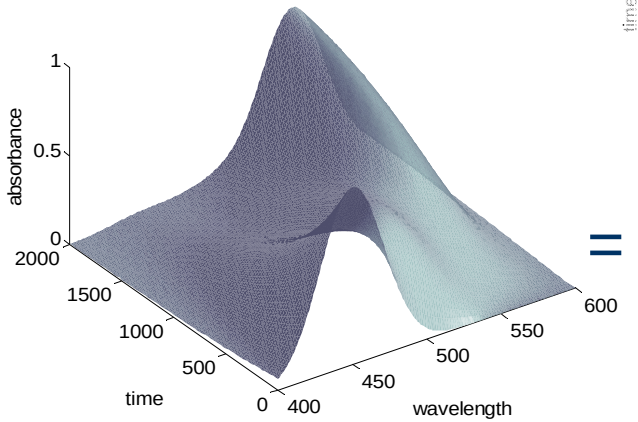


• The number of significant singular(eigen) values and –vectors is the chemical rank of **Y** and a 1st estimate on the number of absorbing species

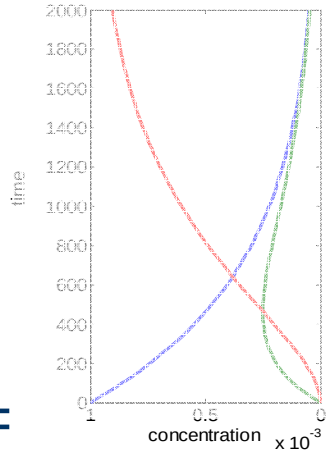
Chemical rank – number of absorbing species

e.g. A→B→C

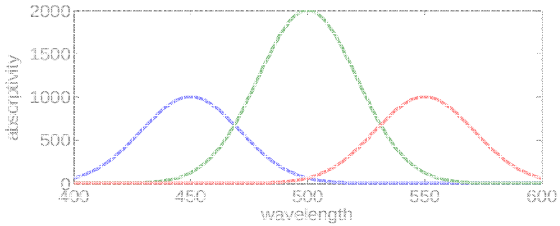


Y

=

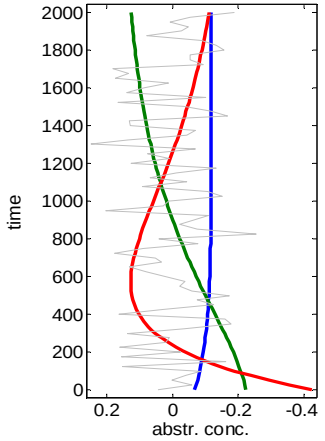


C

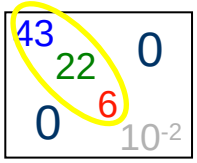


A

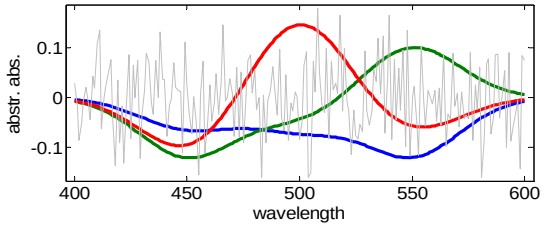
+ **R**



Ū



S̄



V̄

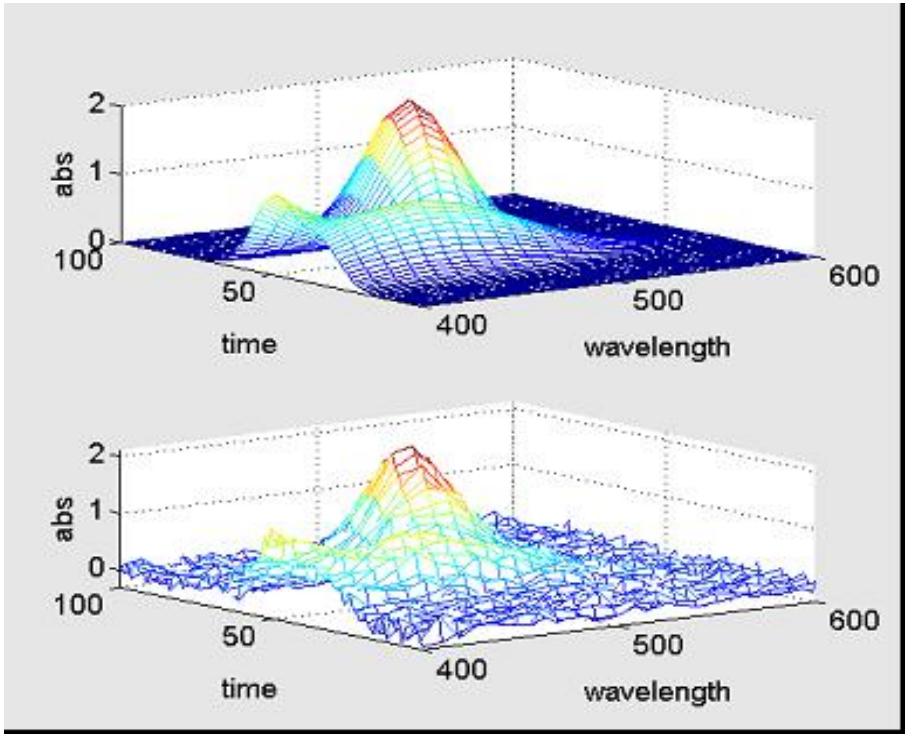
+ $\mathbf{R}_{PCA}$

→ rank(**Y**) = 3

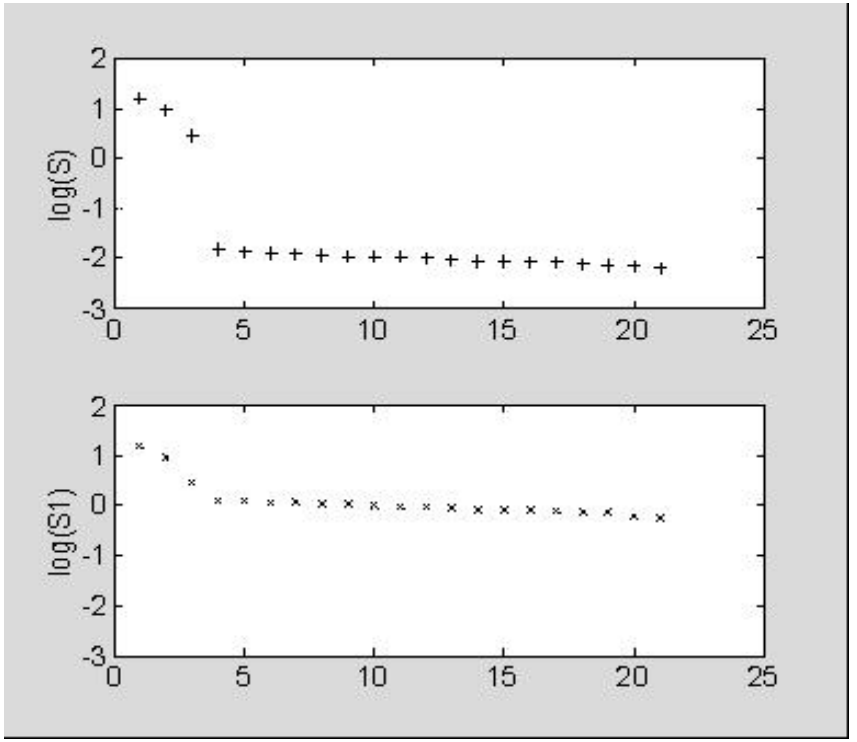
→ 3 absorbing species

Chemical rank – number of absorbing species

Y ($N_t \times N_\lambda$)

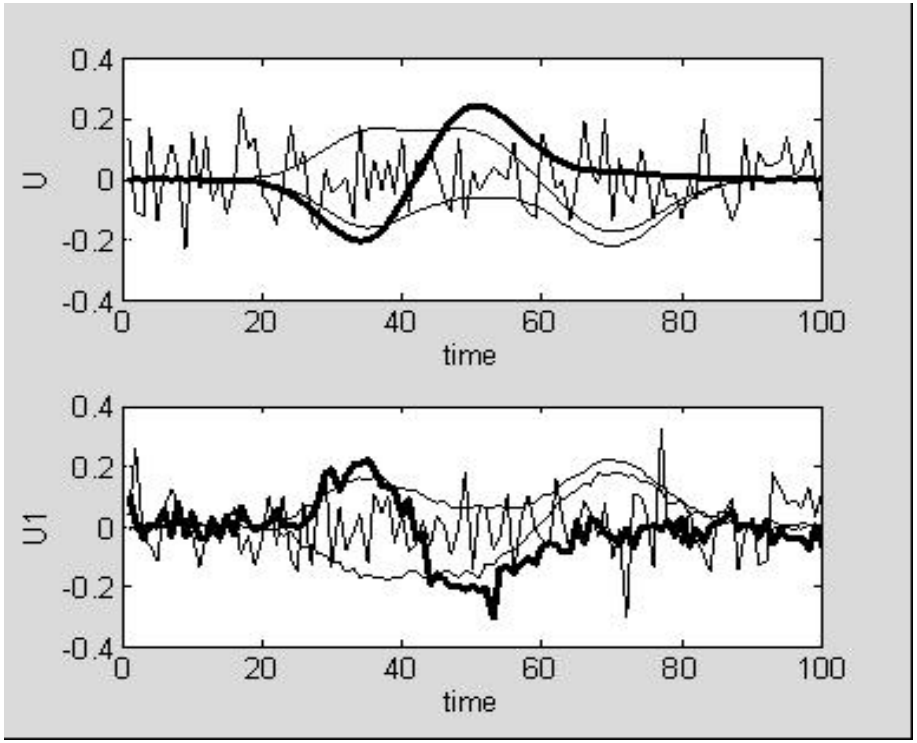


log($s_{i,i}$) vs i



The noise level in the data matrix **Y** determines the drop in the magnitude from significant to insignificant singular values

Chemical rank – number of absorbing species



The noise level in the data matrix **Y** also determines the remaining noise in the significant singular vectors

The signs of the singular vectors can interchange between **U** and **V**)

Geometric interpretations

Geometric interpretations

- e.g. 2 species, A→ B
- the (significant) eigenvectors $\mathbf{v}_{1,:}$ and $\mathbf{v}_{2,:}$ form an orthonormal base in the same ‘plane’ as the pure species spectra $\mathbf{a}_{1,:}$ and $\mathbf{a}_{2,:}$

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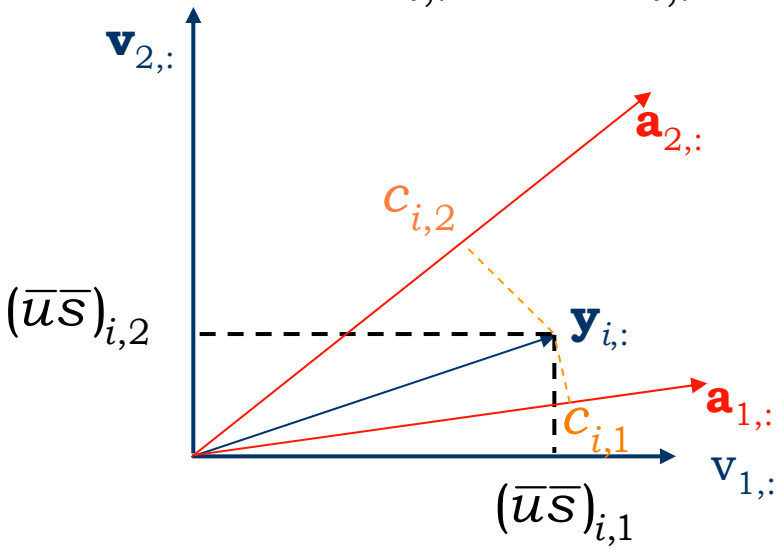
$$\mathbf{y}_{i,:} \approx (\overline{\mathbf{u}}\overline{\mathbf{s}})_{i,:}\overline{\mathbf{V}} \approx \mathbf{c}_{i,:}\mathbf{A}$$



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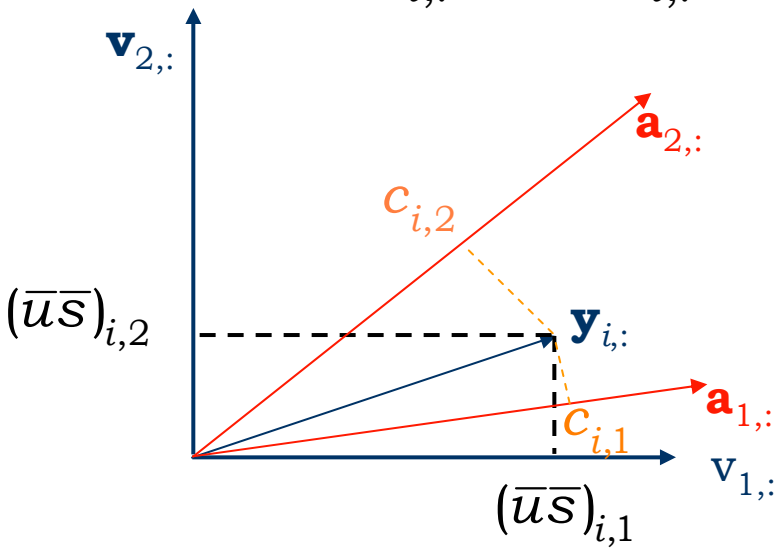

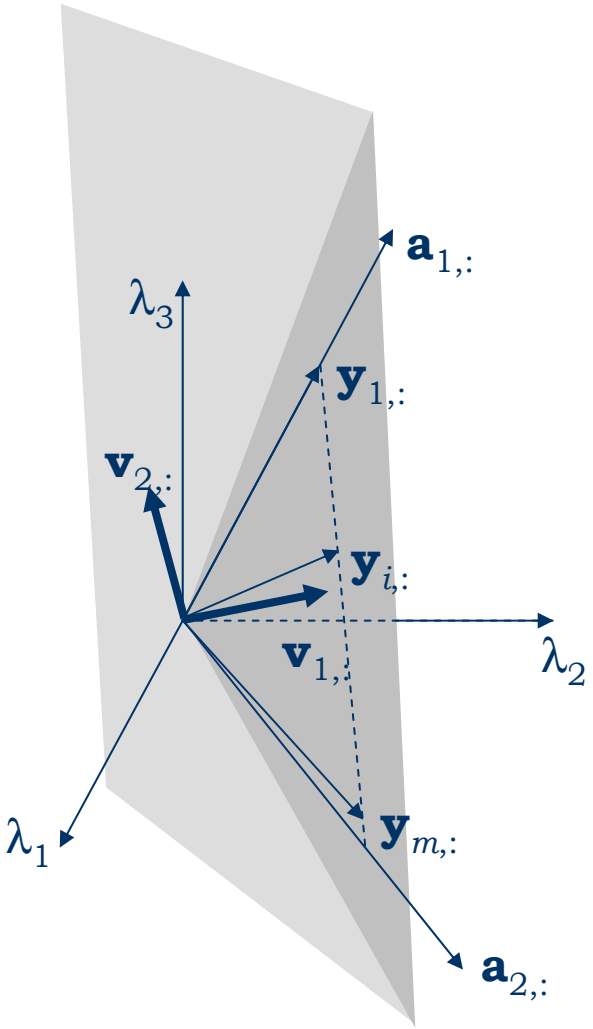

Some noise reduction

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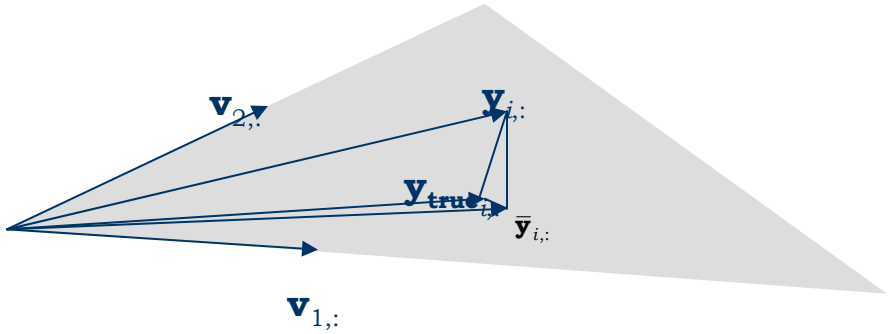
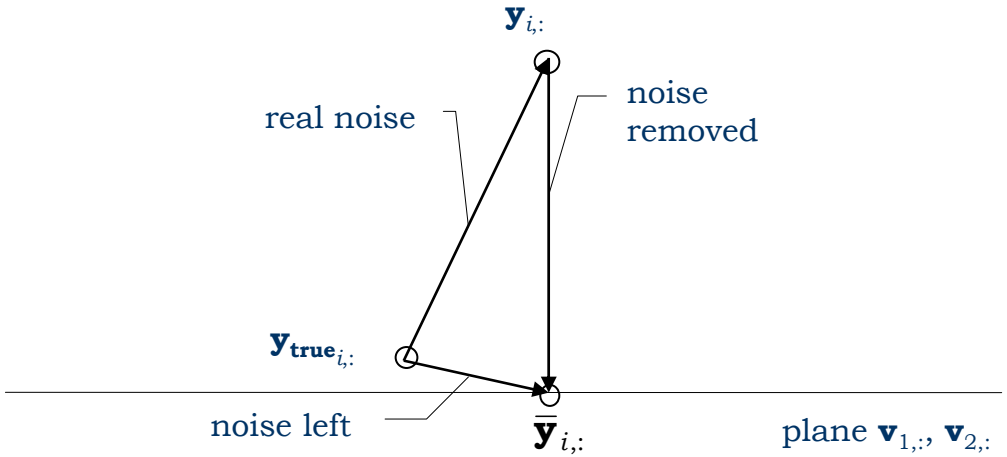
$$\mathbf{Y} = \bar{\mathbf{Y}} + \mathbf{R}_{PCA} = \sum_{i=1}^{N_e} \mathbf{u}_{:,i}\mathbf{s}_{i,i}\mathbf{v}_{i,:} + \sum_{j=N_e+1}^{N_\lambda} \mathbf{u}_{:,j}\mathbf{s}_{j,j}\mathbf{v}_{j,:} = \sum_{k=1}^{N_c} \mathbf{c}_{:,k}\mathbf{a}_{k,:} + \mathbf{R}_{noise}$$





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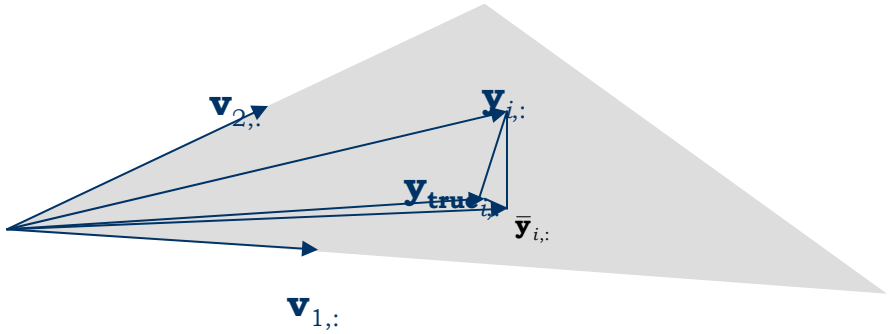

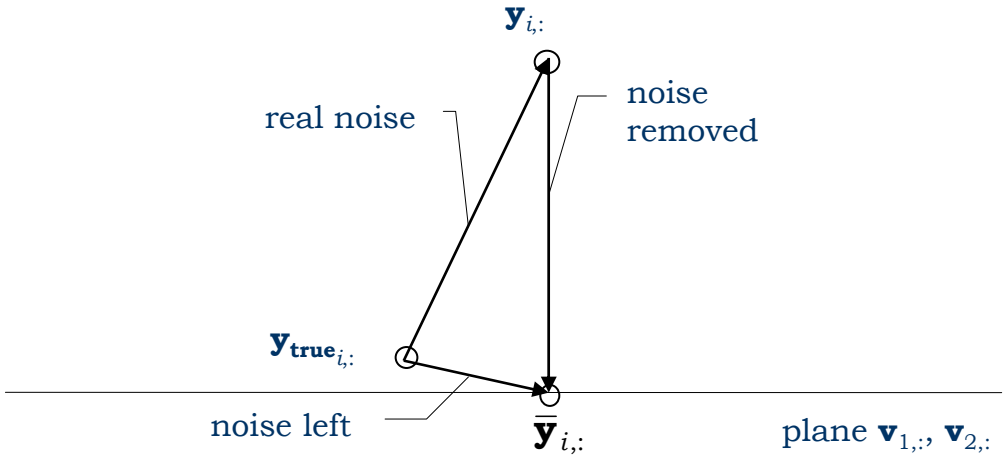

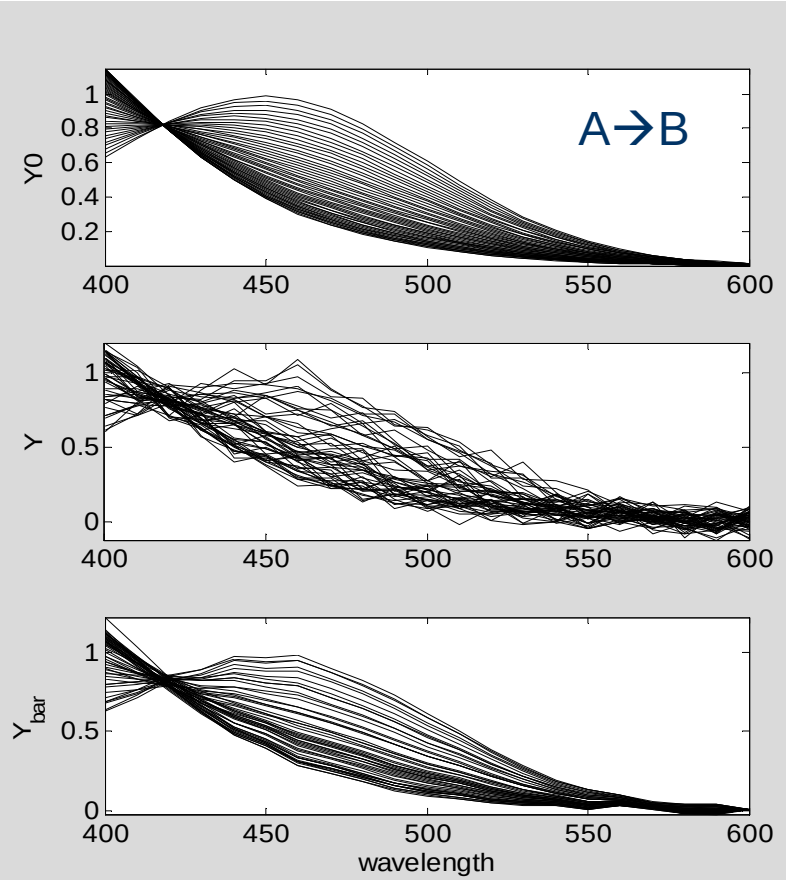

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- linear dependencies in **C** due to the kinetic model are common and sometimes difficult to predict (e.g. $A+B \rightarrow C$)
- linear dependencies in **A** are less common

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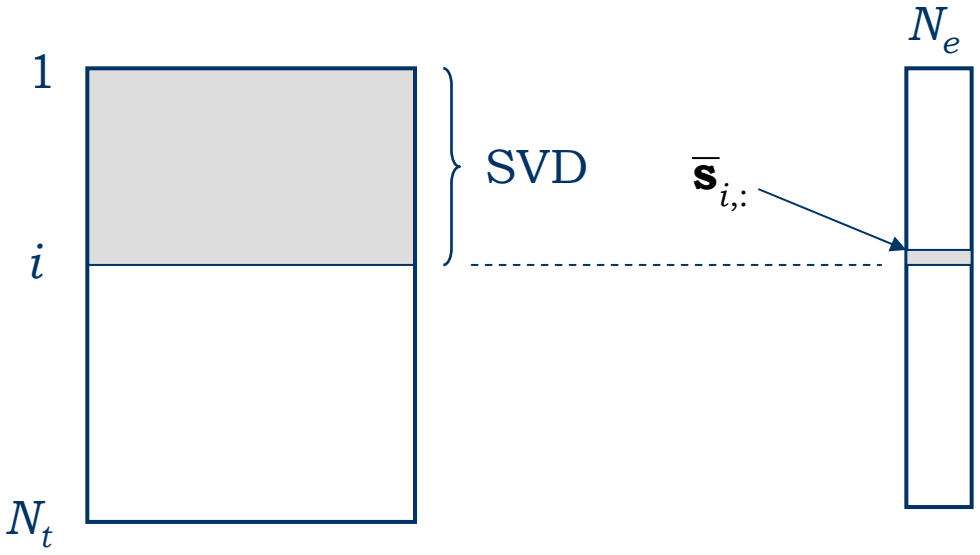
- ideally designed to follow chromatography experiments
  - species appear & disappear sequentially
- capable of roughly following kinetic profiles
  - species can appear & dissappear simultaneously

Evolving Factor Analysis (EFA)

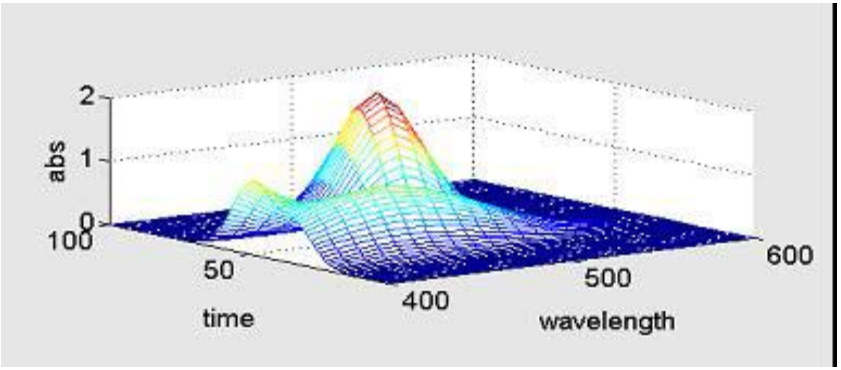
Forward EFA



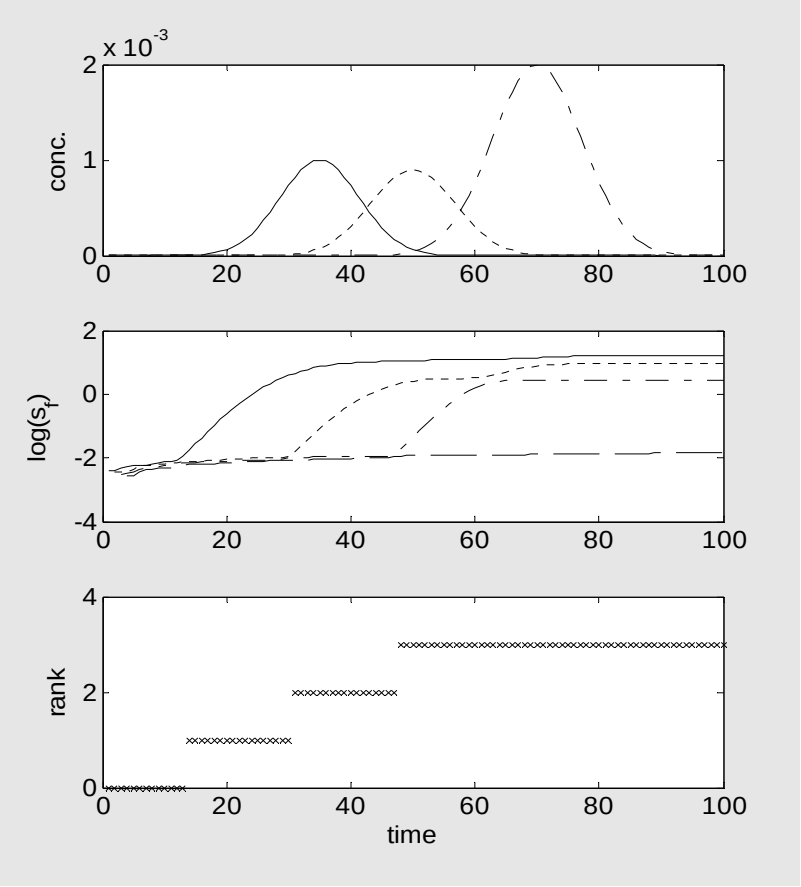
- Repeated rank analysis by SVD in forward direction
- The appearance of a new ‘species’ is indicated by a gradual rise of a new singular value

Evolving Factor Analysis (EFA)

Chromatography

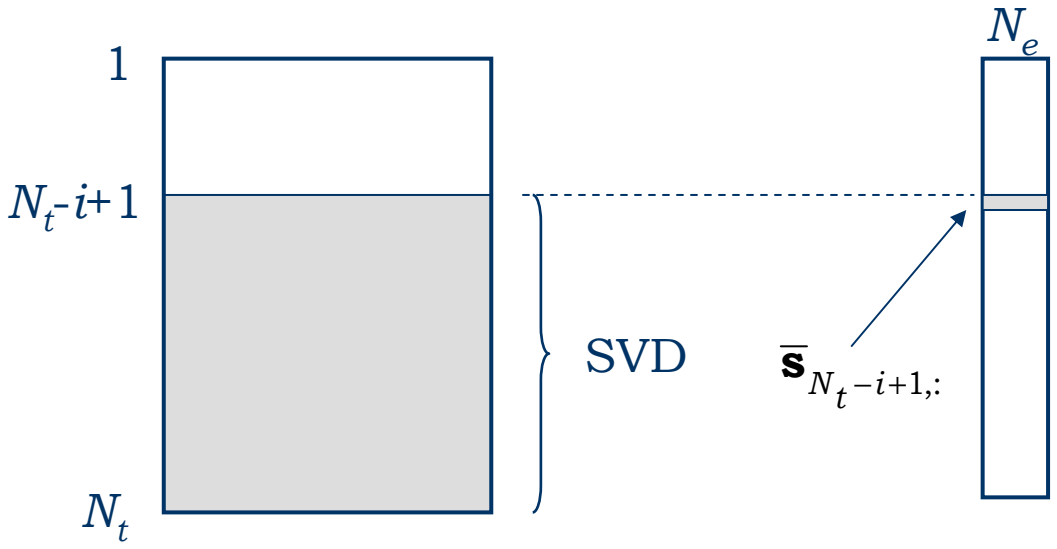


Forward EFA



Evolving Factor Analysis (EFA)

Backward EFA



- Repeated rank analysis by SVD in backward direction
- A ‘disappearing species’ is indicated by a gradual rise of a new singular value

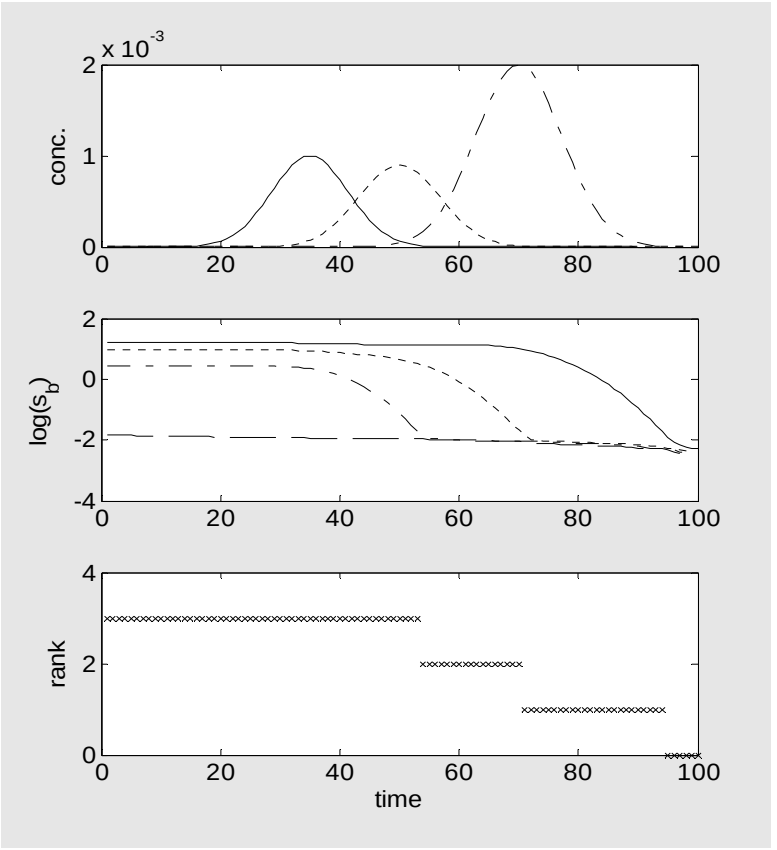
Evolving Factor Analysis (EFA)

Matlab script using a function for forward/backward EFA

```
[t,lam,Y,C,A]=Data_Chrom2;
[ns,nc]=size(C);
ne=nc+1;            % one extra sing. val.
[EFA_f,EFA_b]=EFA(Y,ne);
```

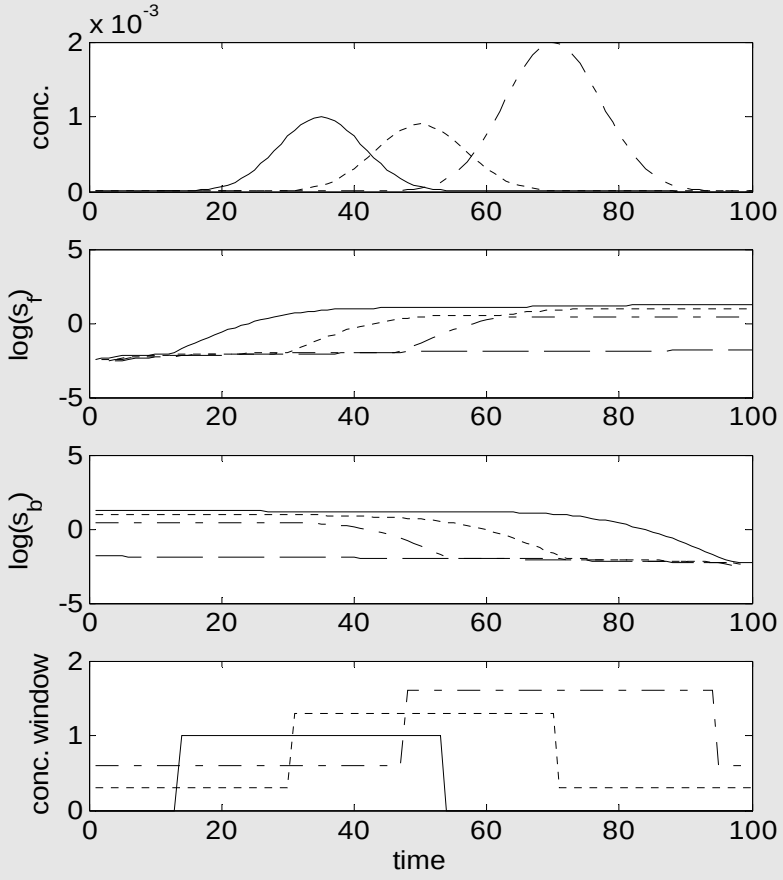
```
function [EFA_f,EFA_b]=EFA(Y,ne)
[ns,nl]=size(Y);
EFA_f=NaN(ns,ne);
EFA_b=NaN(ns,ne);
for i=1:ns
   s_f=svd(Y(1:i,:));                  % forward sv
   s_b=svd(Y(ns-i+1:ns,:));            % backward sv
   EFA_f(i,1:min(i,ne))=s_f(1:min(i,ne))';
   EFA_b(ns-i+1,1:min(i,ne))=s_b(1:min(i,ne))';
end
```

Backward EFA

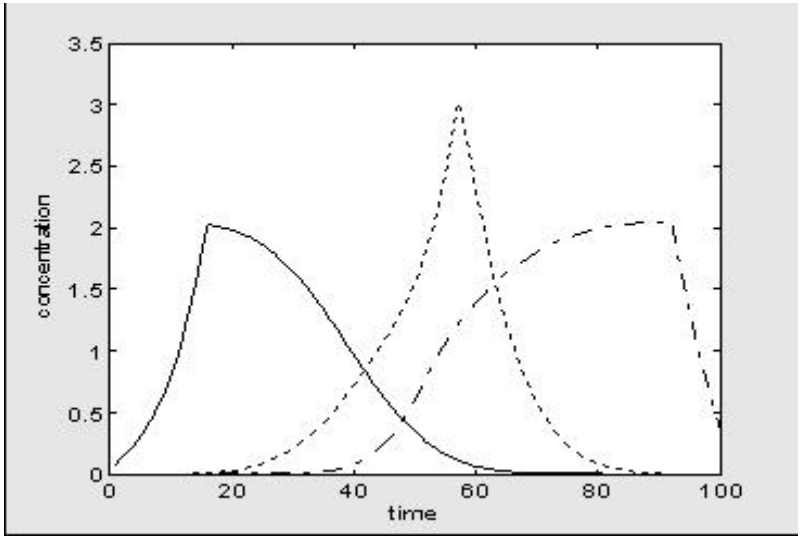


Evolving Factor Analysis (EFA)

Forward and backward EFA



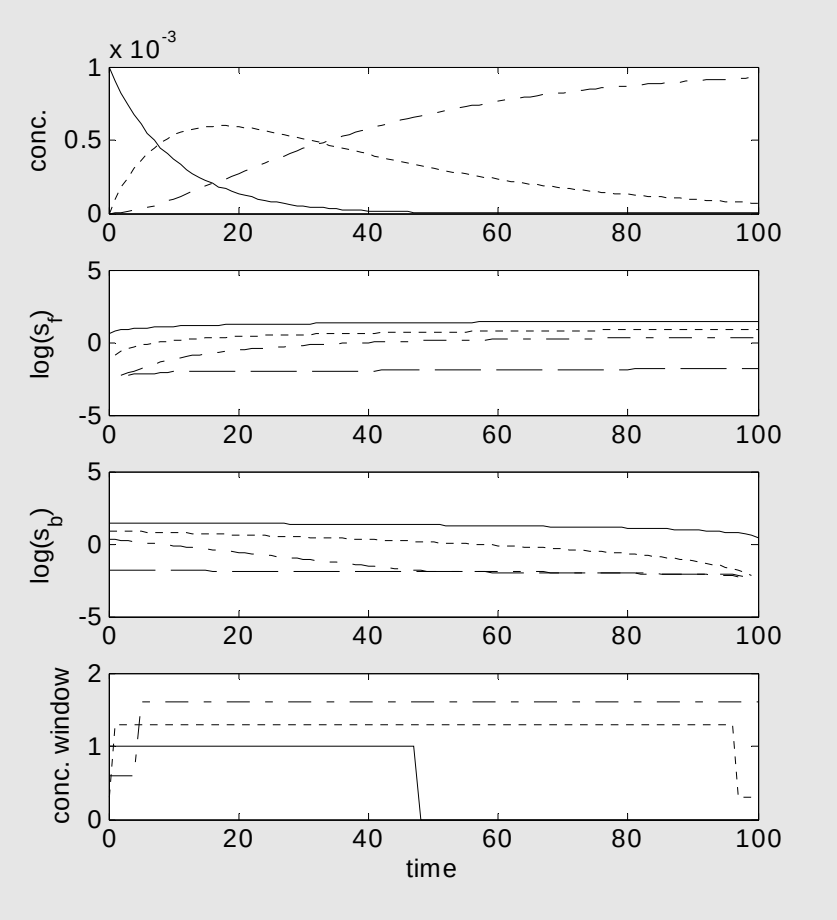
Combined forward/backward EFA results can be used as reasonable initial guesses of concentration profiles for subsequent iterative refinement e.g. by ALS



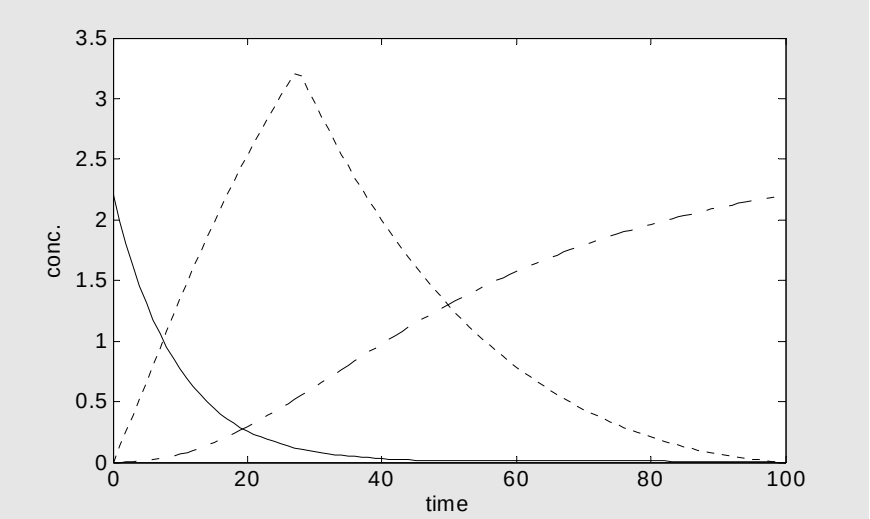

```
% combined SV curves
C=min(EFA_f(:,1:nc),fliplr(EFA_b(:,1:nc)));
```

Evolving Factor Analysis (EFA)

A→B→C, Forward and backward EFA

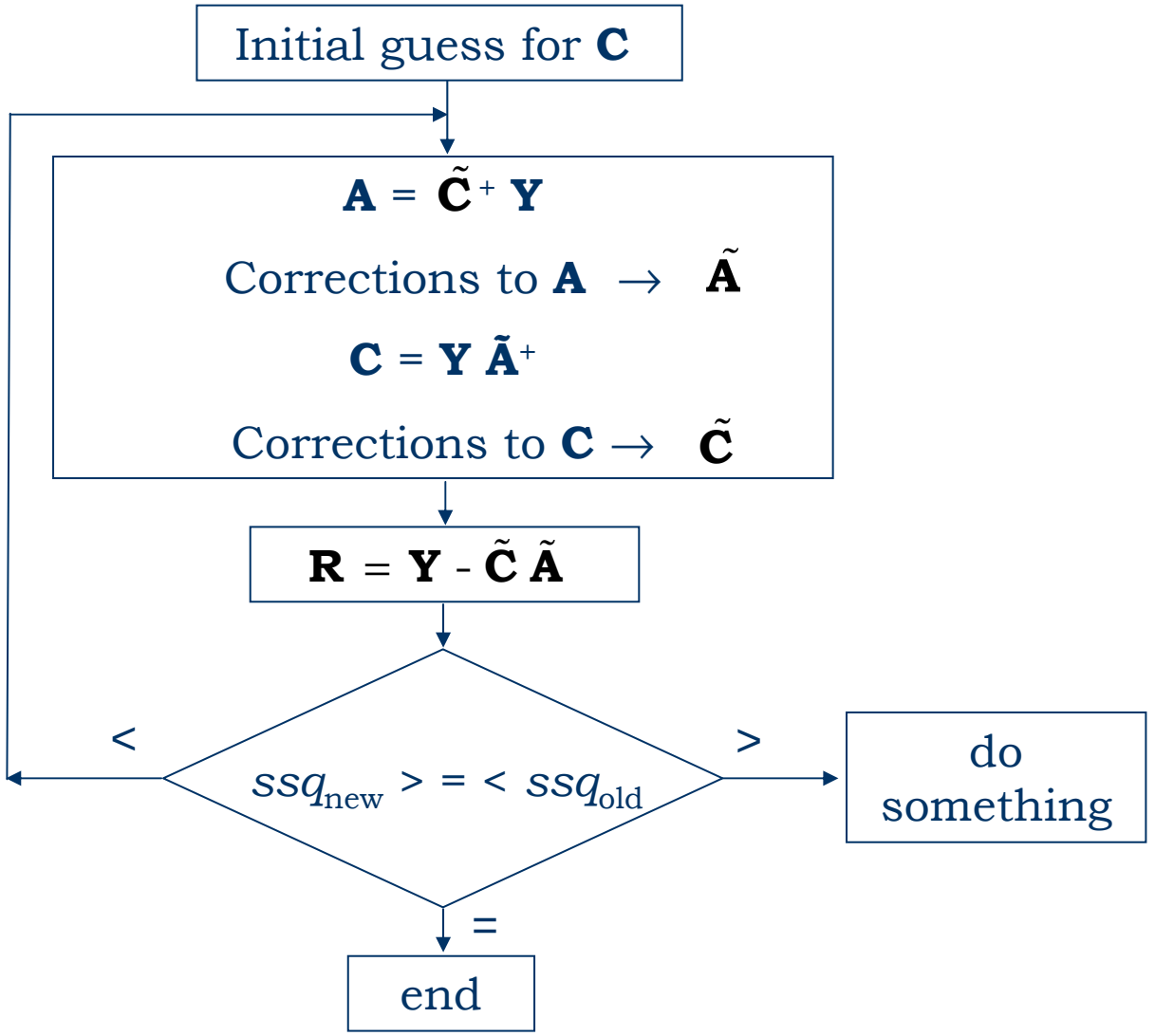


- Combined forward/backward EFA results are not as accurate as in chromatography regarding the appearance and disappearance of species
- But they can still be used as initial guesses of concentration profiles for subsequent iterative refinement e.g. by ALS



Multivariate Curve Resolution by Alternating Least-Squares (MCR-ALS)

- conceptually very simple



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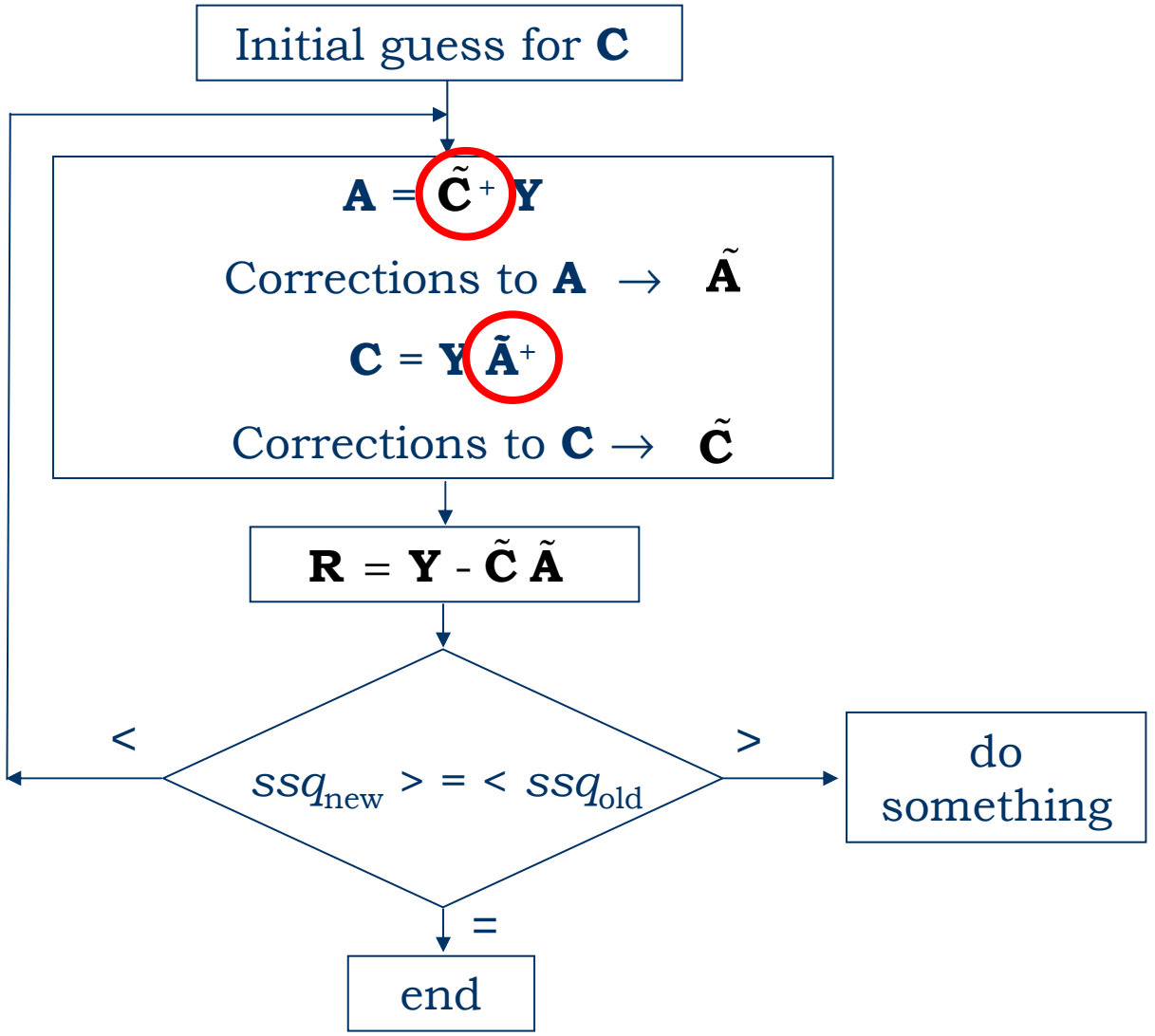

The pseudo inverse and the linear least-squares solution

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$$\mathbf{Y} = \mathbf{C} \times \mathbf{A} + \mathbf{R}$$

$$\mathbf{R} = \mathbf{Y} - \mathbf{C} \times \mathbf{A}$$

$$\mathbf{R} := f(\mathbf{Y}, \mathbf{C}, \mathbf{A})$$

The residuals **R** are a function of **Y** and the two linear parameters **C** & **A**

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The residuals **R** are a function of **Y** and the two linear parameters **C** & **A**

1) **Y** & **C** known: $min \left\| \Sigma\Sigma r_{i,j}^2(\mathbf{A}) \right\|$

R is minimal in the least squares sense if

$$\mathbf{A} = (\mathbf{C}^t\mathbf{C})^{-1}\mathbf{C}^t \times \mathbf{Y} = \mathbf{C}^+ \times \mathbf{Y}$$

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`Matlab: A = C \ Y`

to use the left pseudo inverse of **C**

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Matlab: `A = C \ Y`

to use the left pseudo inverse of C

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R is minimal in the least squares sense if

$$\mathbf{C} = \mathbf{Y} \times \mathbf{A}^t(\mathbf{A}\mathbf{A}^t)^{-1} = \mathbf{Y} \times \mathbf{A}^+$$

The pseudo inverse and the linear least-squares solution

$$\mathbf{Y} = \mathbf{C} \times \mathbf{A} + \mathbf{R}$$

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`Matlab: A = C \ Y`
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R is minimal in the least squares sense if

$$\mathbf{C} = \mathbf{Y} \times \mathbf{A}^t(\mathbf{A}\mathbf{A}^t)^{-1} = \mathbf{Y} \times \mathbf{A}^+$$

`Matlab: C = Y / A`
to use the left pseudo inverse of **C**

Multivariate Curve Resolution by Alternating Least-Squares (MCR-ALS)

```
for it=1:100
    C=norm_max(C);          % normalisation

    [C,A]=constraints_nonneg(Y,C);

    R=Y-C*A;                % residuals
    ssq(it)=sum(sum(R.*R));
end
```

Normalisation to the maximum in each conc. profile

$\mathbf{C}_n = \mathbf{C} \times \square$

$\square$ = diag(max(**C**))$^{-1}$

and

$$\mathbf{A}_n = diag(\max(\mathbf{C})) \times \mathbf{A}$$

```
function [Cn,An]=norm_max(C,A)

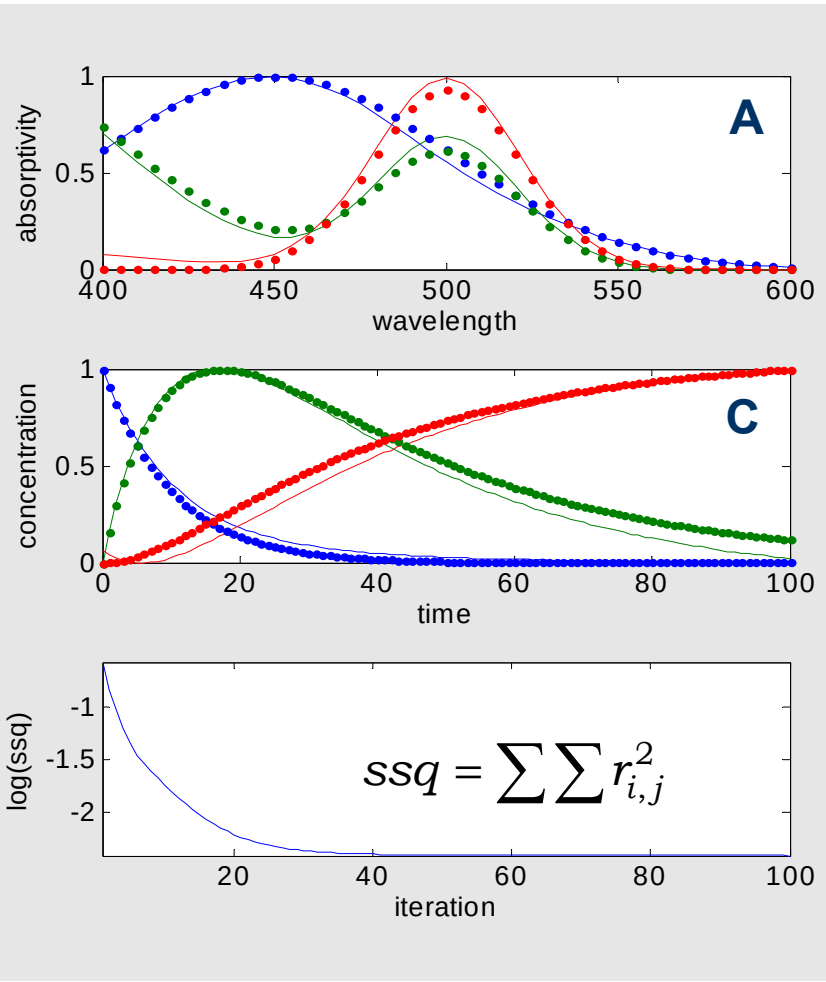
coef=1./max(C);      % norm coeff
Cn=C*diag(coef);     % apply to C
if nargin==2
   An=diag(1./coef)*A; % apply inv coeff to A
end
```

```
function  [C,A]=constraints_nonneg(Y,C)

A=nonneg(Y',C')';    % pos spectra (Andersson)
C=nonneg(Y,A);       % pos conc. (Andersson)
```

A→*B*→*C*

constraints: **C**, **A** >0



Multivariate Curve Resolution by Alternating Least-Squares (MCR-ALS)

A→*B*→*C*

constraints: **C**, **A** >0, known spectrum of *B*

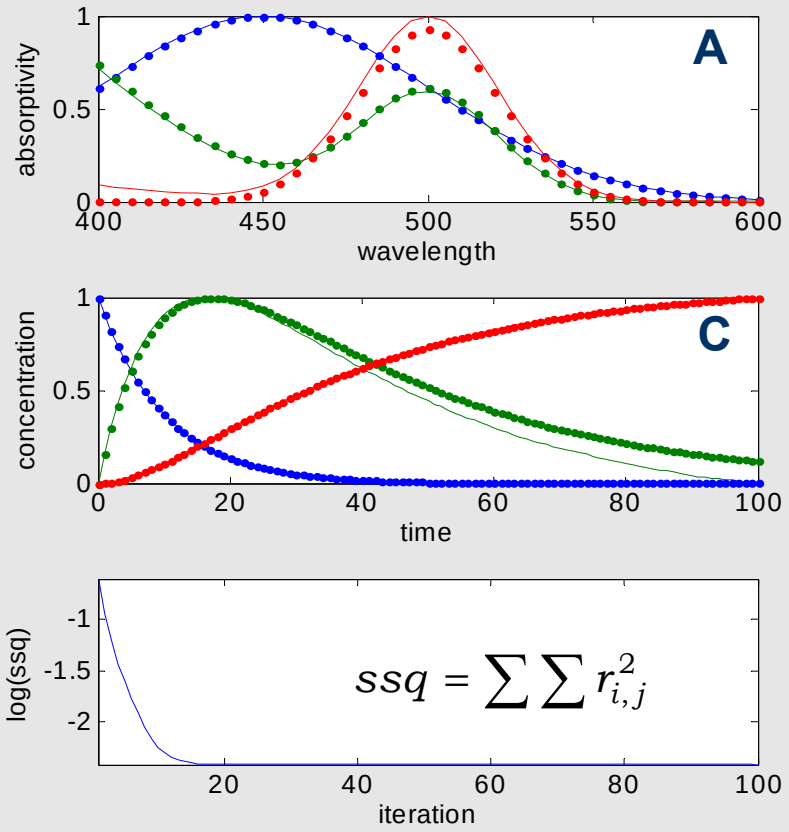
```
for it=1:100
  C=norm_max(C);          % normalisation

  [C,A]=constraints_known_spec_B(Y,C,A_sim);

  R=Y-C*A;                % residuals
  ssq(it)=sum(sum(R.*R));
end
```

```
function [C,A] =
 constraints_nonneg_known_spec_B(Y,C,A_sim)

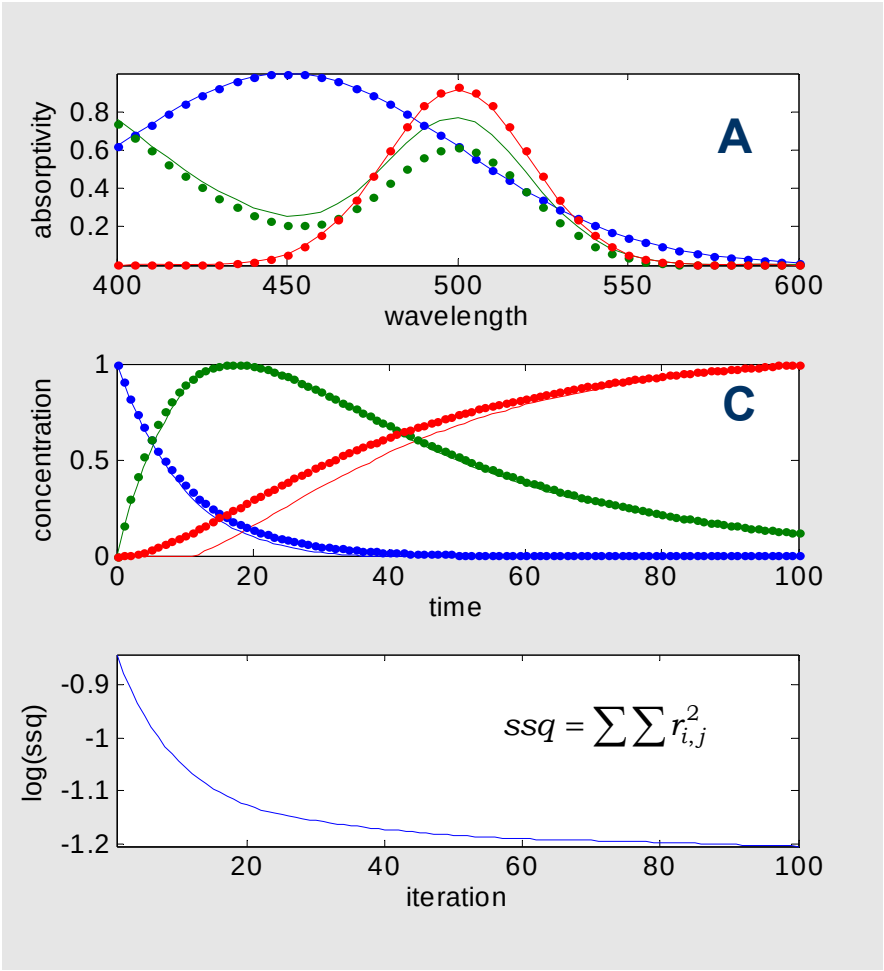
A=nonneg(Y',C')';   % pos spectra (Andersson)
A(2,:)=A_sim(2,:);  % known spectrum of B
C=nonneg(Y,A);      % pos conc. (Andersson)
```



Multivariate Curve Resolution by Alternating Least-Squares (MCR-ALS)

A→*B*→*C*

constraints: **C**, **A** >0, known spectrum of *A* & *C*



Multivariate Curve Resolution by Alternating Least-Squares (MCR-ALS)

```
for it=1:100
    C=norm_closure(C,[],c_tot); % norm. C to Ctot

    [C,A]=constraints_nonneg_known_spec_B(Y,C,A_sim);

    R=Y-C*A;                % residuals
    ssq(it)=sum(sum(R.*R));
end
[C_n,A_n]=norm_closure(C,A,c_tot);
```

Normalisation to the total conc. ($c_{tot}=[A]+[B]+[C]=[A]_0$)

$$\mathbf{C} \,|\, \mathbf{coef} = |\, \mathbf{c}_{tot} \longrightarrow \mathbf{coef} = \mathbf{C}^{+} \times \mathbf{c}_{tot}$$

$$\mathbf{C}_n = \mathbf{C} \times \square \quad \text{diag}(\mathbf{coef})$$

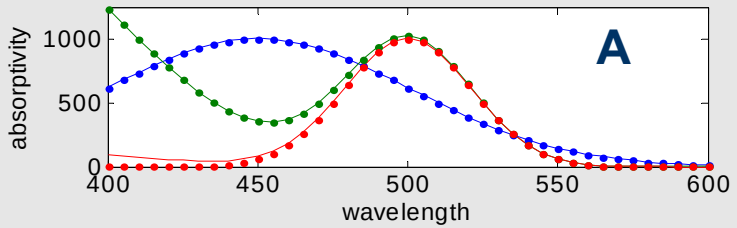
and

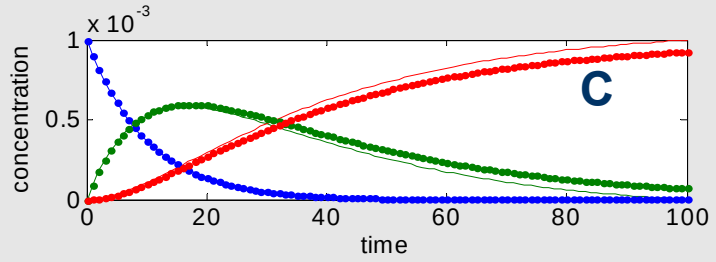
$$\mathbf{A}_n = diag(\mathbf{coef})^{-1} \times \mathbf{A}$$

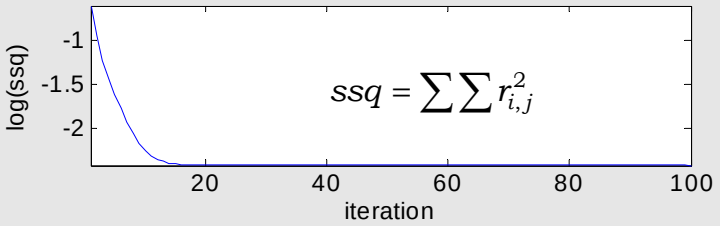
```
function [Cn,An]=norm_closure(C,A,c_tot)

coef=C\(ones(size(C,1),1)*c_tot); % norm. coeff.
Cn=C*diag(coef);                  % apply to C
if ~isempty(A)
   An=diag(1./coef)*A;            % apply inv. to A
end
```

A→B→C
constraints: **C**, **A** >0,
known spectrum of *B, closure*





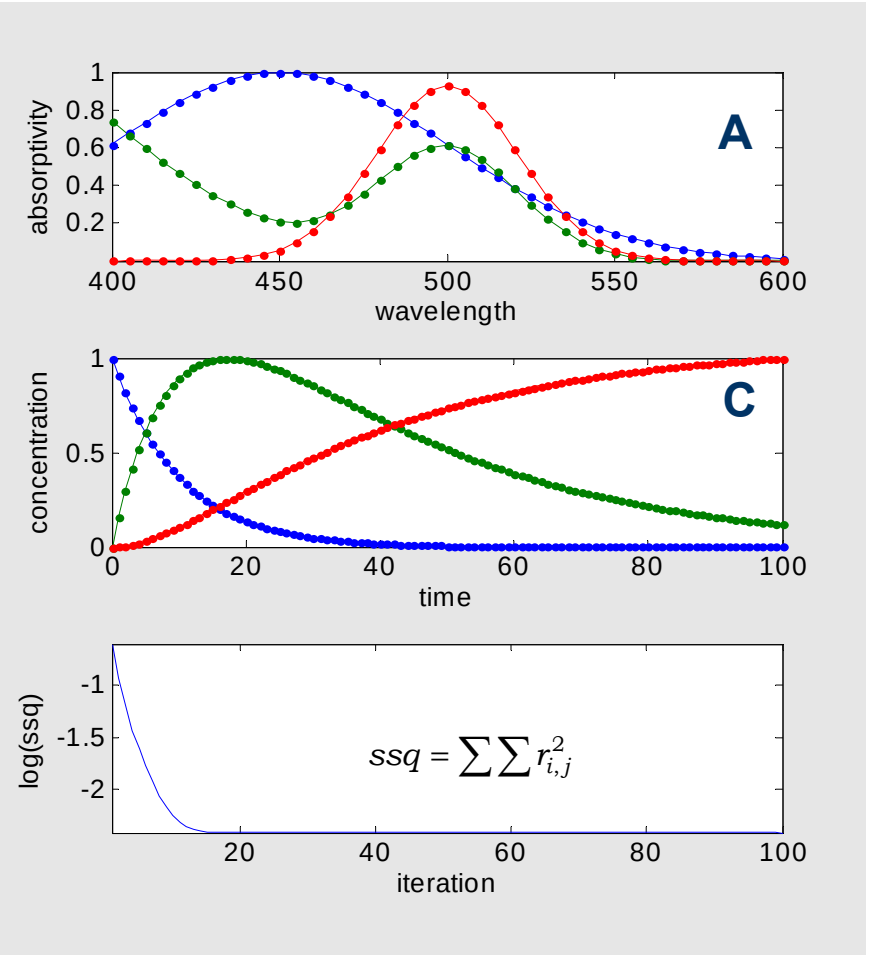


Multivariate Curve Resolution by Alternating Least-Squares (MCR-ALS)

A→*B*→*C*

constraints: **C**, **A** >0, known spectrum of *B* & *C*

As full conversion is not reached for intermediate *B* and product *C*, corresponding known spectra are required for full resolution !



Conclusions: ‘Soft’-Modelling

- Advantages
  - No prior knowledge on the chemical system required
  - Estimation of the number of linearly dependent absorbing species and their approximate evolution from PCA, EFA & ALS
  - Info for the development of a ‘hard’ model
  - ‘Better than nothing’
- Drawbacks
  - No physical model
  - No predictions for other exp. conditions possible
  - Uniqueness of the result is rarely given and difficult to validate

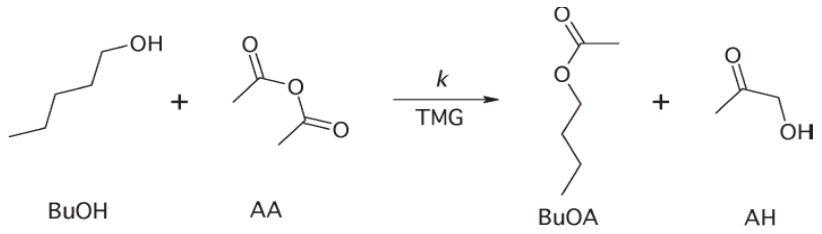
Reading Material

- **Factor Analysis in Chemistry**
  E.R. Malinowski, 3rd ed., Wiley, New York 2002
- **Practical Data Analysis in Chemistry**
  M. Maeder, Y.M. Neuhold, Elsevier, Amsterdam 2007
- **Practical Guide to Chemometrics**
  P. Gemperline (editor), 2nd ed., CRC Press, Boca Raton 2006
- **The Investigation of Organic Reactions and their Mechanisms**
  H. Maskill (editor), Blackwell Publishing, Oxford 2006
- **Evolving factor analysis for the resolution of overlapping chromatographic peaks**
  M. Maeder, Anal. Chem. 59 (1987), 527-530
- **Nonlinear Least-Squares Fitting of Multivariate Absorption Data**
  M. Maeder, A. Zuberbühler. Anal. Chem. 62 (1990), 2220-2224
- **Analyses of 3-way data from equilibrium and kinetic investigations**
  R. Dyson , M. Maeder, Y.M. Neuhold, G. Puxty. Anal. Chim. Act. 490 (2003), 99-108
- **Empirical kinetic modelling of on-line simultaneous infrared and calorimetric measurement using a pareto optimal approach and multi-objective genetic algorithm**
  S.I. Gianoli, G. Puxty, U. Fischer, M. Maeder, K. Hungerbühler. Chemom. Int. Lab. Syst. 85 (2007), 47-62
- **Tutorial on the Fitting of Kinetic Models to Multivariate Spectroscopic Measurements with Non-Linear Regression**
  G. Puxty, M. Maeder, K. Hungerbühler. Chemom. Int. Lab. Syst. 81 (2006), 149-164
- **Data Oriented Process Development: Determination of Reaction Parameters by Small-Scale Calorimetry with in situ Spectroscopy**
  G. Puxty, U. Fischer, M. Jecklin, K. Hungerbühler. Chimia 60 (2006), 605-610

Part 2: Hard Modelling

Introduction

- You follow the reaction between BuOH and the Acetic Acid in the IR range and you get the following absorbance profile :



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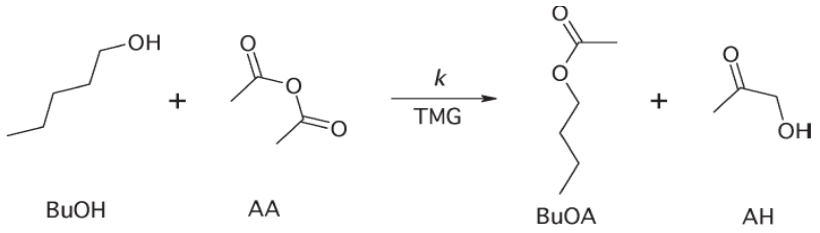

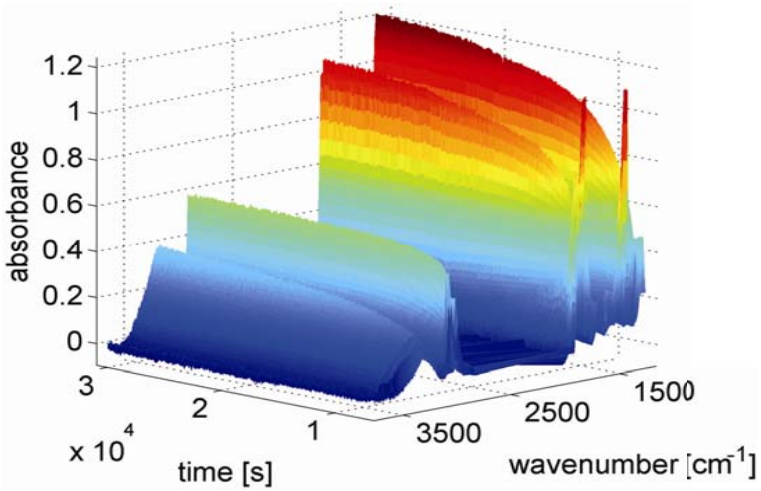

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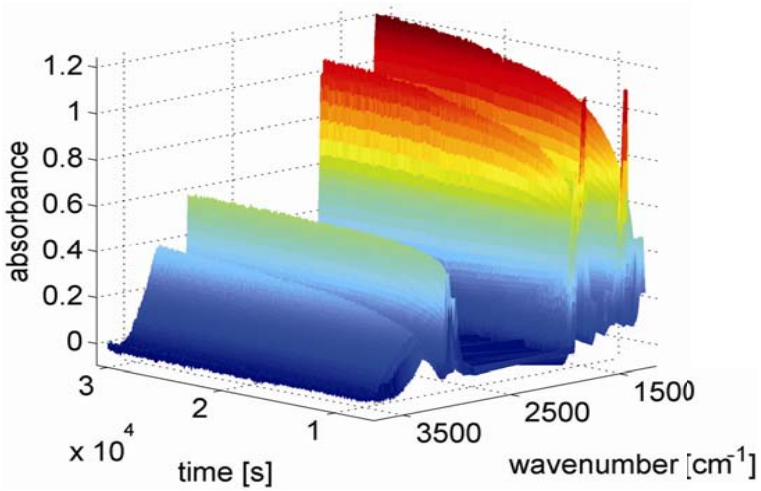
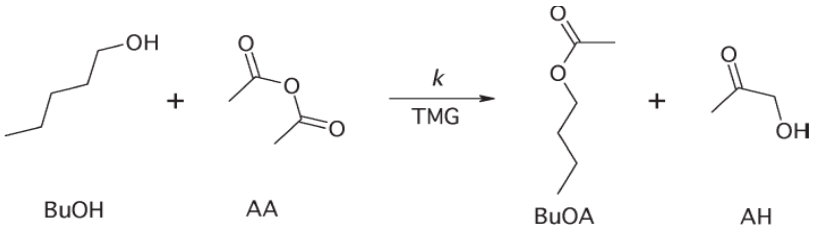
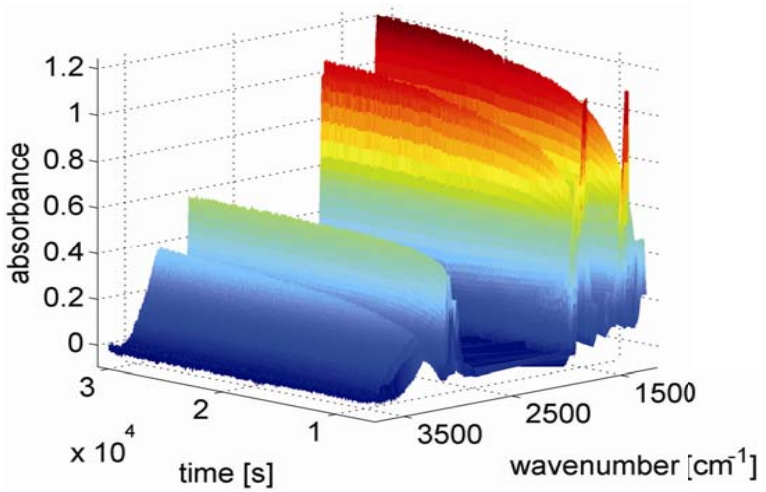




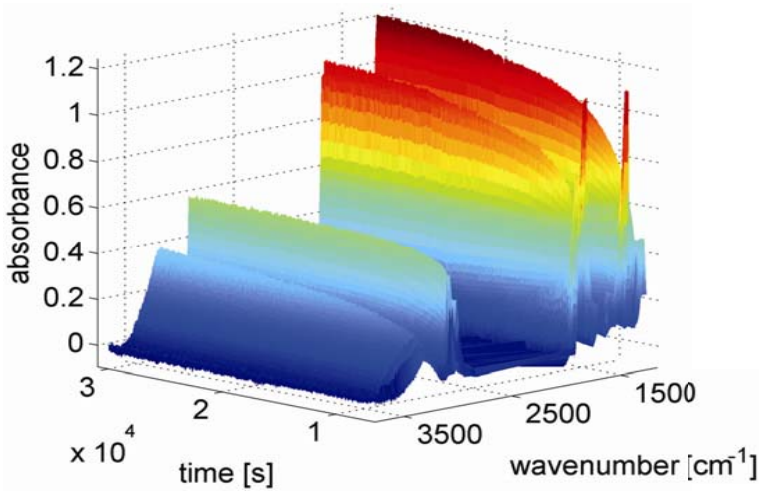
How can the rate constant be extracted from this profile ?

UNIVARIATE approach



UNIVARIATE approach

1. You determine the extinction coefficients of all species with known concentrations



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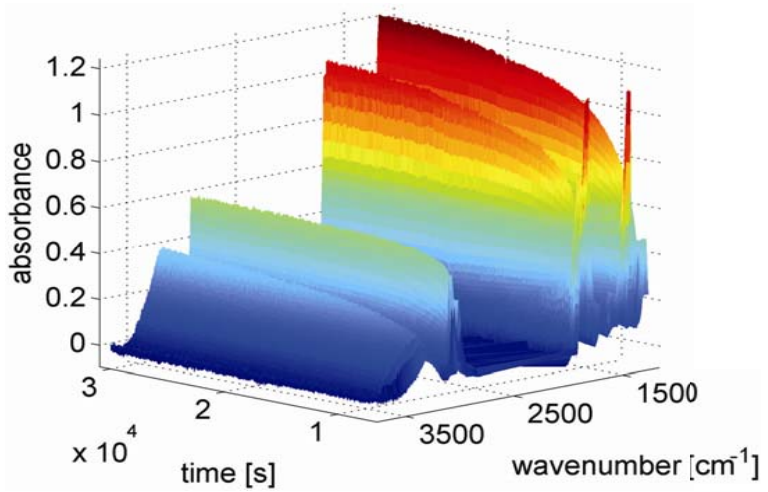
   $\Rightarrow$ all ε_λ



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1. You determine the extinction coefficients of all species with known concentrations

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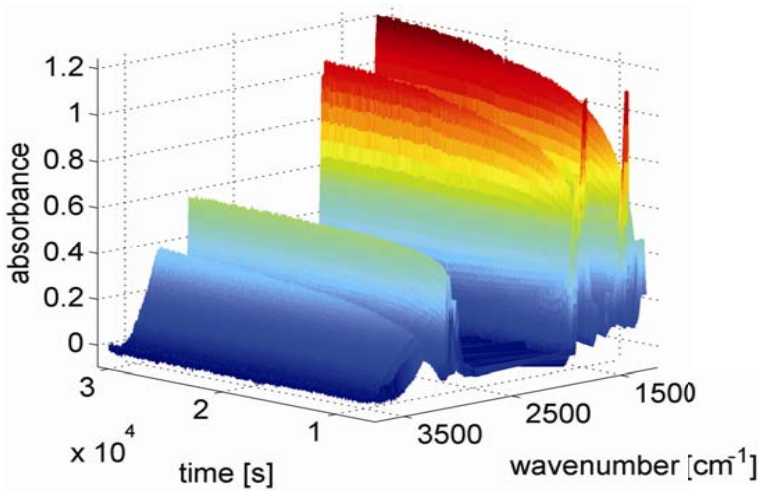
2. You integrate the set of differential equations (2^{nd} order reaction) with the used initial concentrations



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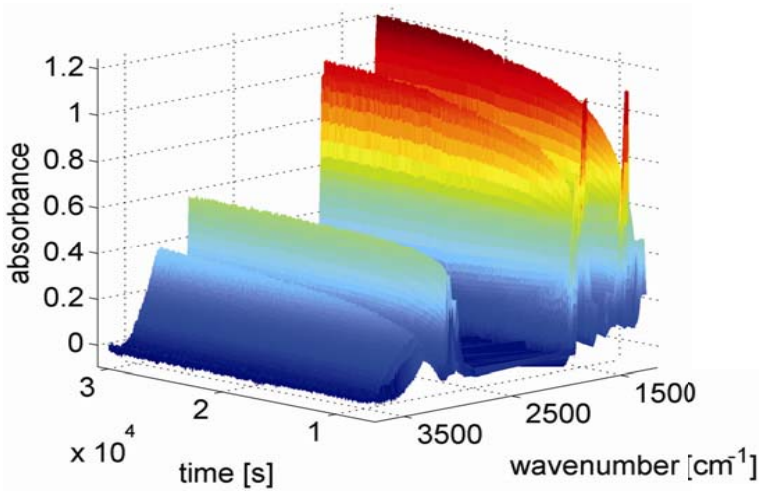

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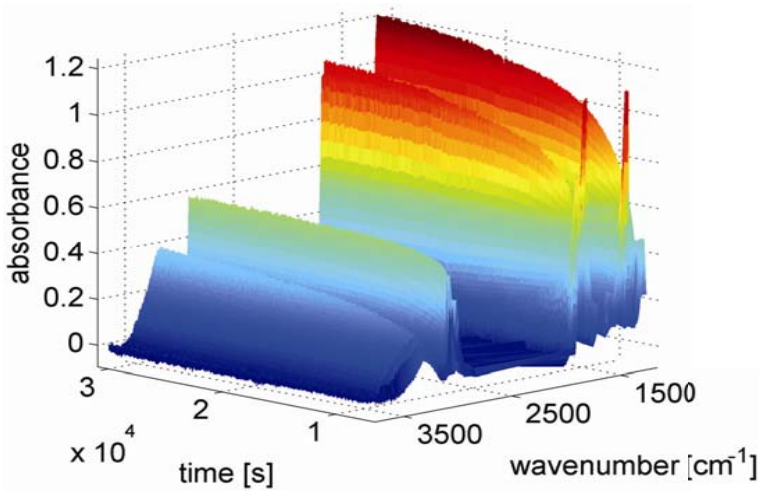
3. You apply Beer's law at one wavelength



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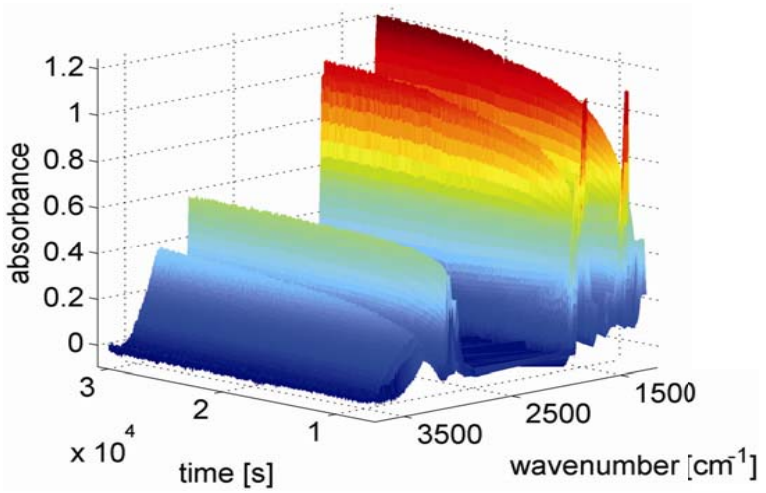
$$abs_\lambda(t) = c_{BuOH}(t,\ k)\cdot\varepsilon_{BuOH,\lambda} + c_{AA,\lambda}(t,\ k)\cdot\varepsilon_{AA,\lambda} + c_{BuOA}(t,\ k)\cdot\varepsilon_{BuOA,\lambda} + c_{HA}(t,\ k)\cdot\varepsilon_{HA,\lambda}$$



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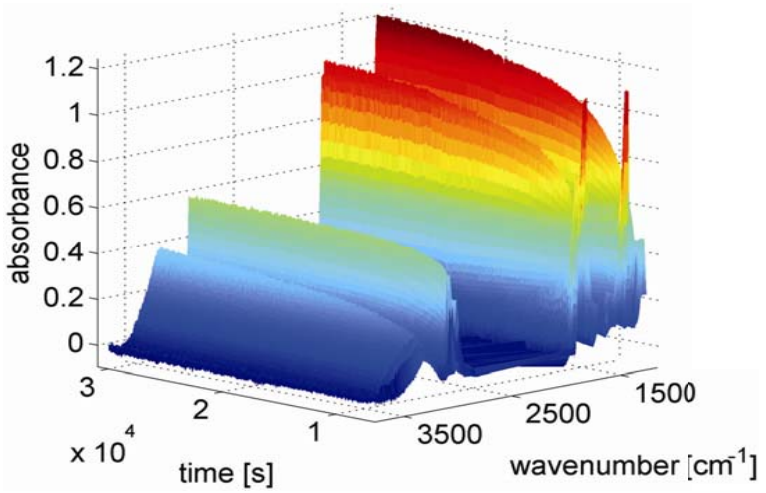
4. You find *k* that best approximate the measured $abs_\lambda(t)$ in the least squares sense



UNIVARIATE approach

1. **CALIBRATION** Determination of extinction coefficients of all species with known concentrations

   $\Rightarrow$ all ε_λ

2. You integrate the set of differential equations (2^{nd} order reaction) with the used initial concentrations

   $\Rightarrow$ all *c(t, k)*

3. You apply Beer's law at one wavelength

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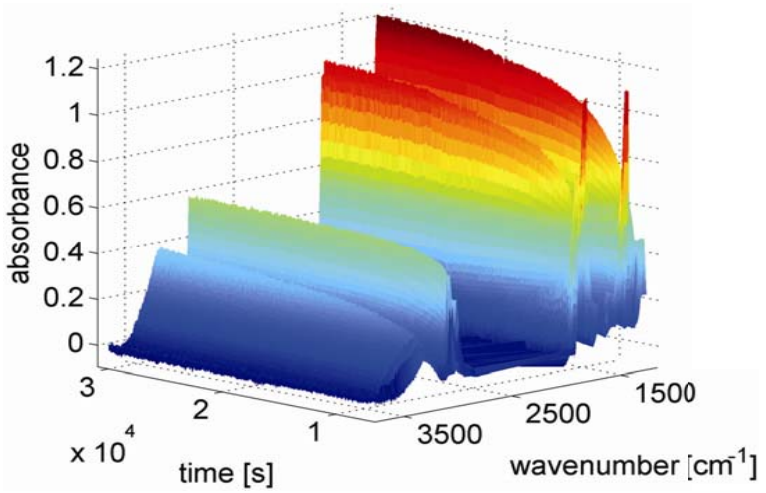

UNIVARIATE approach

1. **CALIBRATION**
   Determination of extinction coefficients of all species with known concentrations
   ⇒ all ε_λ
2. **INTEGRATION OF THE KINETIC MODEL**
   Integration of differential equations (pseudo-first order reaction) with the used initial concentrations
   ⇒ all *c(t, k)*
3. You apply Beer's law at one wavelength

$$abs_\lambda(t) = c_{BuOH}(t,\ k)\cdot\varepsilon_{BuOH,\lambda} + c_{AA,\lambda}(t,\ k)\cdot\varepsilon_{AA,\lambda} + c_{BuOA}(t,\ k)\cdot\varepsilon_{BuOA,\lambda} + c_{HA}(t,\ k)\cdot\varepsilon_{HA,\lambda}$$

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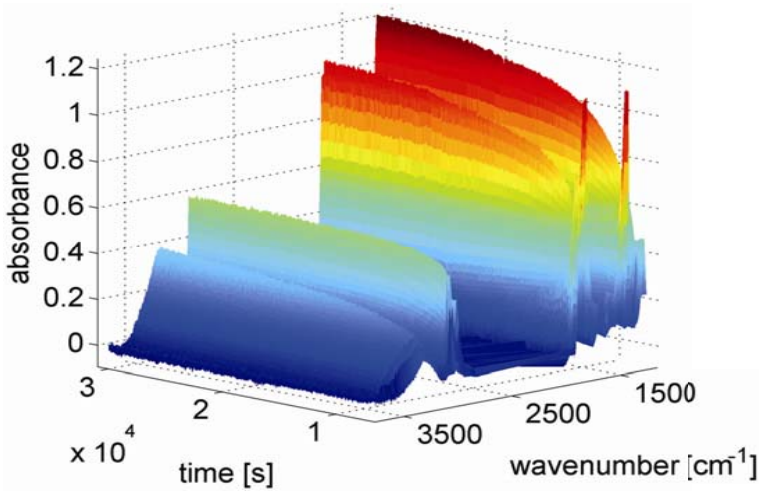

UNIVARIATE approach

1. **CALIBRATION**
   of all species with known concentrations
   $\Rightarrow$ all ε_λ
2. **INTEGRATION OF THE KINETIC MODEL**
   (2nd order reaction) with the used initial concentrations
   $\Rightarrow$ all $c(t, k)$
3. **APPLICATION OF BEER'S LAW IN A UNIVARIATE FORM (= ONE WAVELENGTH)**

$$abs_\lambda(t) = c_{AA}(t, k) \cdot \varepsilon_{AA,\lambda} + c_{AA,\lambda}(t, k) \cdot \varepsilon_{AA,\lambda} + c_{BuOA}(t, k) \cdot \varepsilon_{BuOA,\lambda} + c_{HA}(t, k) \cdot \varepsilon_{HA,\lambda}$$

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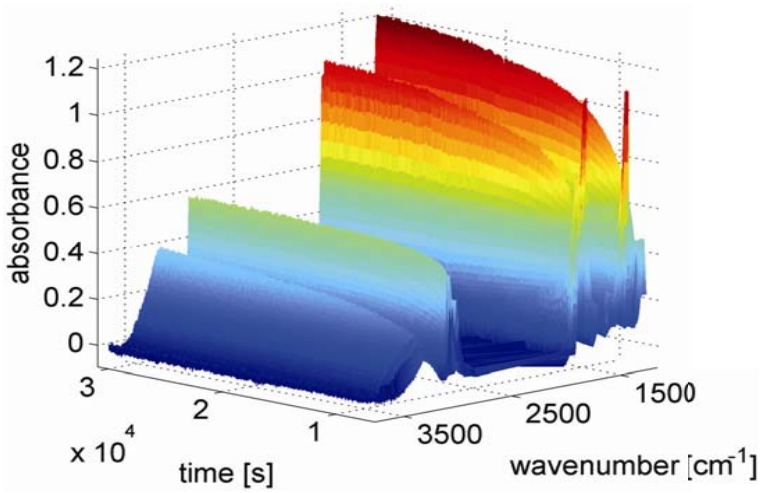

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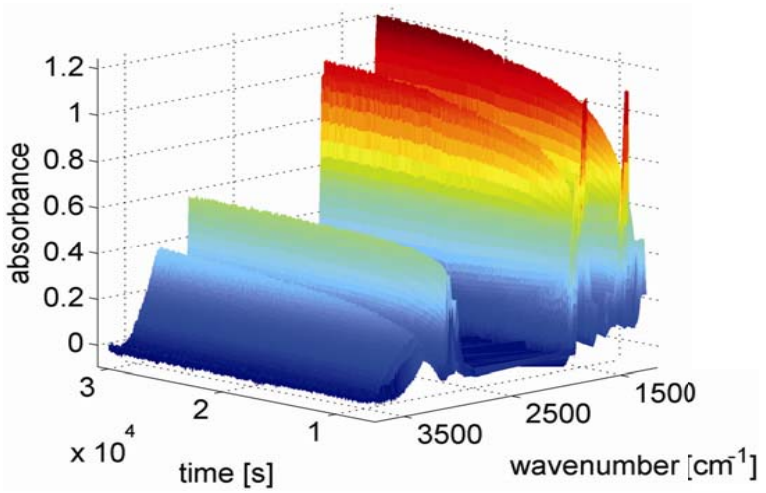
4. **FITTING**
   $abs_\lambda(t)$
   (= NONLINEAR OPTIMISATION)



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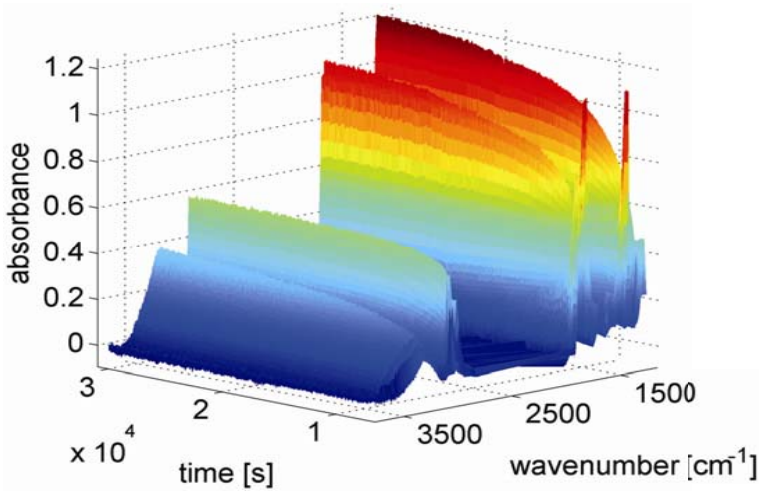

KINETIC HARD-MODELING !!!

UNIVARIATE approach

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4. **FITTING**
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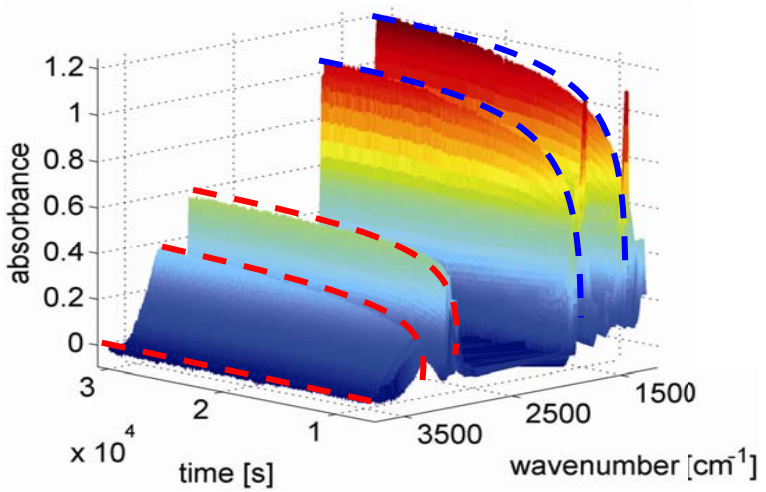

KINETIC HARD-MODELING !!!

Question :
Which wavelength do you follow ?
What about the rest of the spectrum ?

UNIVARIATE approach

1. CALIBRATION function coefficients
of all species with known concentrations

2. INTEGRATION OF THE
KINETIC MODEL

3. APPLICATION OF BEER'S LAW
IN A UNIVARIATE WAY
(= ONE WAVELENGTH)

4. FITTING
(= NONLINEAR OPTIMISATION)

WITH A ***MULTIVARIATE APPROACH*** *THE WHOLE SPECTRUM IS USED TO FIT k*

KINETIC HARD-MODELING !!!

Question :
Which wavelength do you follow ?
What about the rest of the spectrum ?

Presentation of a typical kinetic-modeling algorithm

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The following topics will be addressed :

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- Typical fitted signals
  - Multivariate Spectroscopy (Beer's law)
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1. Divergence problems *(Levenberg-Marquardt modification)*

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  - Univariate Calorimetry
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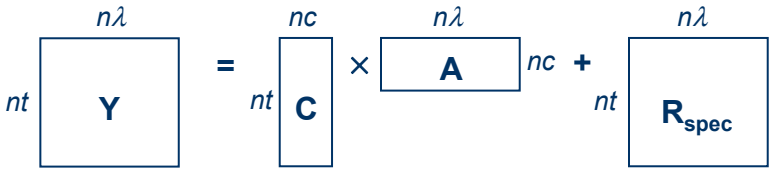
- Newton-Gauss method

Two common problems will also be treated :

1. Divergence problems *(Levenberg-Marquardt modification)*
2. Rank deficiency *(methods of annihilation)*

SPECTROSCOPY and CALORIMETRY

- Spectroscopy (IR, UV, Raman, fluorescence …)



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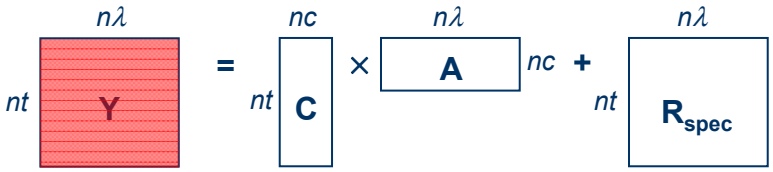
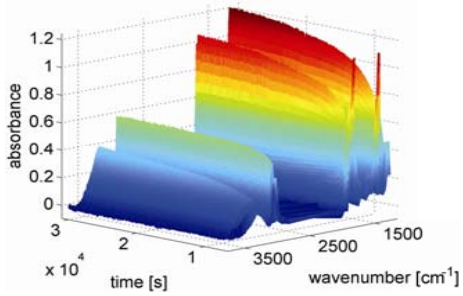
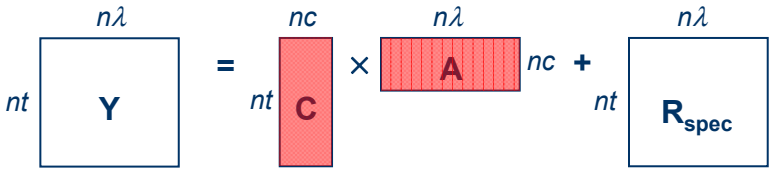


Measurement (Multivariate)

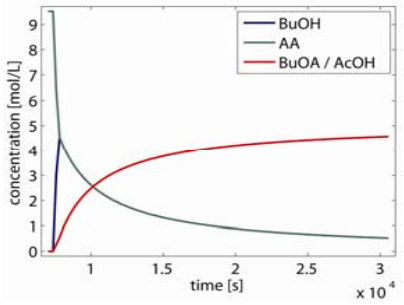


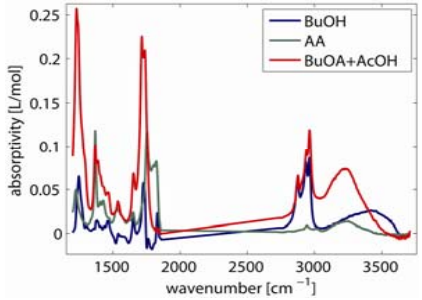
SPECTROSCOPY and CALORIMETRY

- Spectroscopy (IR, UV, Raman, fluorescence …)



Physical model (Beer's law)



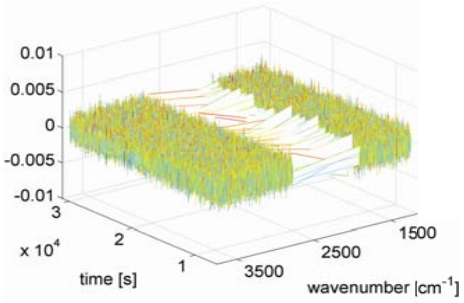


SPECTROSCOPY and CALORIMETRY

- Spectroscopy (IR, UV, Raman, fluorescence …)

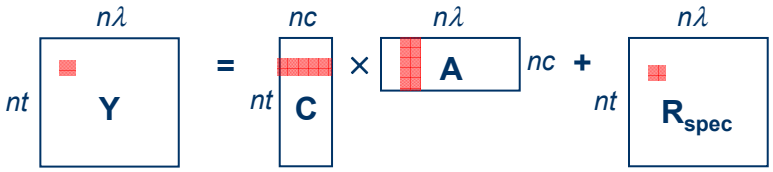
$$\underset{nt}{\boxed{Y}}_{n\lambda} = \underset{nt}{\boxed{C}}_{nc} \times \underset{nc}{\boxed{A}}_{n\lambda} + \underset{nt}{\boxed{R_{spec}}}_{n\lambda}$$

Unmodelled effects (Matrix)



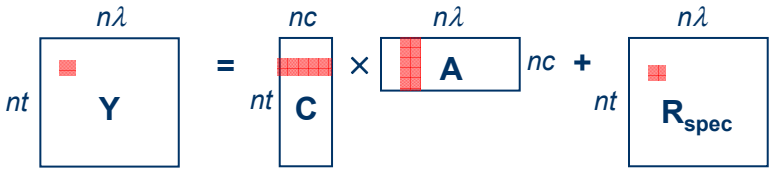
SPECTROSCOPY and CALORIMETRY

- Spectroscopy (IR, UV, Raman, fluorescence …)

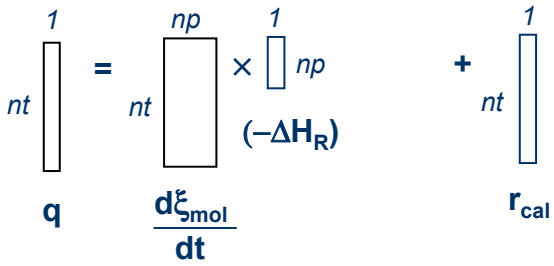


SPECTROSCOPY and CALORIMETRY

- Spectroscopy (IR, UV, Raman, fluorescence …)



- Calorimetry



SPECTROSCOPY and CALORIMETRY

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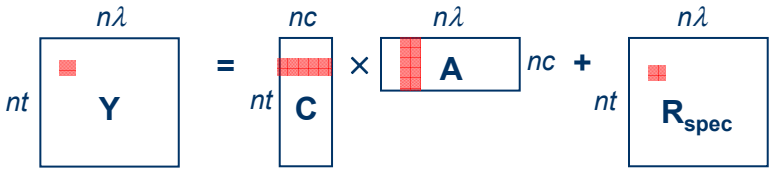
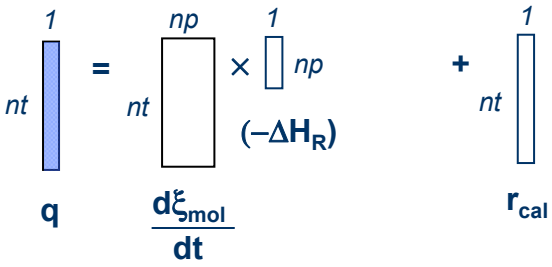
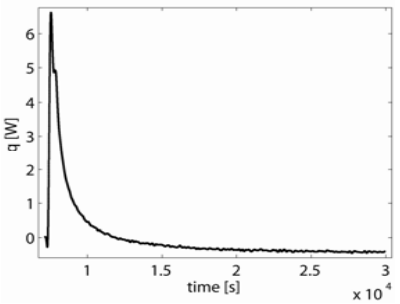


- Calorimetry

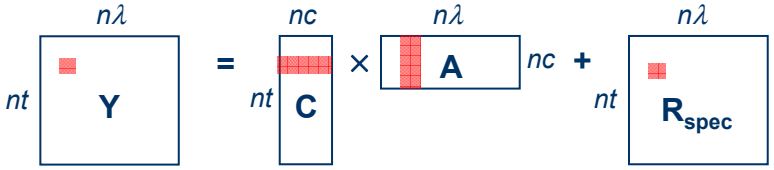


Measurement (Univariate)

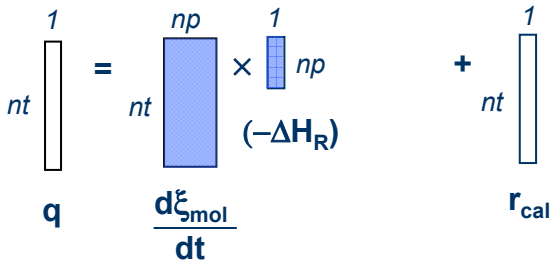


SPECTROSCOPY and CALORIMETRY

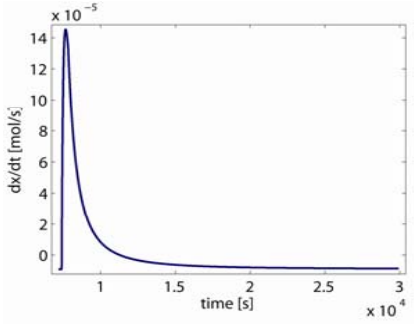
- Spectroscopy (IR, UV, Raman, fluorescence …)



- Calorimetry



Physical model
(Reaction heat balance)



× 43 kJ/mol

SPECTROSCOPY and CALORIMETRY

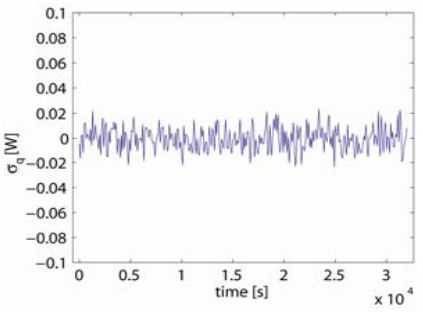
- Spectroscopy (IR, UV, Raman, fluorescence …)

$$\underset{nt \times n\lambda}{\mathbf{Y}} = \underset{nt \times nc}{\mathbf{C}} \times \underset{nc \times n\lambda}{\mathbf{A}} + \underset{nt \times n\lambda}{\mathbf{R}_{spec}}$$

- Calorimetry

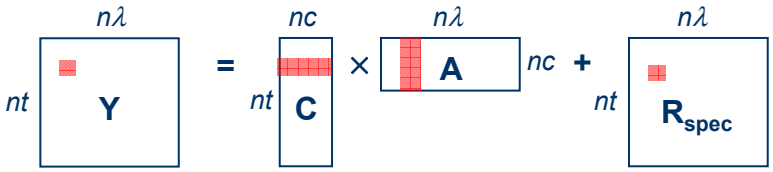
$$\underset{nt \times 1}{\mathbf{q}} = \underset{nt \times np}{\frac{d\xi_{mol}}{dt}} \times \underset{np \times 1}{(-\Delta H_R)} + \underset{nt \times 1}{\mathbf{r}_{cal}}$$

Unmodelled effects (Vector)

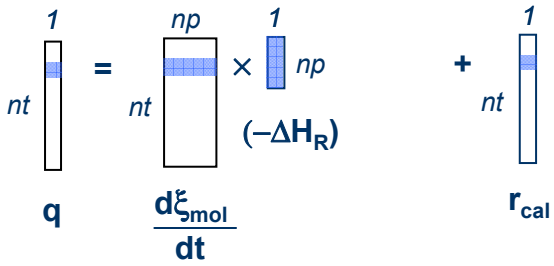


SPECTROSCOPY and CALORIMETRY

- Spectroscopy (IR, UV, Raman, fluorescence …)



- Calorimetry



$$\frac{d\xi_{mol}}{dt} = \mathbf{V}(t) \cdot \mathbf{r}(t)$$

Definition of the optimisation problem

- The residuals are defined as the difference between the measurement and the model

$$\mathbf{Y} = \mathbf{C} \cdot \mathbf{A} + \mathbf{R}_{\mathbf{spec}}$$

$$\mathbf{q} = \frac{\mathbf{d}\xi_{\mathbf{mol}}}{\mathbf{dt}} \cdot \left(-\Delta \mathbf{H}_{\mathbf{R}}\right) + \mathbf{r}_{\mathbf{cal}}$$

Definition of the optimisation problem

- The residuals are defined as the difference between the measurement and the model

$$\mathbf{R}_{\mathbf{spec}} = \mathbf{Y} - \mathbf{C} \cdot \mathbf{A}$$

$$\mathbf{r}_{\mathbf{cal}} = \mathbf{q} - \frac{\mathbf{d}\xi_{\mathbf{mol}}}{\mathbf{dt}} \cdot \left(-\Delta \mathbf{H}_{\mathbf{R}} \right)$$

Definition of the optimisation problem

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$$\mathbf{R}_{spec} = \mathbf{Y} - \mathbf{C} \cdot \mathbf{A}$$

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- As the mass balance part is :

$$\mathbf{C} = f\left(model,\ \mathbf{p}\right)$$

$$\frac{\mathbf{d}\xi_{mol}}{\mathbf{dt}} = g\left(model,\ \mathbf{p}\right)$$

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- As the mass balance part is :

$$\mathbf{C} = f\left(model,\ \mathbf{p}\right)$$

$$\frac{\mathbf{d}\xi_{mol}}{\mathbf{dt}} = g\left(model,\ \mathbf{p}\right)$$

- The optimisation problem is :

minimize $\mathbf{R}_{spec} = f\left(\mathbf{Y}, model,\ \mathbf{p}\right)$
in the least square sense
by changing $\mathbf{p}$

minimize $\mathbf{r}_{cal} = f\left(\mathbf{q}, model,\ \mathbf{p}\right)$
in the least square sense
by changing $\mathbf{p}$

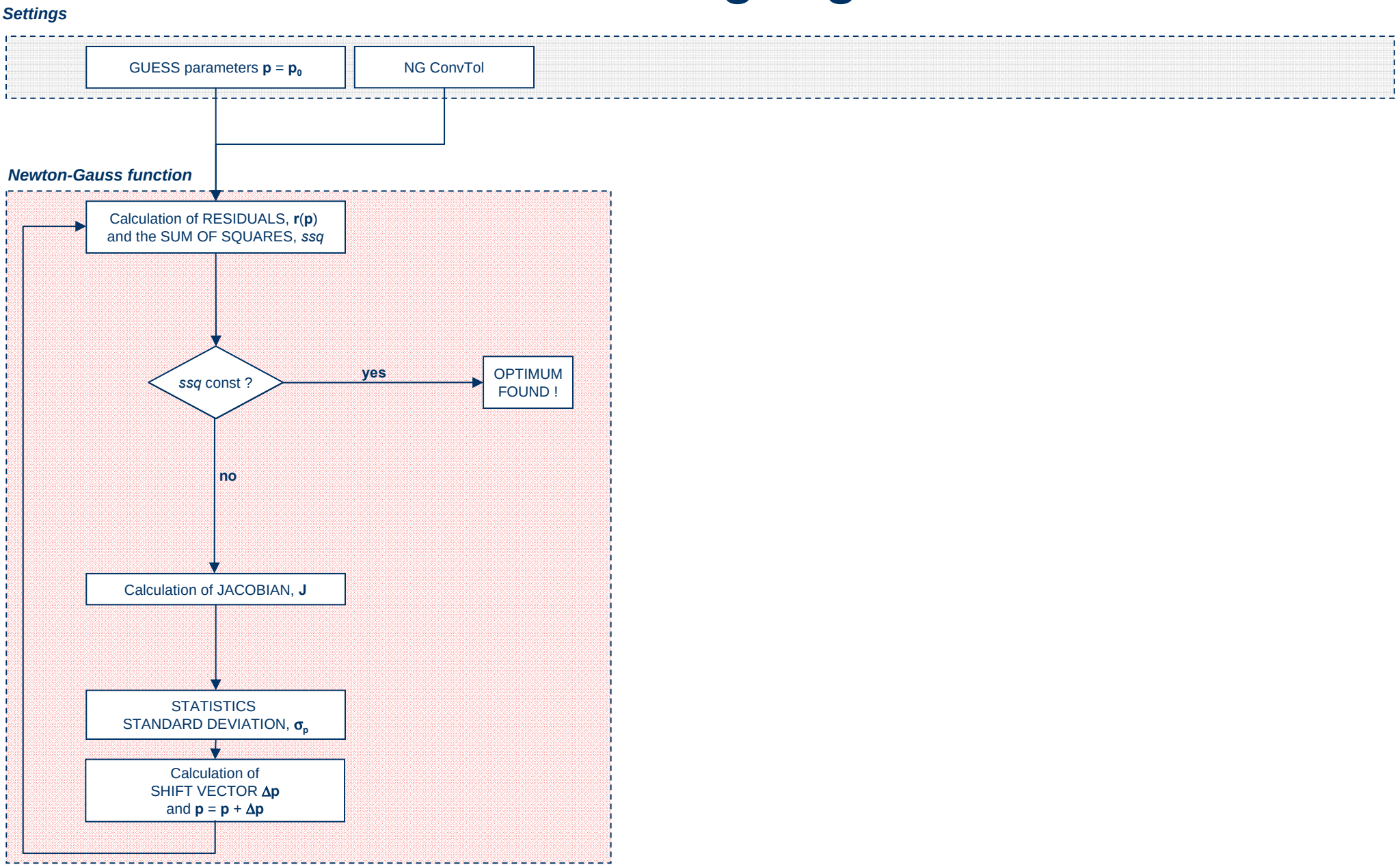
Kinetic modeling algorithm

Kinetic modeling algorithm

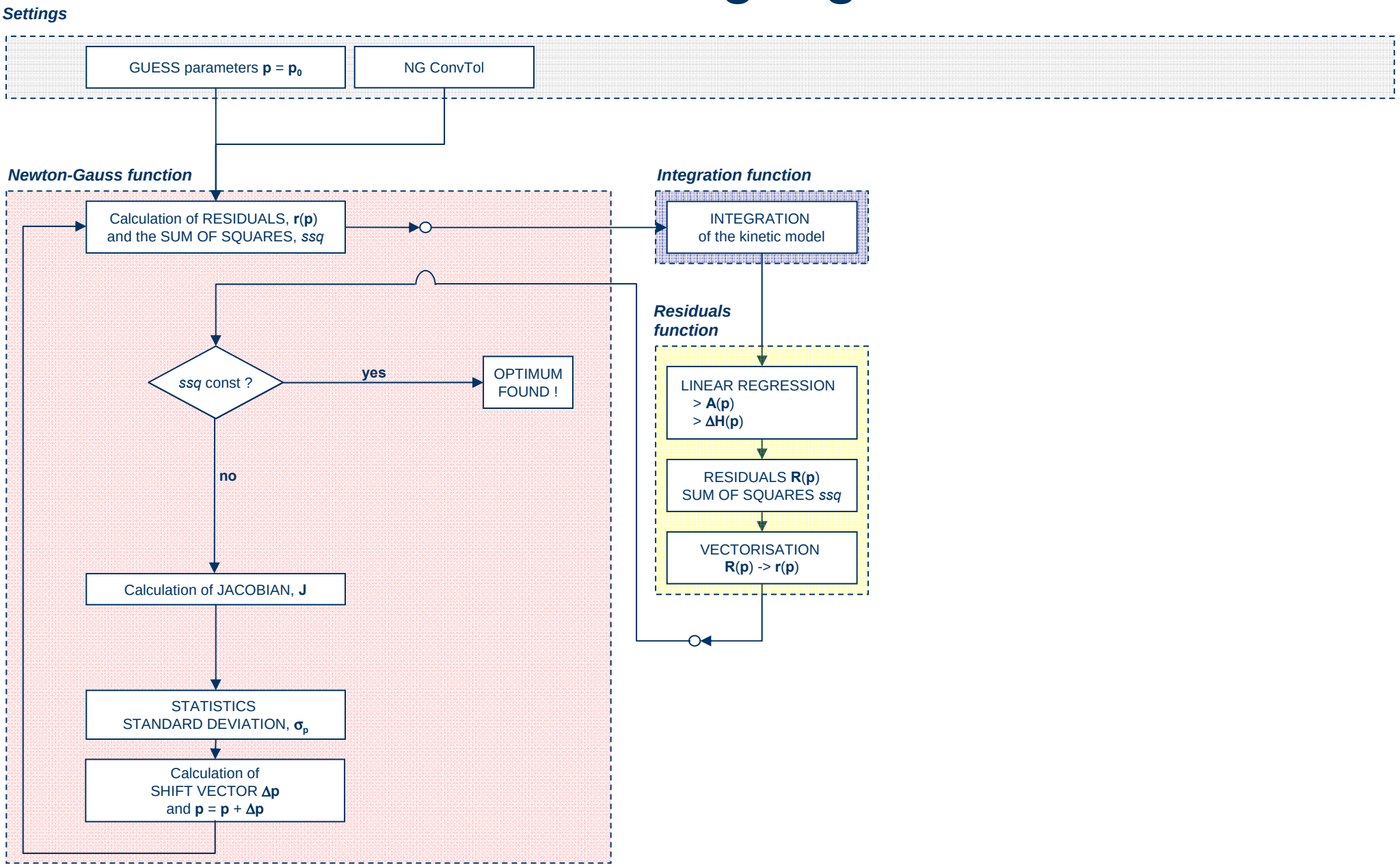
Settings

GUESS parameters $\mathbf{p} = \mathbf{p_0}$

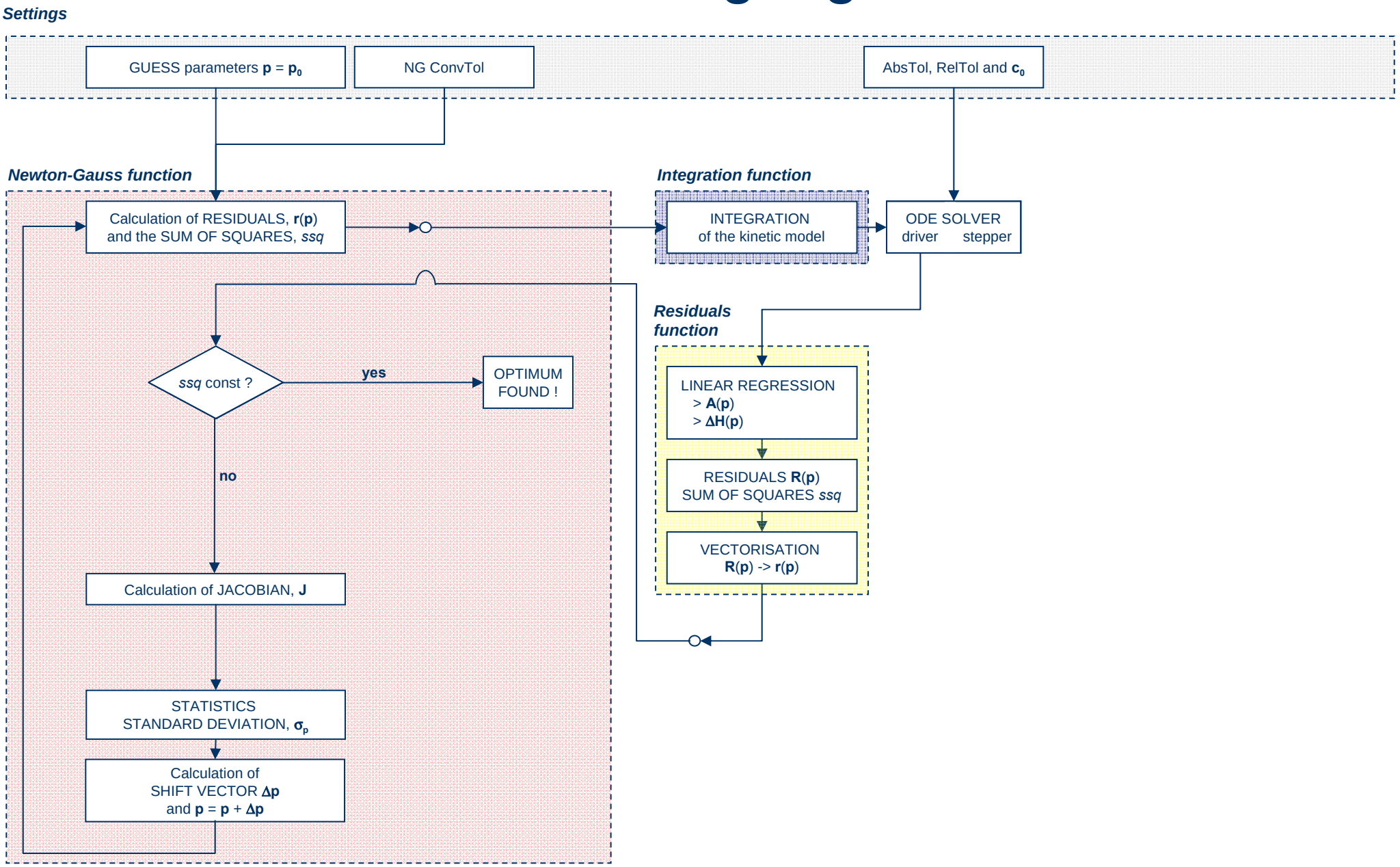
Kinetic modeling algorithm



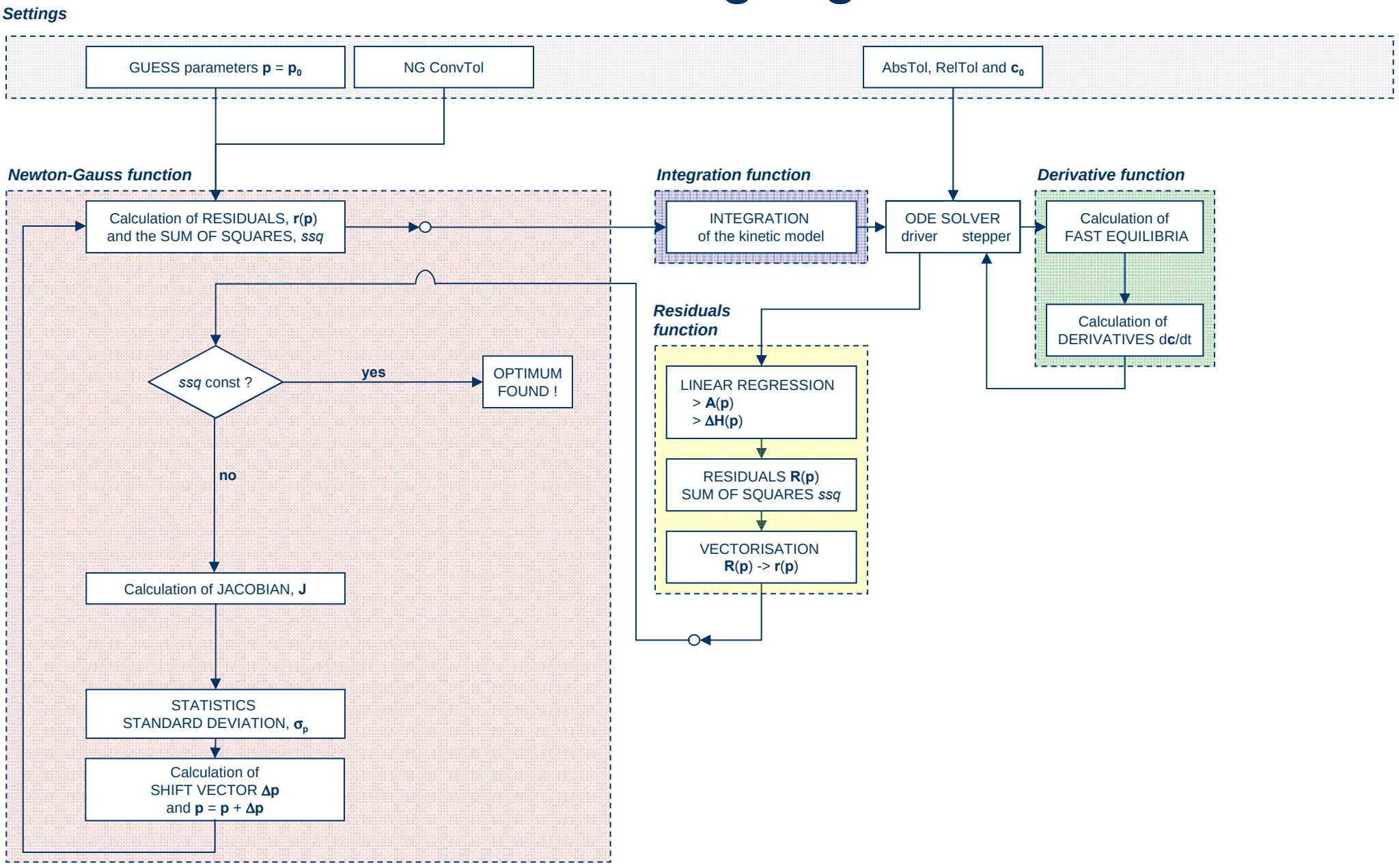
Kinetic modeling algorithm



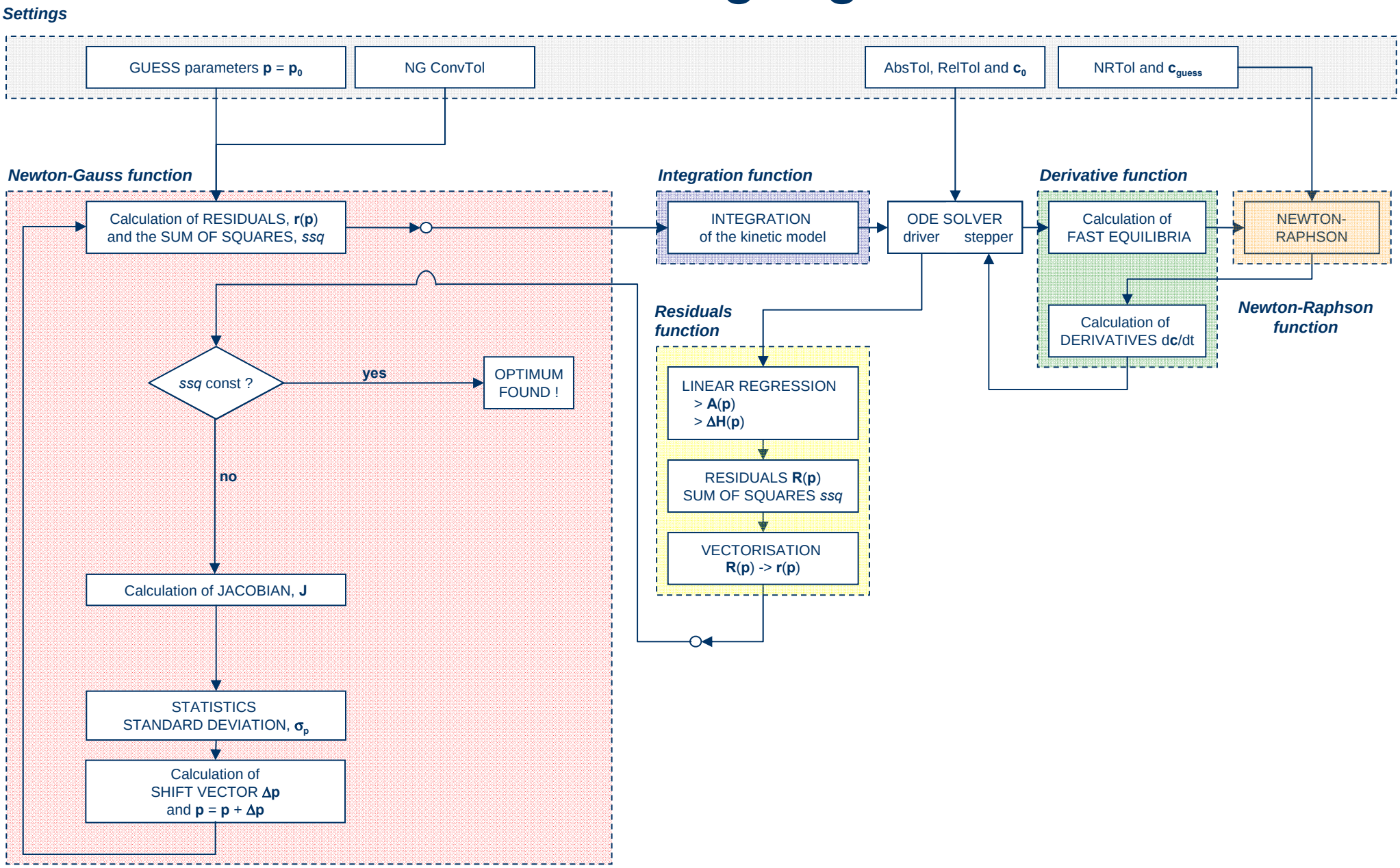
Kinetic modeling algorithm



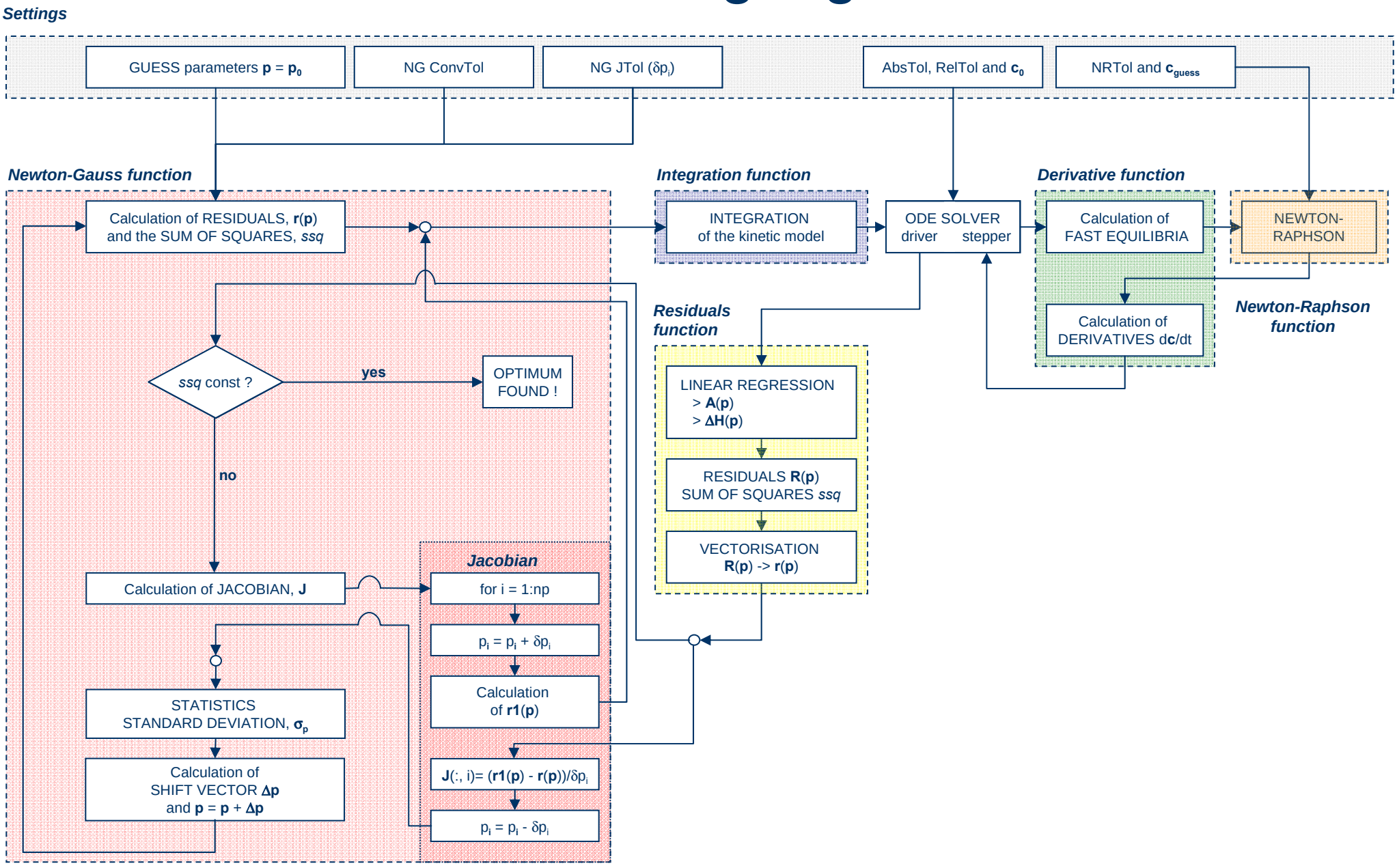
Kinetic modeling algorithm



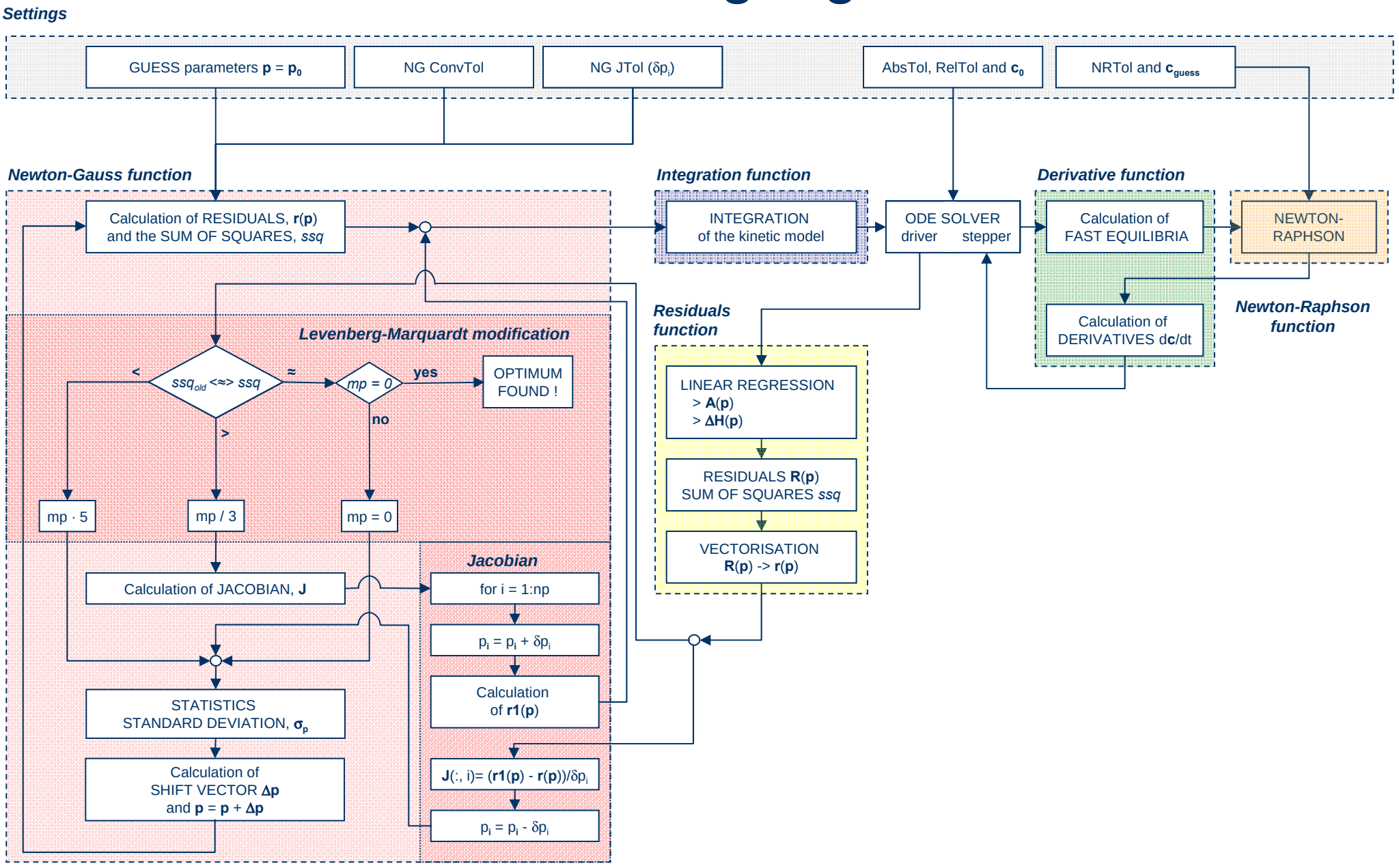
Kinetic modeling algorithm



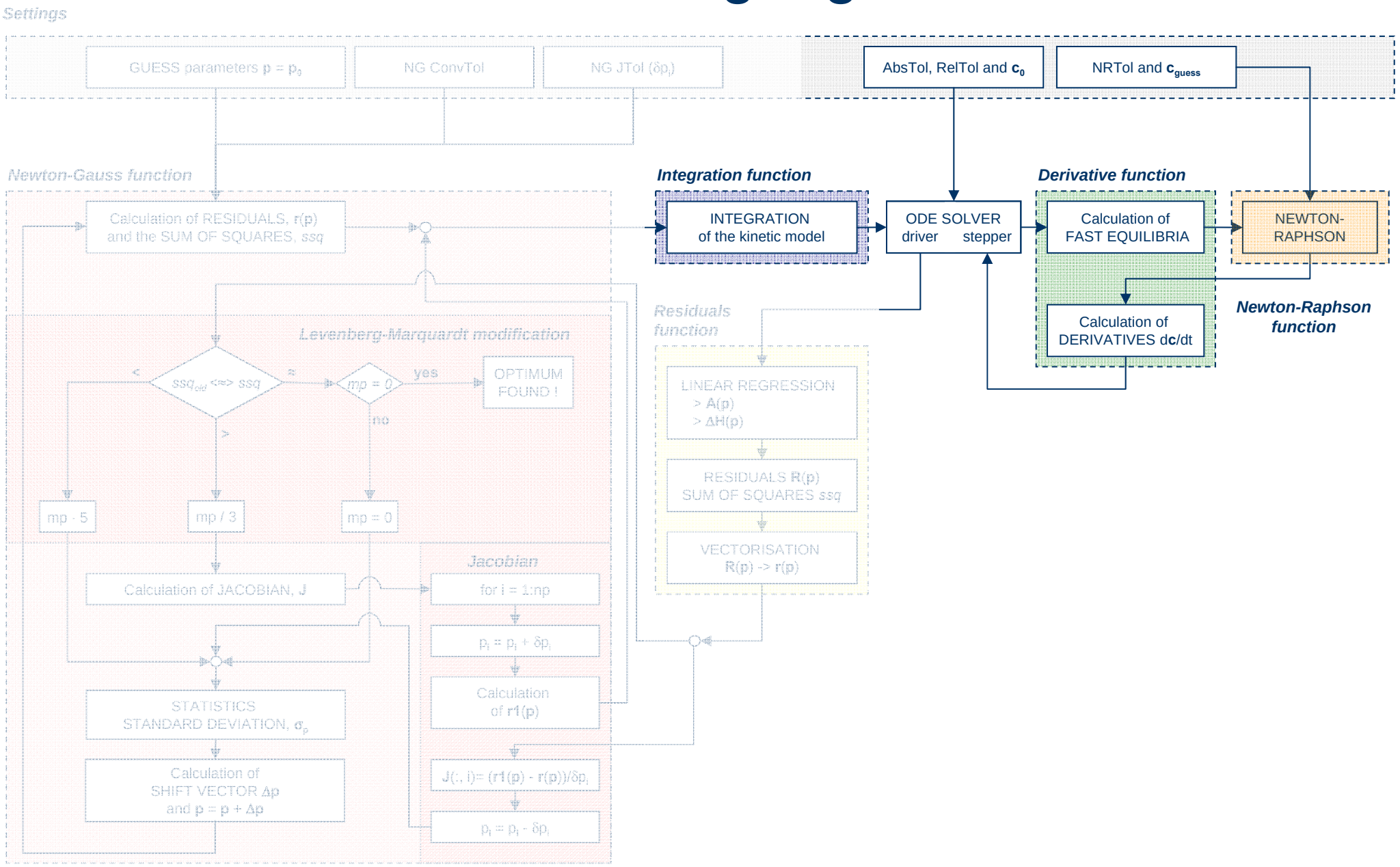
Kinetic modeling algorithm



Kinetic modeling algorithm



Kinetic modeling algorithm



Setup the kinetic model

$$A + B \xrightarrow{k_1} C$$

$$2C \xrightarrow{k_2} D$$

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$\mathbf{X_r}$ =

	A	B	C	D
k_1	1	1	0	0
k_2	0	0	2	0

nc (A, B, C, D)

np (k_1, k_2)

Setup the kinetic model

$$A + B \xrightarrow{k_1} C$$

$$2C \xrightarrow{k_2} D$$

$\mathbf{X_r}$ =

	A	B	C	D
k_1	1	1	0	0
k_2	0	0	2	0

nc (columns) × *np* (rows)

$\mathbf{X_p}$ =

	A	B	C	D
k_1	0	0	1	0
k_2	0	0	0	1

Setup the kinetic model

$$A + B \xrightarrow{k_1} C$$

$$2C \xrightarrow{k_2} D$$

$\mathbf{X_r}$ =

	A	B	C	D
k_1	1	1	0	0
k_2	0	0	2	0

nc (A, B, C, D) × *np* (k_1, k_2)

$\mathbf{X_p}$ =

	A	B	C	D
k_1	0	0	1	0
k_2	0	0	0	1

$\mathbf{X = X_P - X_R}$ =

	A	B	C	D
k_1	-1	-1	1	0
k_2	0	0	-2	1

Setup the kinetic model

$$A + B \xrightarrow{k_1} C$$

$$2C \xrightarrow{k_2} D$$

$\mathbf{X_r}$ = (nc = 4, np = 2)

	A	B	C	D
k_1	1	1	0	0
k_2	0	0	2	0

$\mathbf{X_p}$ =

	A	B	C	D
k_1	0	0	1	0
k_2	0	0	0	1

$\mathbf{X = X_P - X_R}$ =

	A	B	C	D
k_1	-1	-1	1	0
k_2	0	0	-2	1

Rate laws

$$r_j = k_j \prod_{i=1}^{nc} c_i^{\mathbf{X_r}_{j,i}} \qquad for\ j = 1 : np$$

$$r_1 = k_1 \cdot [A]_t \cdot [B]_t$$

$$r_2 = k_2 \cdot [C]_t^2$$

Setup the kinetic model

$$A + B \xrightarrow{k_1} C$$
$$2C \xrightarrow{k_2} D$$

$\mathbf{X_r}$ =

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(nc columns, np rows)

$\mathbf{X_p}$ =

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$$r_2 = k_2 \cdot [C]_t^2$$

Derivatives

$$\frac{dc_i}{dt} = \sum_{j=1}^{np} \mathbf{X}_{j,i} \cdot r_j \qquad for\ i = 1:nc$$

$$\frac{d[A]_t}{dt} = \frac{d[B]_t}{dt} = -r_1$$
$$\frac{d[C]_t}{dt} = r_1 - 2 \cdot r_2$$
$$\frac{d[D]_t}{dt} = r_2$$

Setup the kinetic model

$$A + B \xrightarrow{k_1} C$$
$$2C \xrightarrow{k_2} D$$

$\mathbf{X_r}$ =

	A	B	C	D
k_1	1	1	0	0
k_2	0	0	2	0

(*nc* columns, *np* rows)

$\mathbf{X_p}$ =

	A	B	C	D
k_1	0	0	1	0
k_2	0	0	0	1

$\mathbf{X = X_P - X_R}$ =

	A	B	C	D
k_1	-1	-1	1	0
k_2	0	0	-2	1

Rate laws

$$r_j = k_j \prod_{i=1}^{nc} c_i^{\mathbf{X_r}_{j,i}} \qquad for\ j = 1 : np$$

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Derivatives

$$\frac{dc_i}{dt} = \sum_{j=1}^{np} \mathbf{X}_{j,i} \cdot r_j \qquad for\ i = 1 : nc$$

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$$\frac{d[D]_t}{dt} = r_2$$

Most of the time : no analytical solution for this system of ODEs
→ **Numerical integration**

Dosing events

$$A + B \xrightarrow{k_1} C$$

$$2C \xrightarrow{k_2} D$$

In case of dosing, the set of ODEs is modified accordingly :

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Rate laws $r_j = k_j \prod_{i=1}^{ns} c_i^{\mathbf{X_r}_{j,i}}$ *for* $j = 1 : np$

$$r_1 = k_1 \cdot [A]_t \cdot [B]_t$$
$$r_2 = k_2 \cdot [C]_t^{\,2}$$

Dosing events

$A + B \xrightarrow{k_1} C$

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In case of dosing, the set of ODEs is modified accordingly :

Rate laws

$$r_j = k_j \prod_{i=1}^{ns} c_i^{\mathbf{X_r}_{j,i}} \qquad for\ j = 1:np$$

$$r_1 = k_1 \cdot [A]_t \cdot [B]_t$$
$$r_2 = k_2 \cdot [C]_t^{\,2}$$

Derivatives

$$\frac{dc_i}{dt} = \left(\sum_{j=1}^{np} \mathbf{X}_{j,i} \cdot r_j \right) + \frac{F}{V_t} \cdot \left(c_i^{feed} - c_i \right)$$

$$\frac{dV}{dt} = F \qquad for\ i = 1:nc$$

$$\frac{d[A]_t}{dt} = -r_1 + \frac{F}{V_t} \cdot \left([A]^{feed} - [A]_t \right)$$
$$\frac{d[B]_t}{dt} = -r_1 + \frac{F}{V_t} \cdot \left([B]^{feed} - [B]_t \right)$$
$$\frac{d[C]_t}{dt} = r_1 - 2 \cdot r_2 + \frac{F}{V_t} \cdot \left([C]^{feed} - [C]_t \right)$$
$$\frac{d[D]_t}{dt} = r_2 + \frac{F}{V_t} \cdot \left([D]^{feed} - [D]_t \right)$$
$$\frac{dV}{dt} = F$$

Dosing events

$A + B \xrightarrow{k_1} C$

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$$r_j = k_j \prod_{i=1}^{ns} c_i^{\mathbf{X_r}_{j,i}} \qquad for\ j = 1:np$$

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Derivatives

$$\frac{dc_i}{dt} = \left(\sum_{j=1}^{np} \mathbf{X}_{j,i} \cdot r_j \right) + \frac{F}{V_t} \cdot \left(c_i^{feed} - c_i \right)$$

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$$\frac{d[C]_t}{dt} = r_1 - 2 \cdot r_2 + \frac{F}{V_t} \cdot \left([C]^{feed} - [C]_t \right)$$
$$\frac{d[D]_t}{dt} = r_2 + \frac{F}{V_t} \cdot \left([D]^{feed} - [D]_t \right)$$
$$\frac{dV}{dt} = F$$

The dosing event adds an additional ODE and a term to the ODEs of all species

NB : $+\frac{F}{V_t} \cdot A_{in}$ = added material

$-\frac{F}{V_t} \cdot A$ = dilution phenomenon

Numerical integration of the model

- First Approach : Euler's method

$$c_i(t+\Delta t) \approx c_i(t) + \left(\frac{dc_i}{dt}\right)_t \cdot \Delta t$$

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Applied to our specific example *without dosing*

$$A + B \xrightarrow{k_1} C$$
$$2C \xrightarrow{k_2} D$$

$$[C]_{t+\Delta t} \approx [C]_t + \left(\frac{d[C]_t}{dt}\right)_t \cdot \Delta t$$
$$= [C]_t + \left(k_1 \cdot [A]_t \cdot [B]_t - k_2 \cdot [C]_t^{\,2}\right) \cdot \Delta t$$

Numerical integration of the model

- First Approach : Euler's method

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Applied to our specific example *without dosing*

$$A + B \xrightarrow{k_1} C$$
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$$[C]_{t+\Delta t} \approx [C]_t + \left(\frac{d[C]_t}{dt}\right)_t \cdot \Delta t$$
$$= [C]_t + \left(k_1 \cdot [A]_t \cdot [B]_t - k_2 \cdot [C]_t^{\,2}\right) \cdot \Delta t$$

- Nowadays, more sophisticated integration methods exist (e.g. ode45) with a ***stepsize control***

Stepsize control and stiff problems

- In stepsize controlled ODE solvers, the stepsize is adjusted at each step to meet the user-specified accuracy

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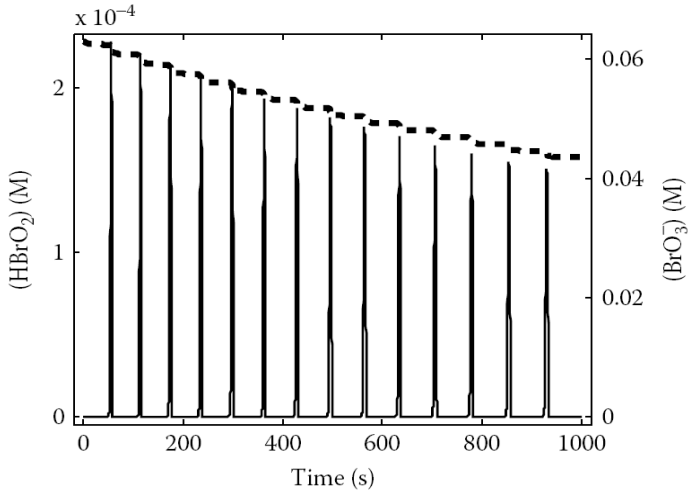
- In stepsize controlled ODE solvers, the stepsize is adjusted at each step to meet the user-specified accuracy

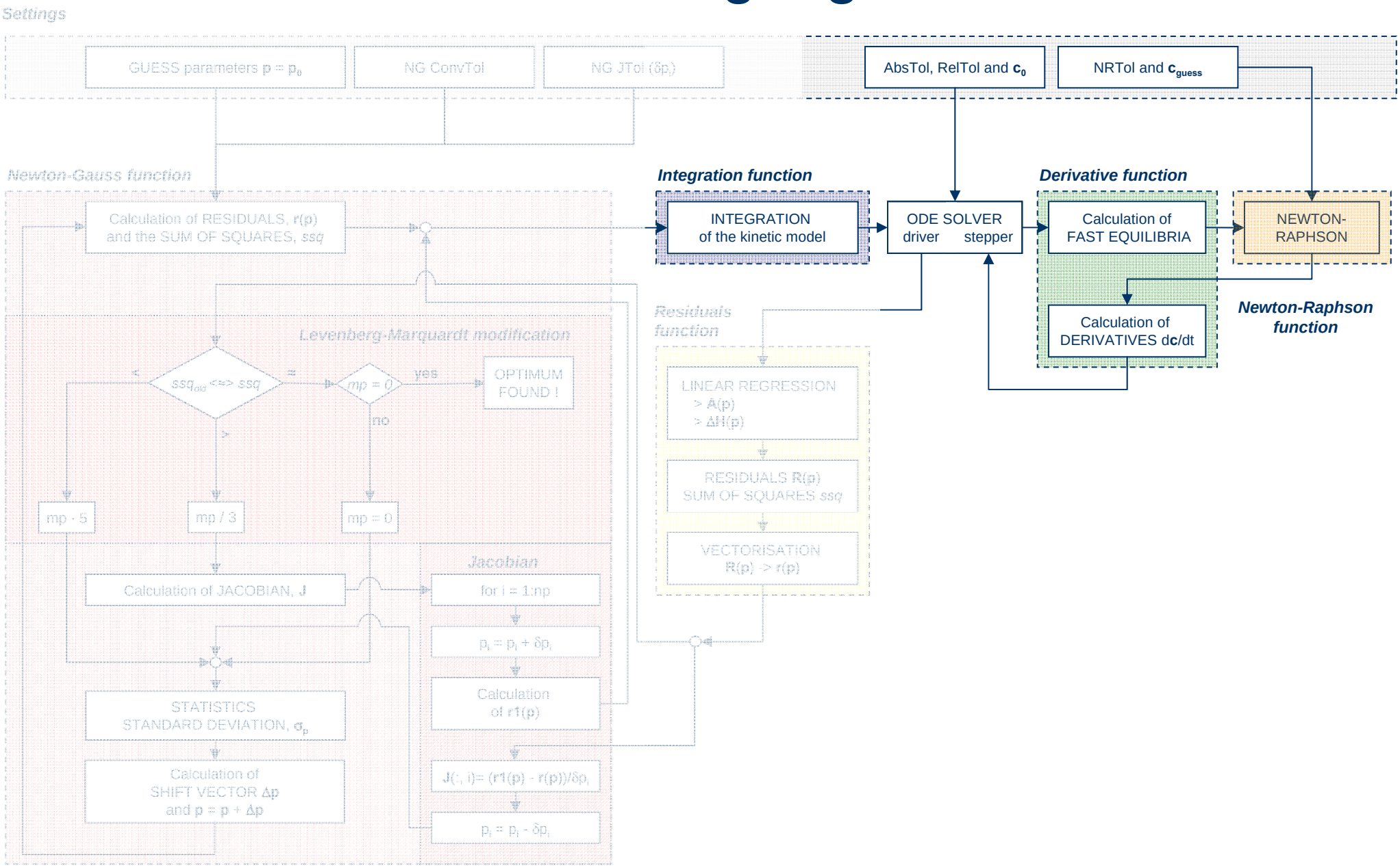
- The accuracy is measured with absolute (*AbsTol*) and relative (*RelTol*) tolerance's values.

- For some kinetic models, the slopes of the concentration profiles are dramatically different (stiff problem) and require the use of a **stiff ODE solver** (eg. ode15s)



Gemperline, P. (2006). Practical Guide to Chemometrics (2nd edition), Taylor and Francis, Boca Raton, USA.

Kinetic modeling algorithm



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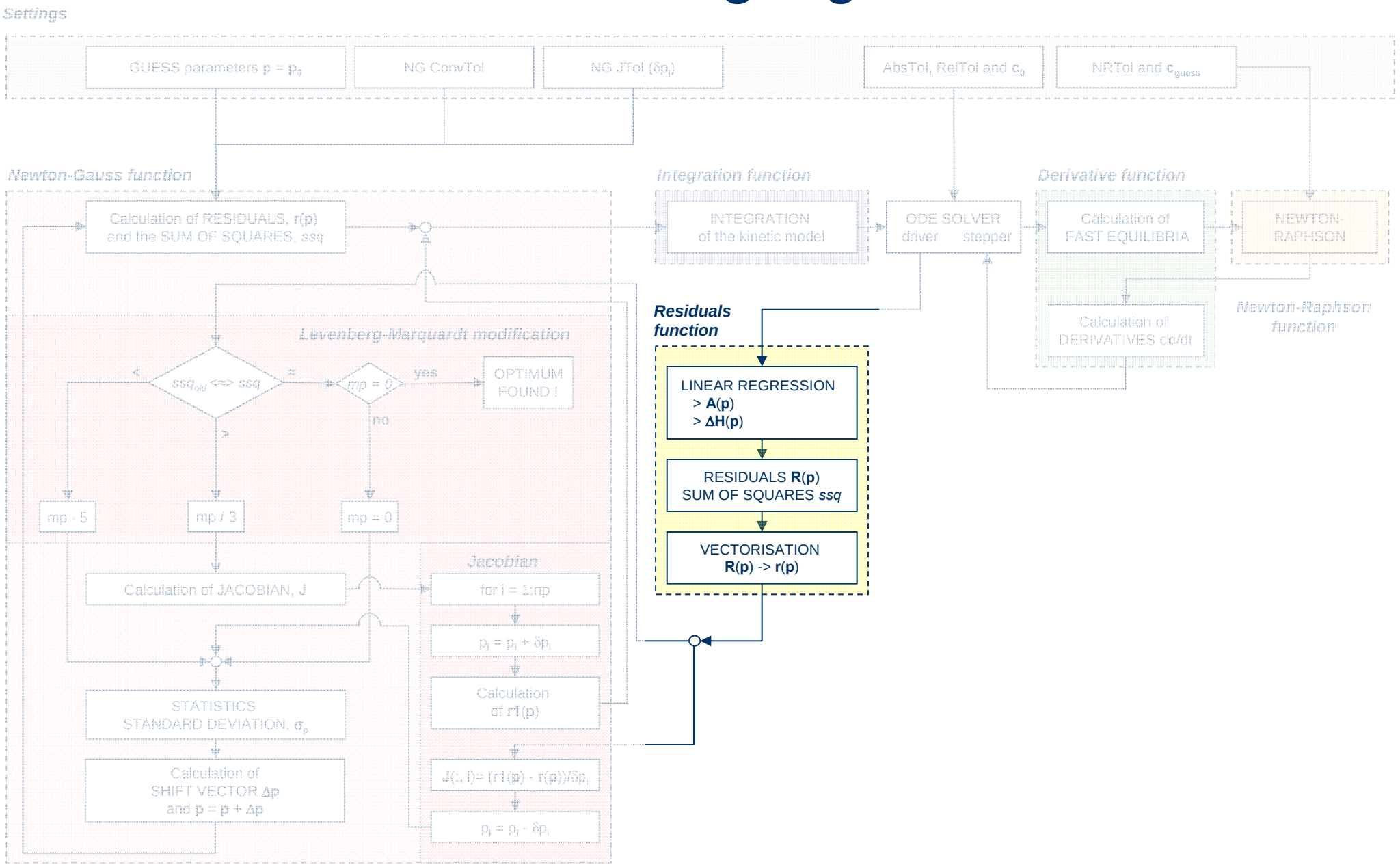

Linear and Nonlinear parameters

GENERAL CONSIDERATION

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- If $S(\mathbf{p})$ is a measured signal depending on the parameters vector **p**

  Linear parameters are defined as: $\left(\frac{\partial S(\mathbf{p})}{\partial p_i}\right)_{p_{j\neq i}} \neq f(p_i)$

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APPLIED TO KINETIC MODELING :

- **C** and **A** are LINEAR parameters with respect to **Y (Beer's law)** $\mathbf{Y} = \mathbf{C} \cdot \mathbf{A}$
- $\mathbf{d\xi_{mol}/dt}$ and $\mathbf{\Delta H_r}$ are LINEAR parameters with respect to **q (Reaction heat balance)** $\mathbf{q} = \frac{\mathbf{d\xi_{mol}}}{\mathbf{dt}} \cdot \left(-\mathbf{\Delta H_R}\right)$
- Rate constants are NONLINEAR parameters with respect to **C** and $\mathbf{d\xi_{mol}/dt}$ so are they for **Y** and for **q**

Separation of linear parameters – LINEAR REGRESSION

- At each iteration, the linear parameters are calculated in one step as the best linear estimate in the least squares sense

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Pure spectra
(from Spectroscopy)

$$\mathbf{A} = \mathbf{C}^{+} \cdot \mathbf{Y} = \left(\mathbf{C}^{t} \cdot \mathbf{C}\right)^{-1} \cdot \mathbf{C}^{t} \cdot \mathbf{Y}$$

Enthalpies
(from Calorimetry)

$$\Delta \mathbf{H}_{R} = -\left(\frac{d\xi_{mol}}{dt}\right)^{+} \cdot \mathbf{q} = -\left(\left(\frac{d\xi_{mol}}{dt}\right)^{t} \cdot \left(\frac{d\xi_{mol}}{dt}\right)\right)^{-1} \cdot \left(\frac{d\xi_{mol}}{dt}\right)^{t} \cdot \mathbf{q}$$

Remarks :

- The above formula is a multidimensional linear regression in a matrix notation
- The *superscript* + is meant for the PSEUDO-INVERSE.
  As the matrices are not square, the inverse is not defined.

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THIS LINEAR REGRESSION MAKES ANY CALIBRATION OF THE ABSORPTIVITIES REDUNDANT !

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Calorimetry :	$\mathbf{r}_{\mathbf{cal}} = \mathbf{q} - \mathbf{q}_{\mathbf{calc}} = \mathbf{q} - \frac{d\xi_{mol}}{dt} \cdot (-\Delta H_R)$	$ssq_{cal} = \sum_{i=1}^{nt} \mathbf{r}_{\mathbf{cal}}(i)$

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ADVANCED PROBLEM :

- Combination of the signals : $ssq_{total} = w \cdot ssq_{spec} + (1-w) \cdot ssq_{cal} \quad , \quad w = ?$

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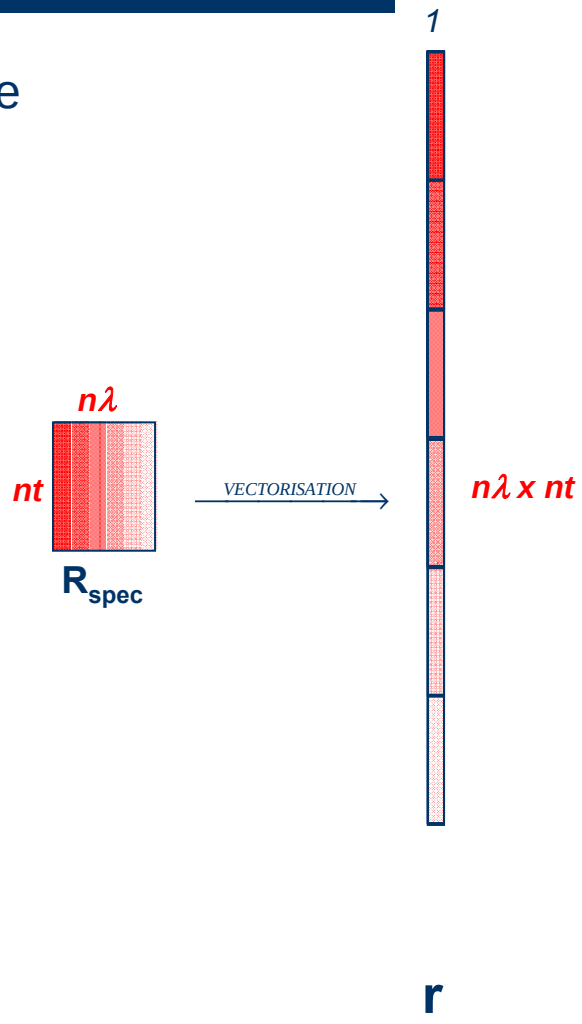

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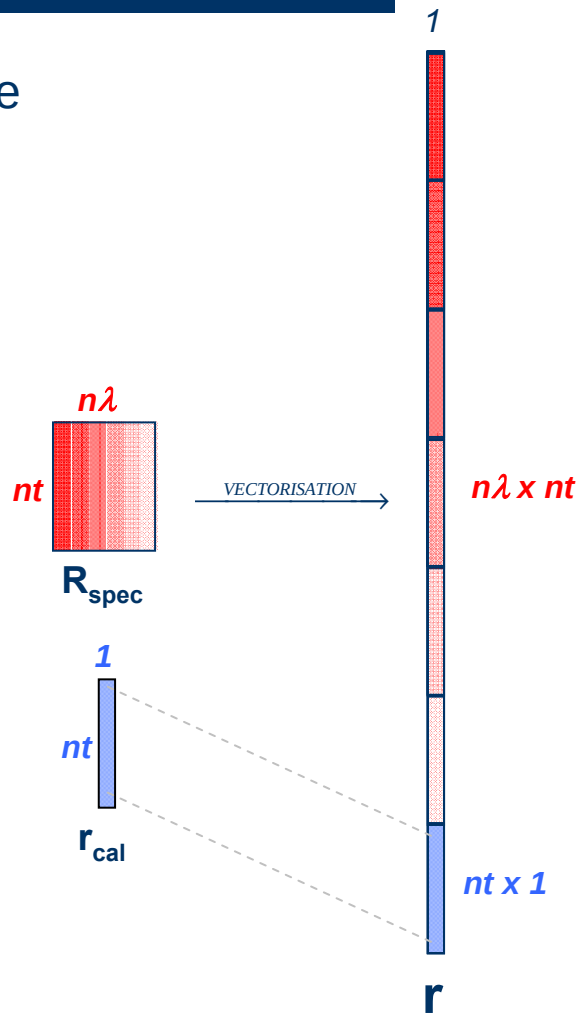

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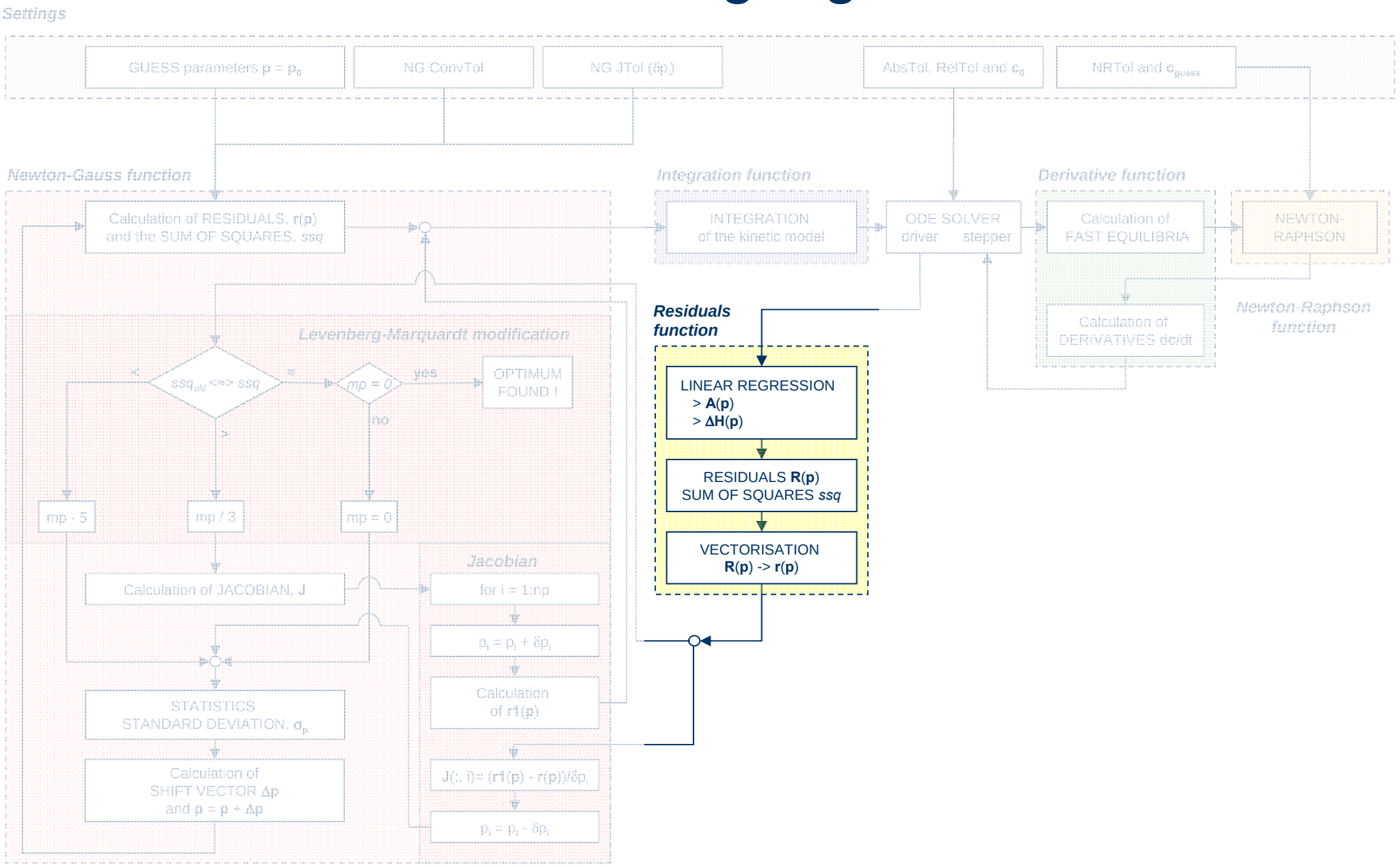

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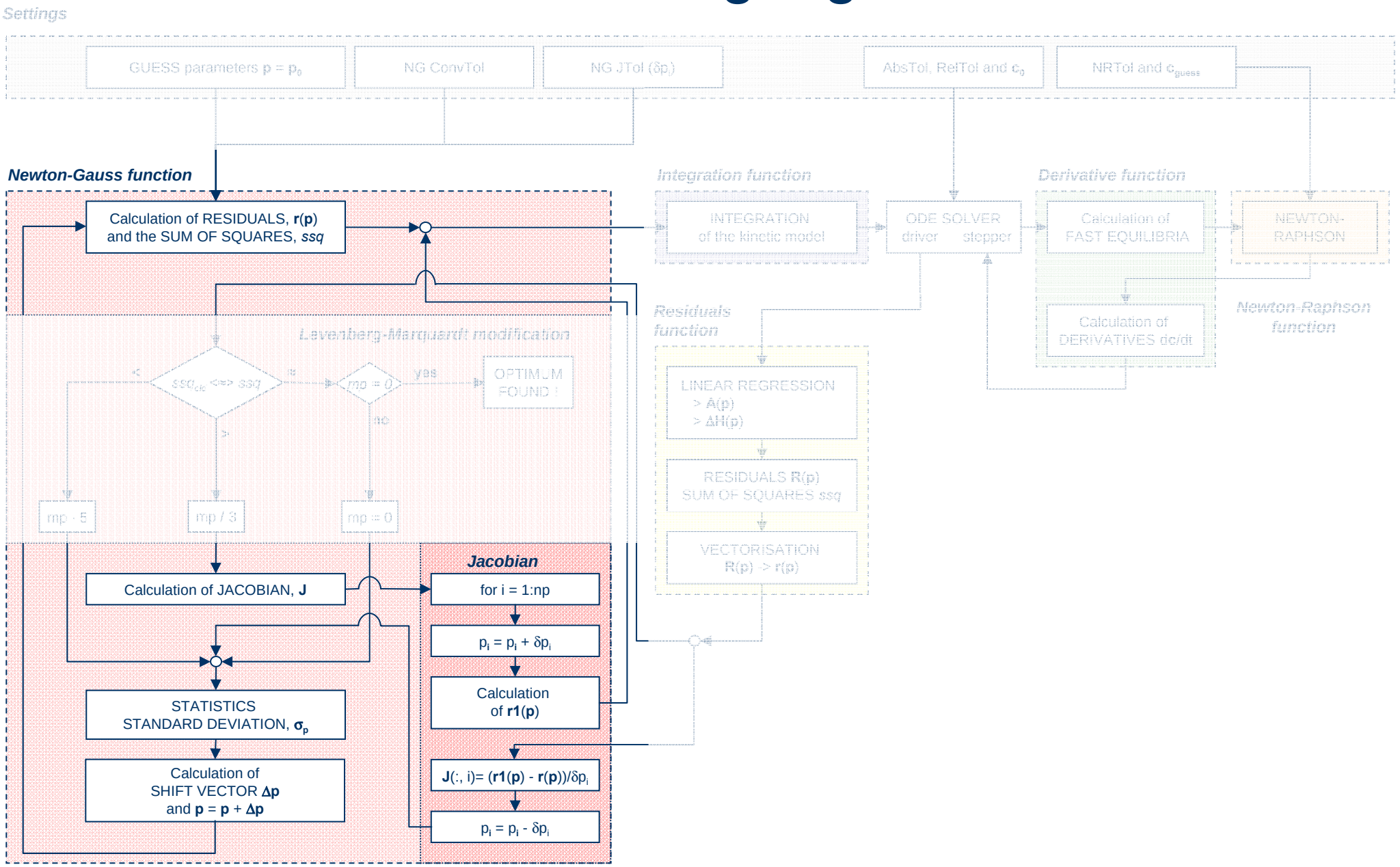

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The Newton-Gauss algorithm requires the calculation of the Jacobian

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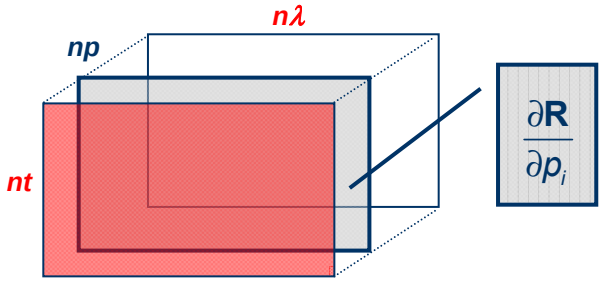

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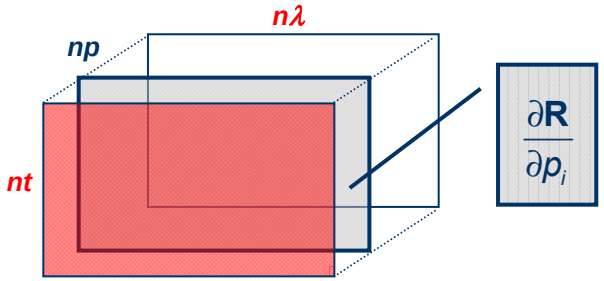

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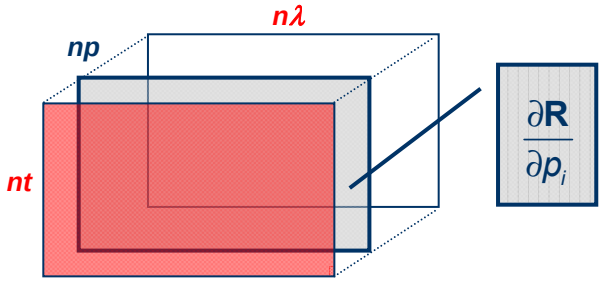

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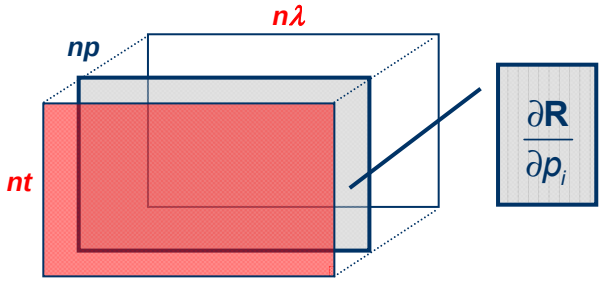

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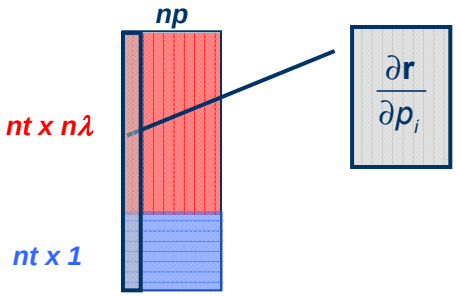

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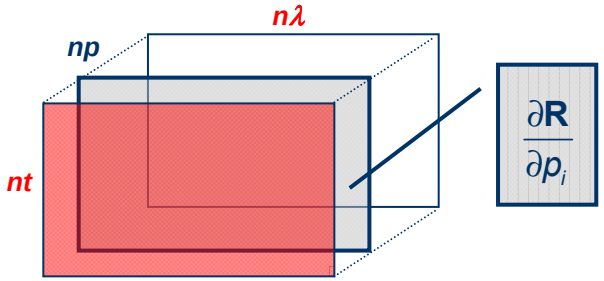

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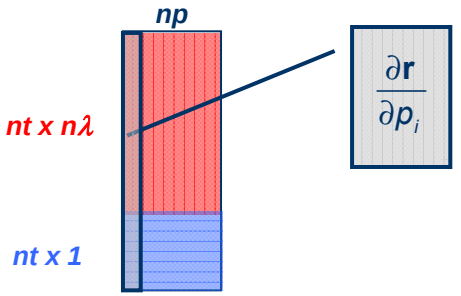

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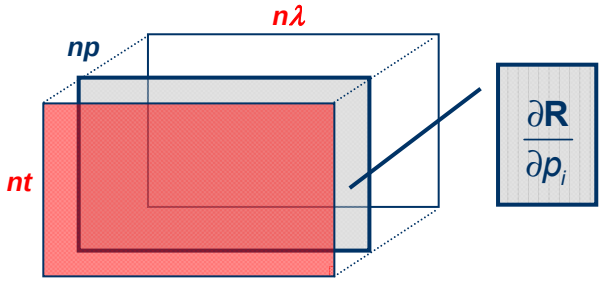

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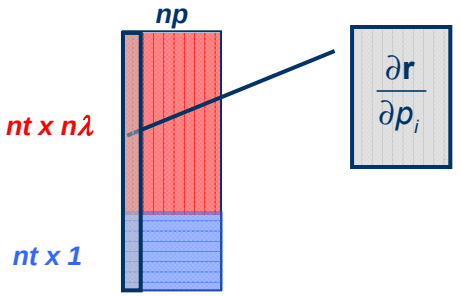

MATRIX ☺

The Jacobian is computed using a forward finite difference

$$\frac{\partial \mathbf{r}(\mathbf{p})}{\partial p_i} = \frac{\mathbf{r}(\mathbf{p} + \delta p_i) - \mathbf{r}(\mathbf{p})}{\delta p_i} \quad \text{with } \delta p_i \approx 10^{-6} \cdot p_i$$

The Hessian (statistics)

$$\mathbf{H} = \mathbf{J}^{\mathbf{t}} \cdot \mathbf{J}$$

- Knowing that $\mathbf{H} = \mathbf{J}^{\mathbf{t}} \cdot \mathbf{J}$ the shift vector can be re-written as:

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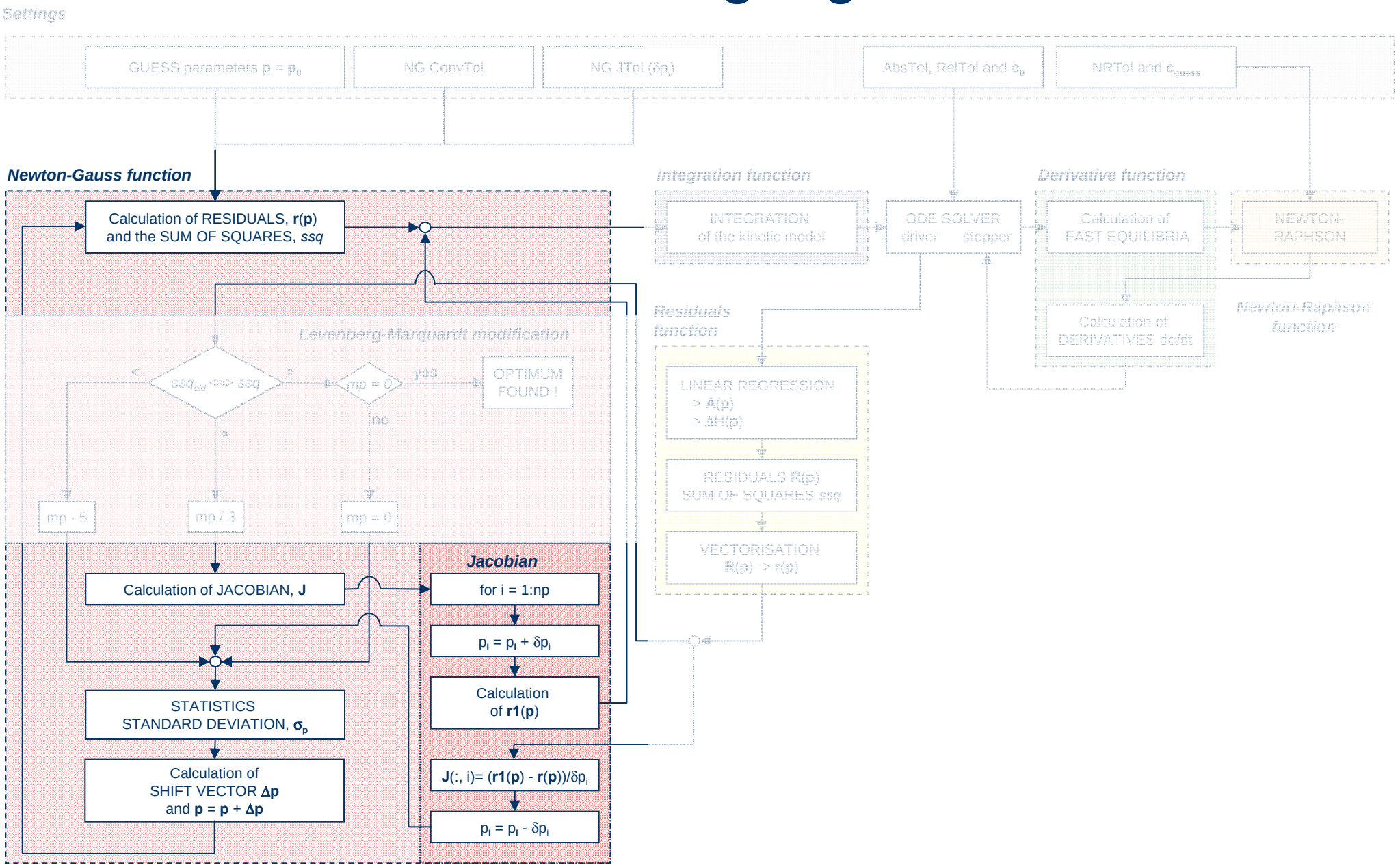
This allows the calculation of the
Standard Error of each parameter :

$$\sigma_{\mathbf{p}} = \sigma_{\mathbf{r}} \cdot \sqrt{diag\left(H^{-1}\right)} \qquad with \quad \sigma_{\mathbf{r}} = \sqrt{\frac{ssq}{df}} \approx \sigma_{\mathbf{Y}}$$

Kinetic modeling algorithm



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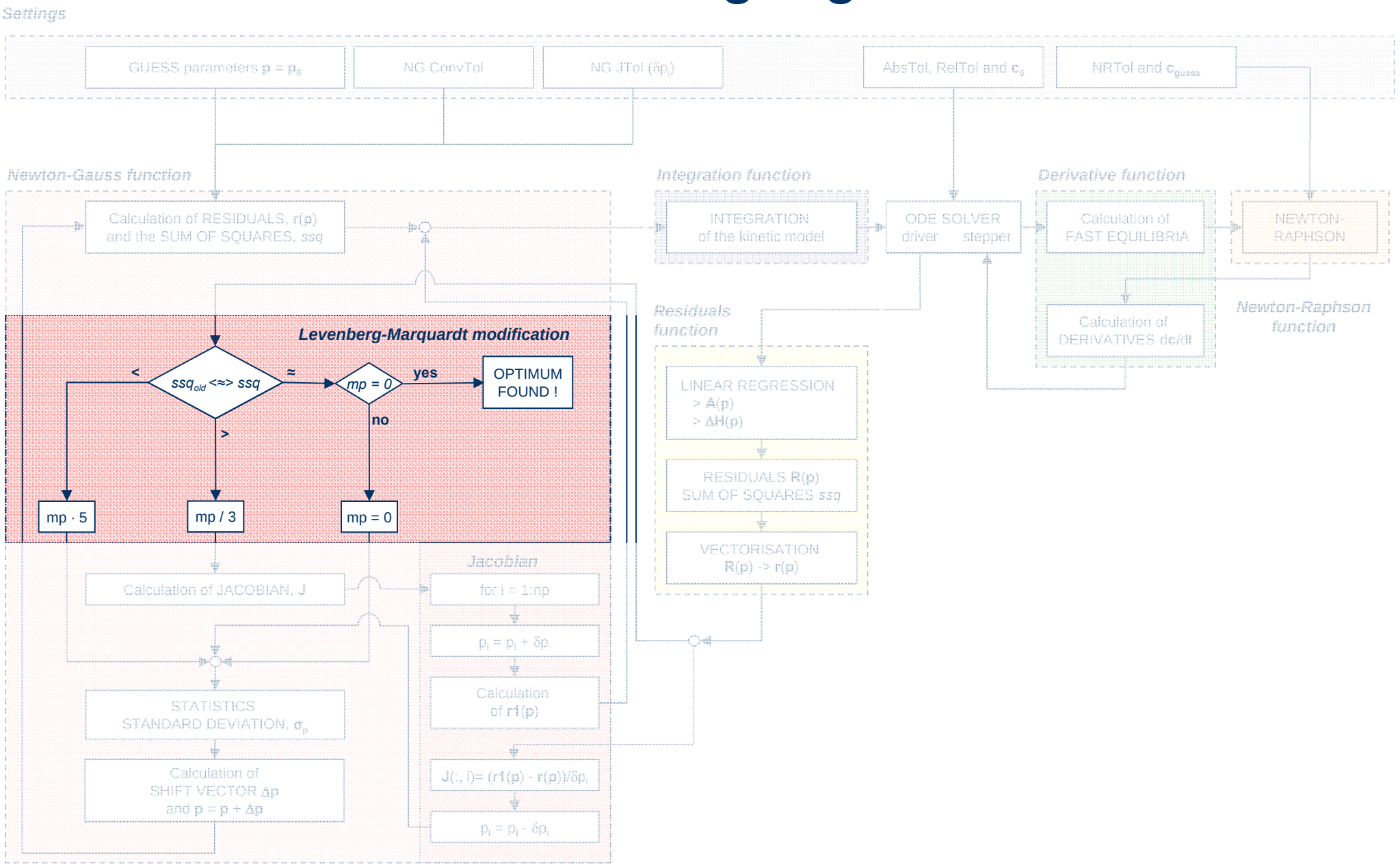

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Inverse Hessian method
(Newton-Gauss)

Is there a way to switch progressively from one method to the other ?

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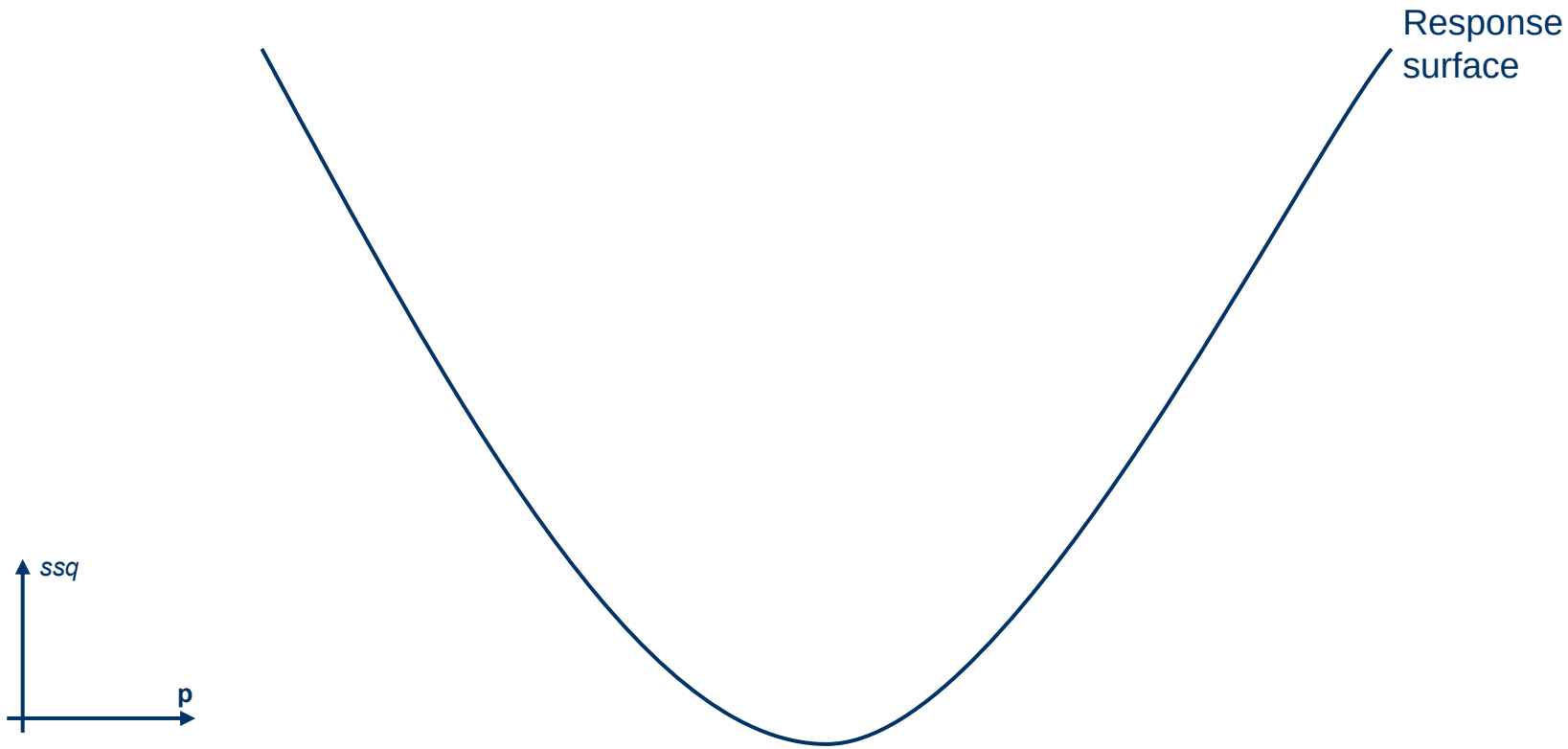
Steepest Descent Method

Geometrical interpretation on the response surface



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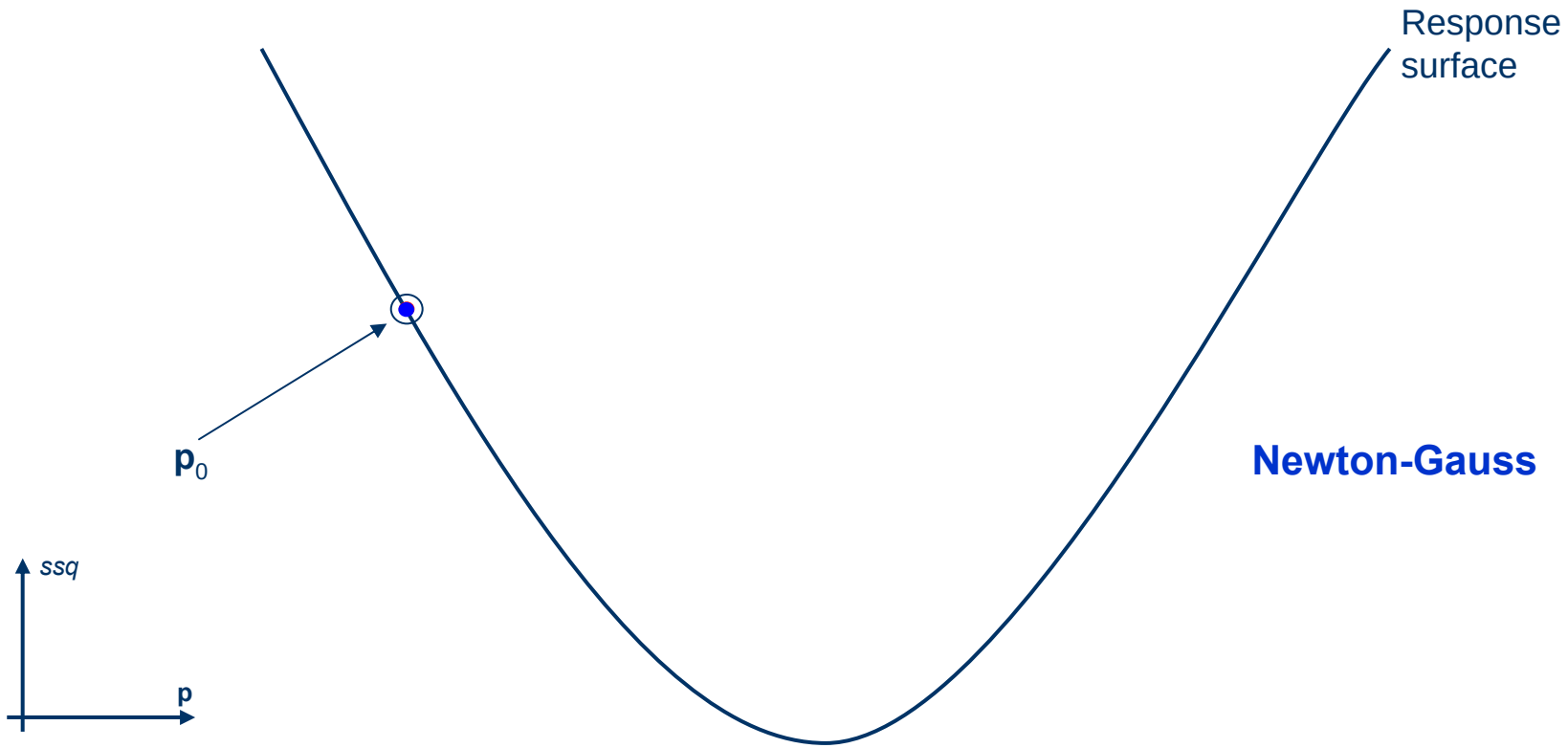
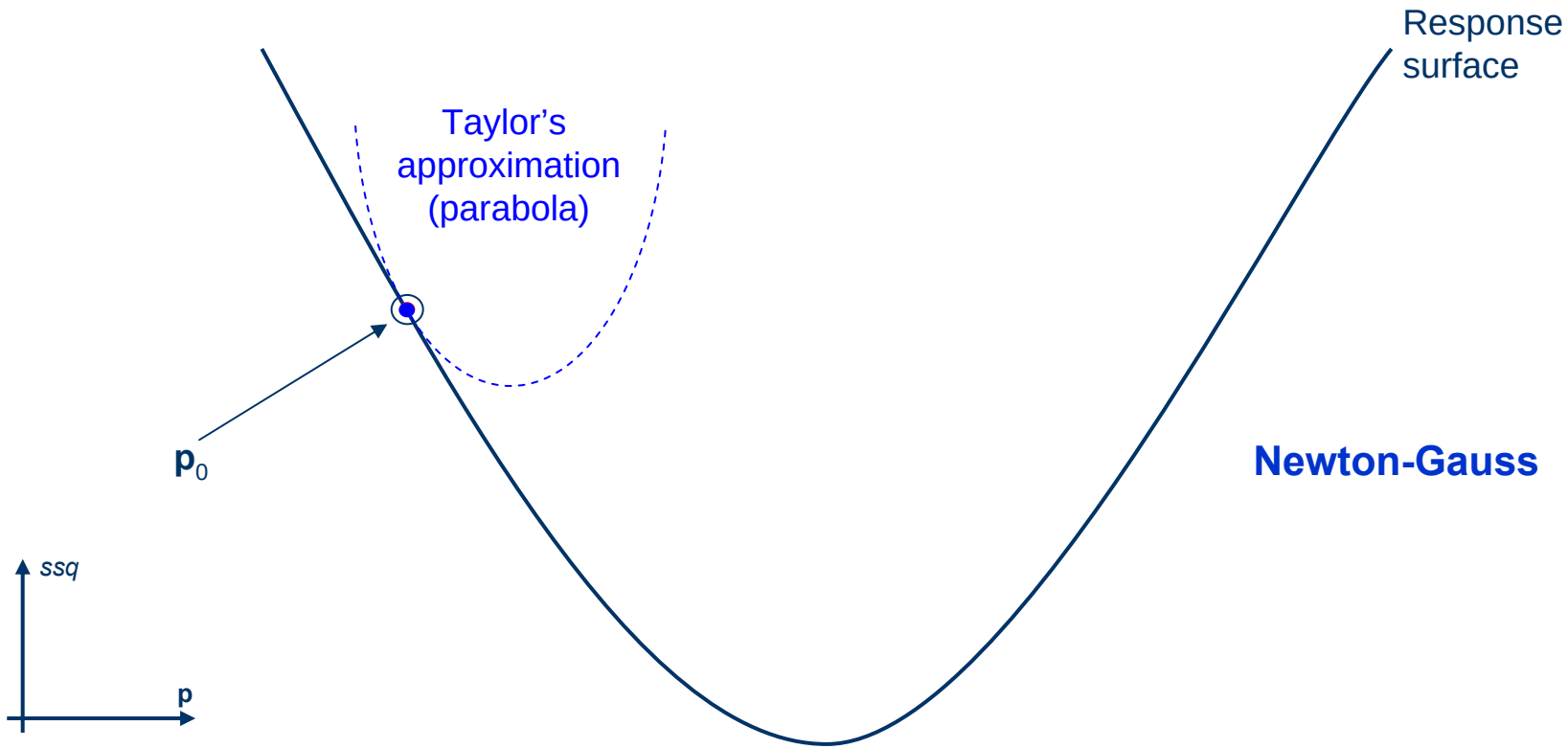
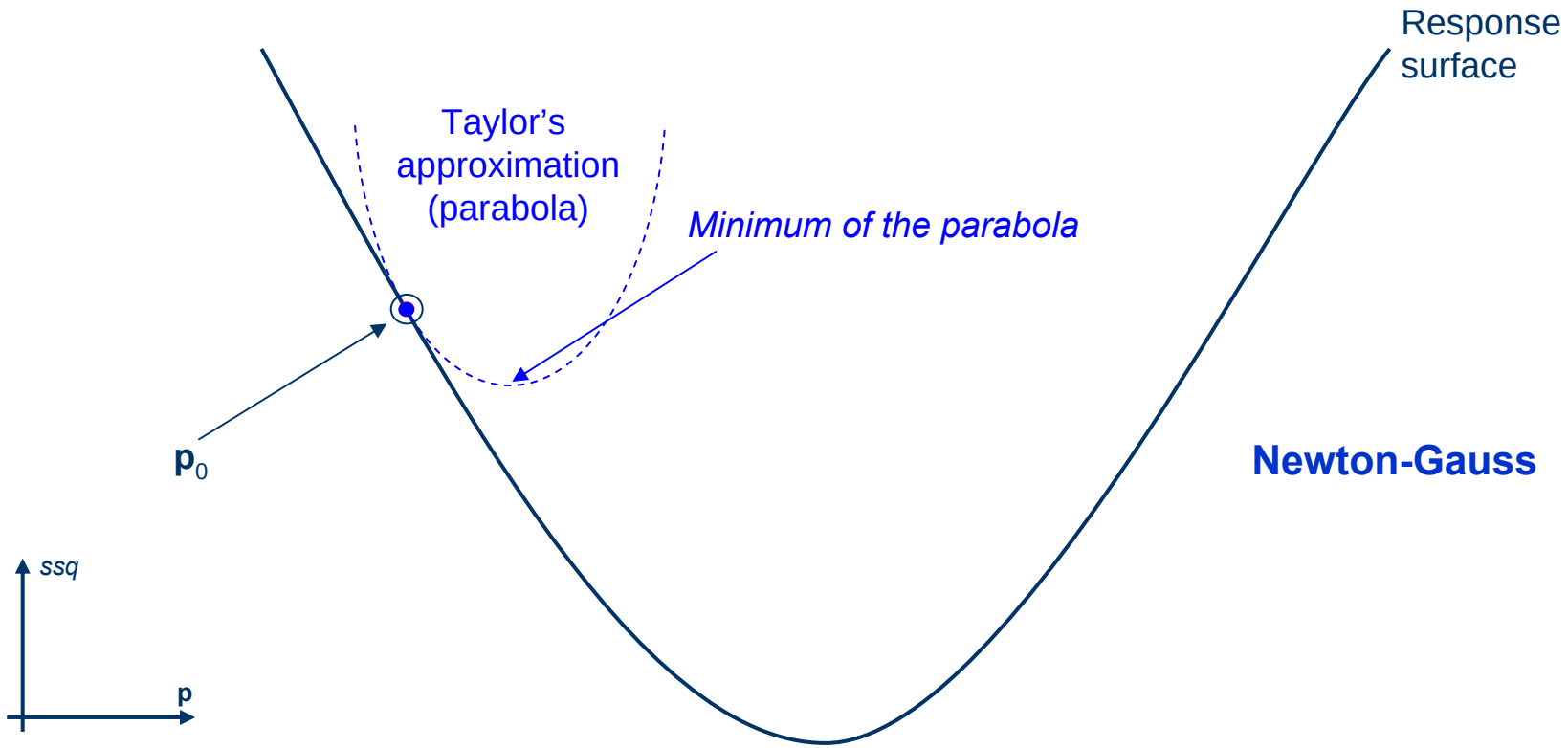


Newton-Gauss

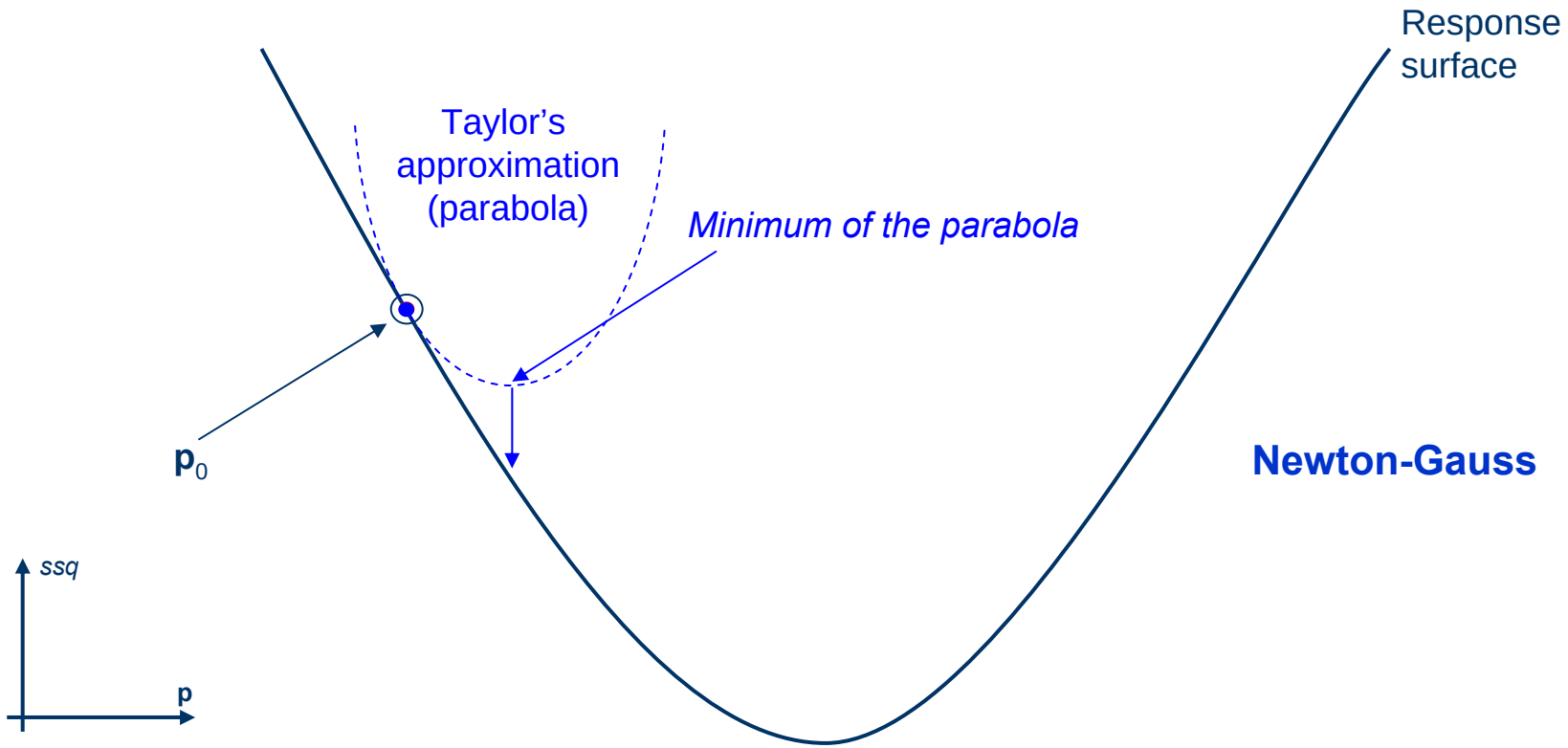
Geometrical interpretation on the response surface



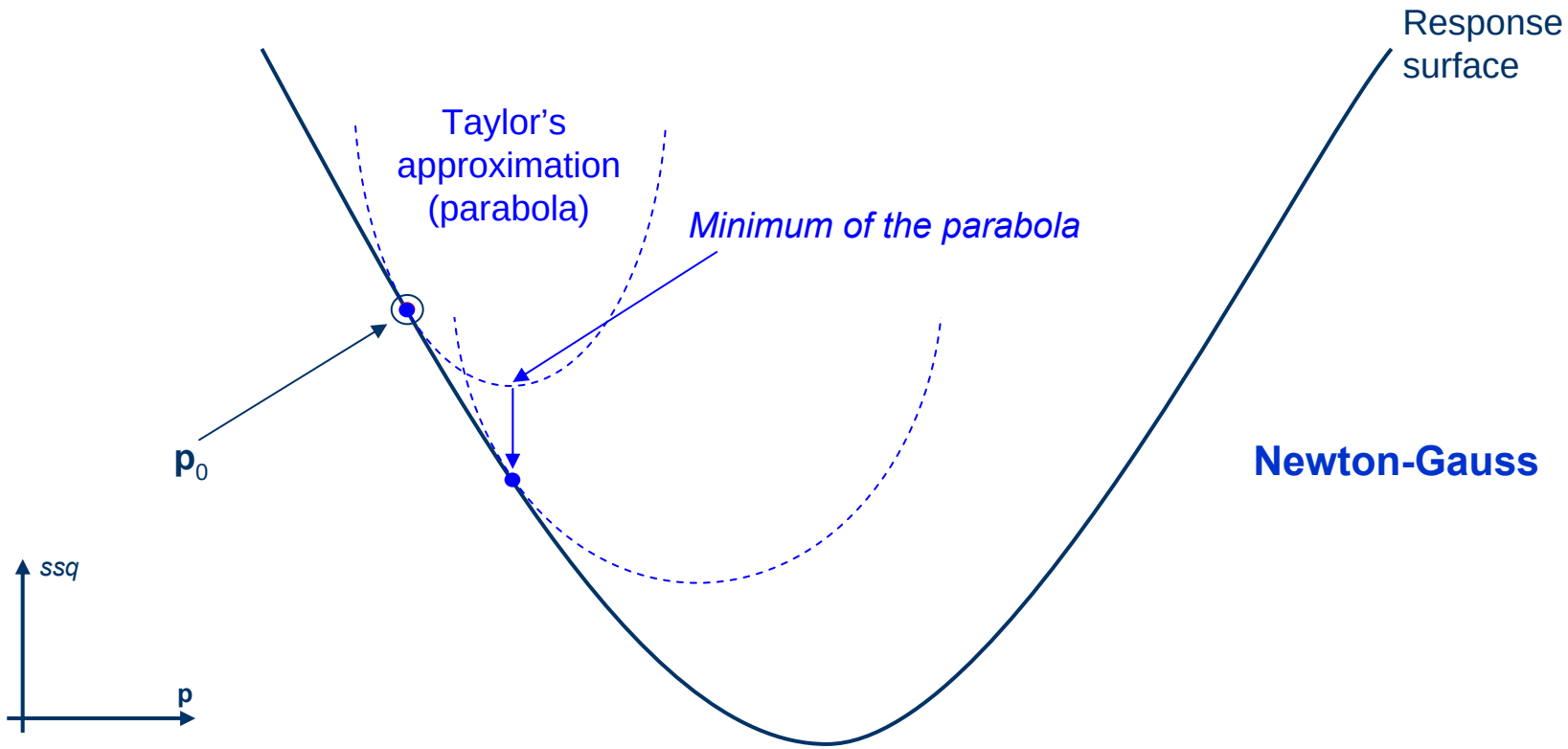
Geometrical interpretation on the response surface



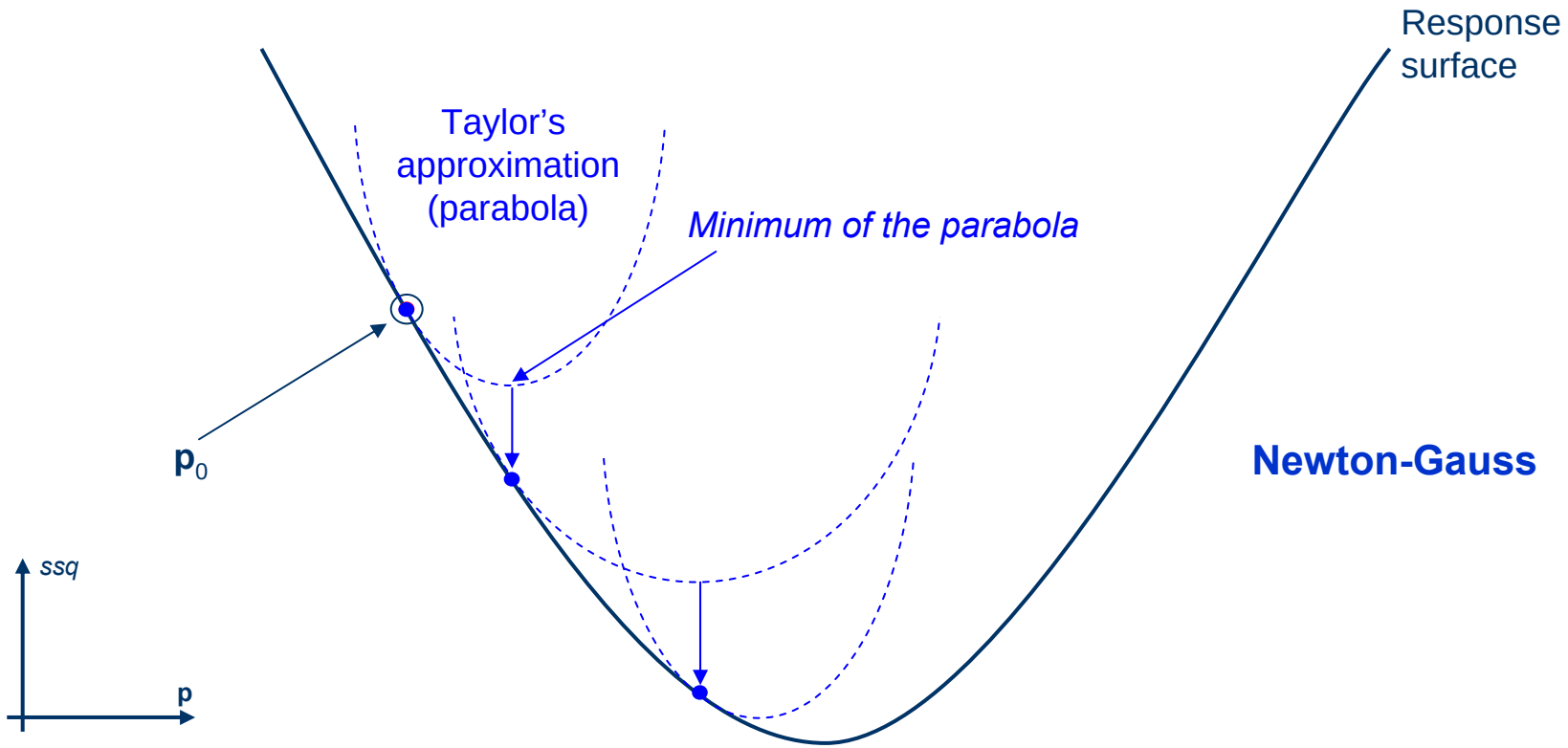
Geometrical interpretation on the response surface



Geometrical interpretation on the response surface



Geometrical interpretation on the response surface



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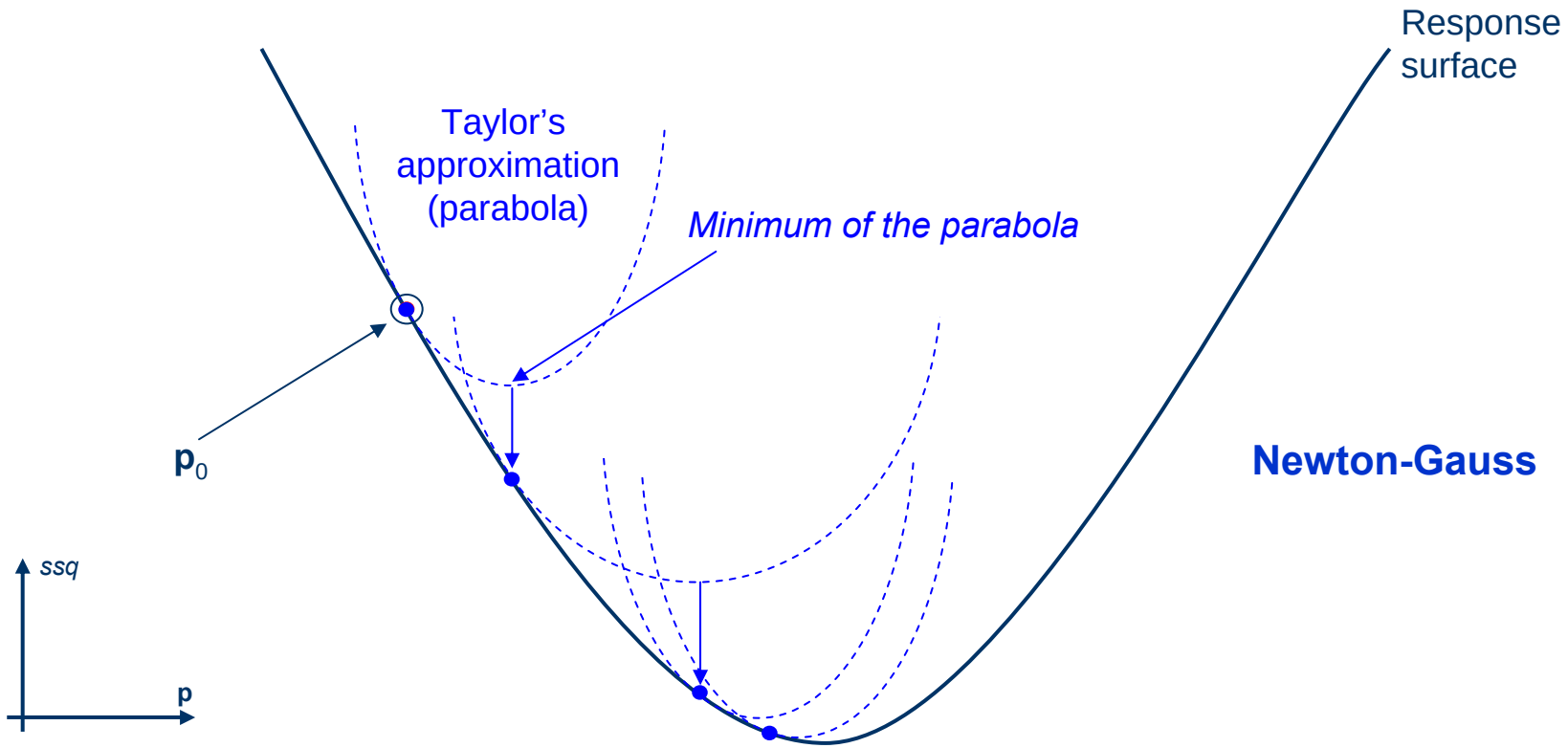

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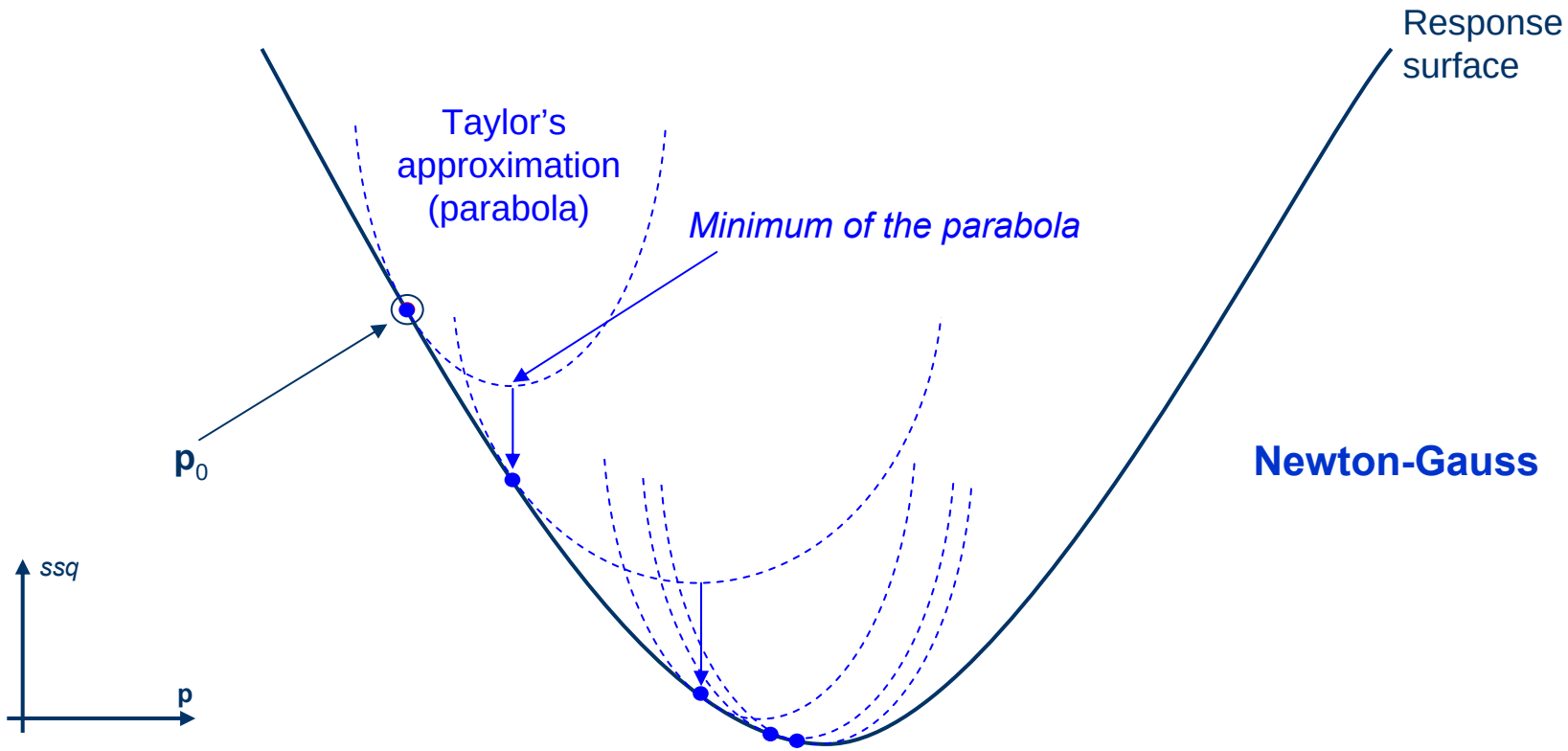

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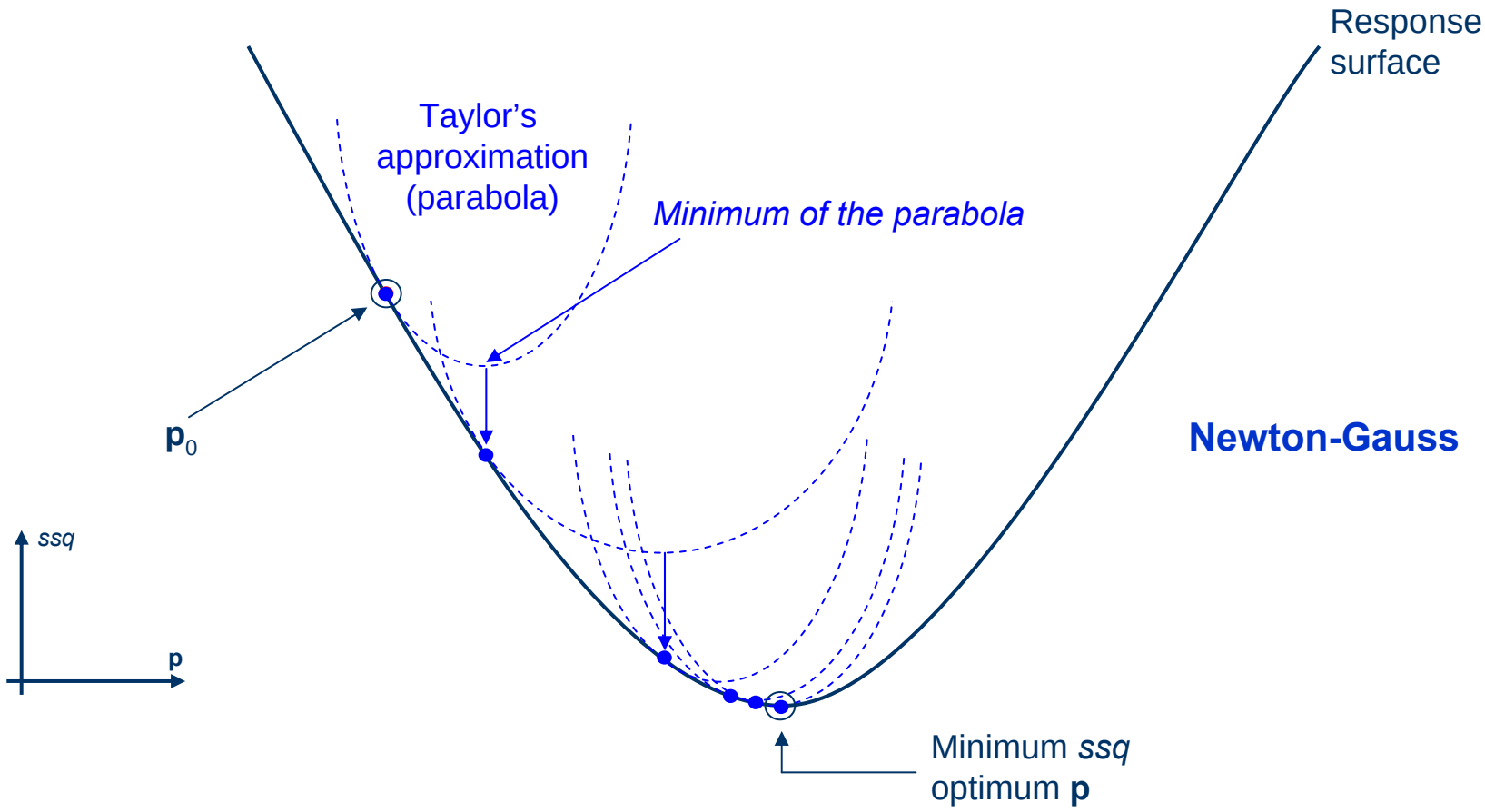

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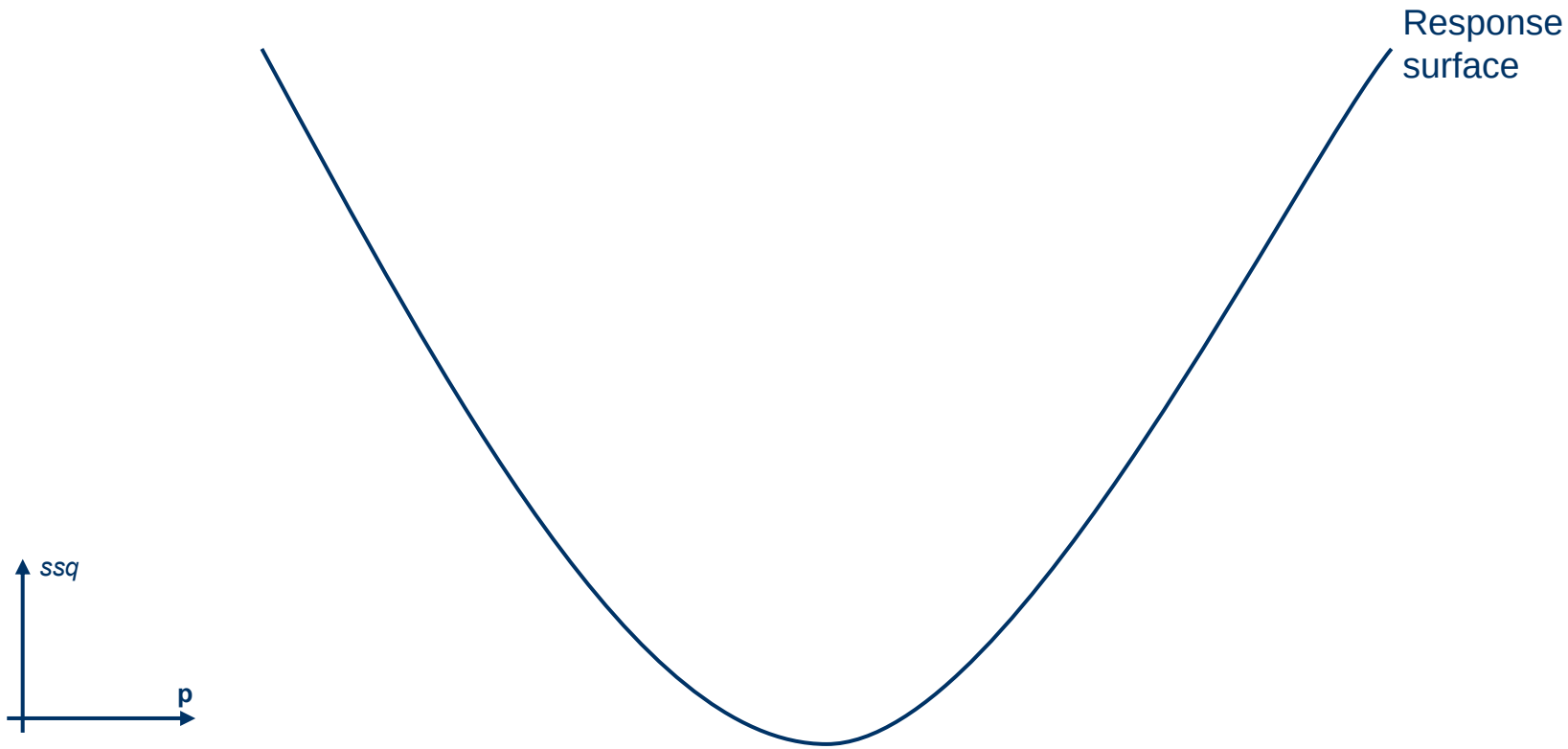

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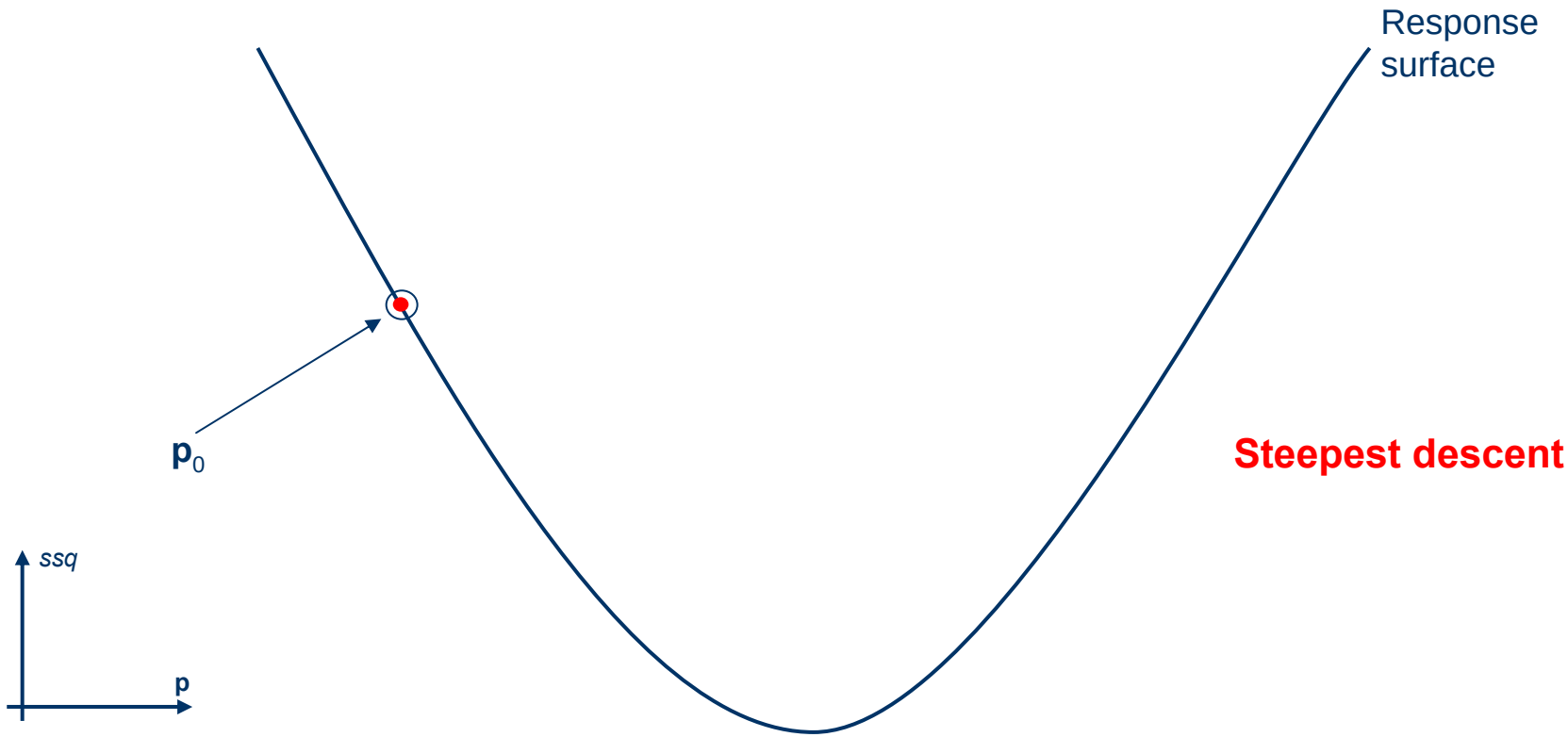

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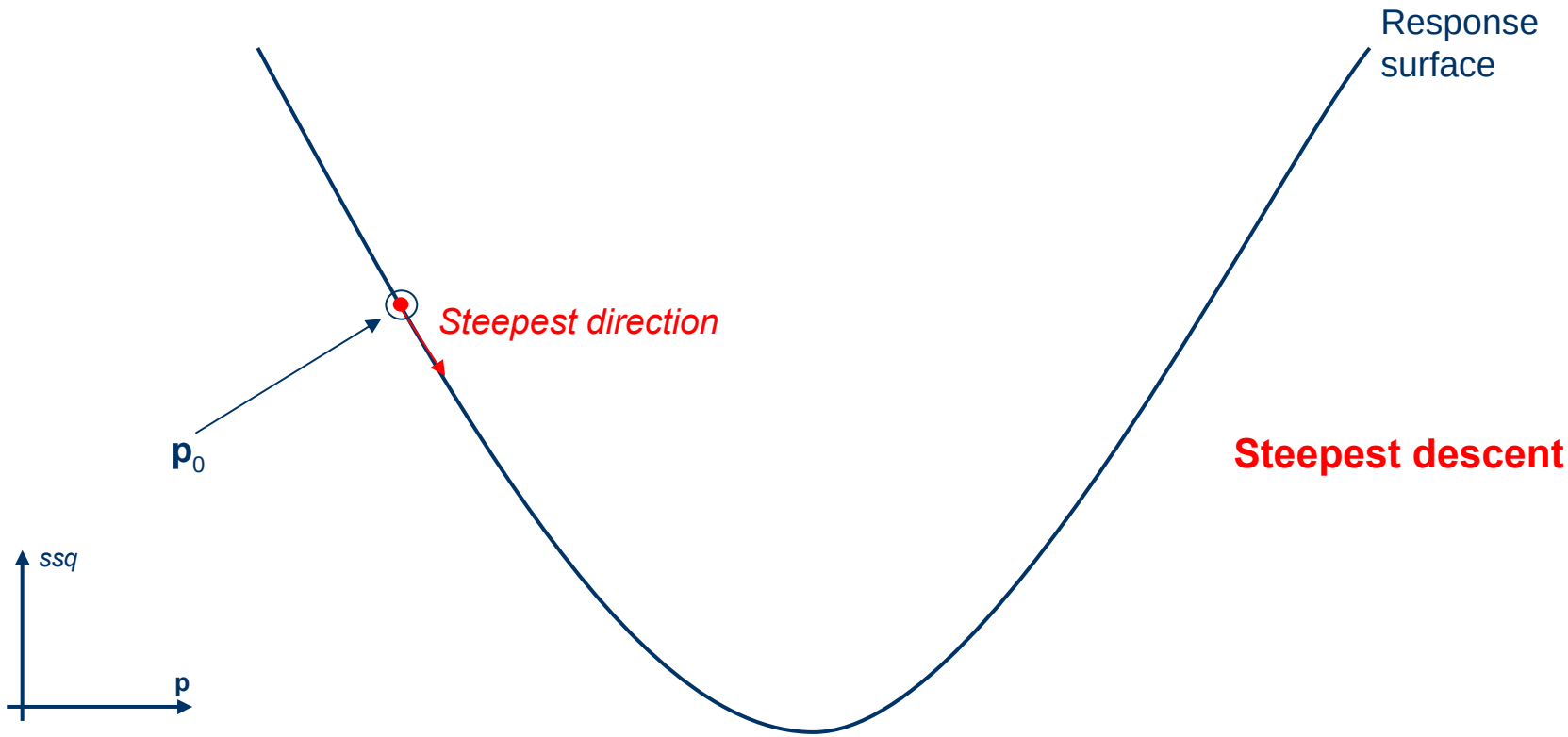
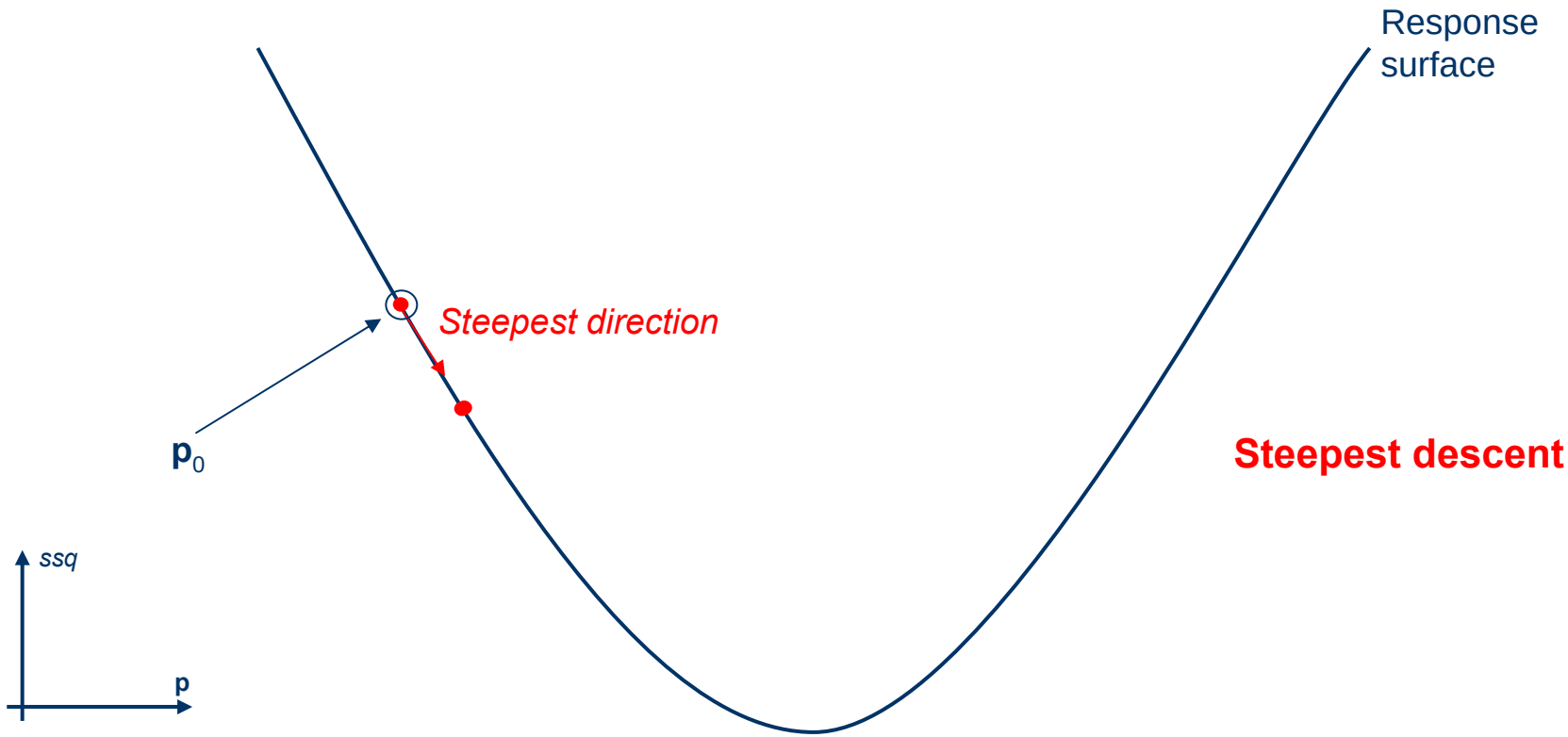


Geometrical interpretation on the response surface



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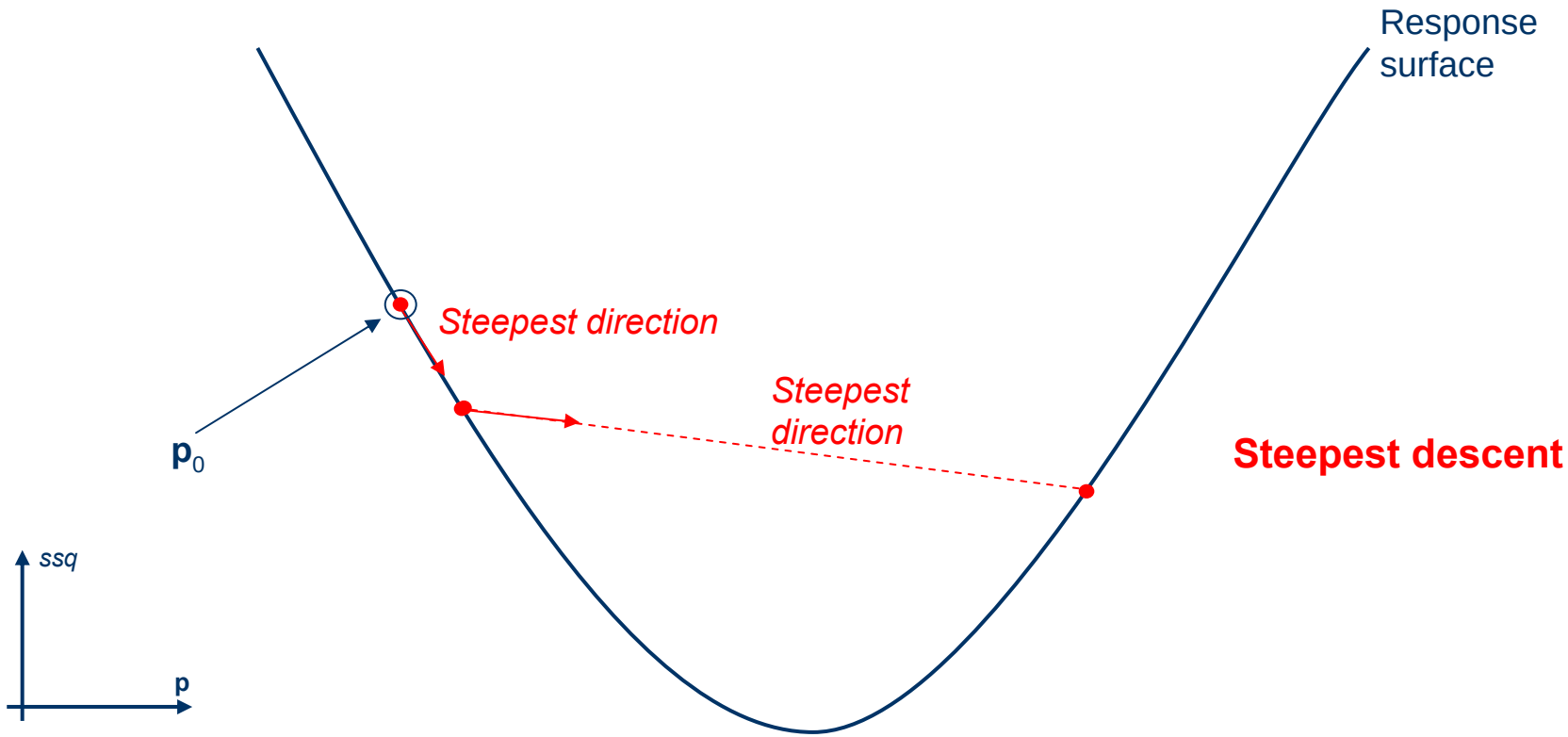
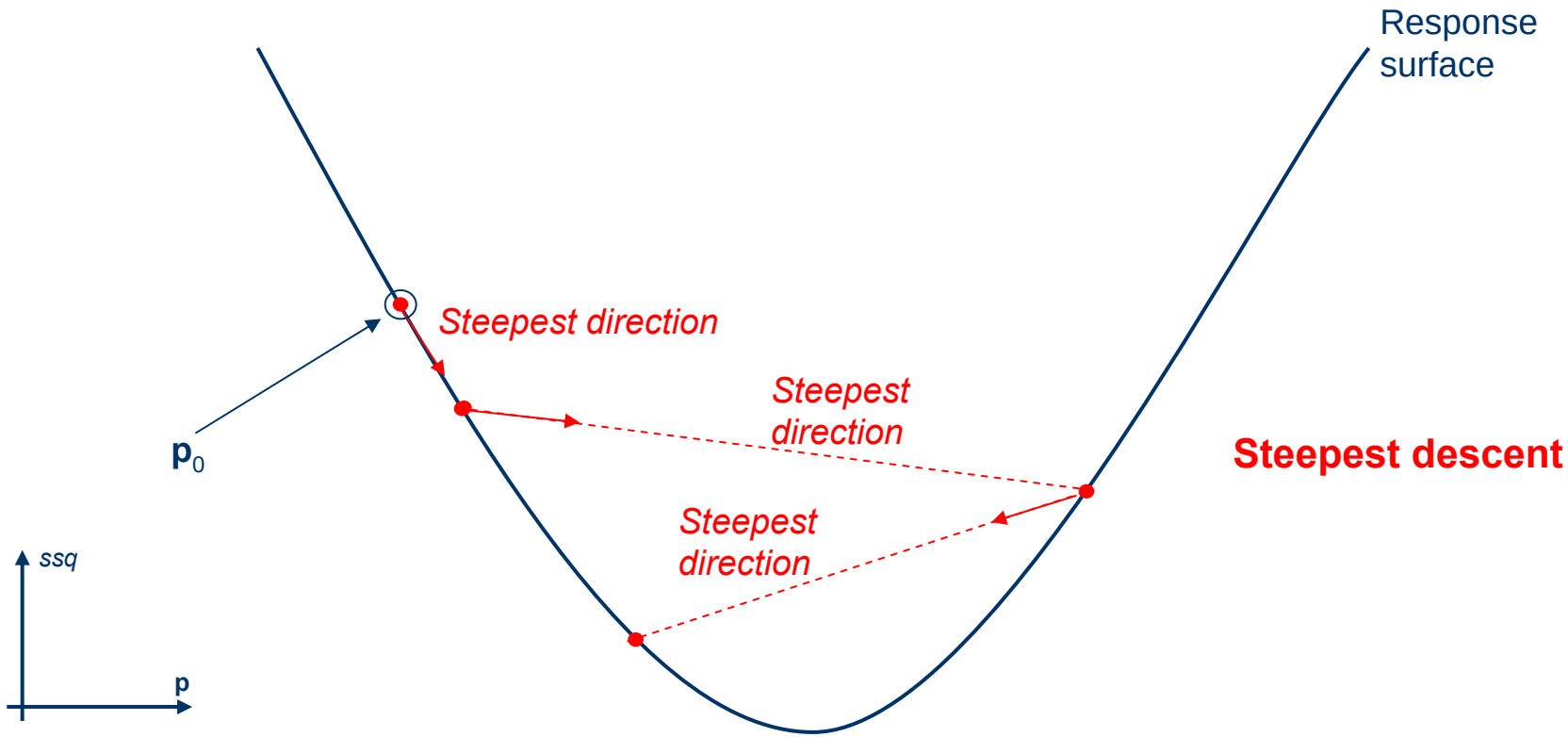


Geometrical interpretation on the response surface



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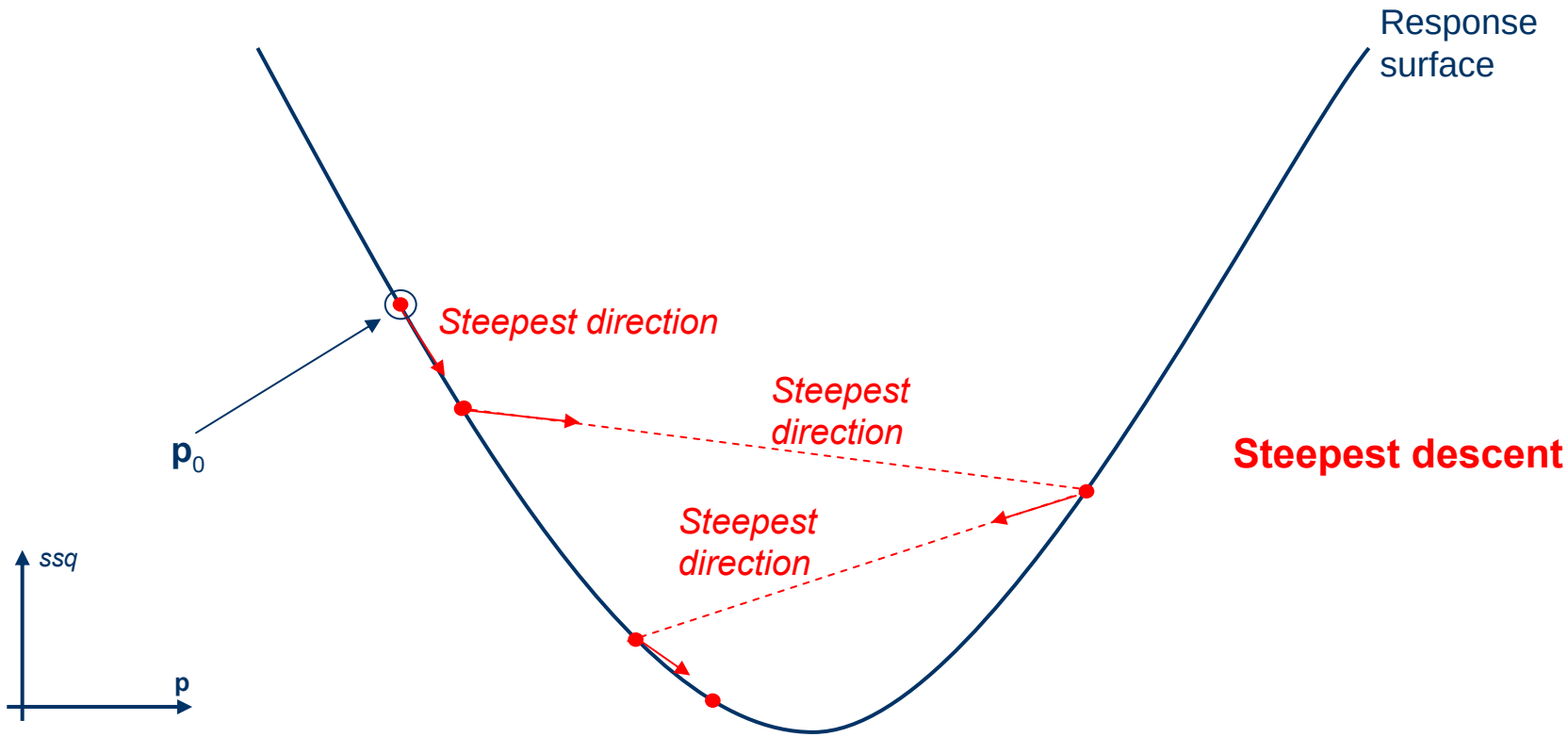

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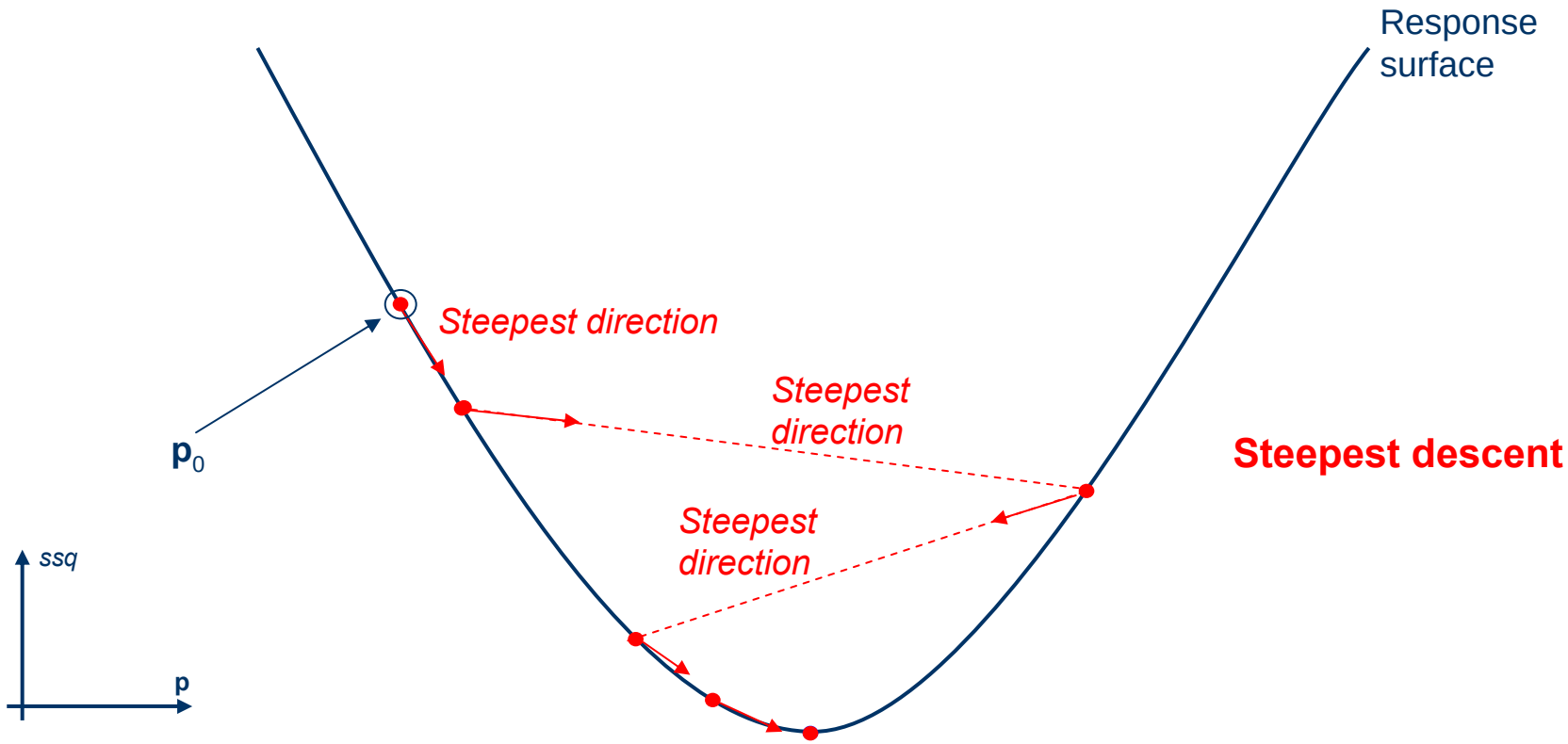
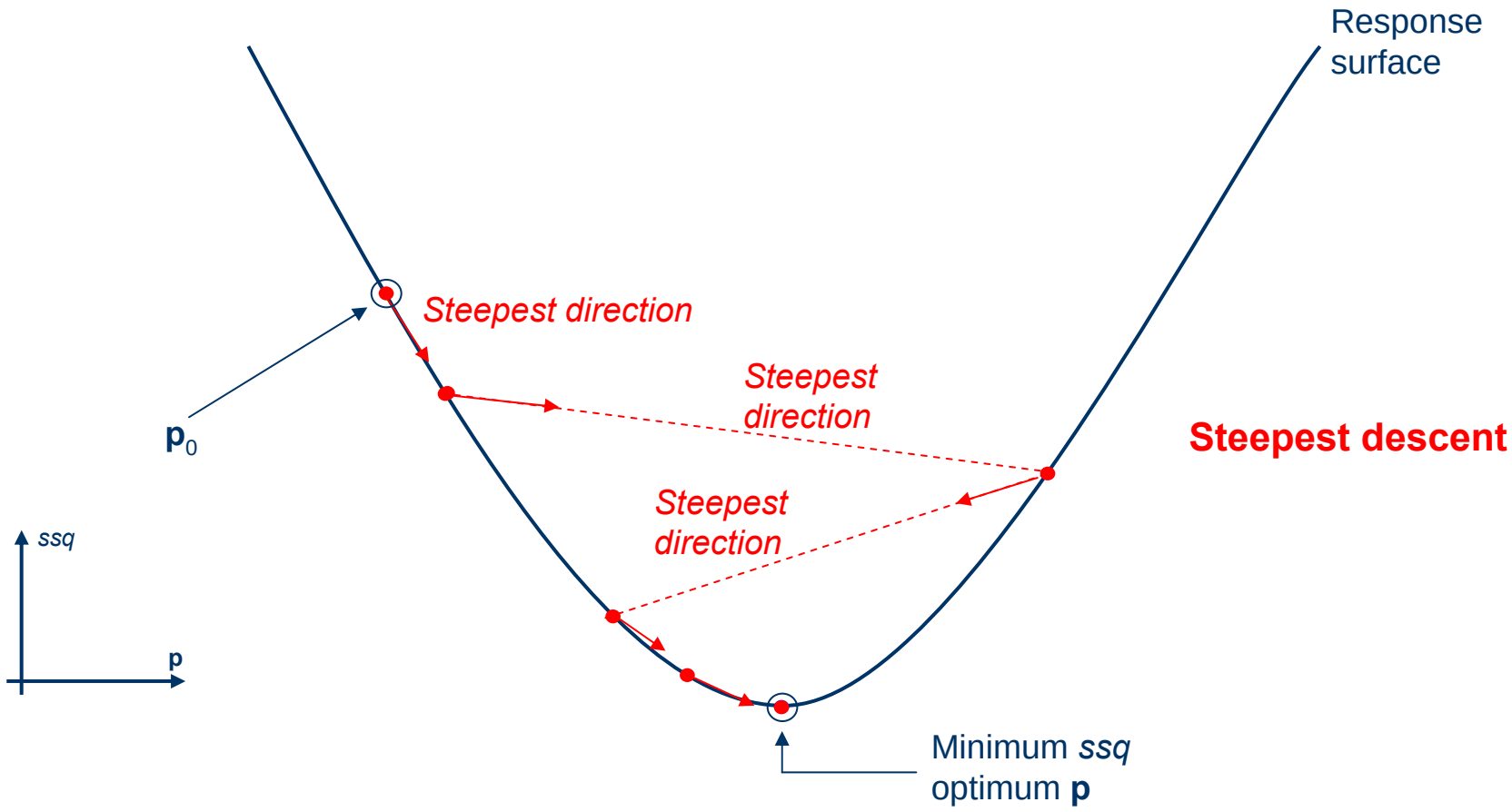
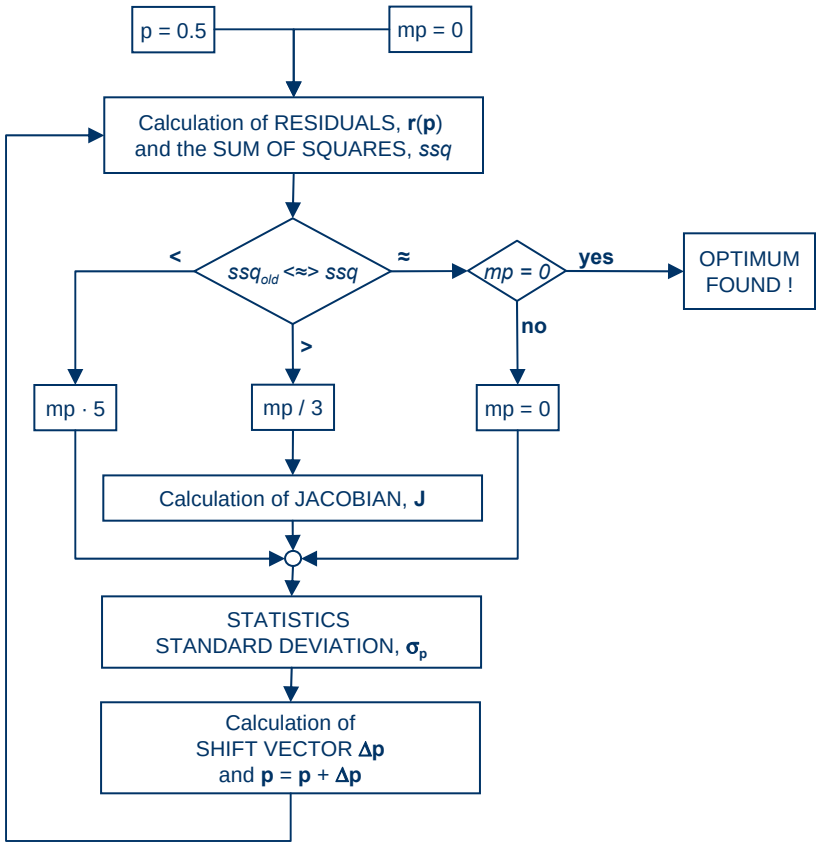


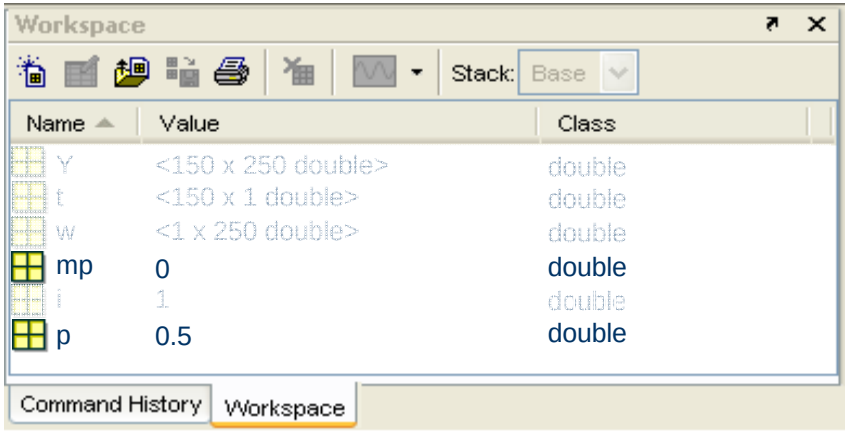
Geometrical interpretation on the response surface

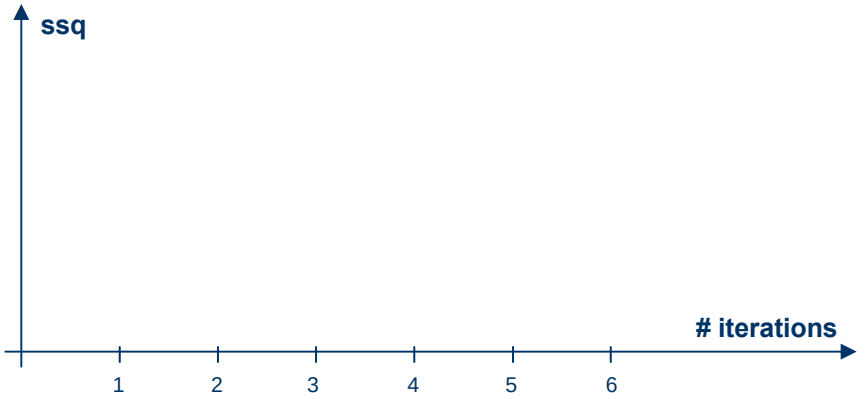


The NG/LM algorithm

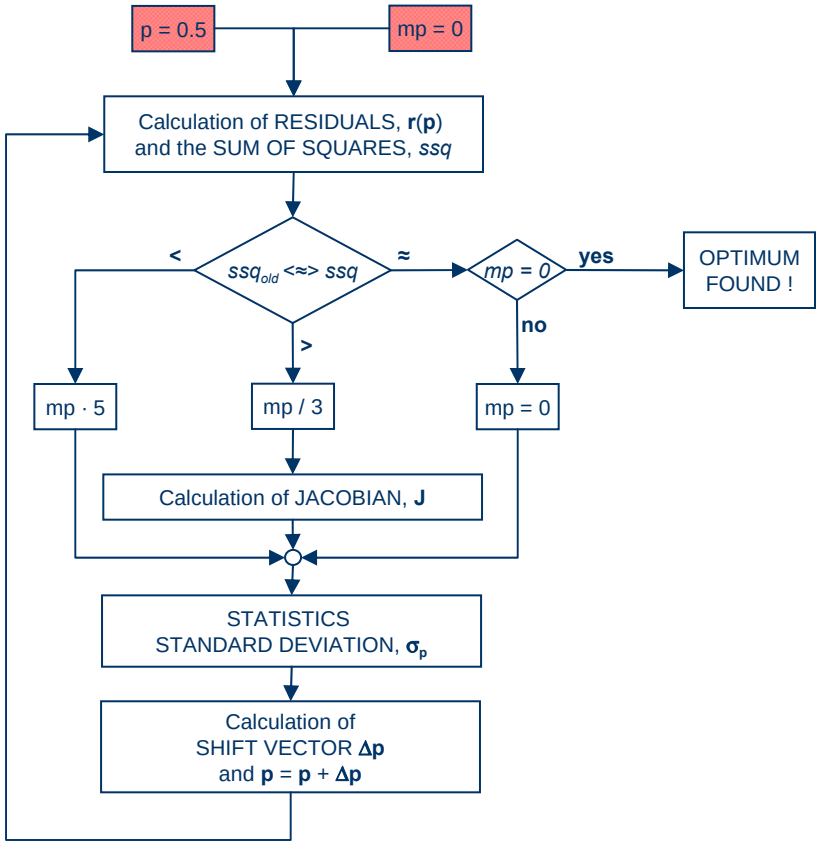


Current method :



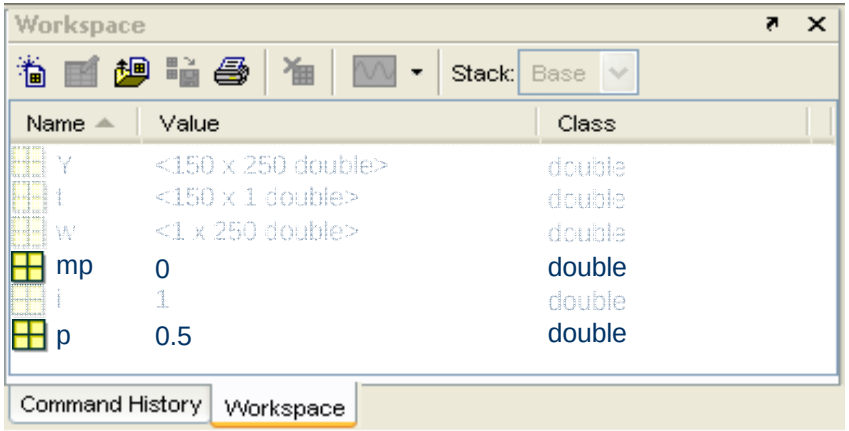


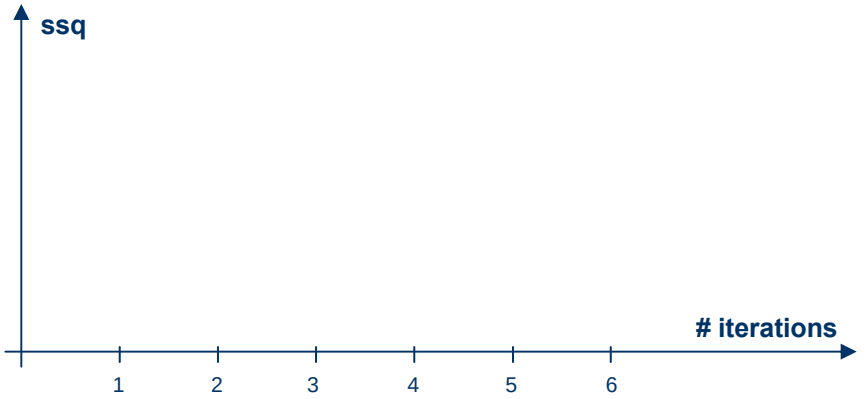
The NG/LM algorithm



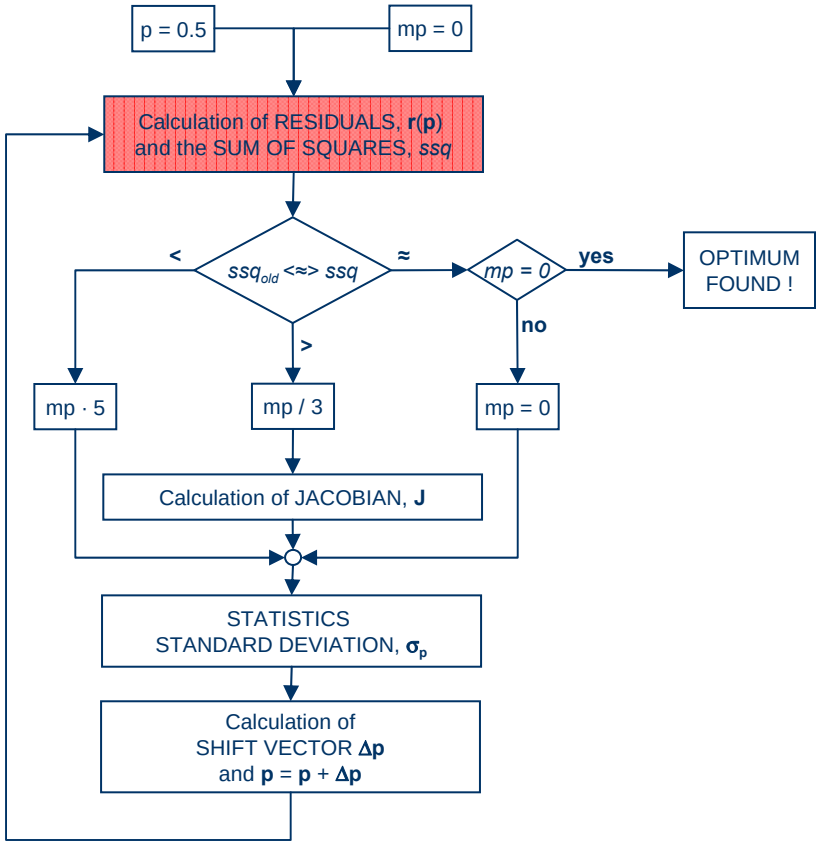
Current method :

Newton-Gauss method



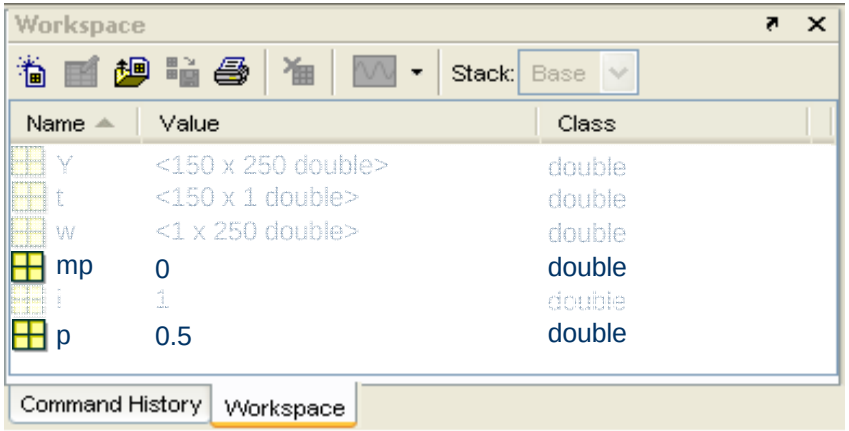


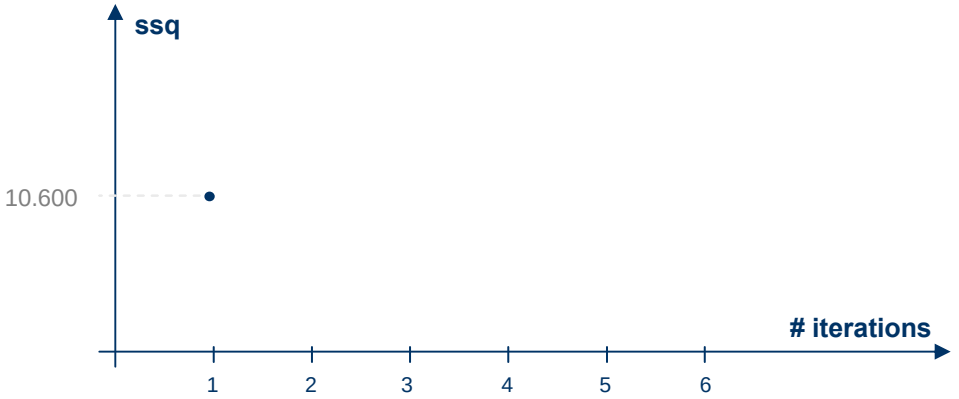
The NG/LM algorithm



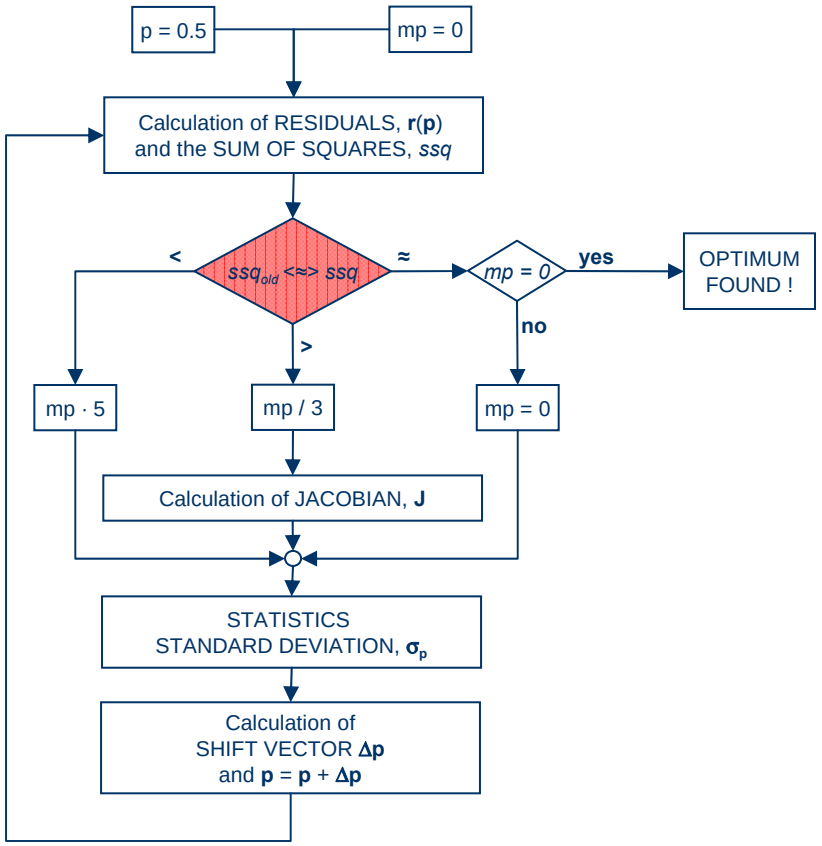
Current method :

Newton-Gauss method



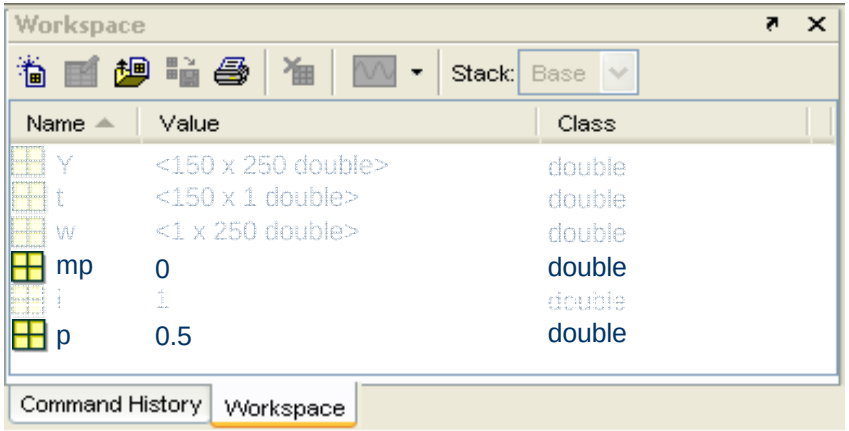


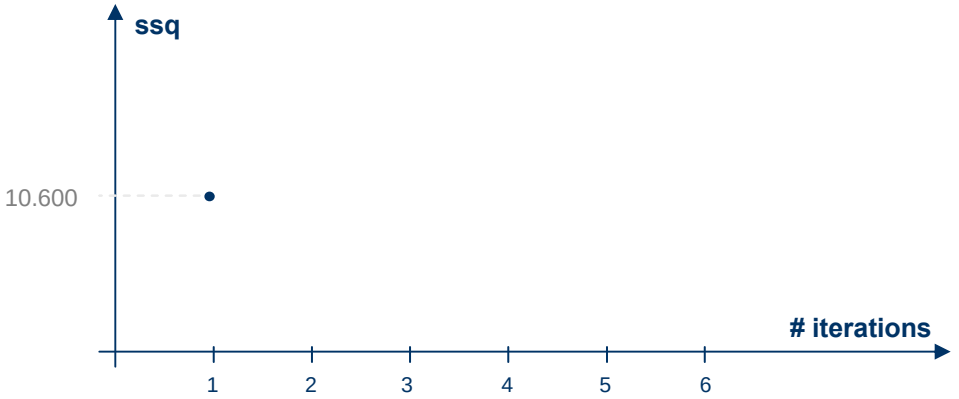
The NG/LM algorithm



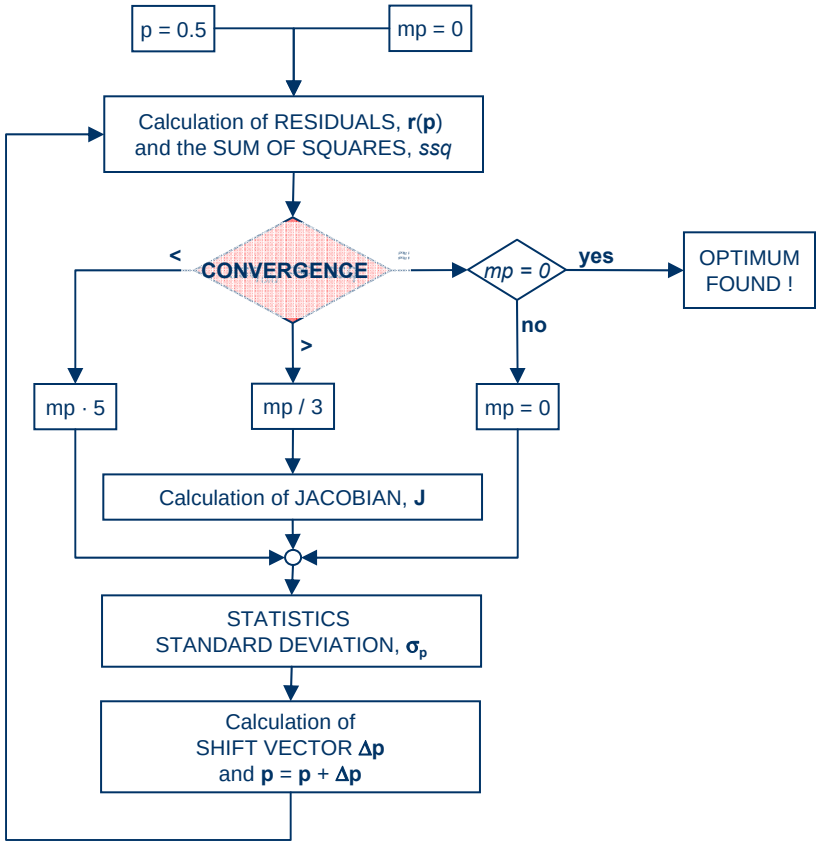
Current method :

Newton-Gauss method



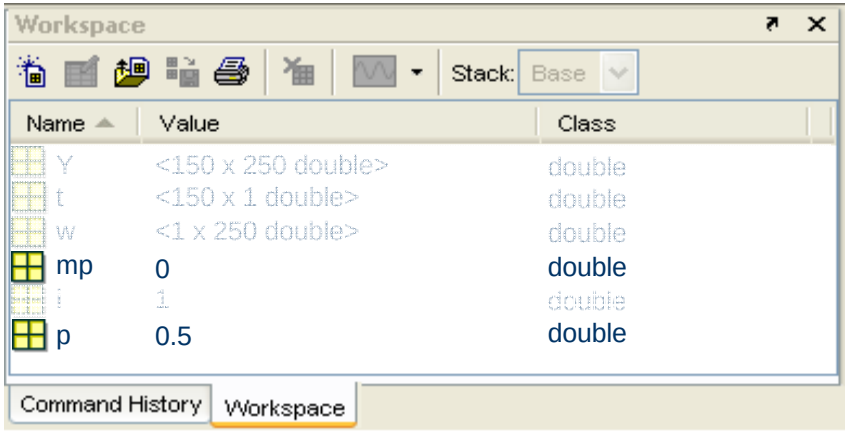


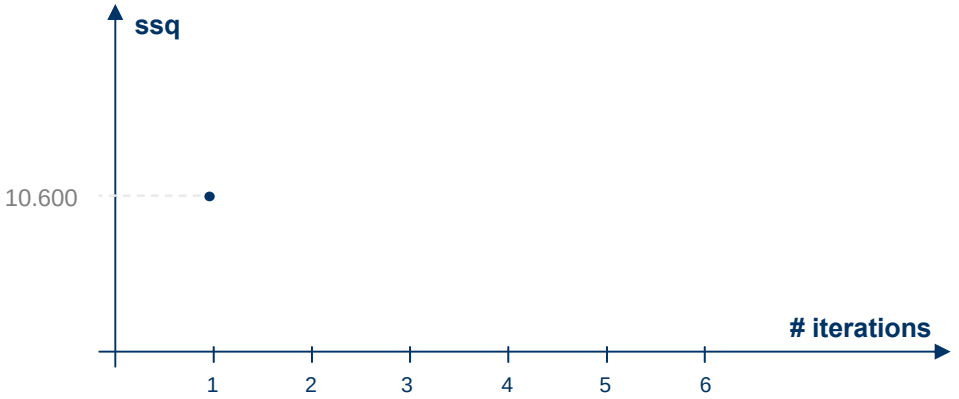
The NG/LM algorithm



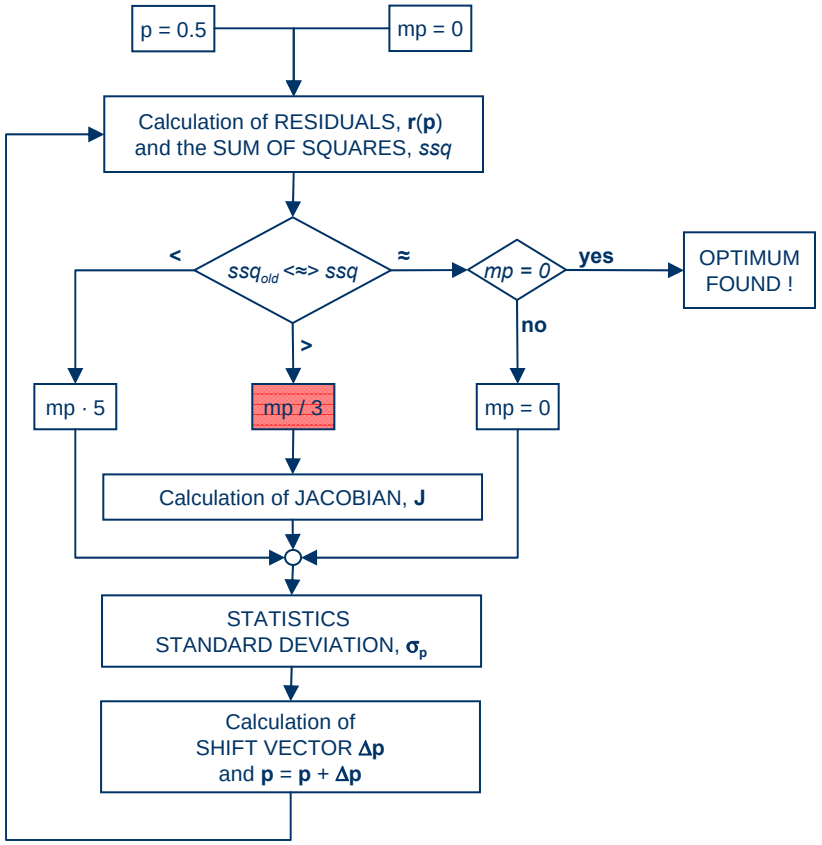
Current method :

Newton-Gauss method



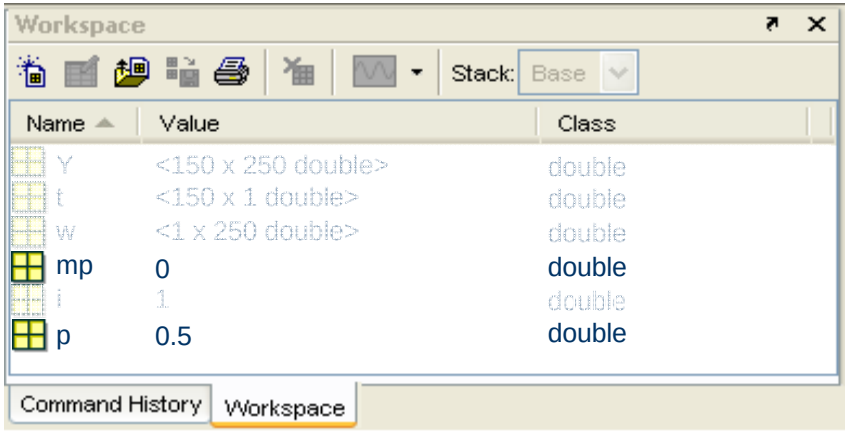


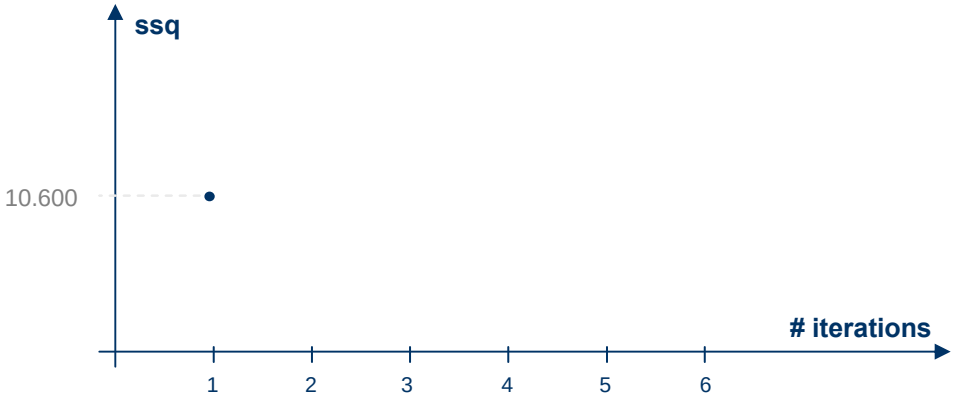
The NG/LM algorithm



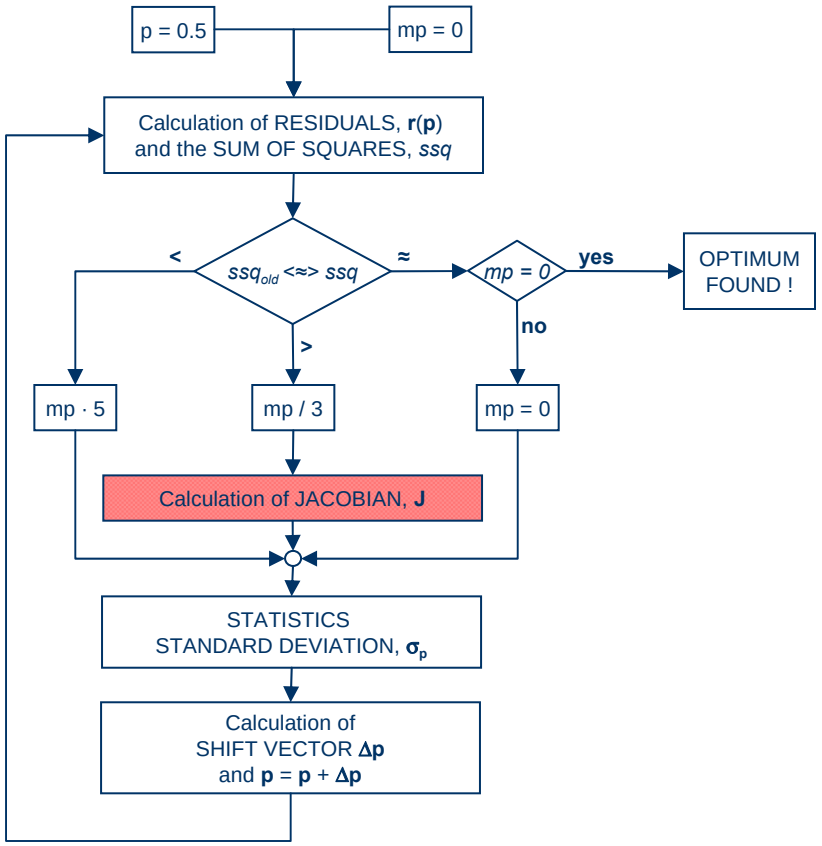
Current method :

Newton-Gauss method



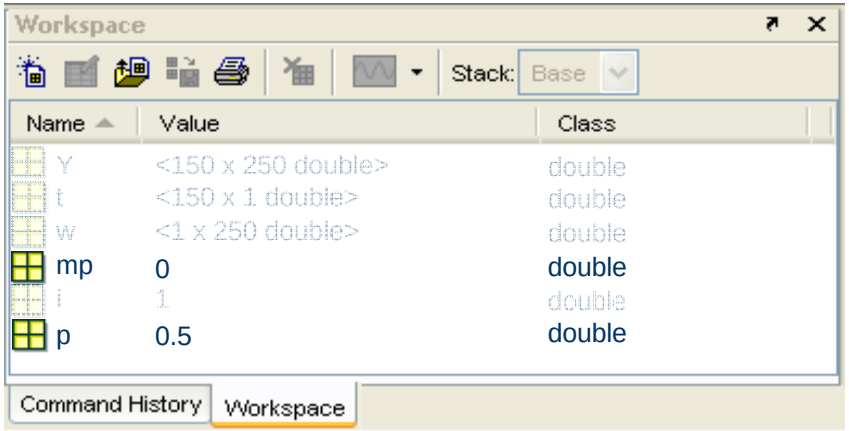


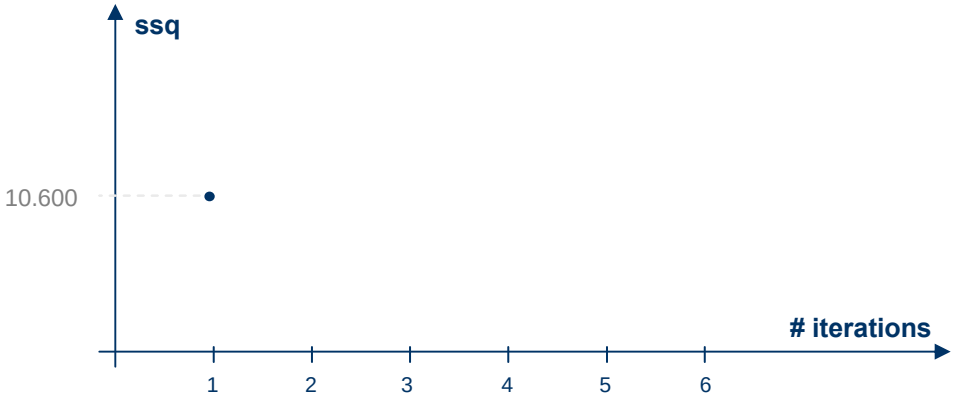
The NG/LM algorithm



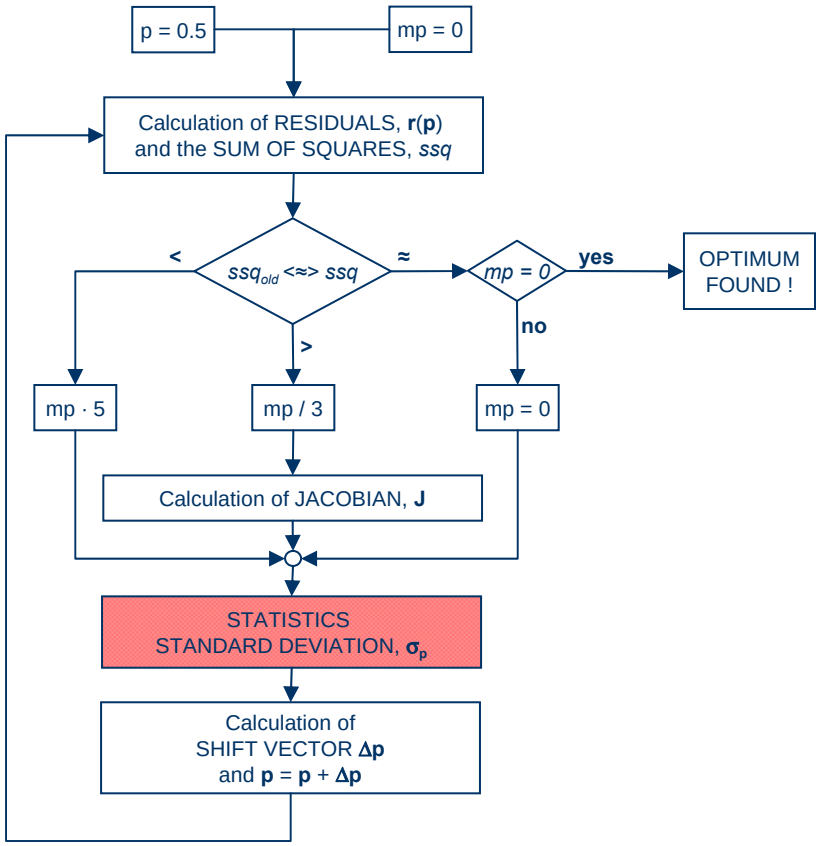
Current method :

Newton-Gauss method



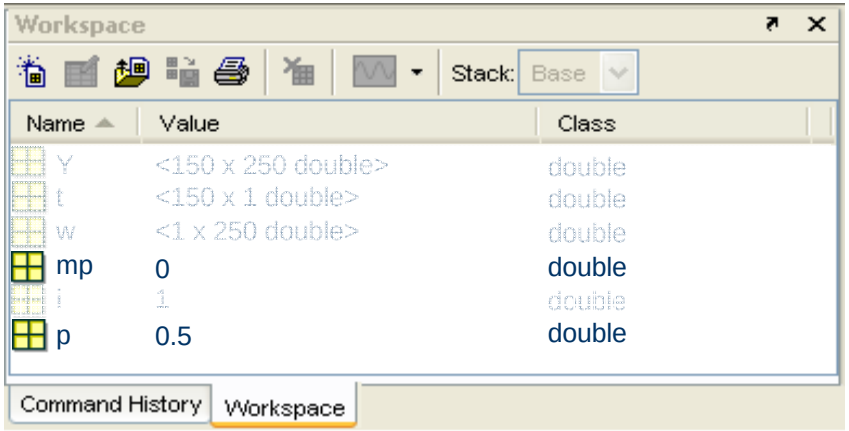


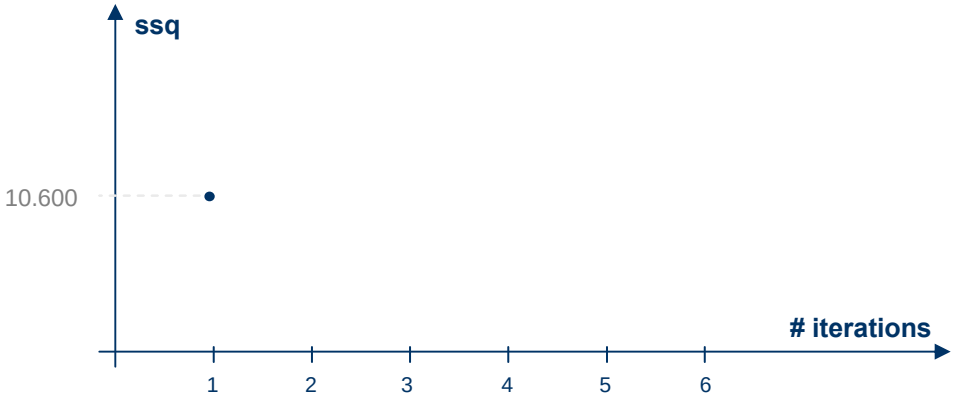
The NG/LM algorithm



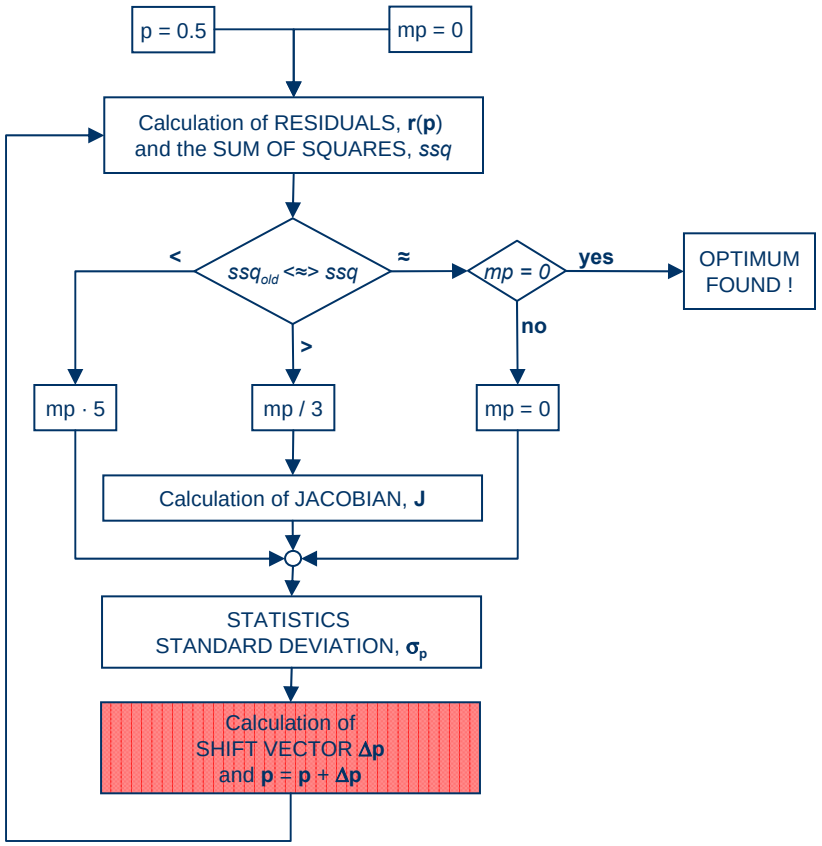
Current method :

Newton-Gauss method



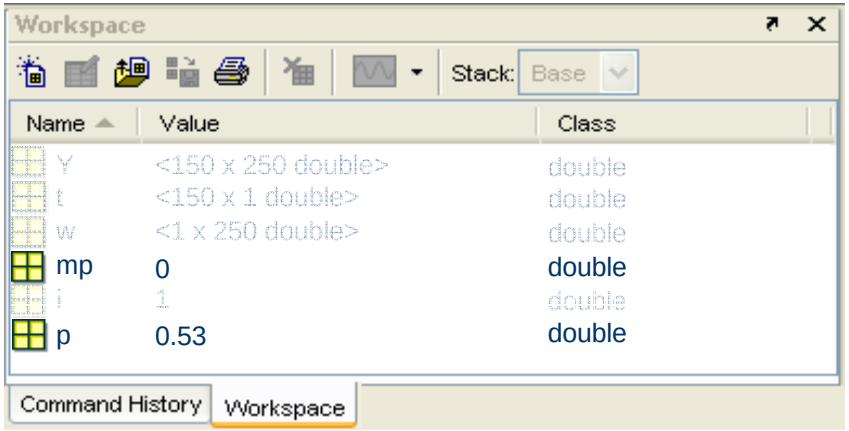


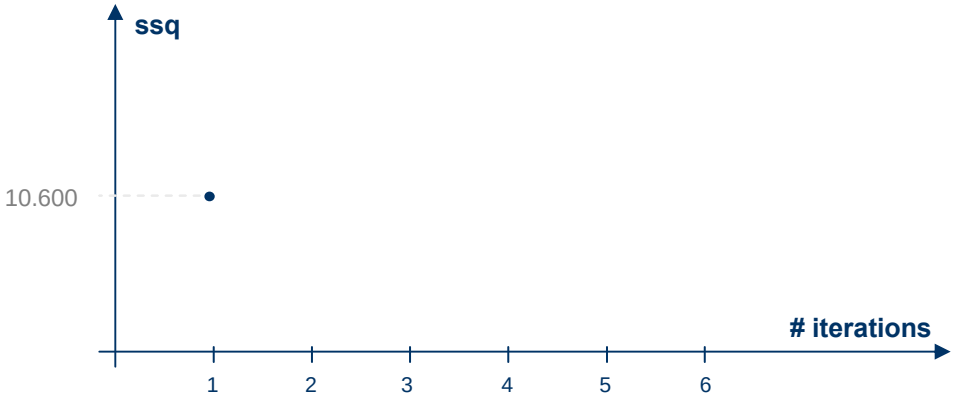
The NG/LM algorithm



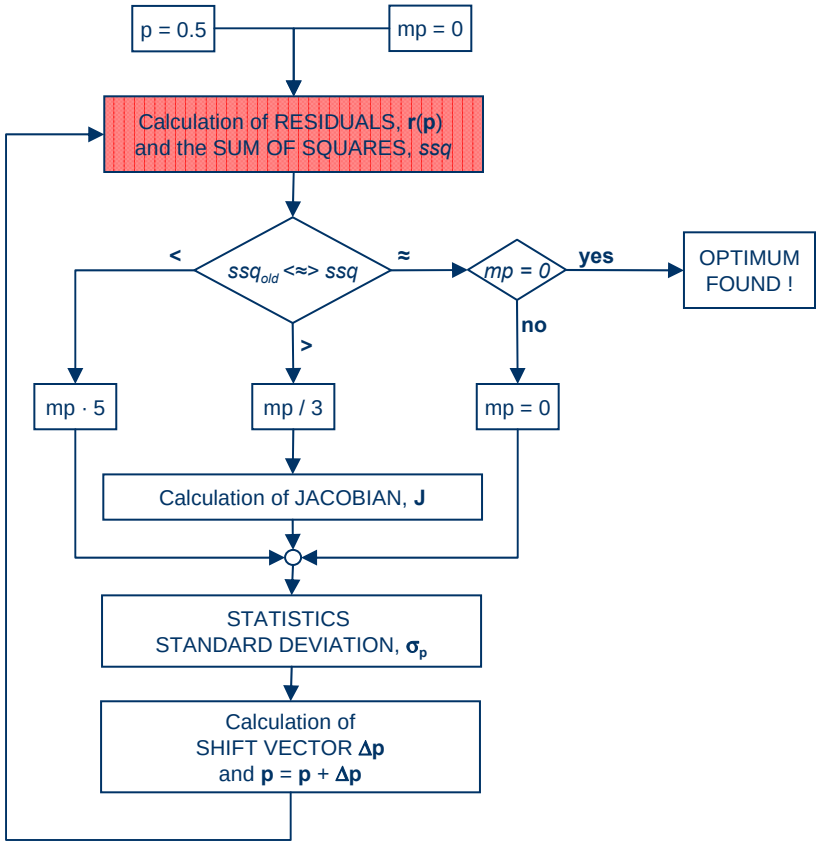
Current method :

Newton-Gauss method



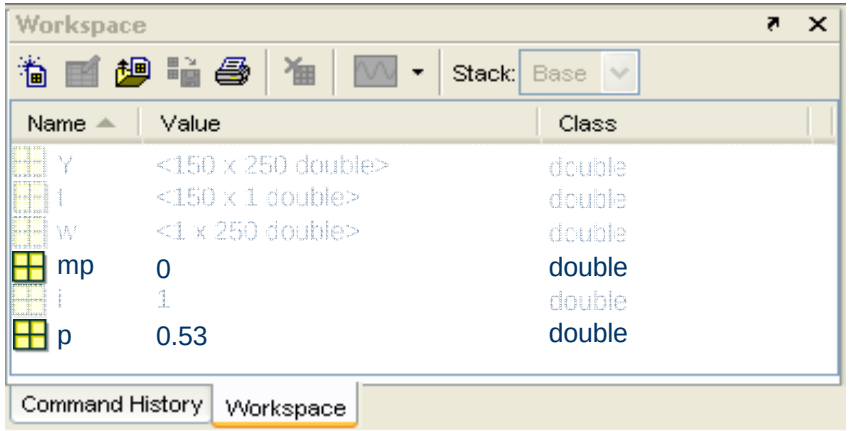


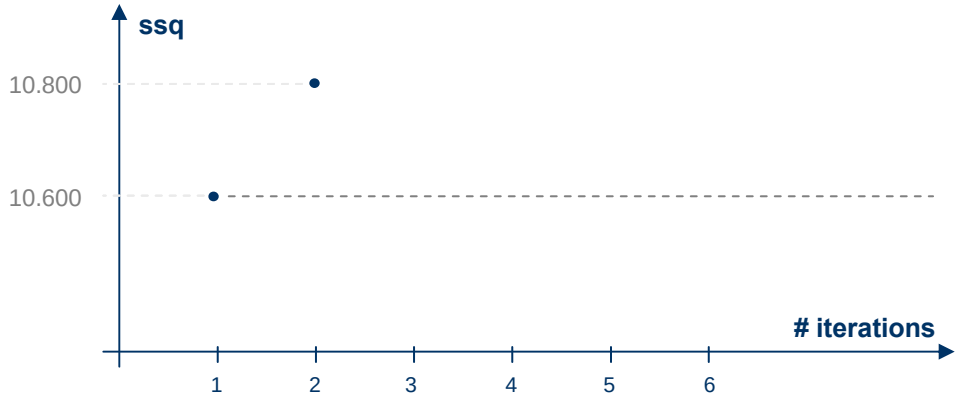
The NG/LM algorithm



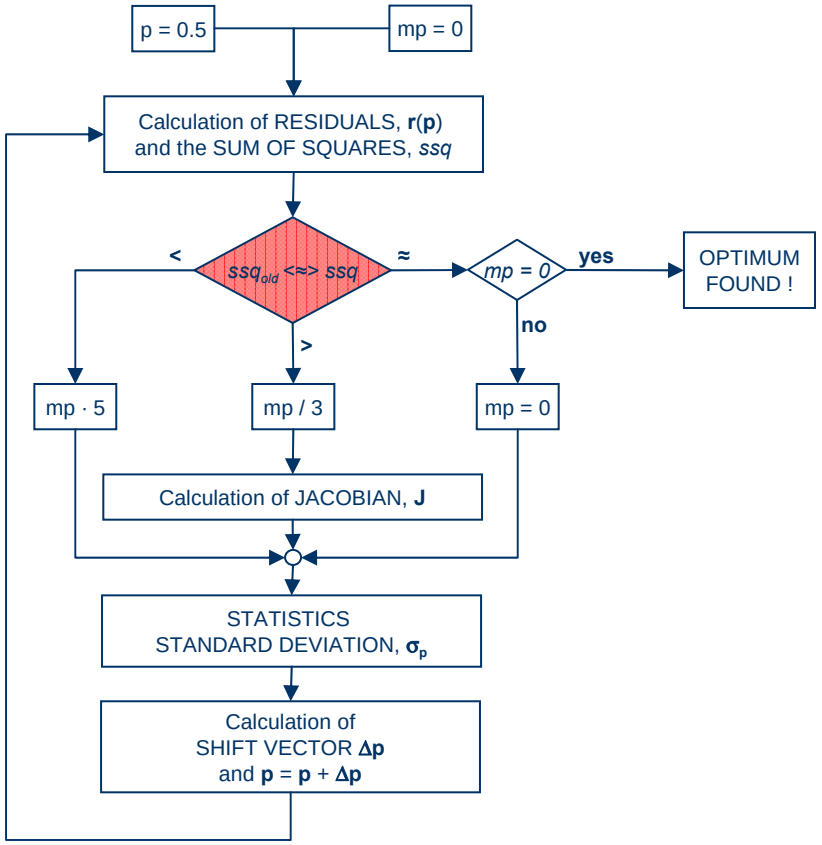
Current method :

Newton-Gauss method



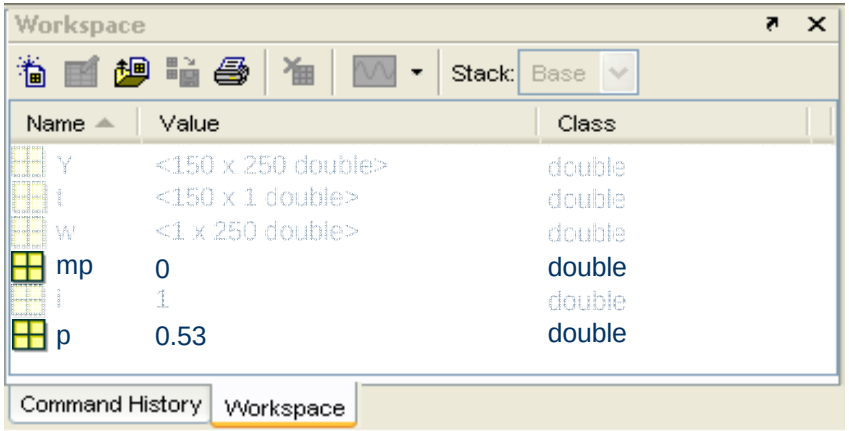


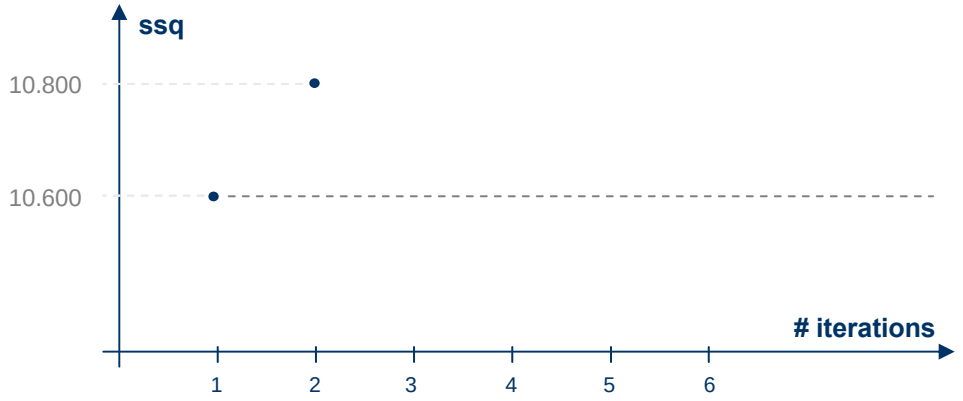
The NG/LM algorithm



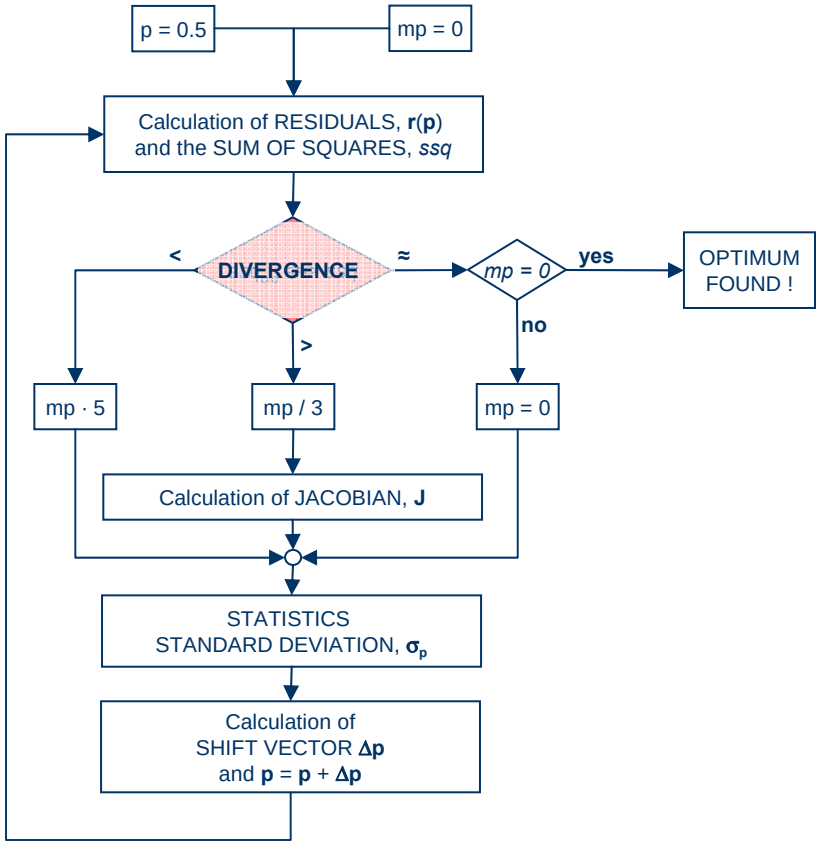
Current method :

Newton-Gauss method



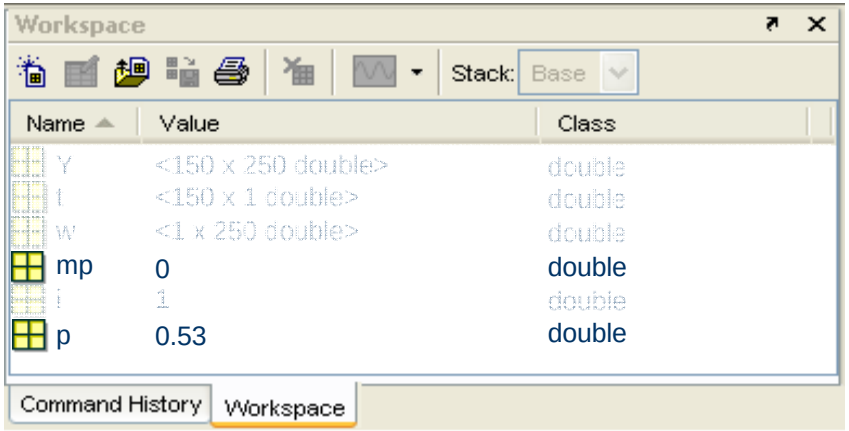


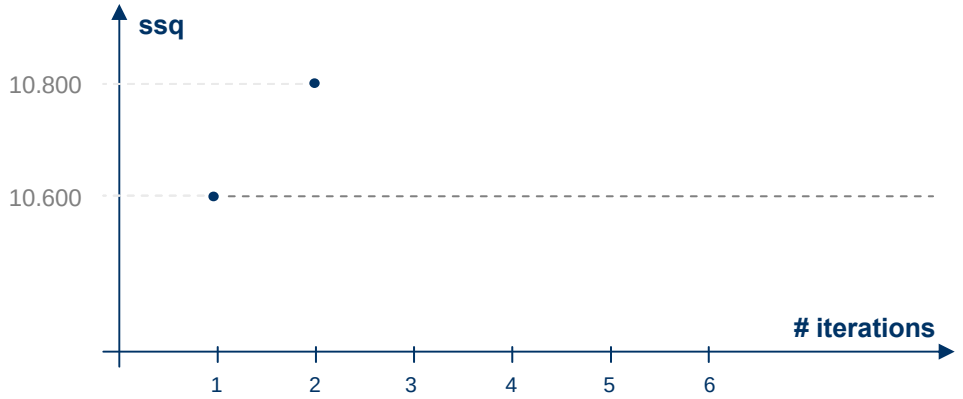
The NG/LM algorithm



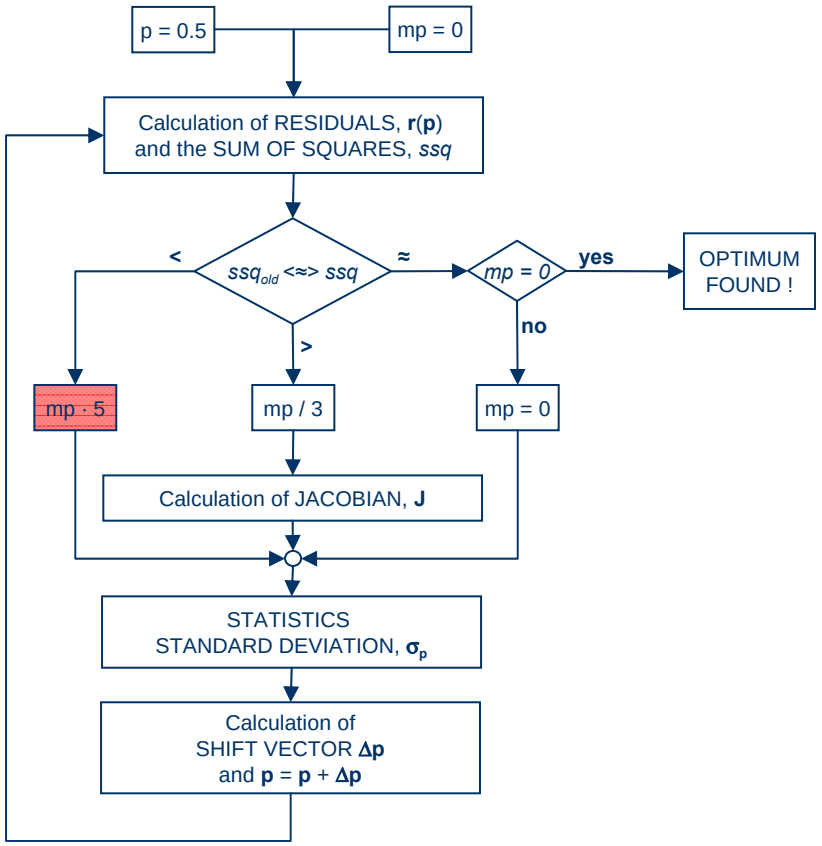
Current method :

NG/Levenberg-Marquardt method



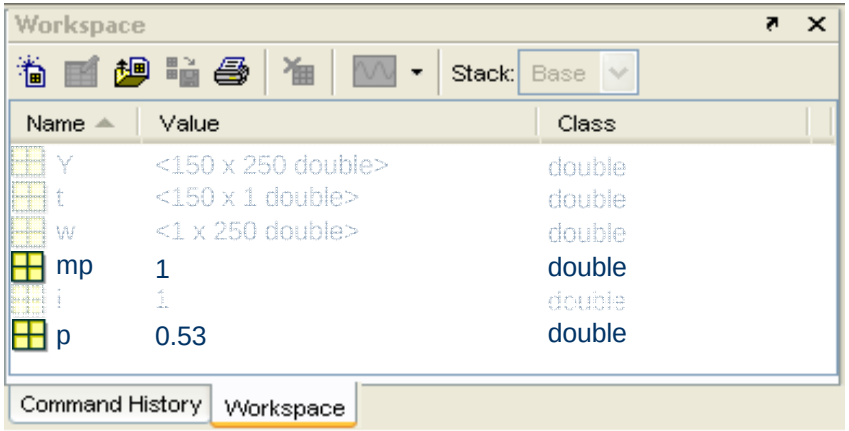


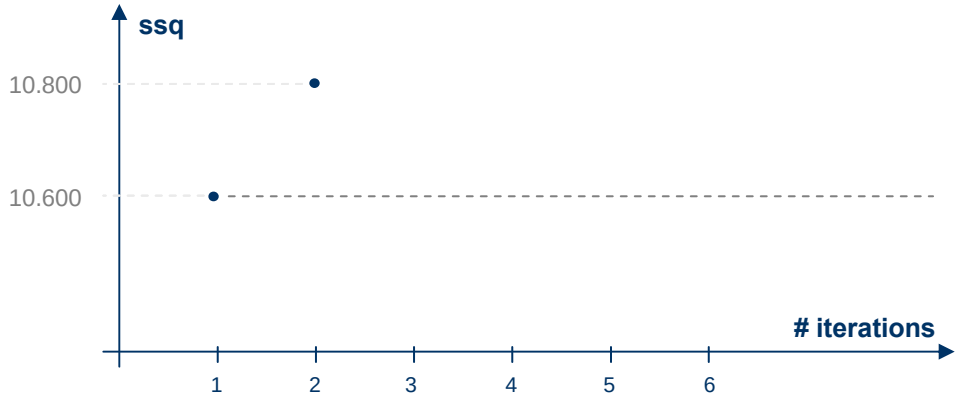
The NG/LM algorithm



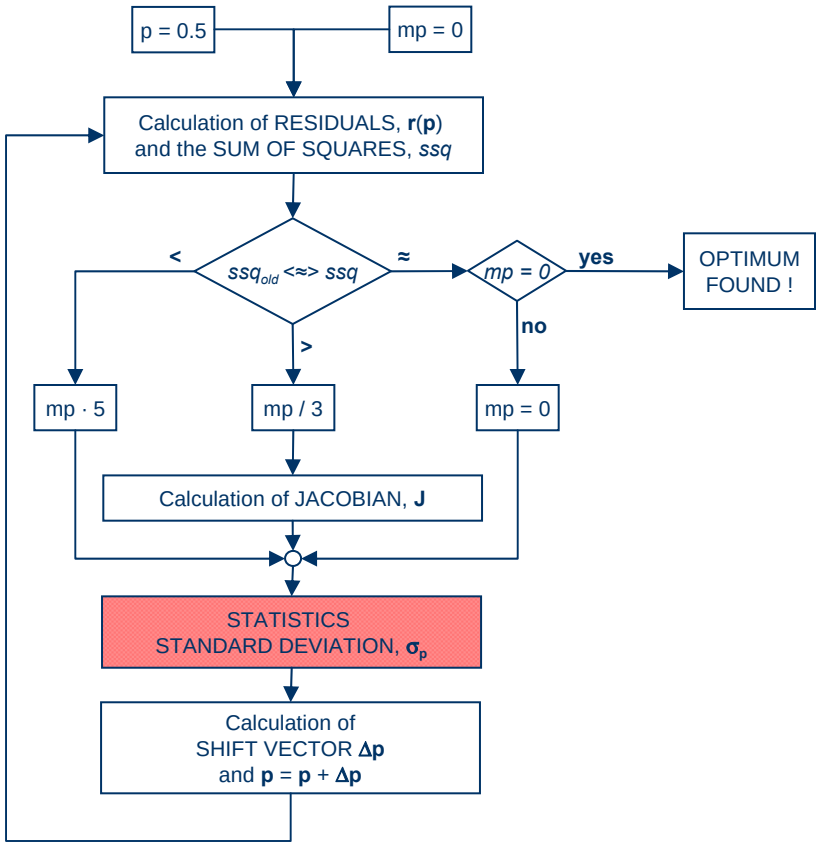
Current method :

NG/Levenberg-Marquardt method



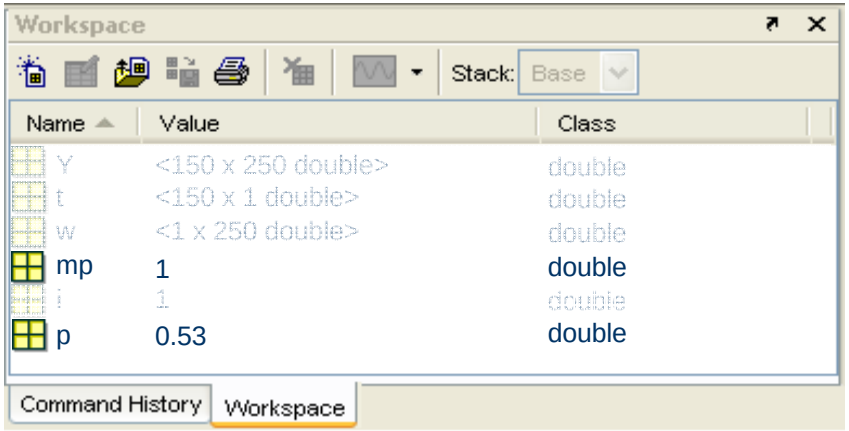


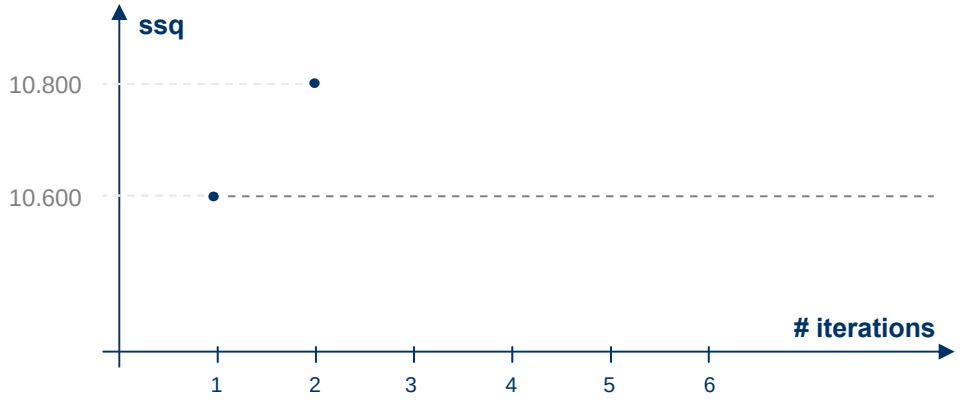
The NG/LM algorithm



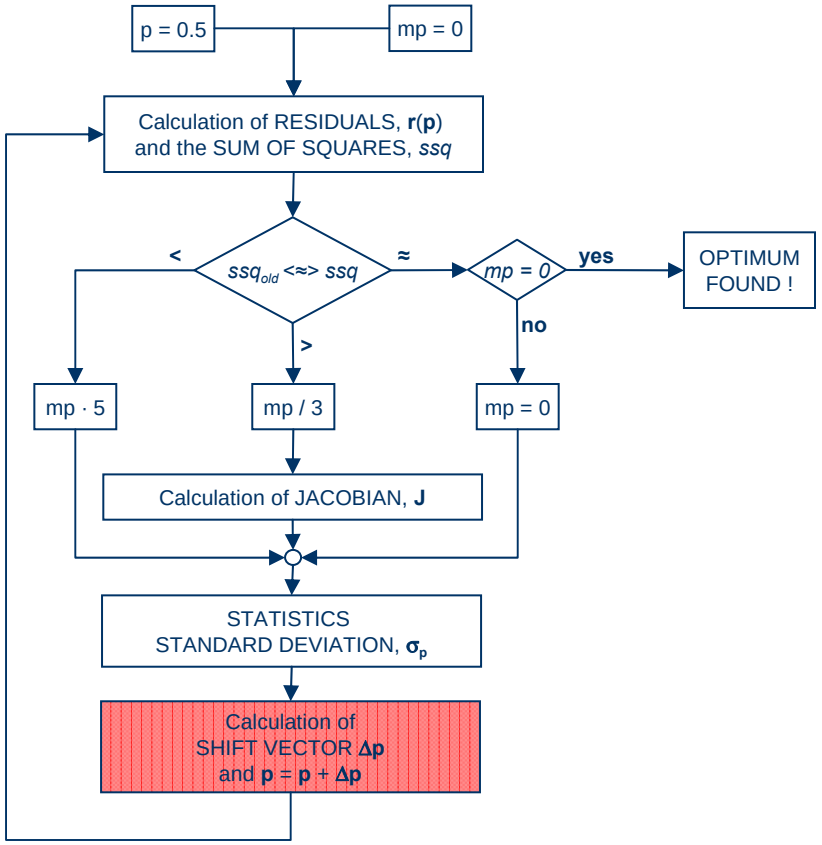
Current method :

NG/Levenberg-Marquardt method





The NG/LM algorithm



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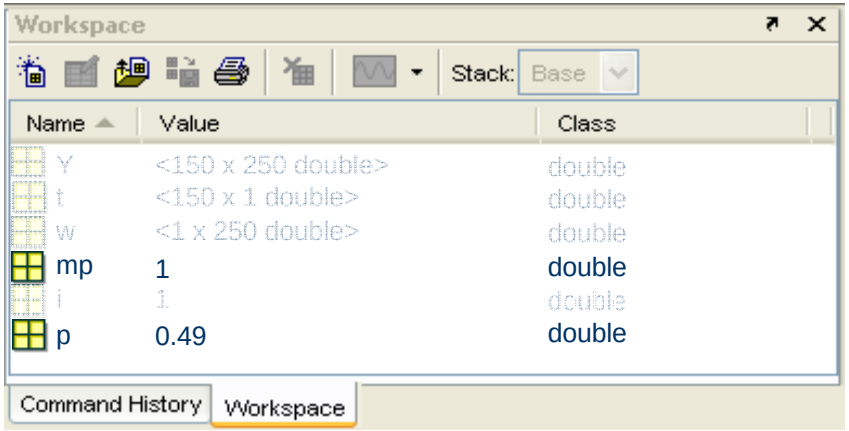

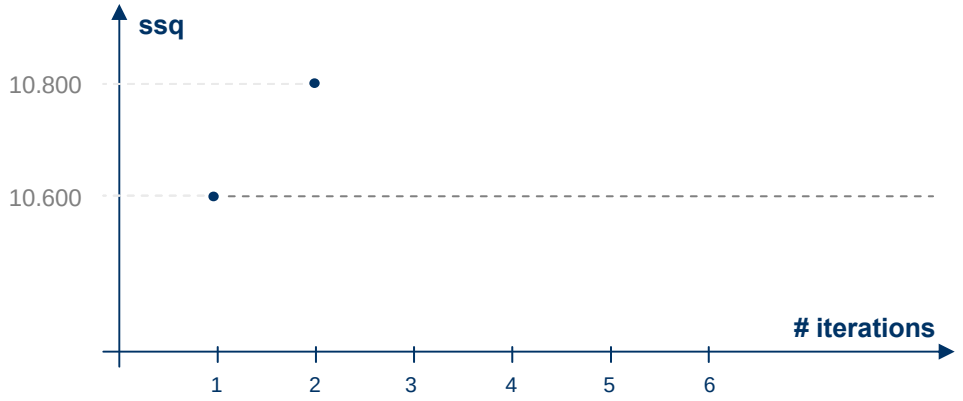

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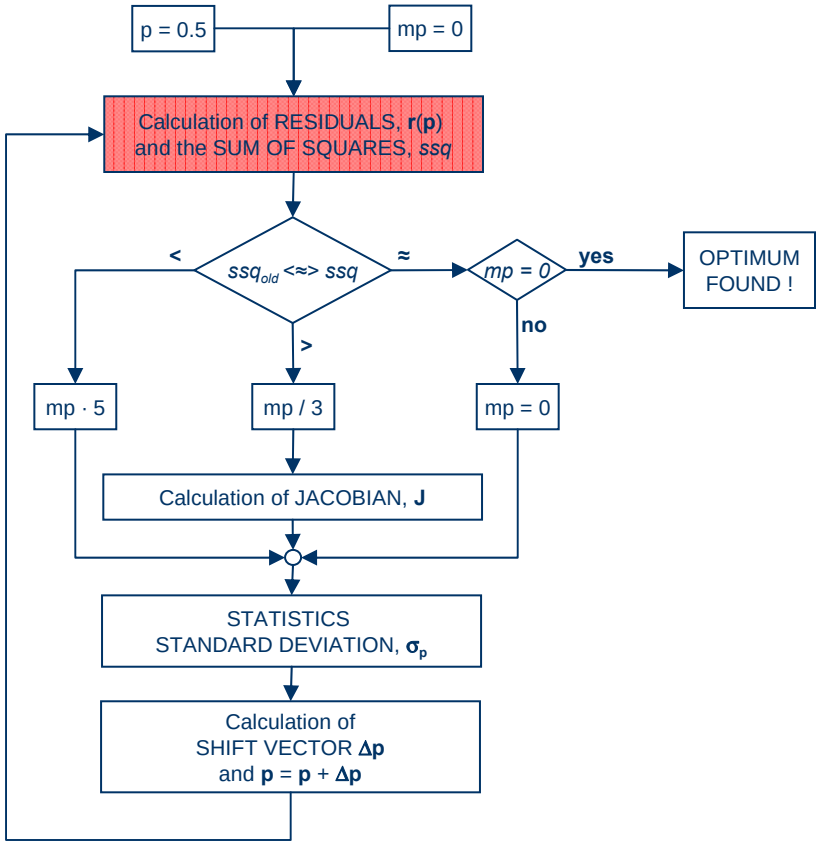

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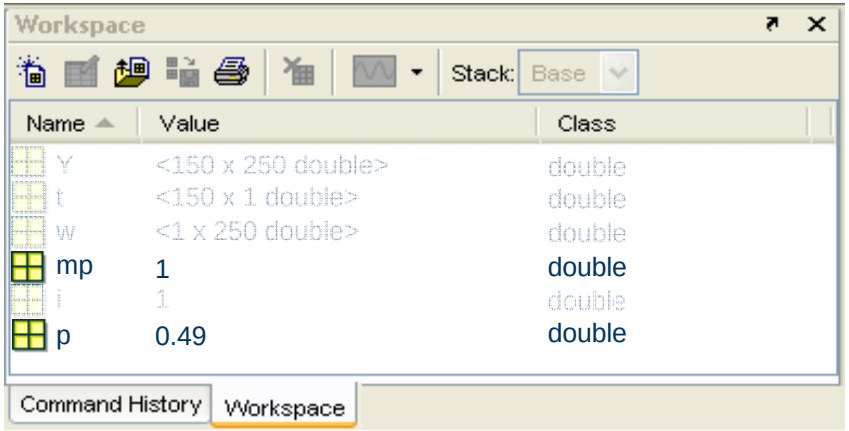

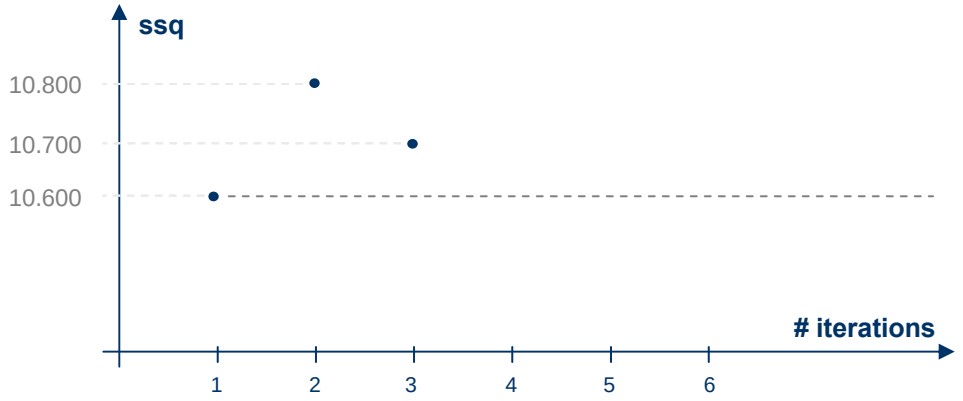

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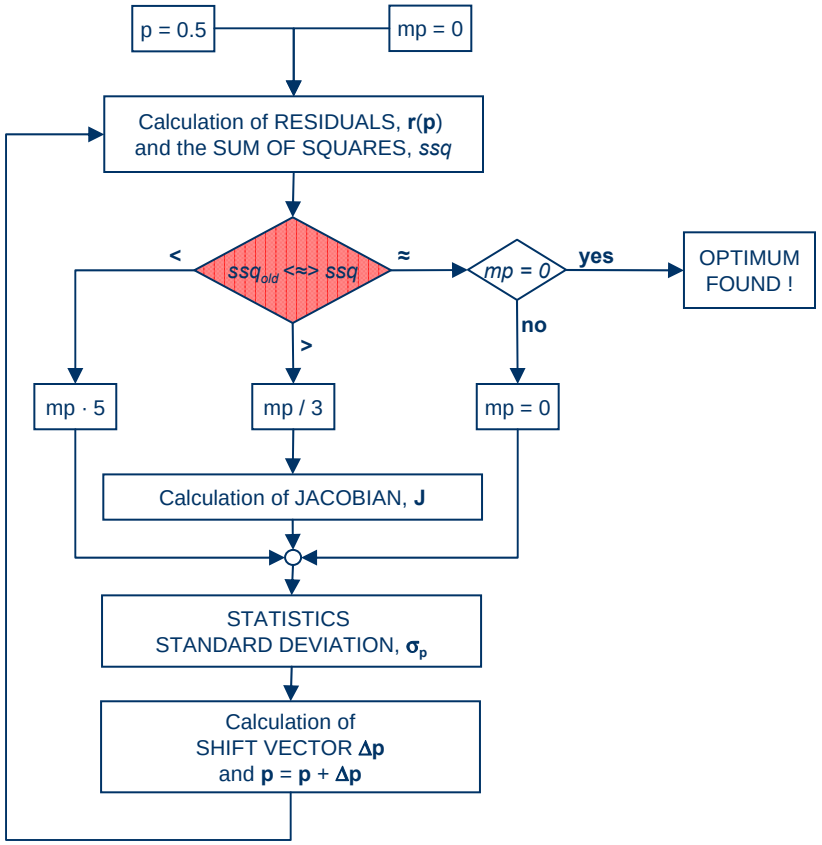

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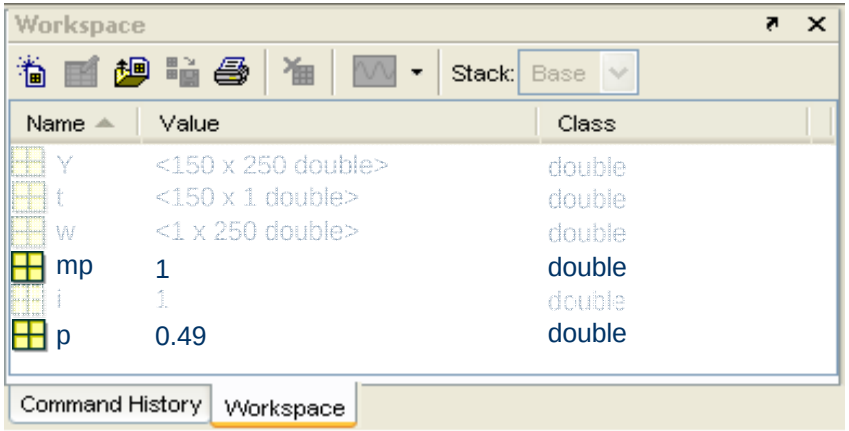

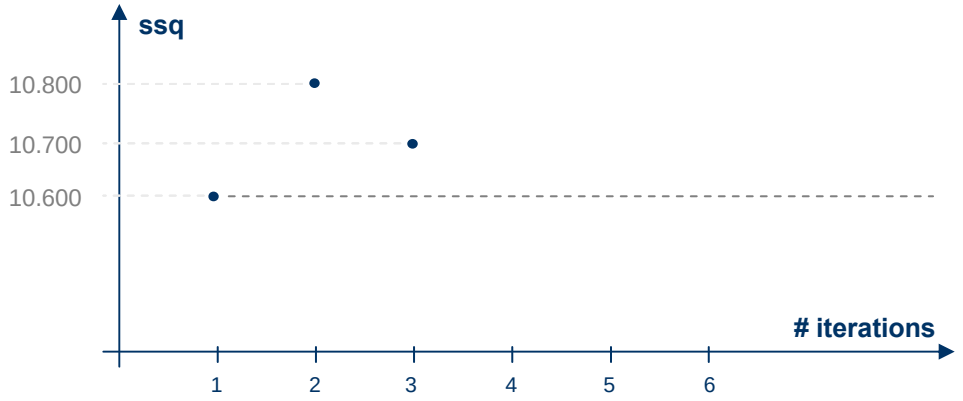

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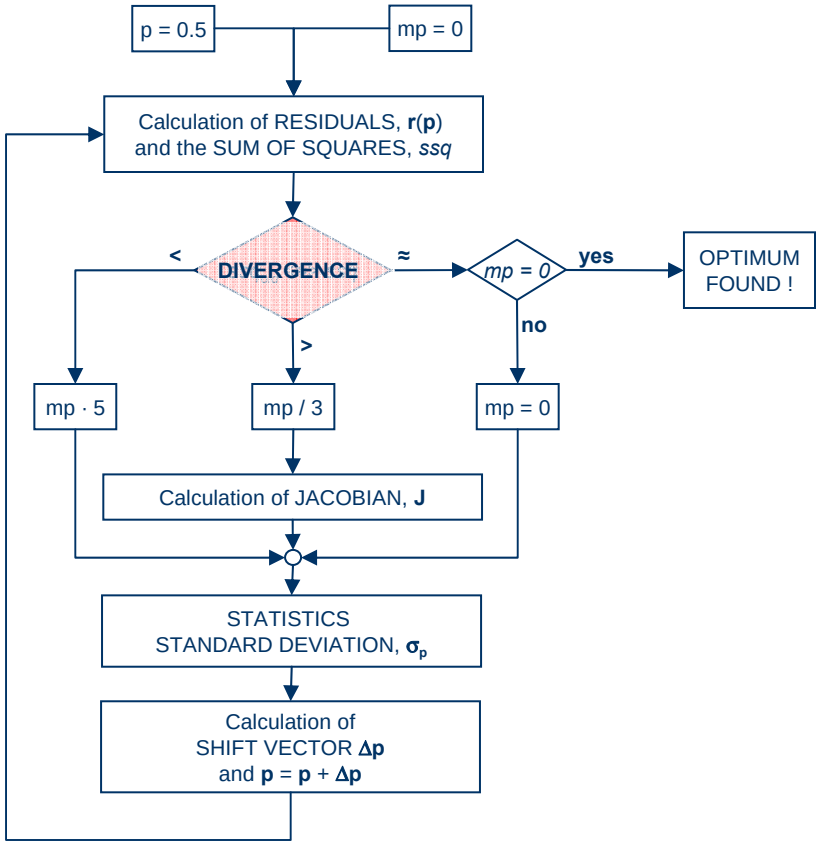

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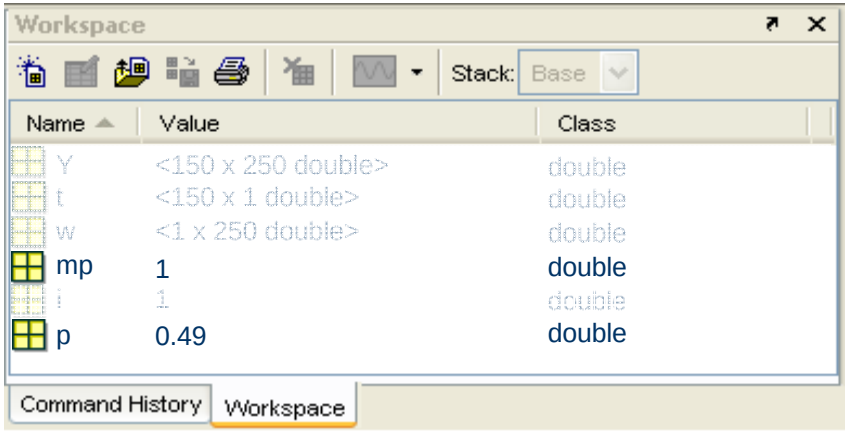

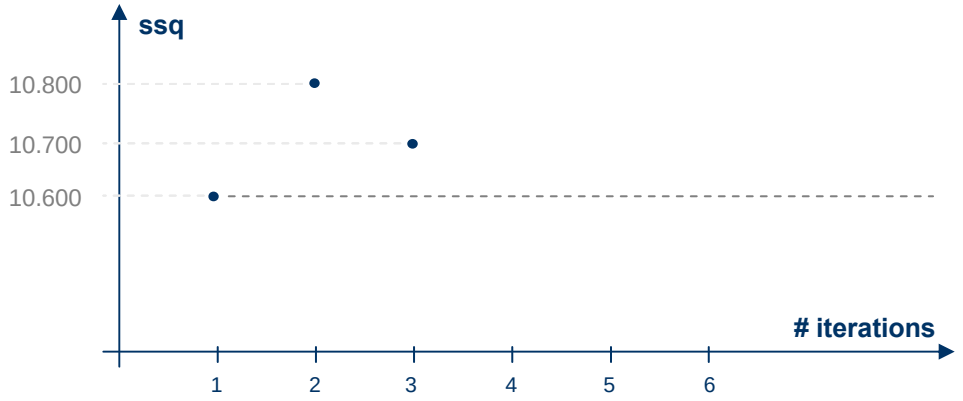

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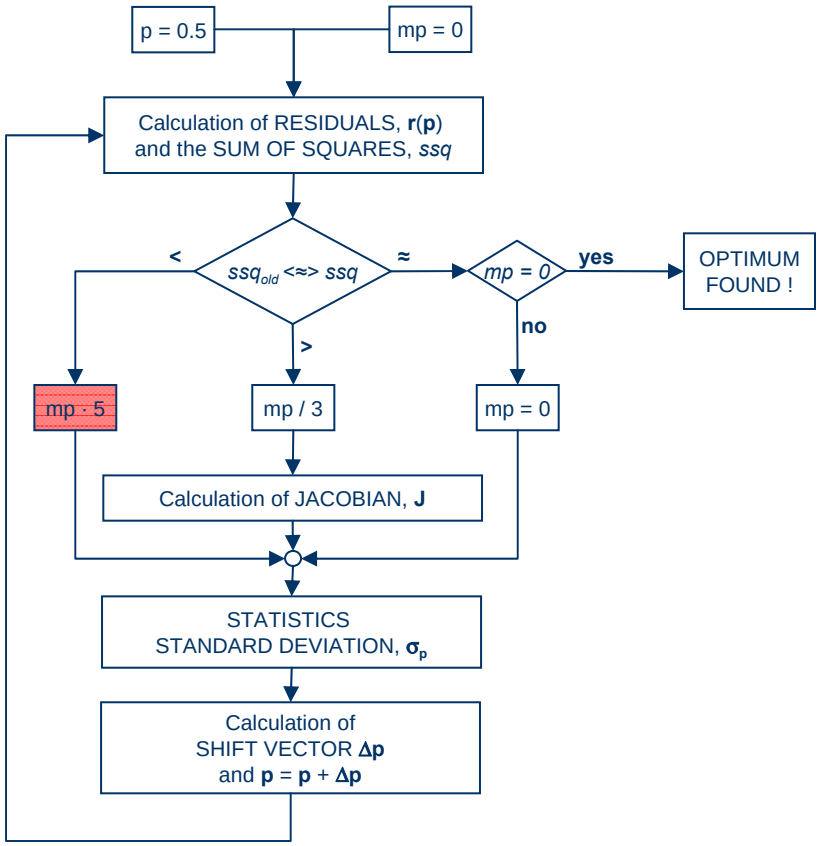

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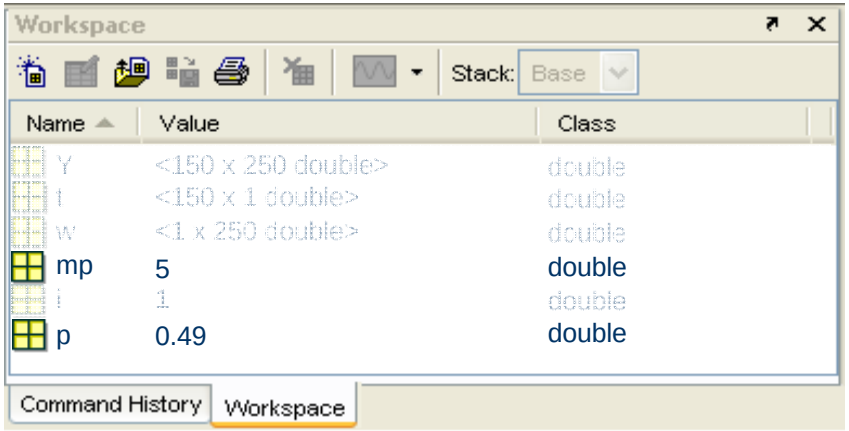

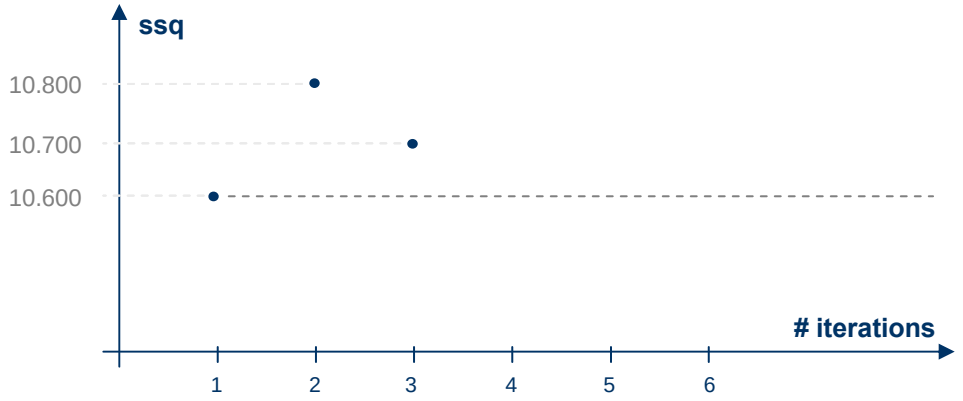

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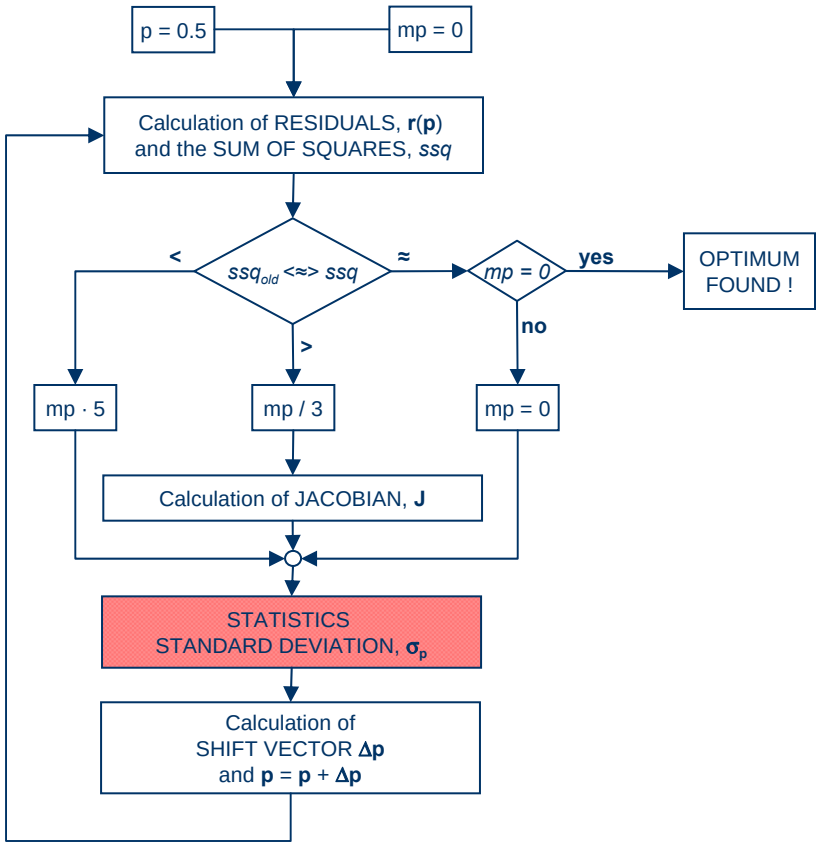

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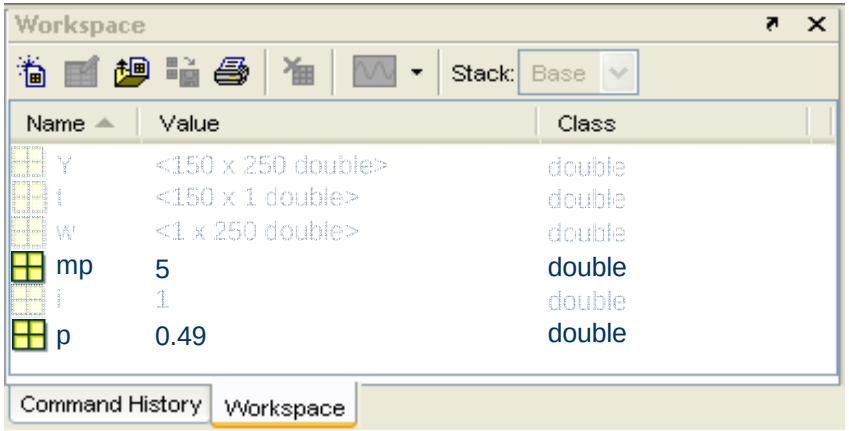
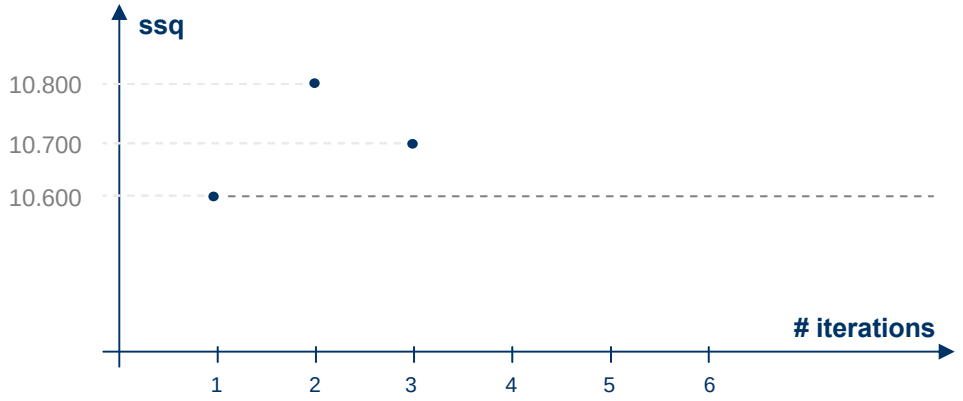
NG/Levenberg-Marquardt method





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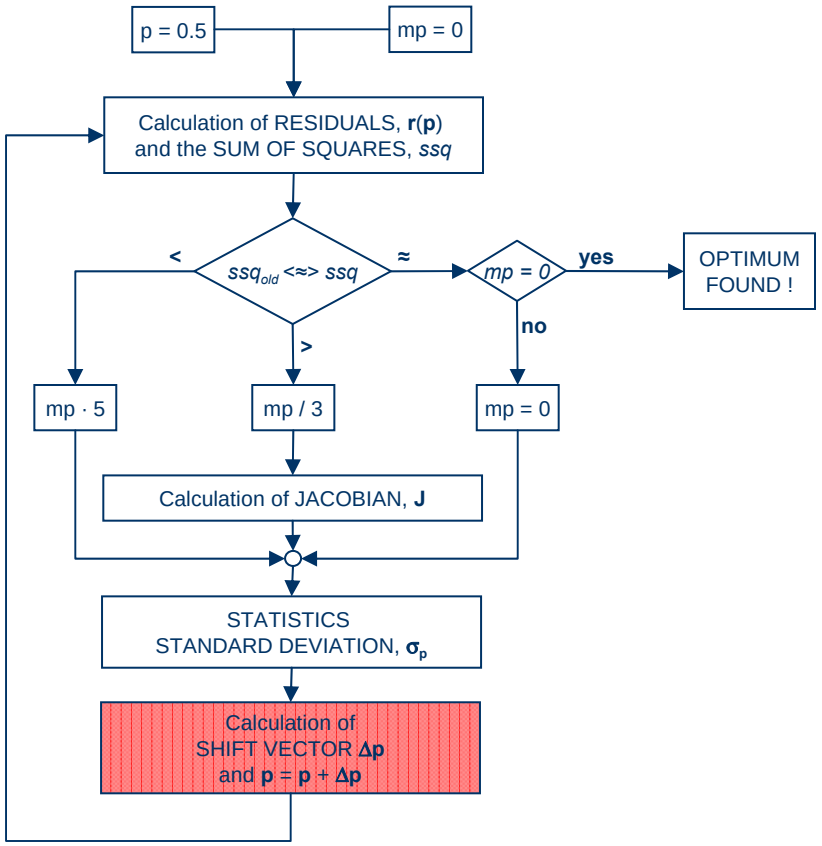

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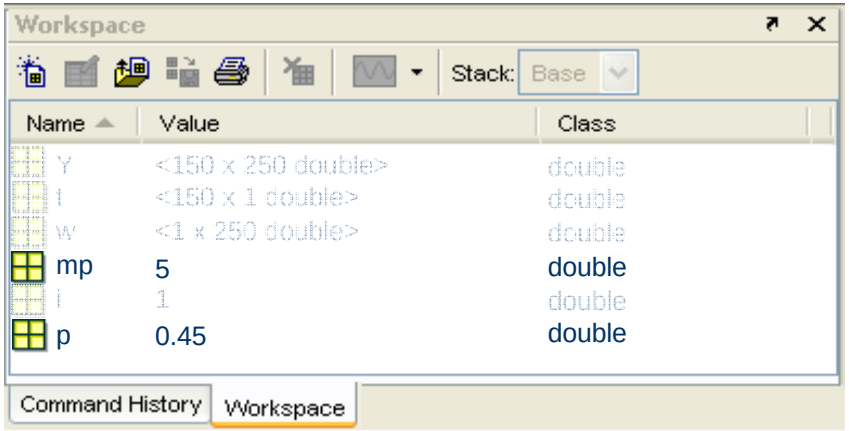

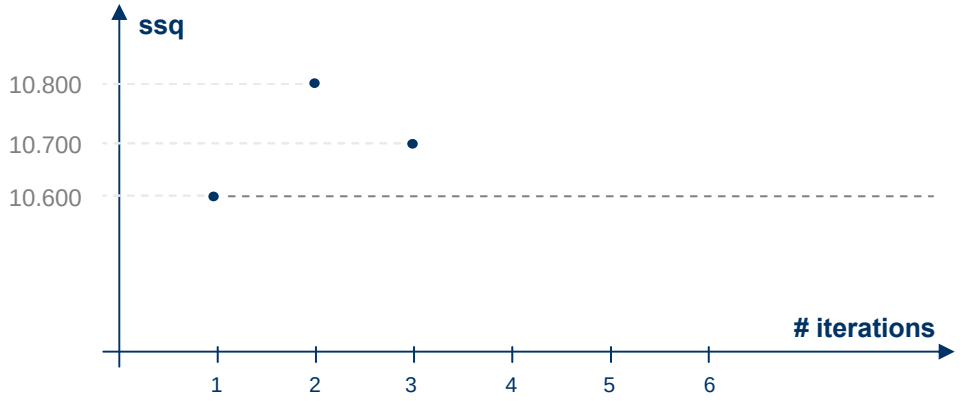

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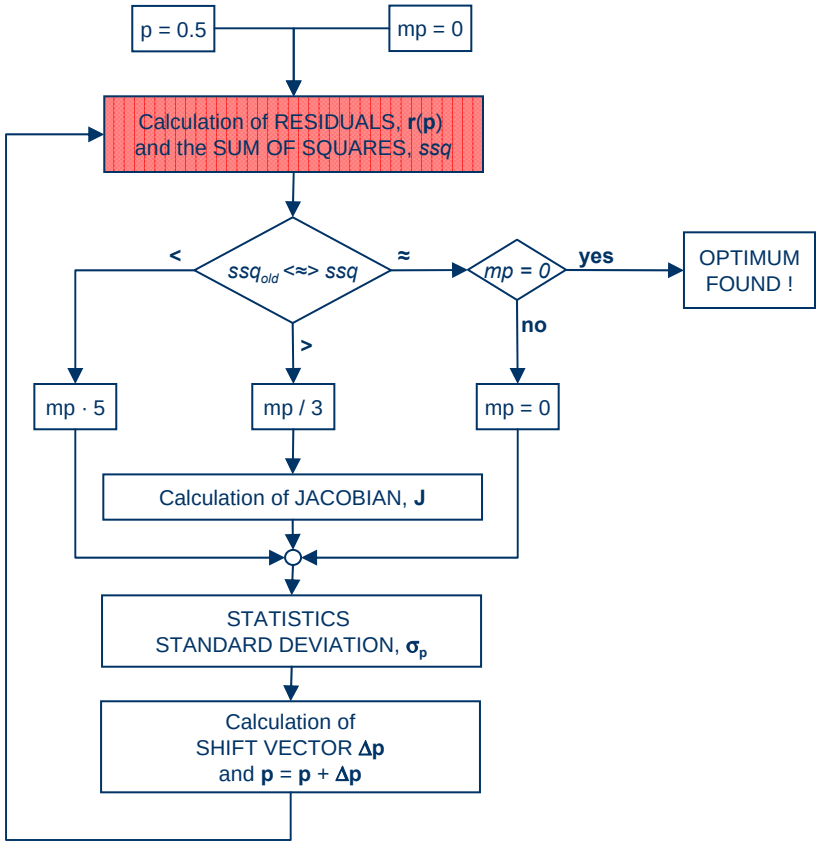

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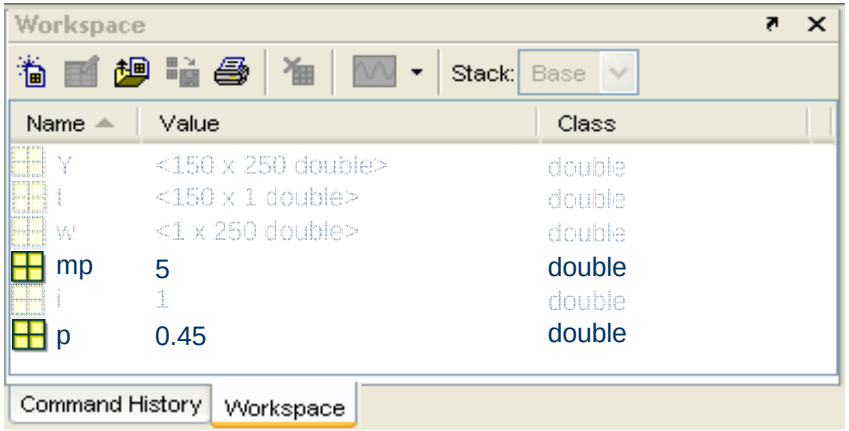

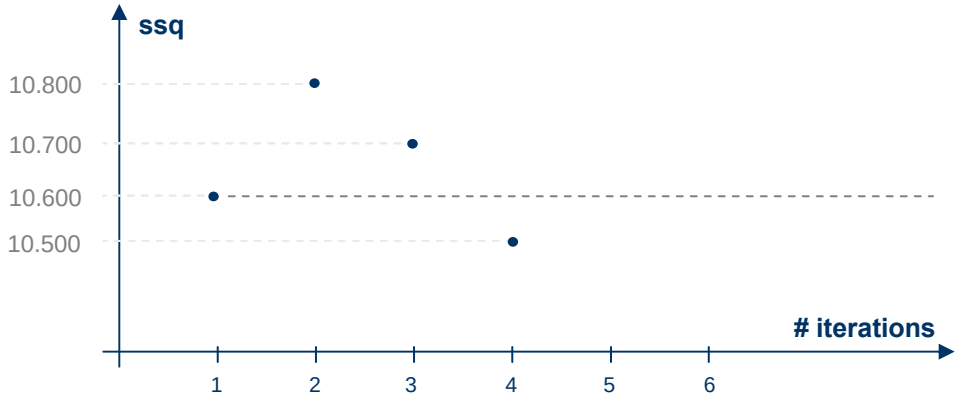

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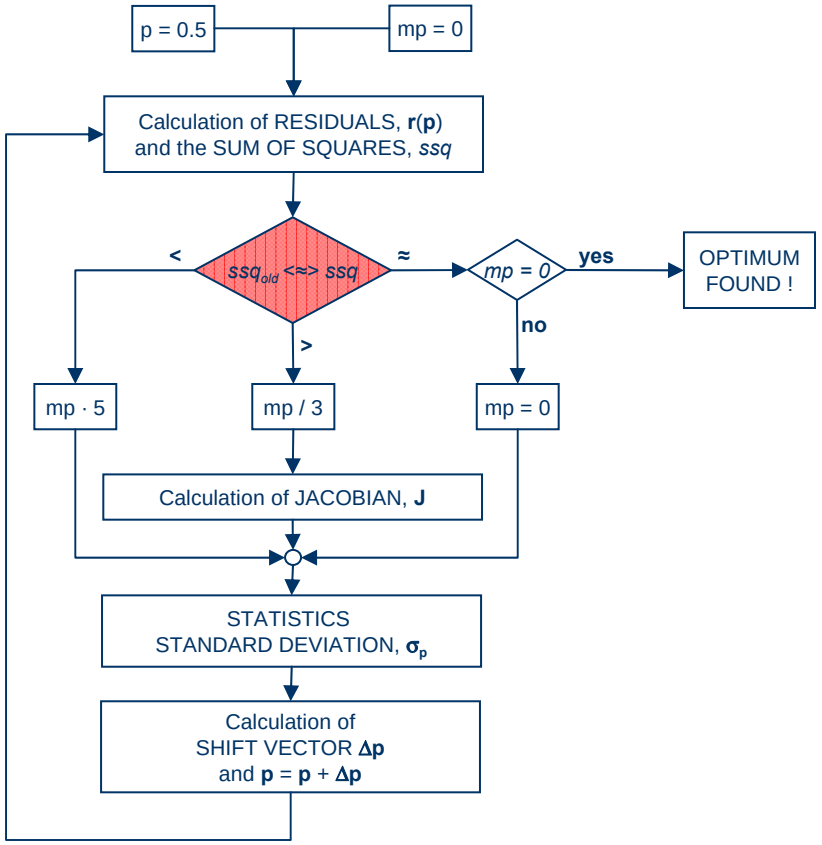

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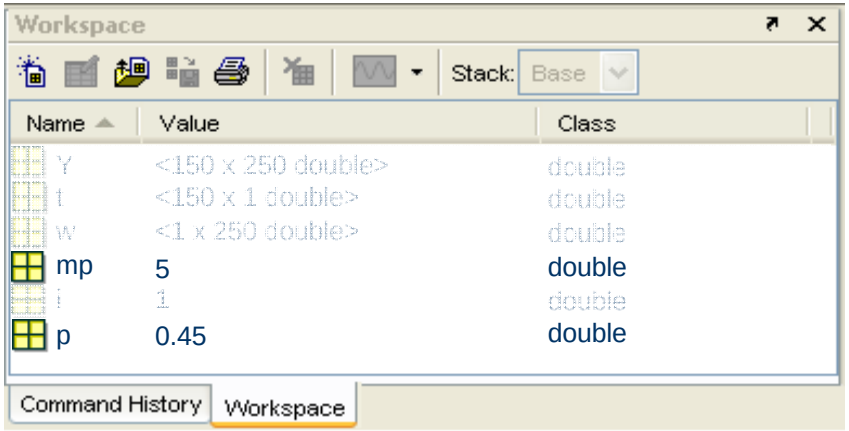

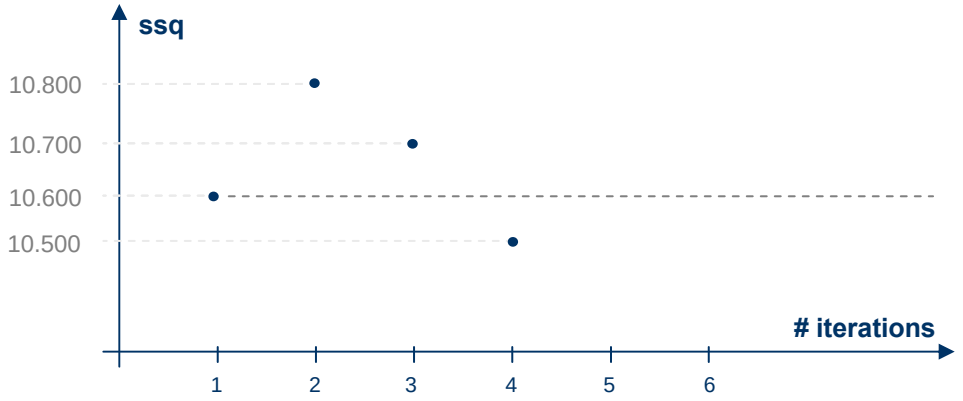

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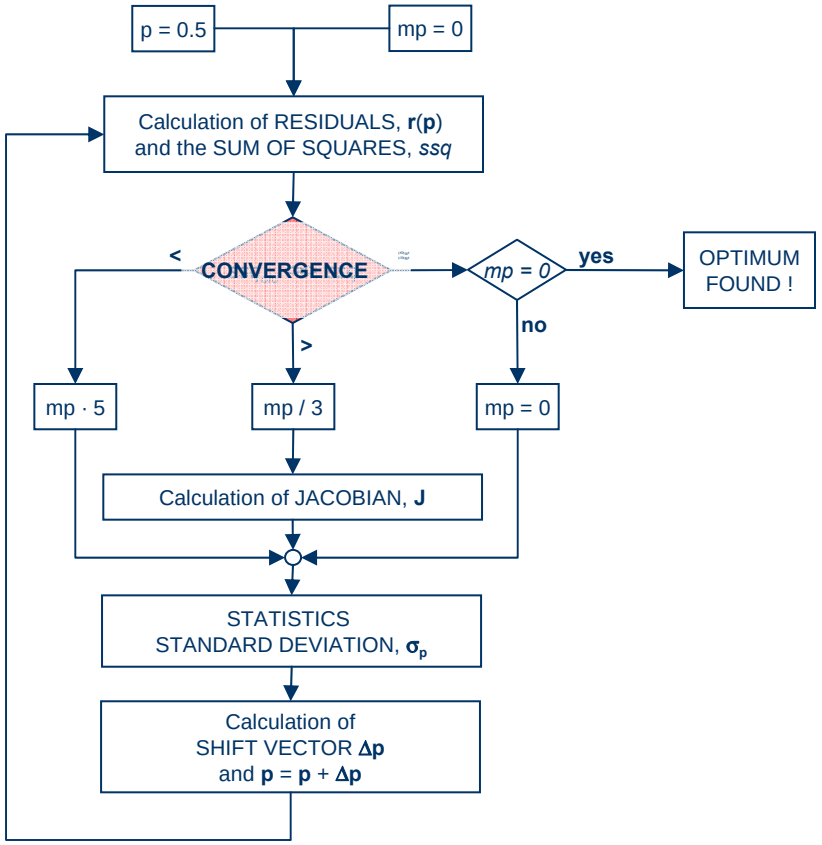

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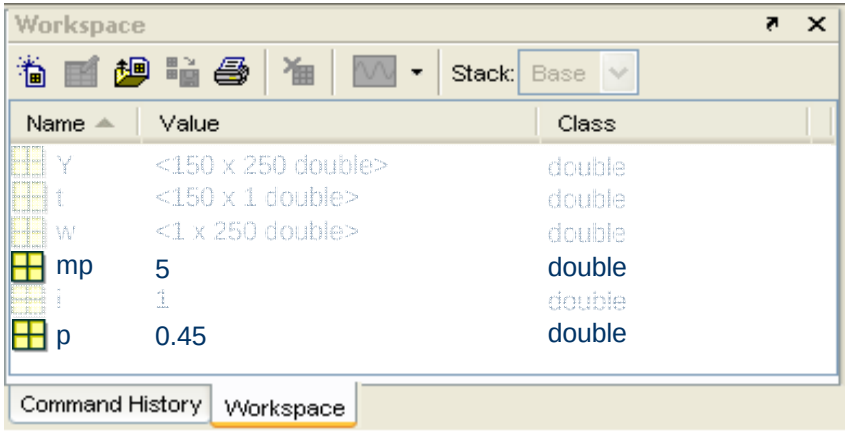

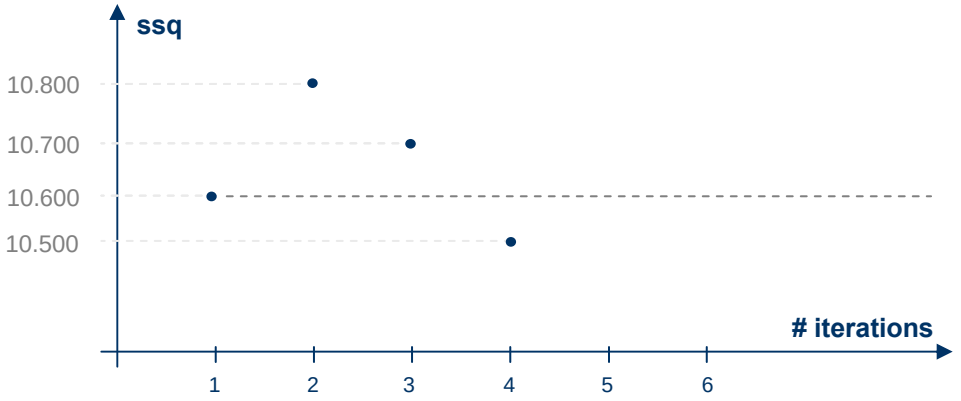

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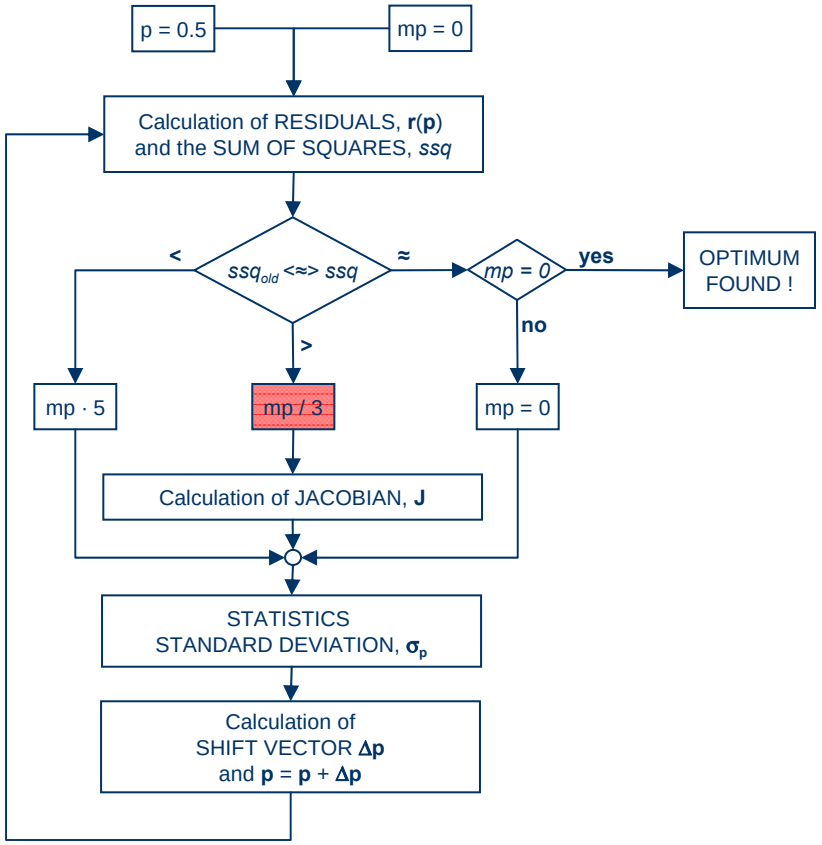

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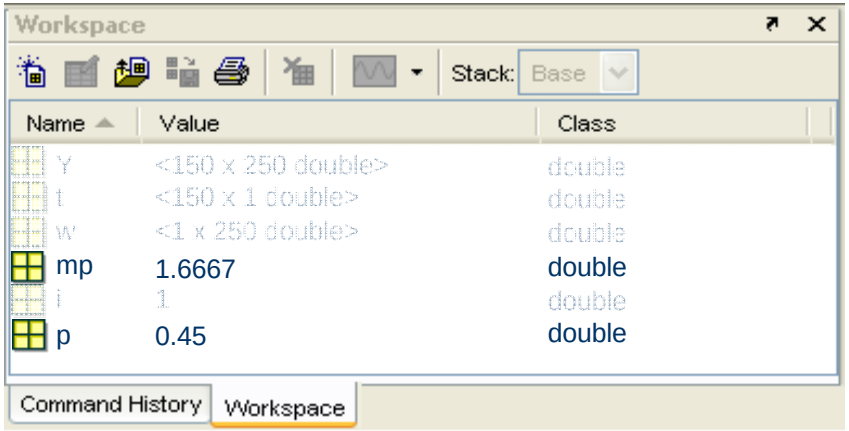

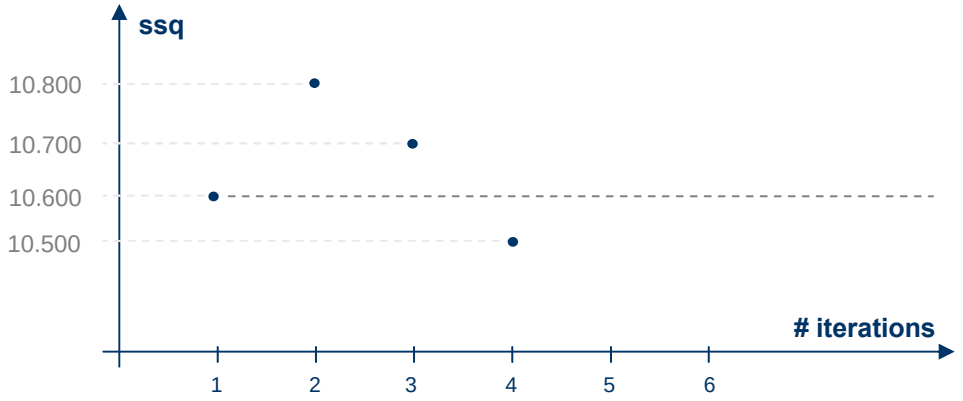

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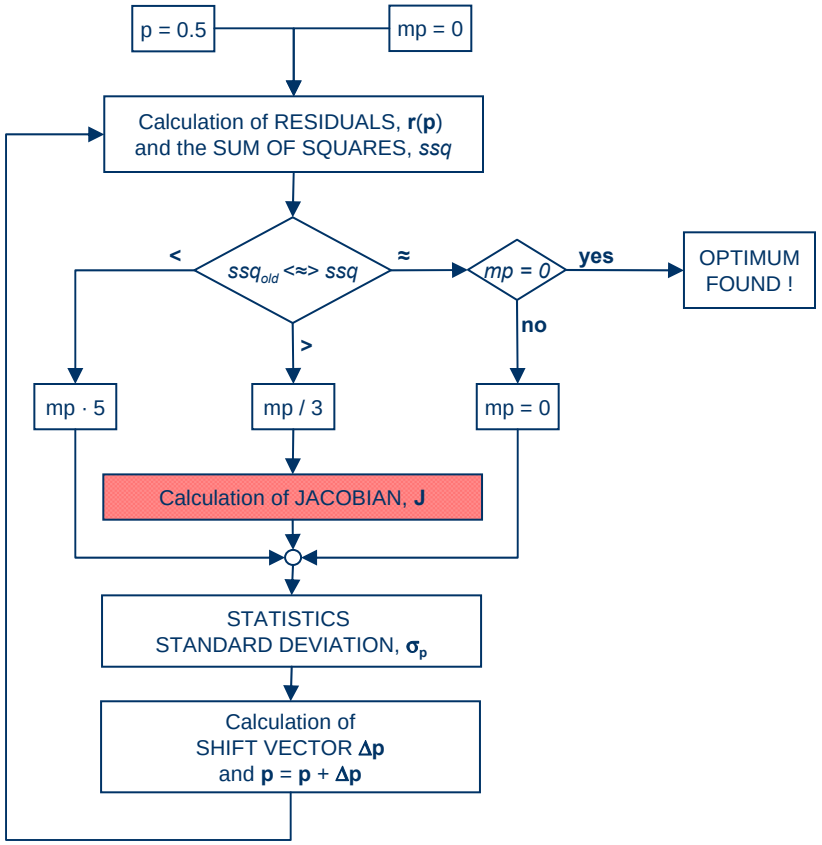

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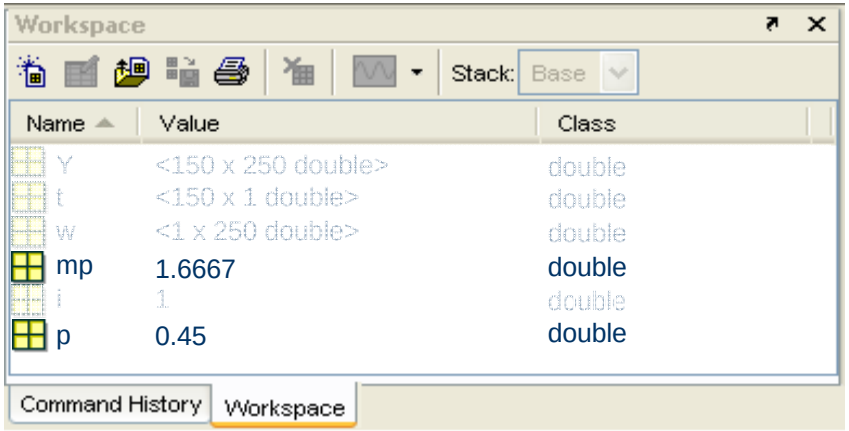

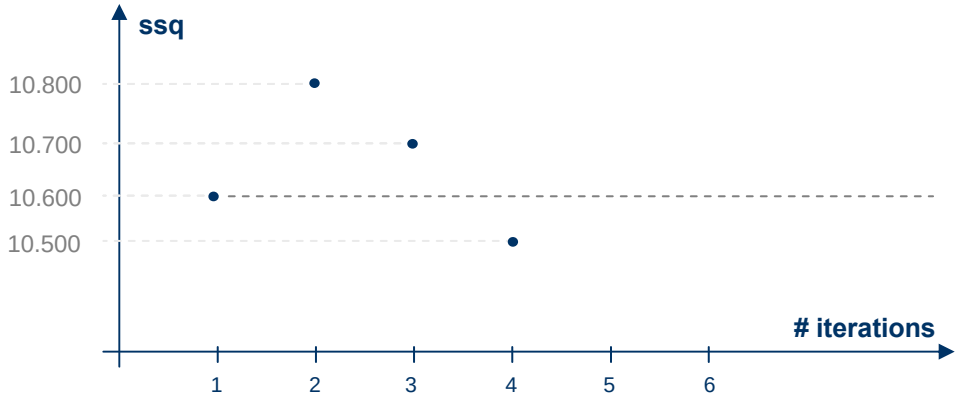

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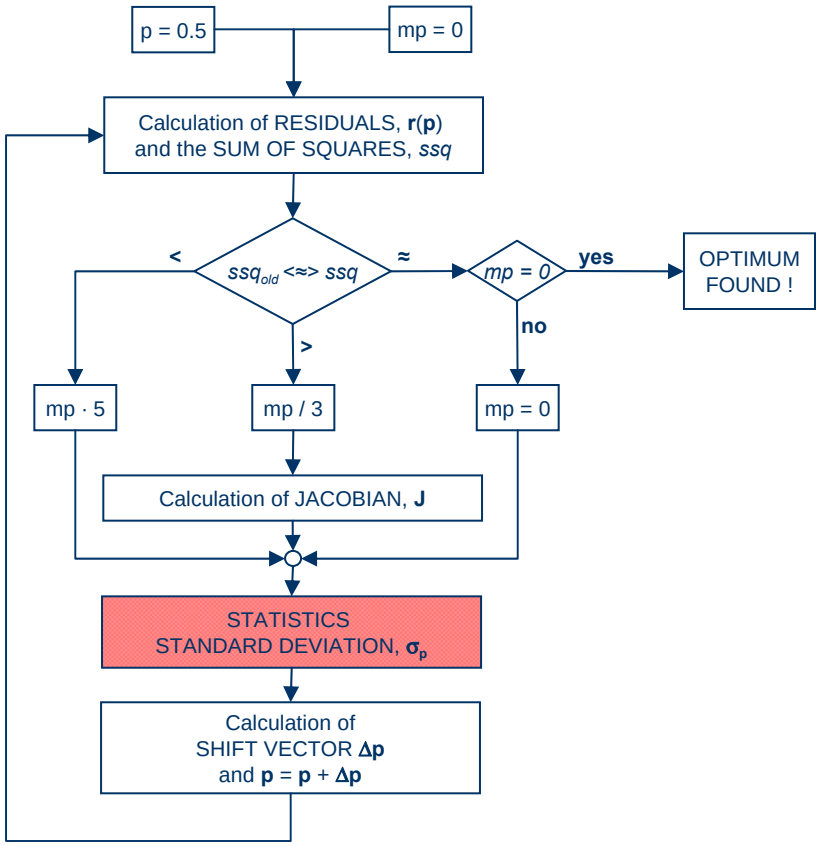

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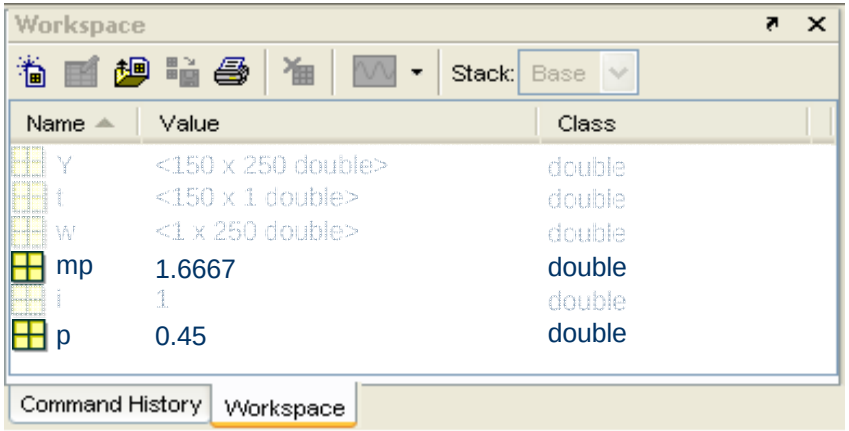

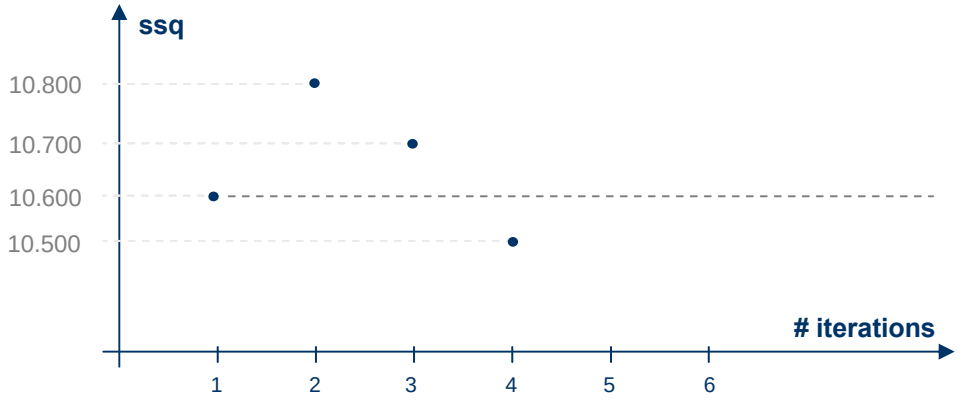

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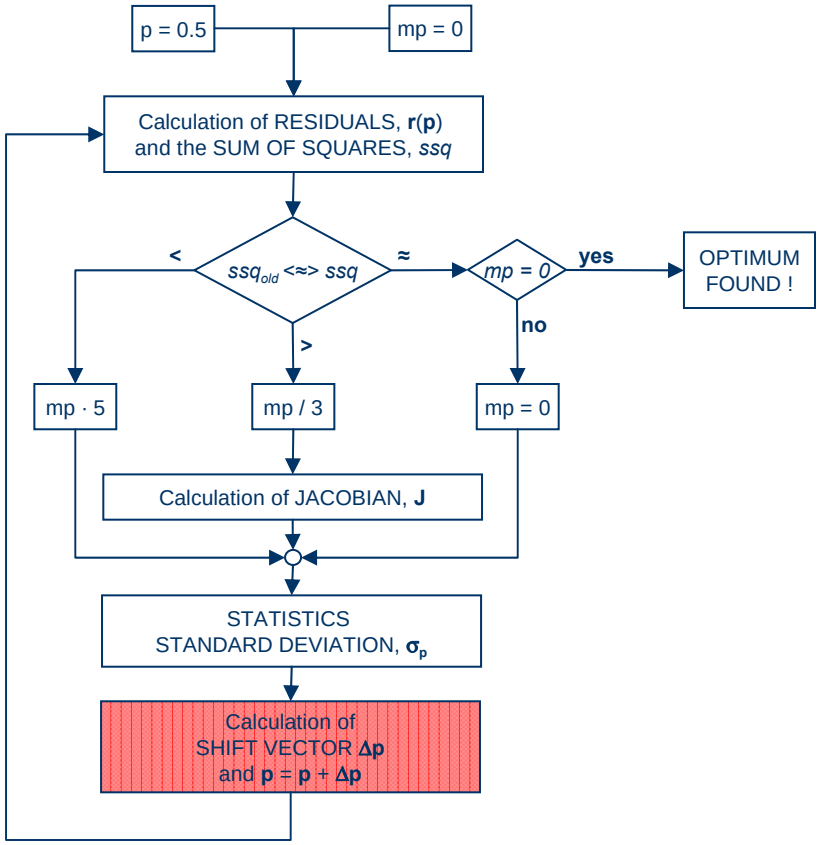

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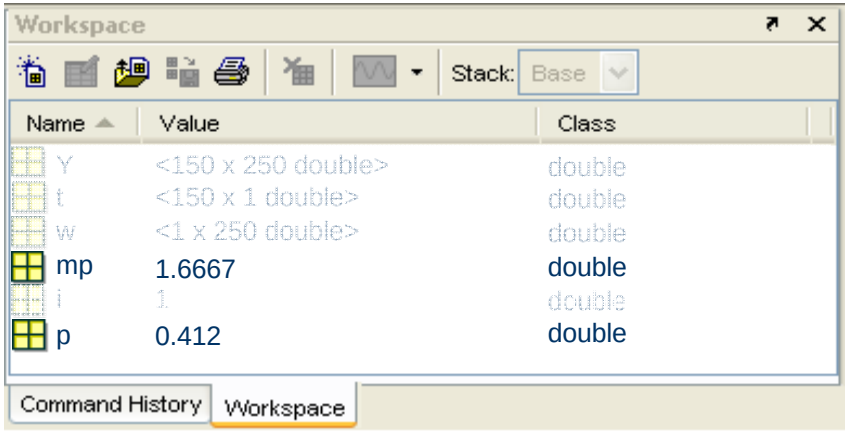

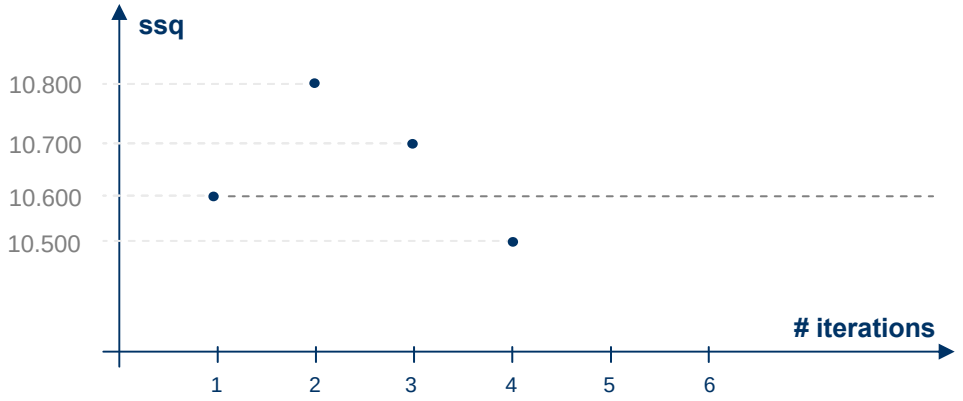

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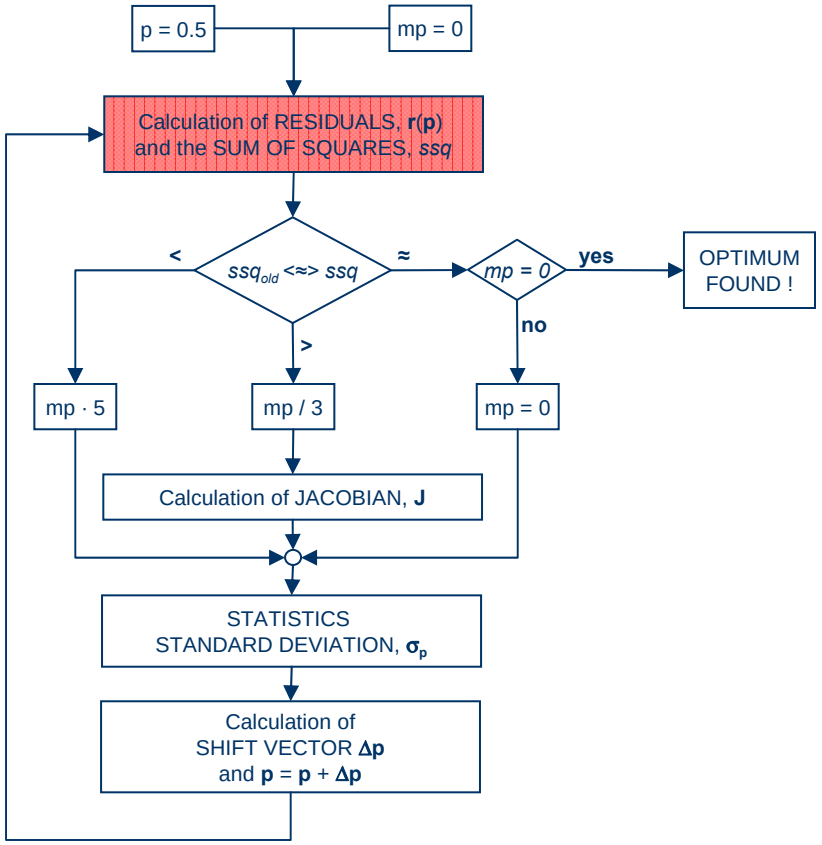
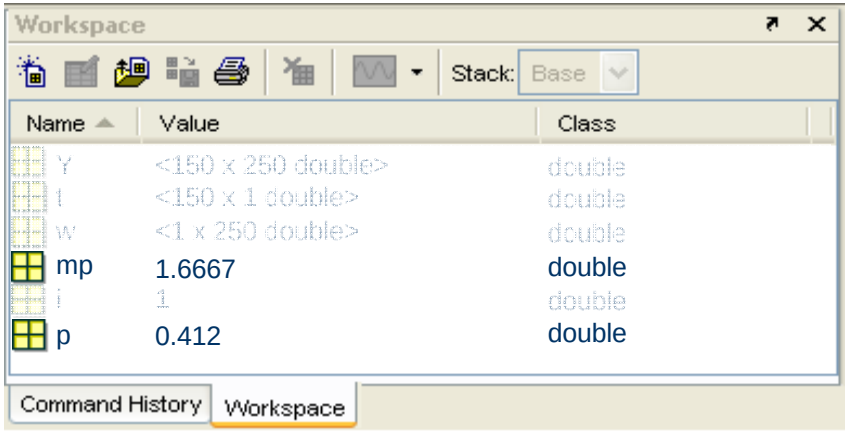
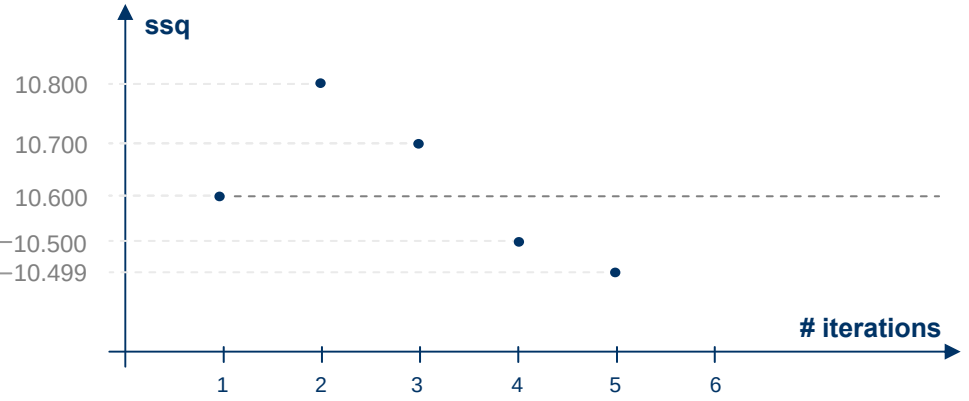


$$\frac{10.500-10.499}{10.500}=9.523\cdot10^{-5}<10^{-4}$$

Current method :

NG/Levenberg-Marquardt method





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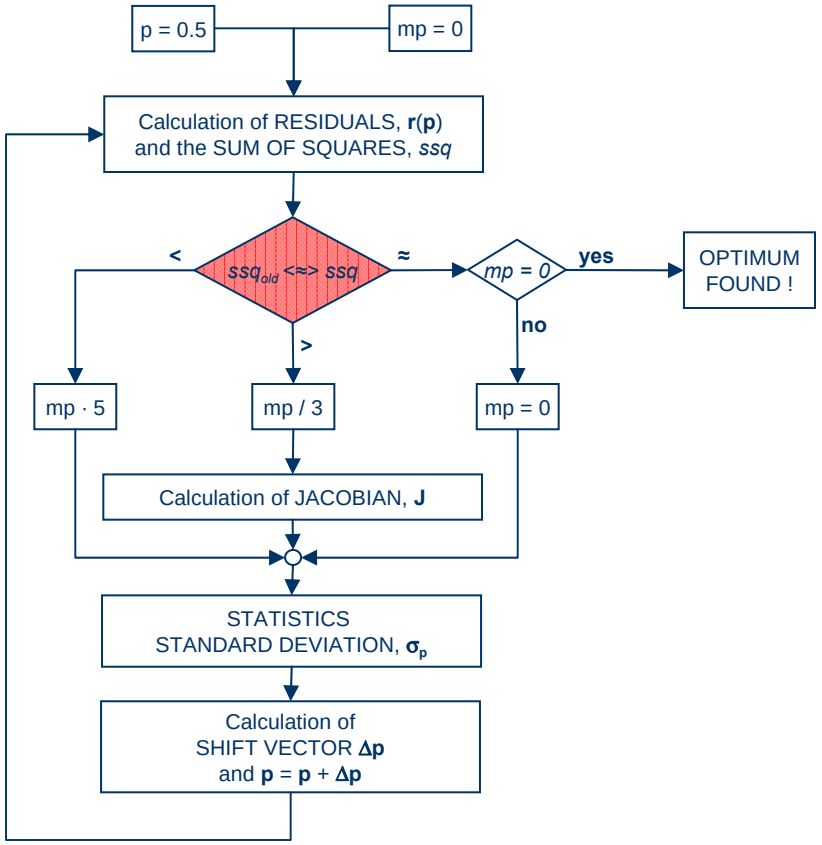

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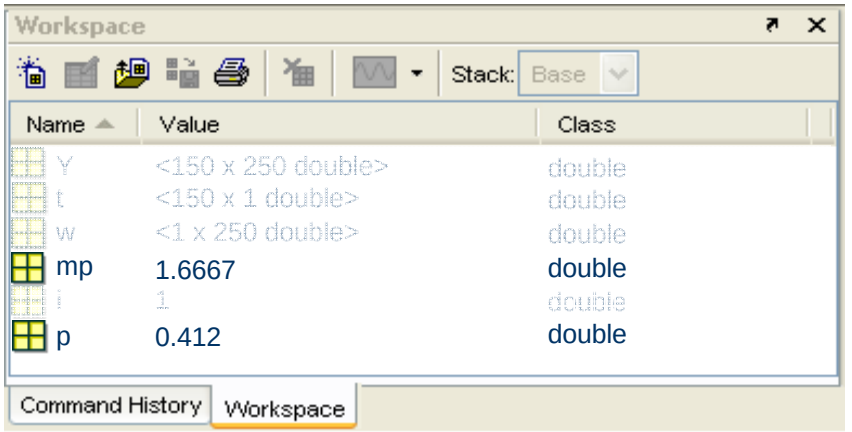
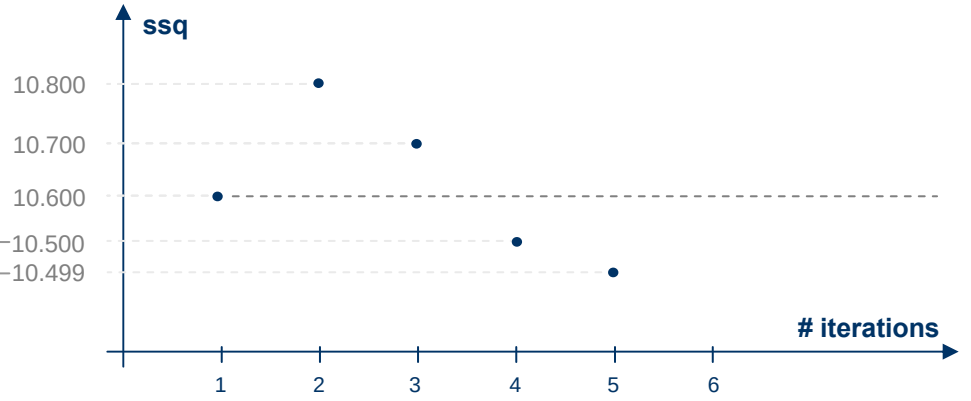
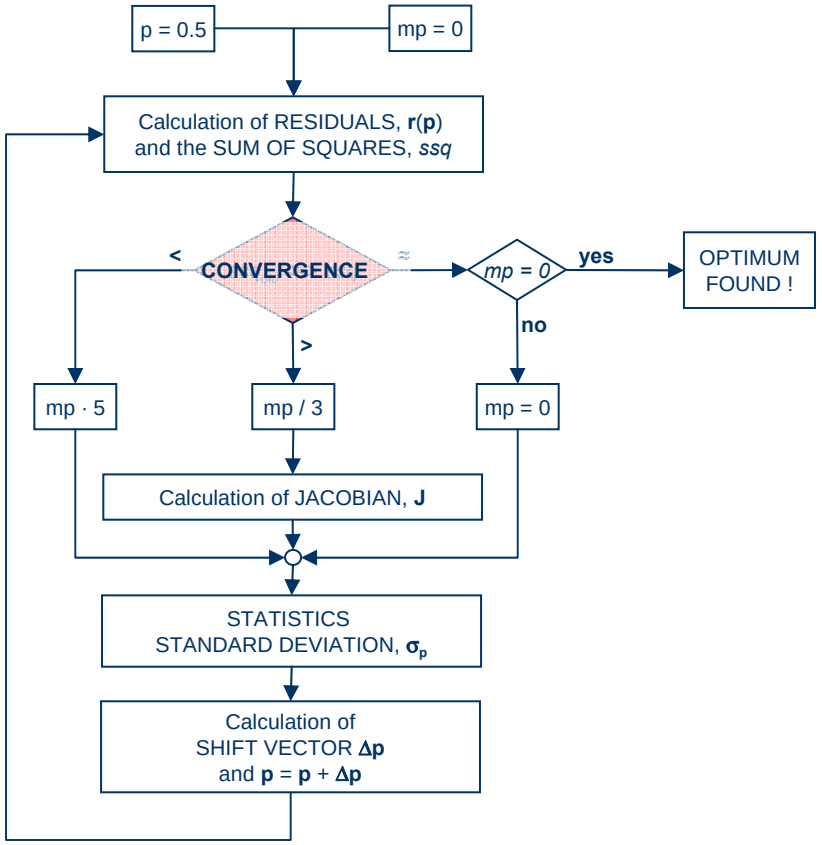




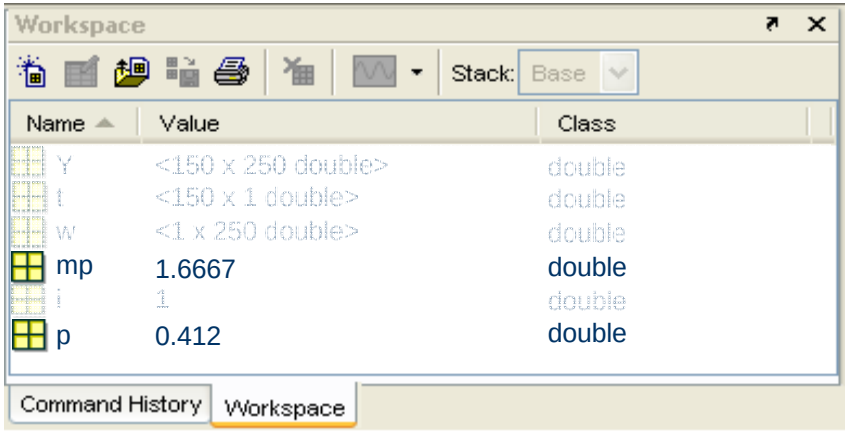
The NG/LM algorithm

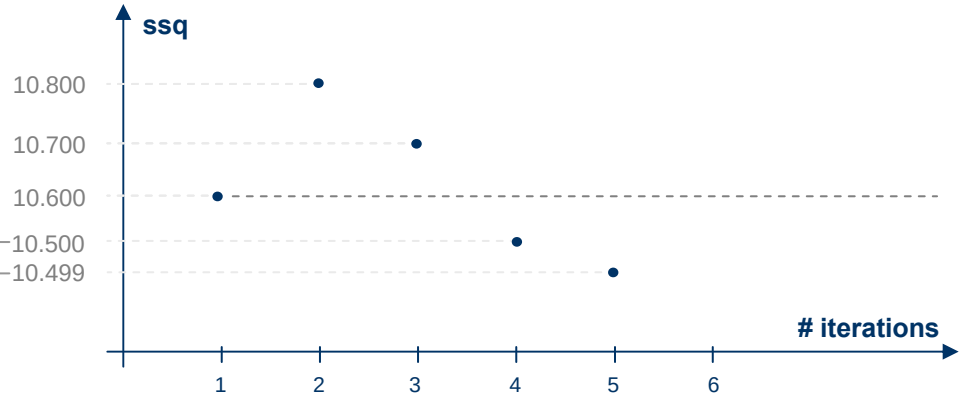


$$\frac{10.500-10.499}{10.500}=9.523\cdot10^{-5}<10^{-4}$$

Current method :

NG/Levenberg-Marquardt method





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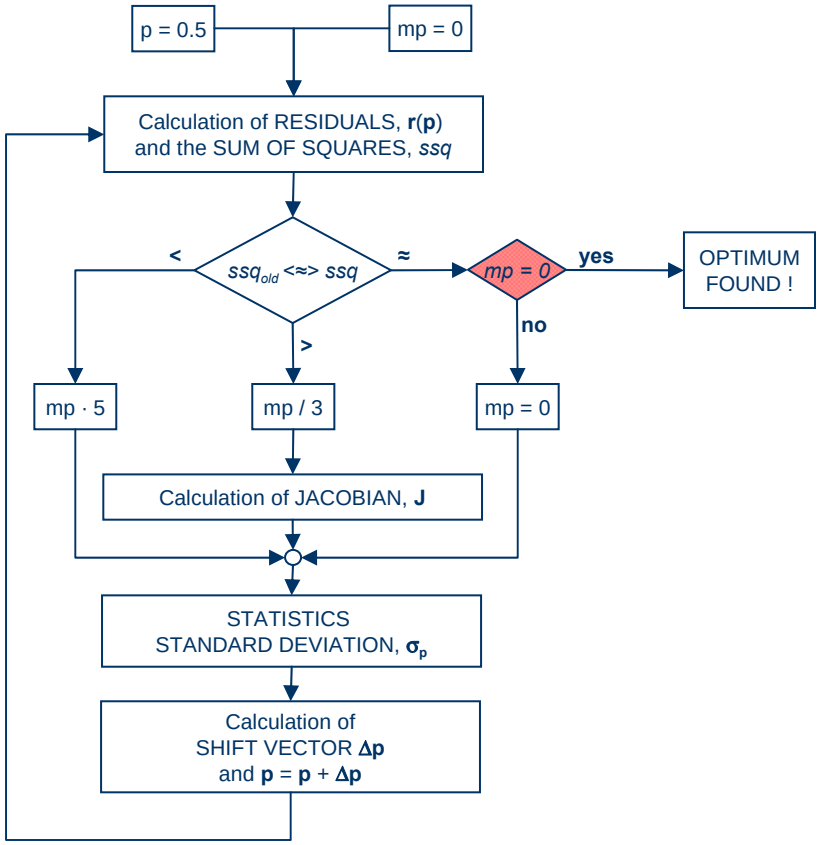

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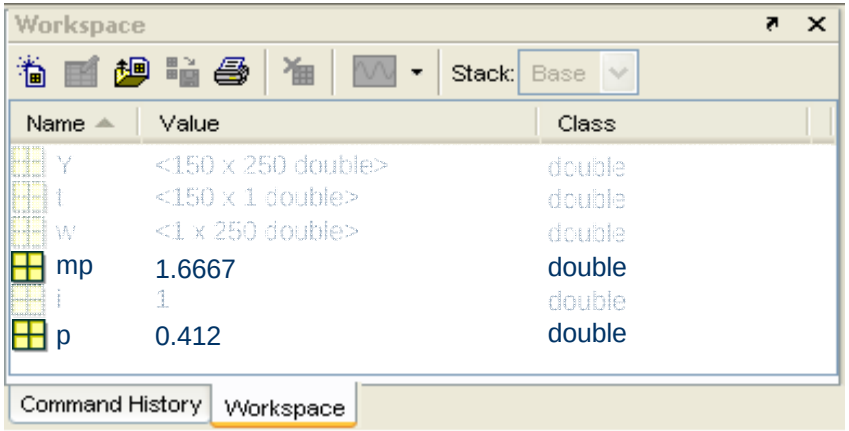

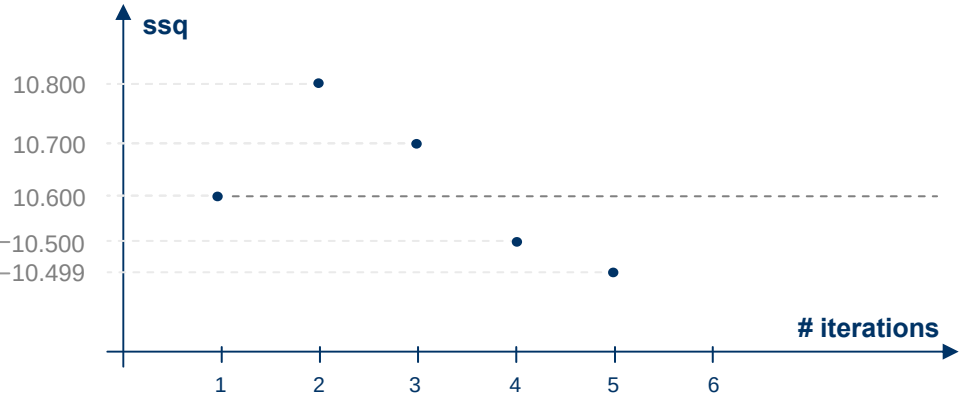

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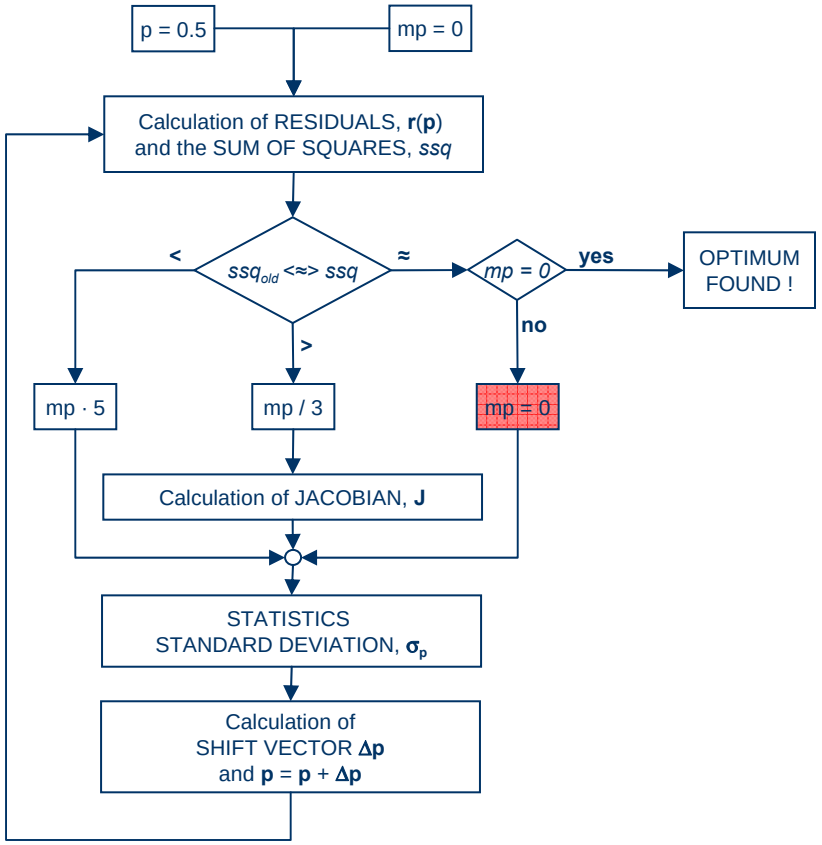

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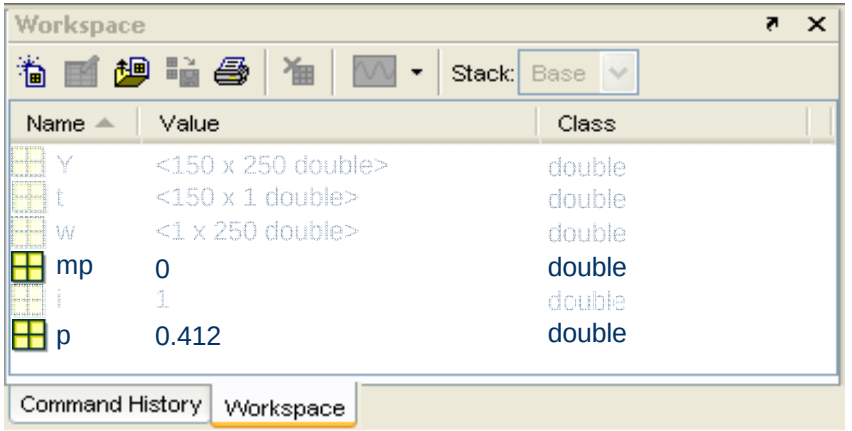
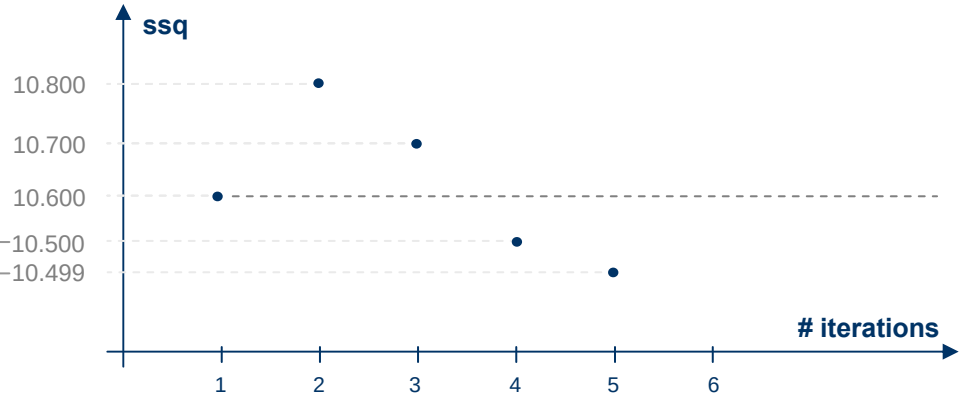
Newton-Gauss method





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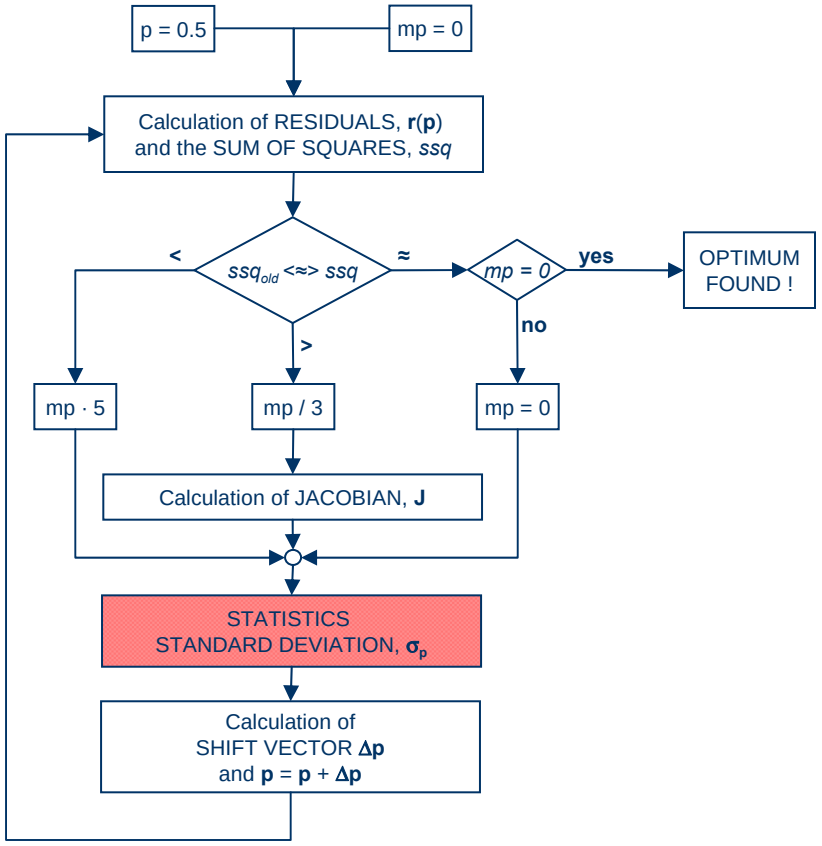

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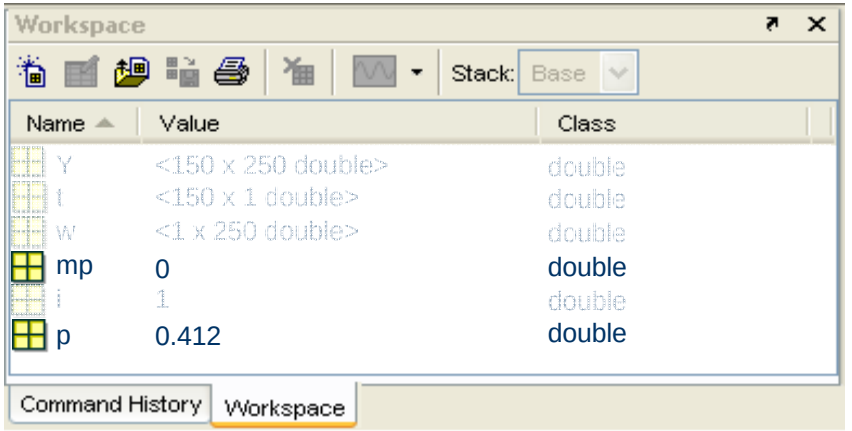

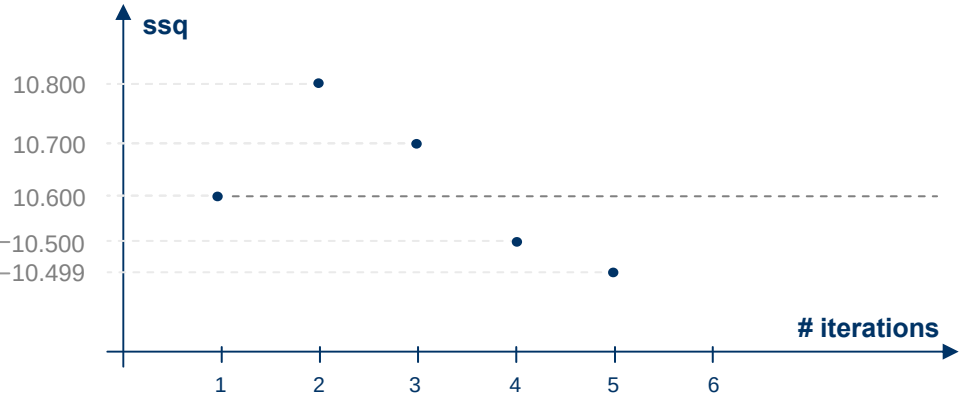

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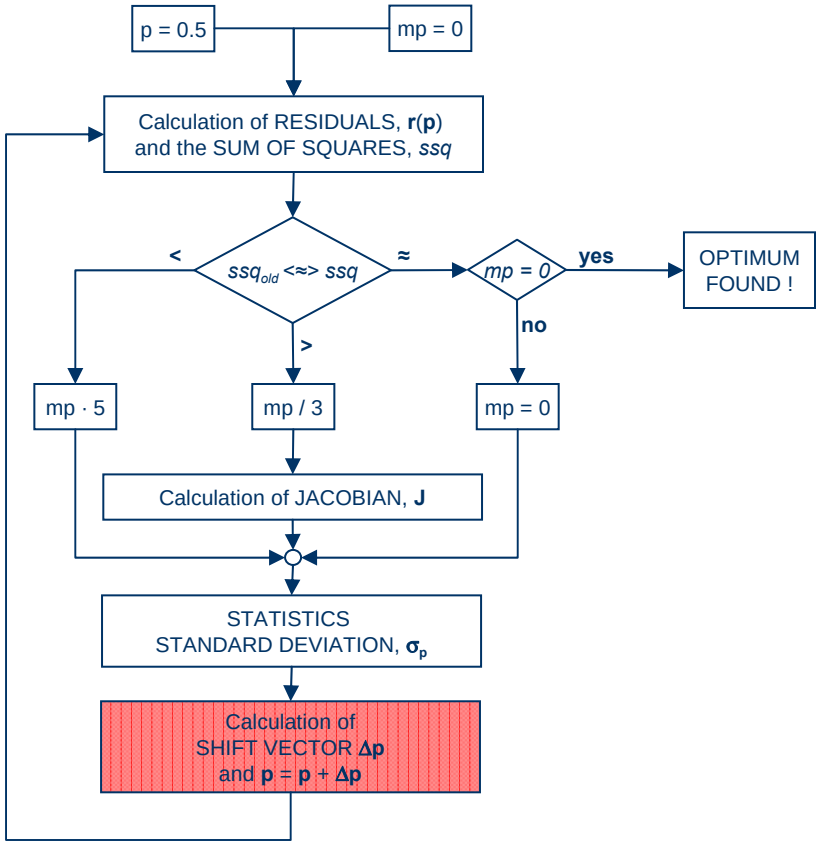

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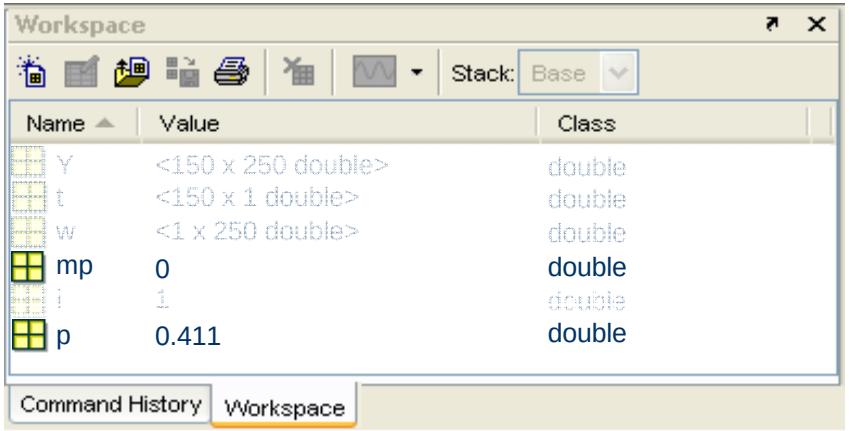

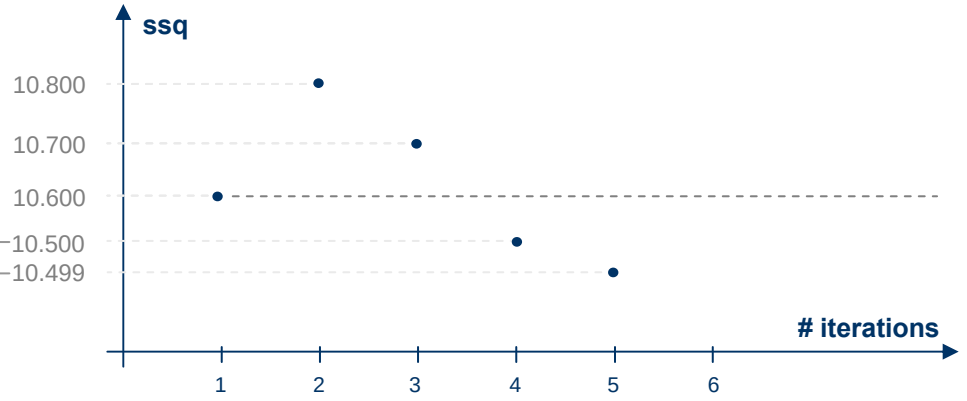

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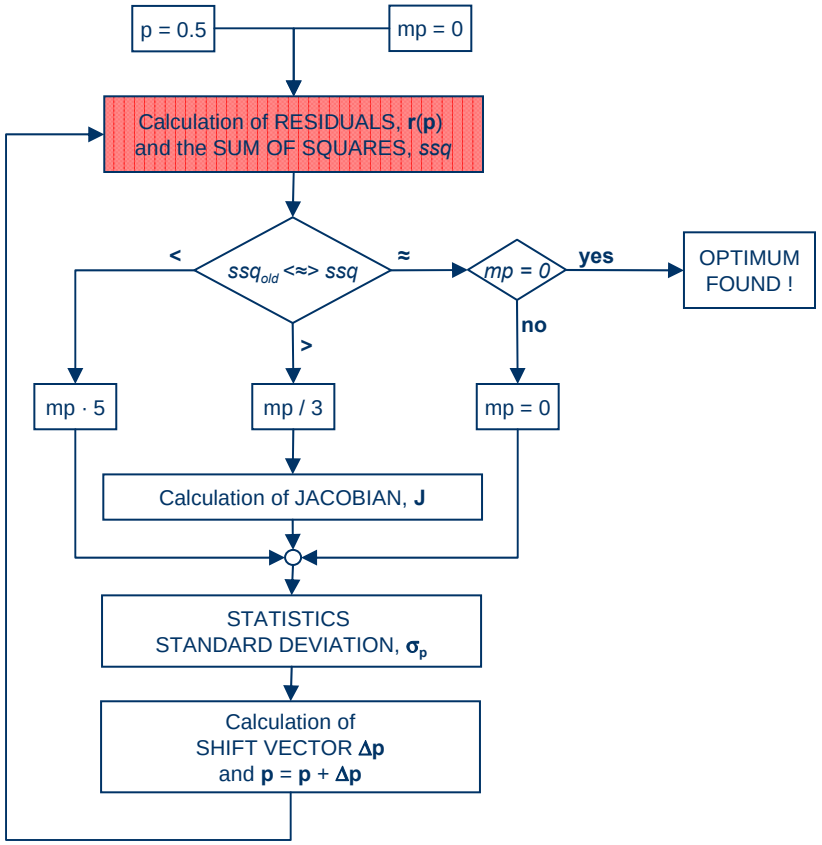

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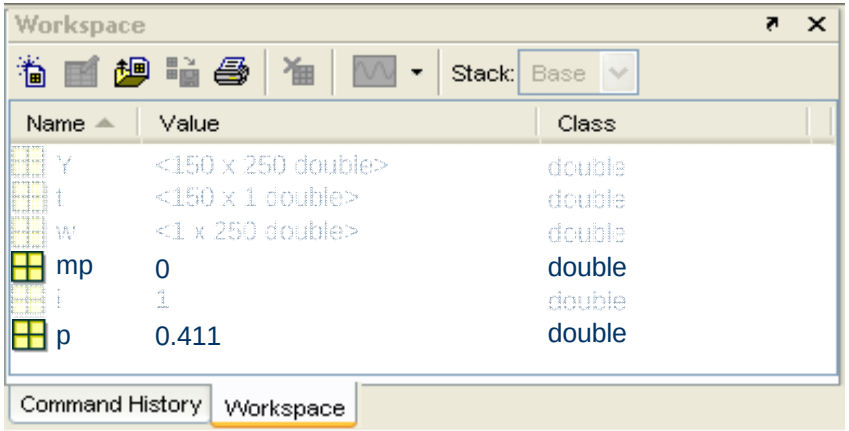

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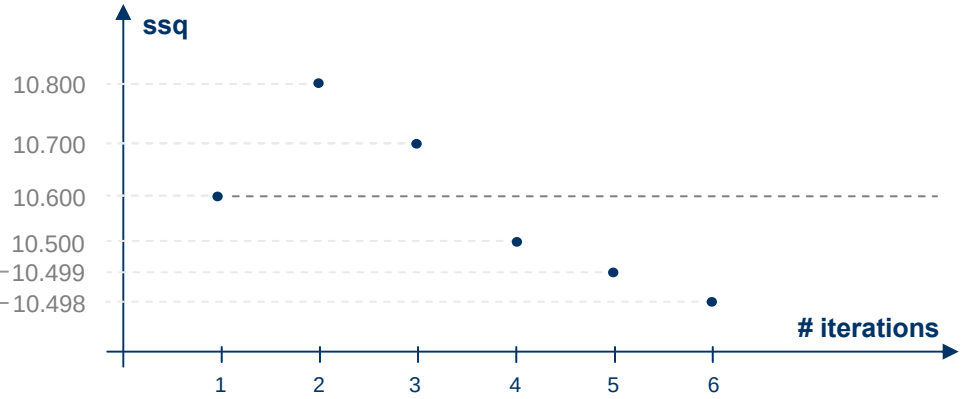

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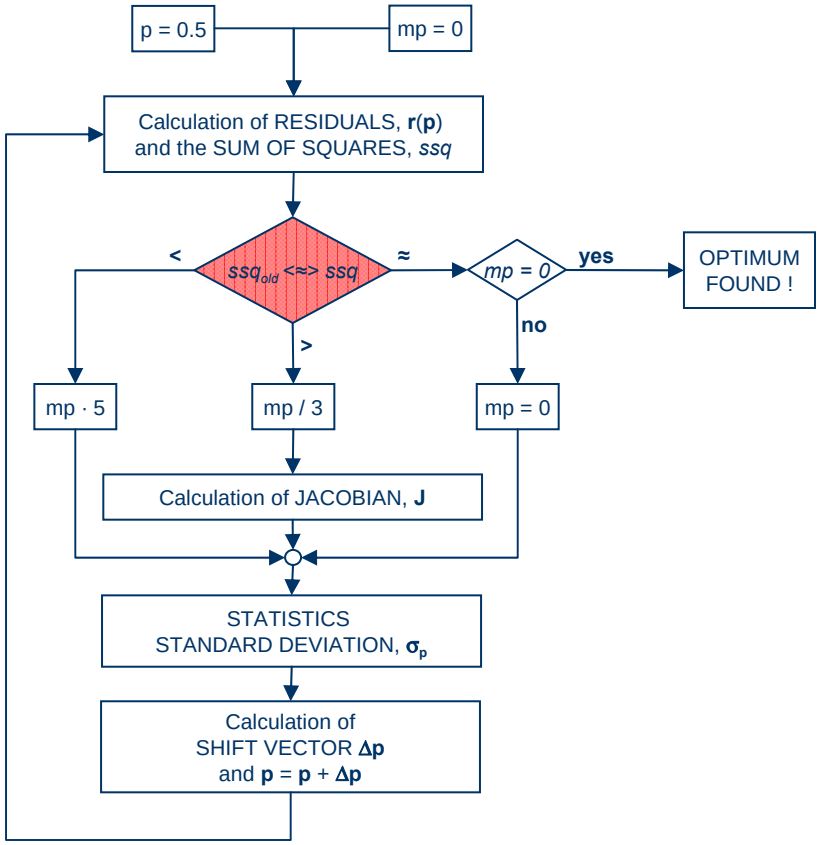

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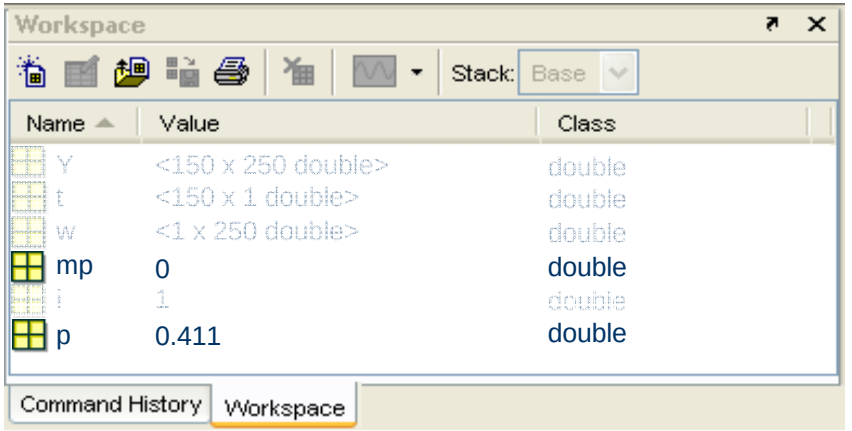

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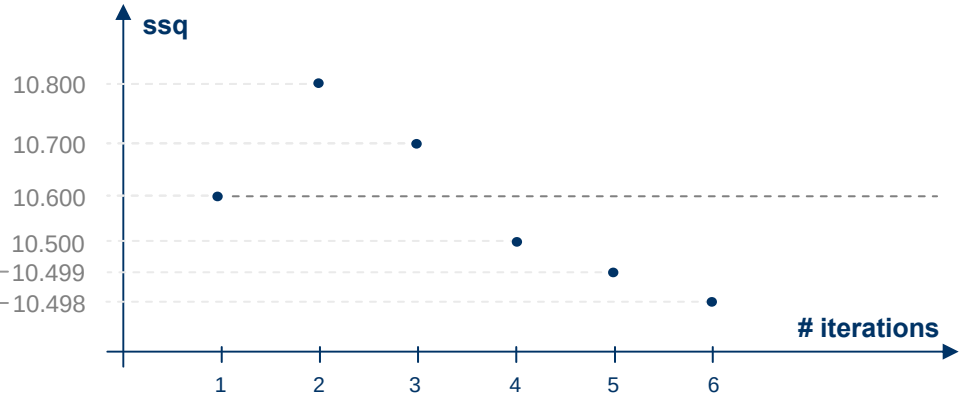

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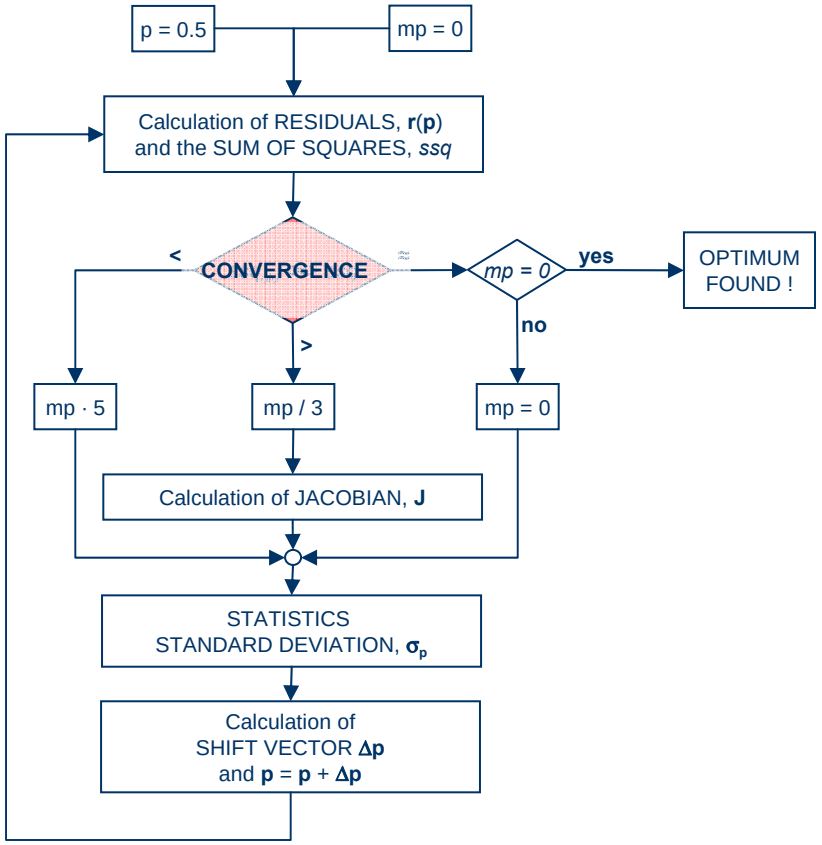

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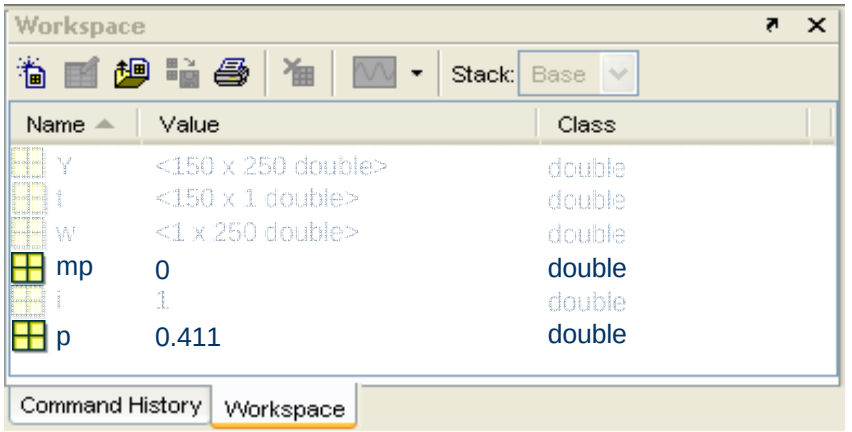

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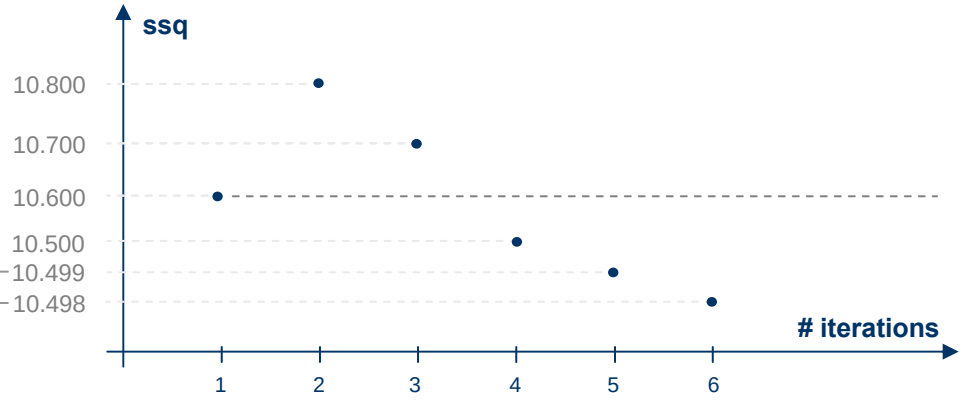

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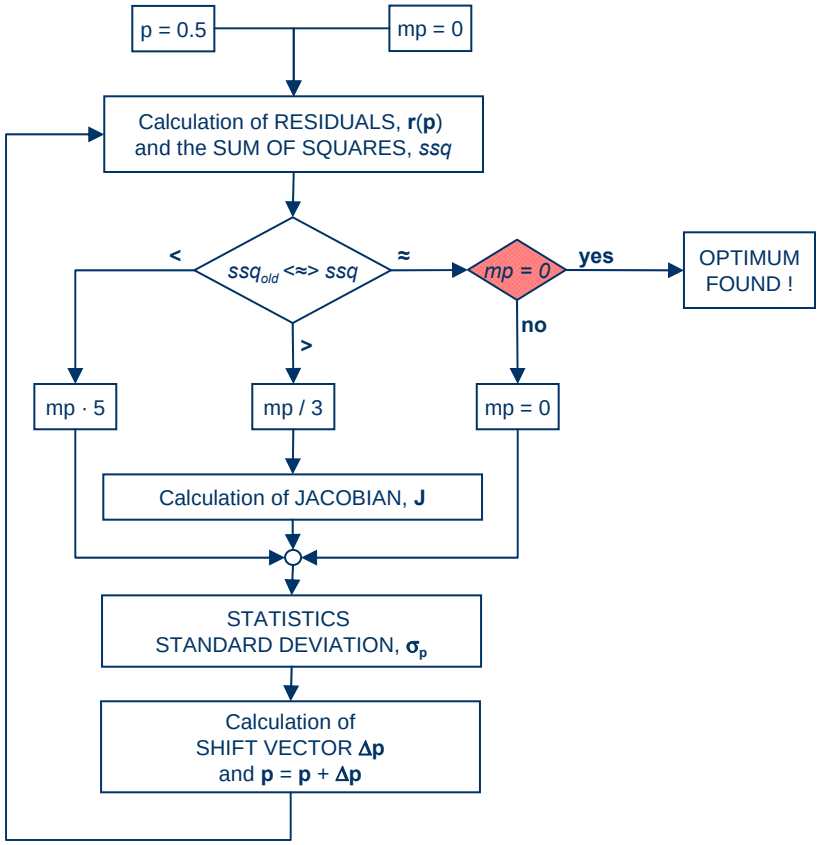

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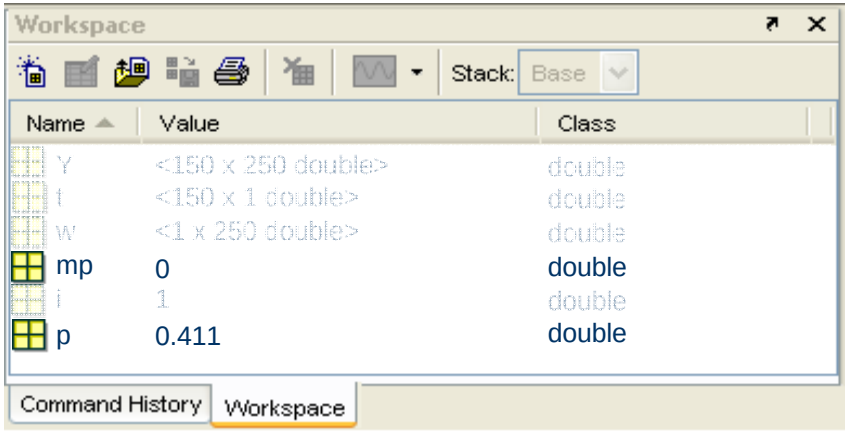

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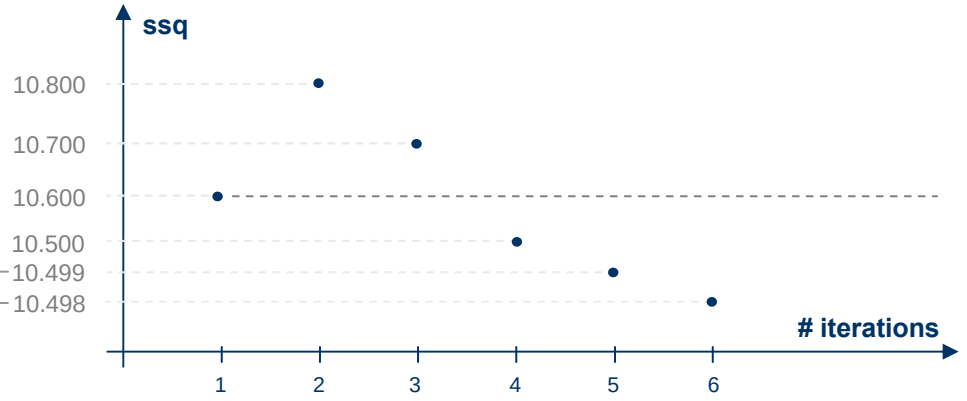

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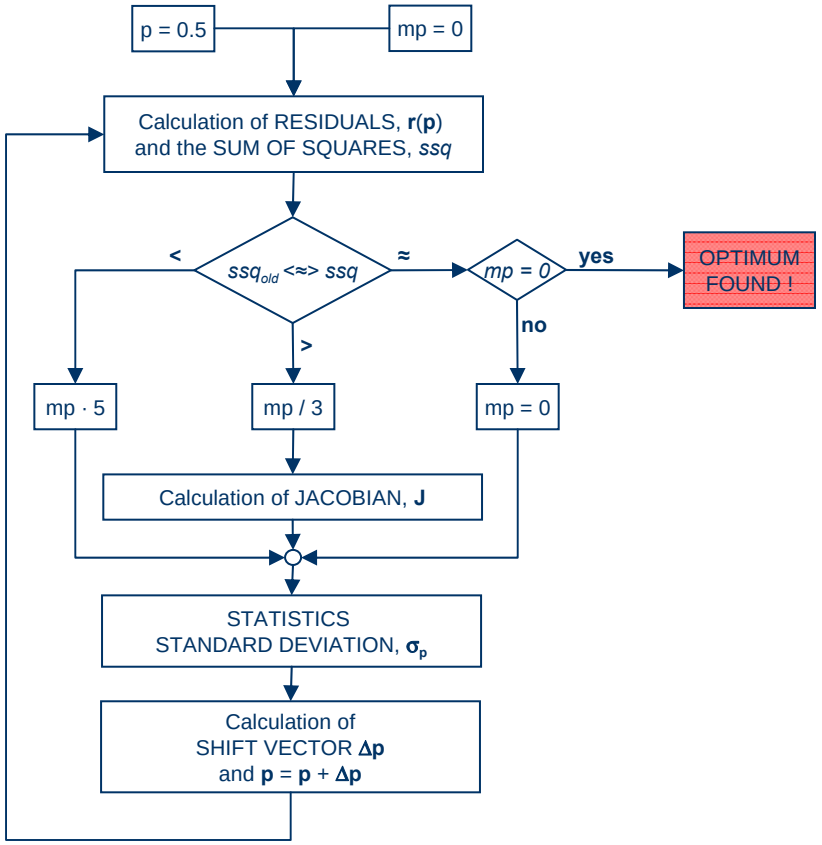

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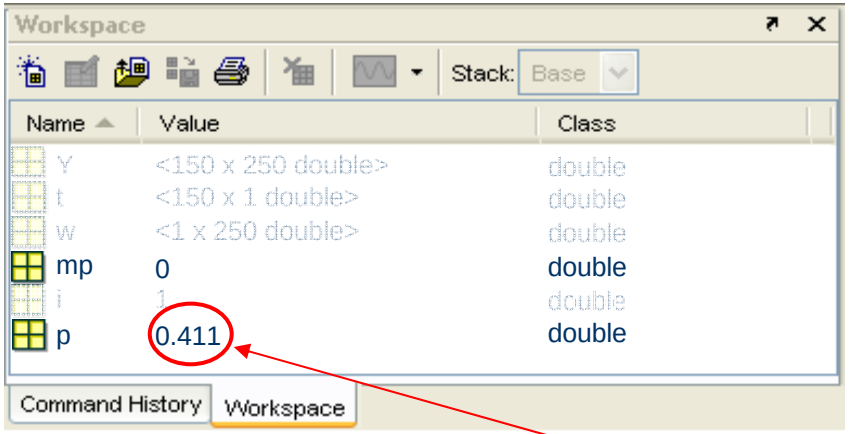

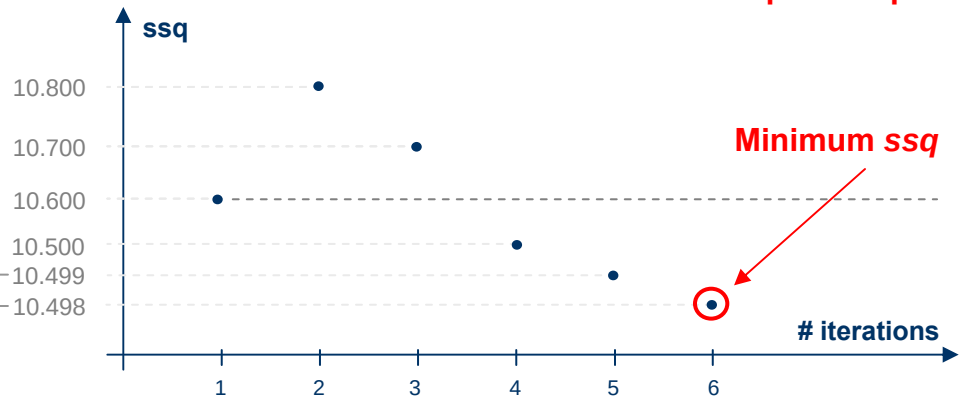

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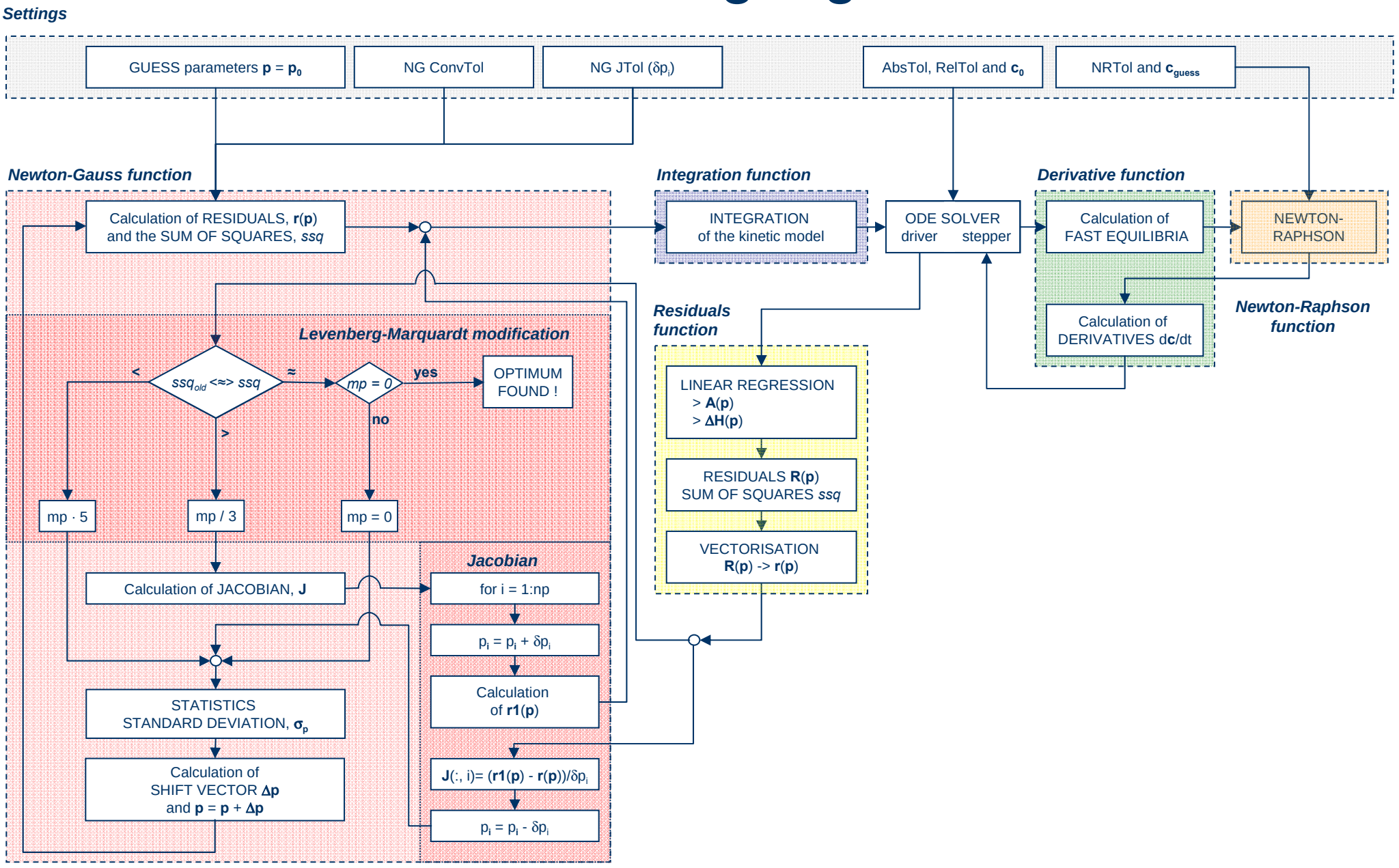
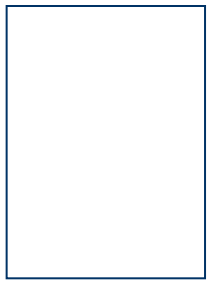
Kinetic modeling algorithm



Rank Deficiency
of the concentration profile

$\mathbf{C} =$ 

The maximum *rank* of **C** is :

$$rank_{max}(\mathbf{C}) = \min(nt, nc)$$

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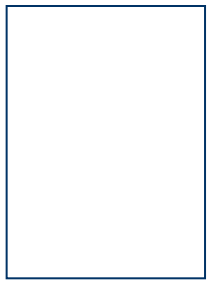
$\mathbf{C} =$ 

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The pseudo inverse $(\mathbf{C}^{+})$ only exists if : $rank(\mathbf{C}) = rank_{max}(\mathbf{C})$

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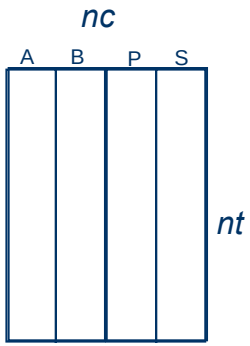
$\mathbf{C} =$ 

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Example : $A + B \xrightarrow{k} P + S \quad (in\ batch\ conditions)$

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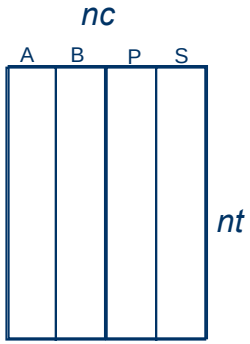
$\mathbf{C} =$ 

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- *Maximum possible rank ?*
- *Number of independant species in stoichiometric conditions ($A_0 = B_0 = 1$) ?*
- *And in non-stoichiometric conditions ($A_0 = 1$, $B_0 = 0.5$) ?*

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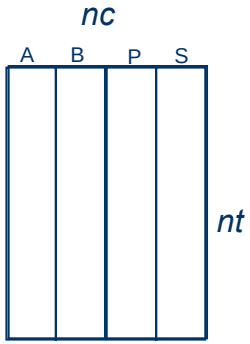
$\mathbf{C} =$ 

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Example : $A + B \xrightarrow{k} P + S \quad (in\ batch\ conditions)$

- *Maximum possible rank ?*

  $4\ species \Rightarrow rank_{max}(\mathbf{C}) = 4$

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Rank Deficiency
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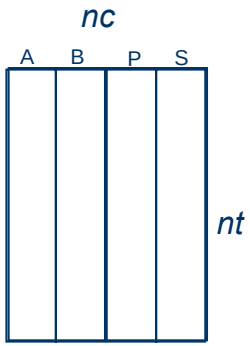
$\mathbf{C} =$ 

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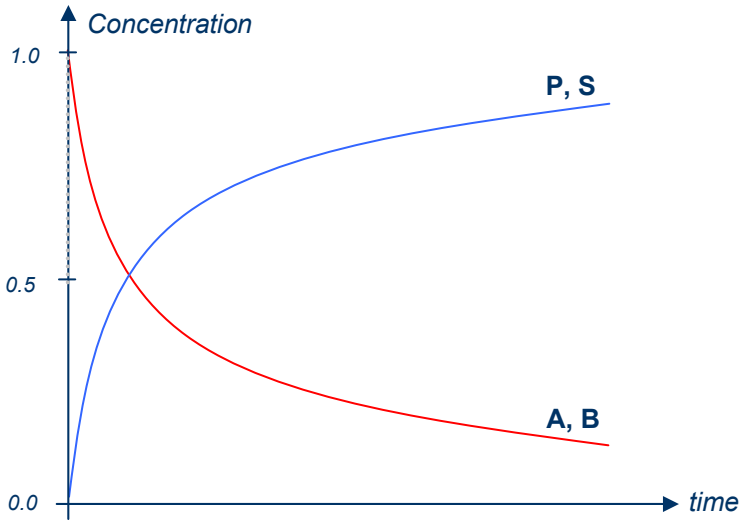

$[A]_t$ and $[B]_t$ are identical
$[P]_t$ and $[S]_t$ are identical

1 species
1 species
→ 2 independant species

Rank Deficiency
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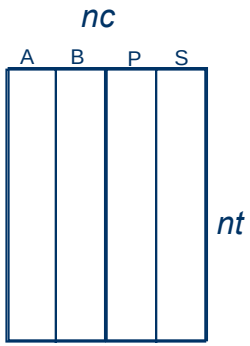
$\mathbf{C} =$ 

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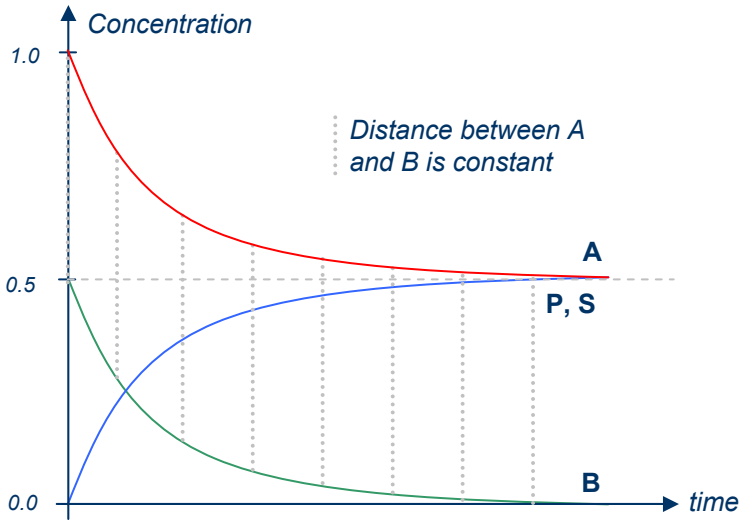
- *And in non-stoichiometric conditions ($A_0 = 1$, $B_0 = 0.5$) ?*

  $rank(\mathbf{C}) = 2 \quad (still!)$



[A]$_t$ *and* ***[B]$_t$*** *disappear at the same rate*
[P]$_t$ *and* ***[S]$_t$*** *are identical*

1 species
1 species → *2 independant species*

Annihilation of Rank Deficiency

5 ways to break the rank deficiency :

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- **Model Reduction** : set the dependant species as colorless (non-absorbing)

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- Extend the wavelength-time domain to resolve linear dependencies
  (**Tri-linear Measurements**)

  Example : Coupling chromatrography to UV

Annihilation of Rank Deficiency

5 ways to break the rank deficiency :

- **Model Reduction** : set the dependant species as colorless (non-absorbing)

  ⇒ *Rates constants will be correct*
  Absorption spectra will be wrong (mixed pure spectra)

- Provide **Known Spectra**

- Work under **Semi-Batch Conditions**

- Use concentration dependant measurements
  (**Second Order Global Analysis**)

- Extend the wavelength-time domain to resolve linear dependencies
  (**Tri-linear Measurements**)

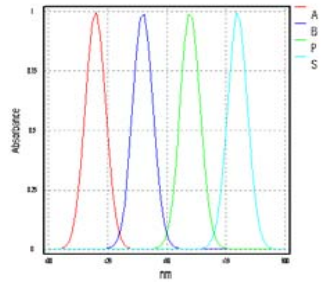
  Example : Coupling chromatography to UV

This last method is not addressed here, because it is highly complex

Reduced model

$$A(1) + B(0.5) \xrightarrow{k=0.5} P + S$$

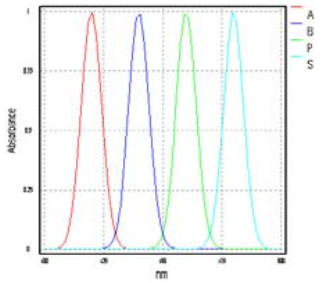
Simulated absorption spectra



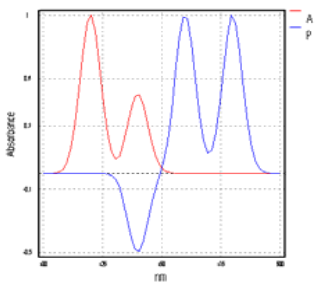
Reduced model

$$A(1) + B(0.5) \xrightarrow{k=0.5} P + S$$

Simulated absorption spectra

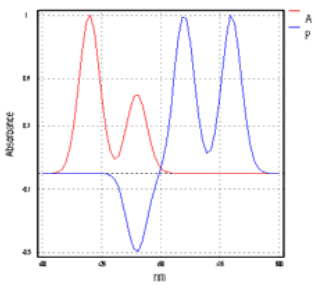


Fitted absorption spectra (mixed)



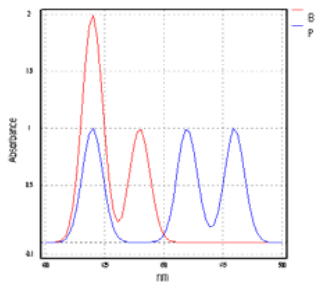
Colored species : ***A, S***

$k_{fitted} = 0.5$



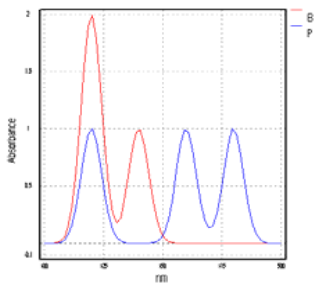
A, P

$k_{fitted} = 0.5$



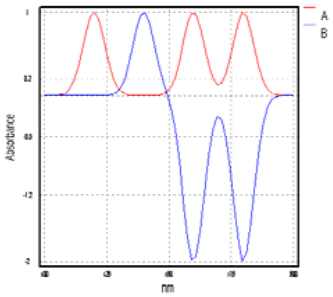
B, P

$k_{fitted} = 0.5$



B, S

$k_{fitted} = 0.5$



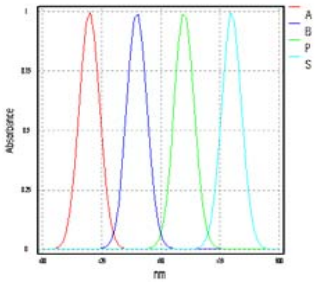
A, B

$k_{fitted} = 0.5$

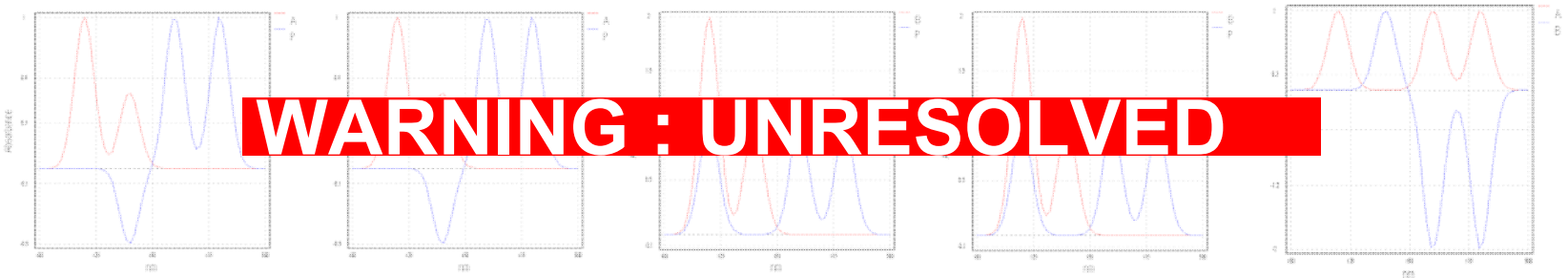
Reduced model

$$A(1) + B(0.5) \xrightarrow{k=0.5} P + S$$

Simulated absorption spectra



Fitted absorption spectra (mixed)



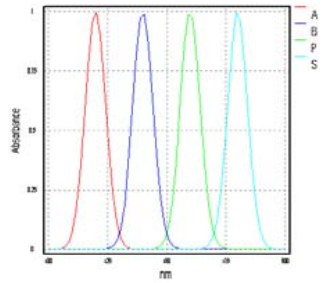
WARNING : UNRESOLVED

Colored species :	***A, S***	***A, P***	***B, P***	***B, S***	***A, B***
	$k_{fitted} = 0.5$	$k_{fitted} = 0.5$	$k_{fitted} = 0.5$	$k_{fitted} = 0.5$	$k_{fitted} = 0.5$

Add known spectra

$$A(1) + B(0.5) \xrightarrow{k=0.5} P + S$$

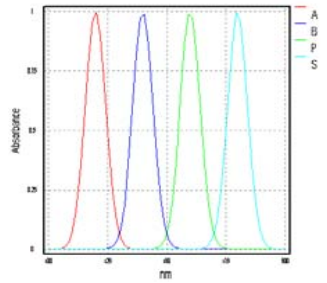
Simulated absorption spectra



Add known spectra

$$A(1) + B(0.5) \xrightarrow{k=0.5} P + S$$

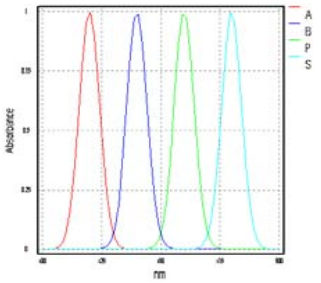
Simulated absorption spectra



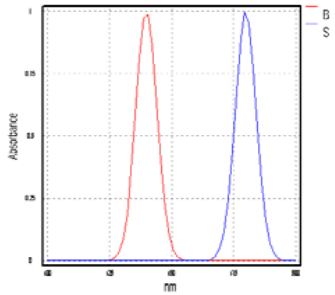
2 species are dependent

Add known spectra

Simulated absorption spectra



$$A(1) + B(0.5) \xrightarrow{k=0.5} P + S$$

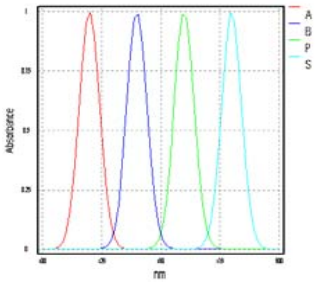


2 species are dependent

Let's provide 2 pure spectra : those of B and S

Add known spectra

Simulated absorption spectra



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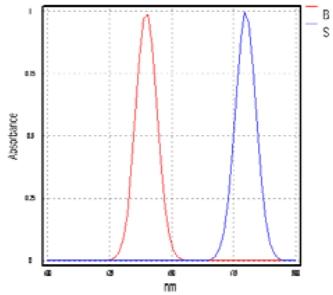


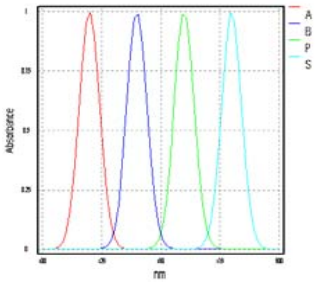
2 species are dependent

Let's provide 2 pure spectra : those of B and S

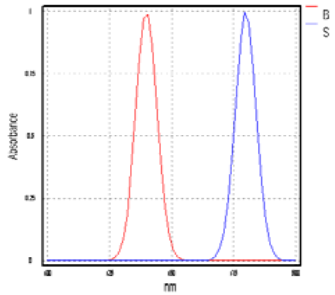
All species are set colored

Add known spectra

Simulated absorption spectra



$$A(1) + B(0.5) \xrightarrow{k=0.5} P + S$$

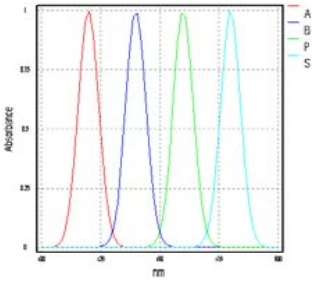


2 species are dependent

Let's provide 2 pure spectra : those of B and S

All species are set colored

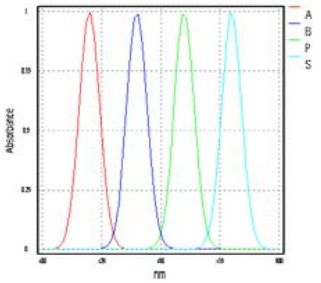
Fitted absorption spectra :



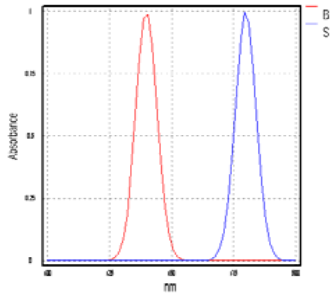
$k_{fitted} = 0.5$

Add known spectra

Simulated absorption spectra



$$A(1) + B(0.5) \xrightarrow{k=0.5} P + S$$



2 species are dependent

Let's provide 2 pure spectra : those of B and S

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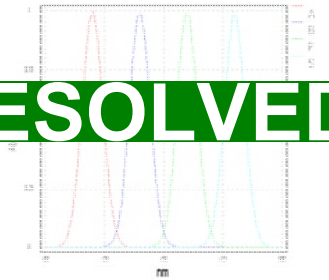


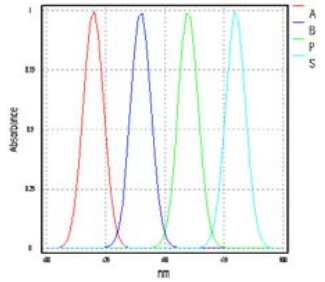
RESOLVED

$k_{fitted} = 0.5$

Work under semi-batch conditions

$$A + B \xrightarrow{k=0.5} P + S$$

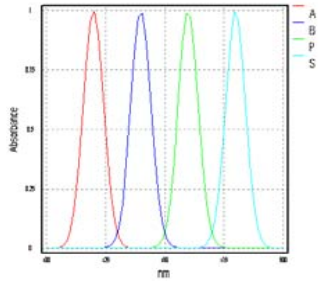
Simulated absorption spectra



Work under semi-batch conditions

$$A + B \xrightarrow{k=0.5} P + S$$

Simulated absorption spectra

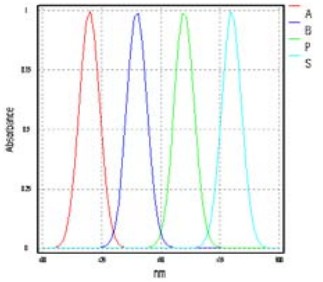


Dose A into B will break the rank deficiency between A and B but not between P and S !!!

Work under semi-batch conditions

$$A + B \xrightarrow{k=0.5} P + S$$

Simulated absorption spectra

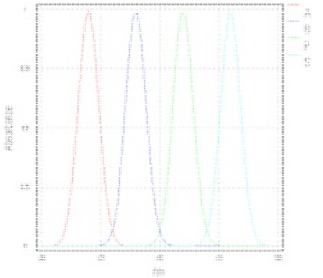


Dose A into B will break the rank deficiency between A and B but not between P and S !!!

To break both rank deficiencies, one has to dose :

- *A and P into B or*
- *A and S into B or*
- *B and P into A or*
- *B and S into A*

Fitted absorption spectra :

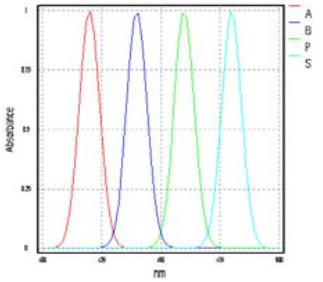


$k_{fitted} = 0.5$

Work under semi-batch conditions

$$A + B \xrightarrow{k=0.5} P + S$$

Simulated absorption spectra

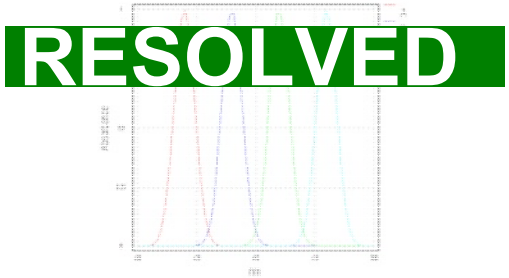


Dose A into B will break the rank deficiency between A and B but not between P and S !!!

To break both rank deficiencies, one has to dose :

- *A and P into B or*
- *A and S into B or*
- *B and P into A or*
- *B and S into A*

Fitted absorption spectra :

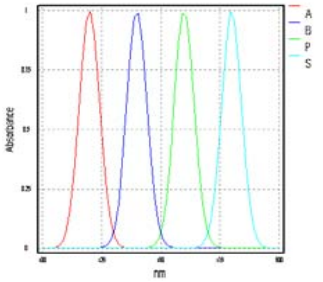


$k_{fitted} = 0.5$

Work under semi-batch conditions

$$A + B \xrightarrow{k=0.5} P + S$$

Simulated absorption spectra



Dose A into B will break the rank deficiency between A and B but not between P and S !!!

To break both rank deficiencies, one has to dose :

- *A and P into B or*
- *A and S into B or*
- *B and P into A or*
- *B and S into A*

Fitted absorption spectra :

RESOLVED

OR

PARTIALLY RESOLVED

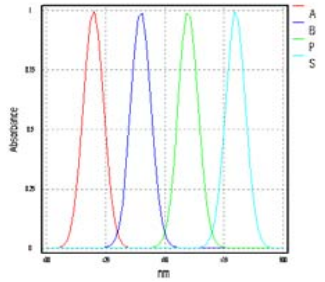
$k_{fitted} = 0.5$

NB : in practice, one dose A in B or B in A and set one of the two products (P or S) as uncolored. In such case, one spectrum of the two products is unresolved

Second Order Global Analysis

$$A + B \xrightarrow{k=0.5} P + S$$

Simulated absorption spectra



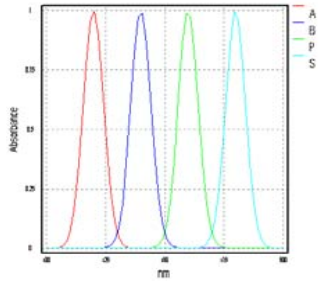
Second Order Global Analysis

$$A + B \xrightarrow{k=0.5} P + S$$

Let's make 3 Concentration dependent measurements

	A_0	B_0
# 1	*1*	*1*
# 2	*0.5*	*1*
# 3	*1*	*0.5*

Simulated absorption spectra



Second Order Global Analysis

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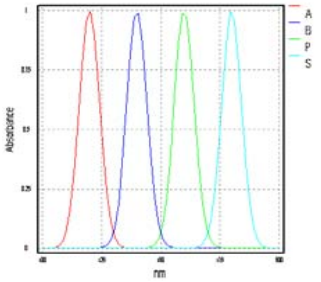
Simulated absorption spectra



Let's make 3 Concentration dependent measurements

	A_0	B_0
# 1	*1*	*1*
# 2	*0.5*	*1*
# 3	*1*	*0.5*

Hypothesis of Global Spectra :
The 3 sets of experiment share the same pure spectra

$$\begin{bmatrix} Y_{\#1} \\ Y_{\#2} \\ Y_{\#3} \end{bmatrix} = \begin{bmatrix} C_{\#1} \\ C_{\#2} \\ C_{\#3} \end{bmatrix} + \begin{bmatrix} A_{global} \end{bmatrix} + \begin{bmatrix} R_{\#1} \\ R_{\#2} \\ R_{\#3} \end{bmatrix}$$

$$Y_{tot} \qquad C_{tot} \qquad R_{tot}$$

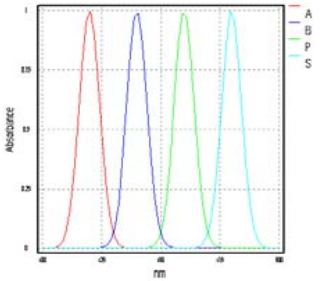
Second Order Global Analysis

$$A + B \xrightarrow{k=0.5} P + S$$

Let's make 3 Concentration dependent measurements

	A_0	B_0
# 1	*1*	*1*
# 2	*0.5*	*1*
# 3	*1*	*0.5*

Simulated absorption spectra



Hypothesis of Global Spectra :
The 3 sets of experiment share the same pure spectra

$$\begin{bmatrix} Y_{\#1} \\ Y_{\#2} \\ Y_{\#3} \\ Y_{tot} \end{bmatrix} = \begin{bmatrix} C_{\#1} \\ C_{\#2} \\ C_{\#3} \\ C_{tot} \end{bmatrix} \times A_{global} + \begin{bmatrix} R_{\#1} \\ R_{\#2} \\ R_{\#3} \\ R_{tot} \end{bmatrix}$$

Rank deficiency is broken in C_{tot}

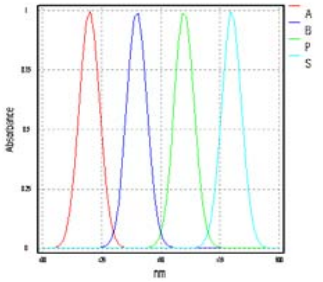
Second Order Global Analysis

$$A + B \xrightarrow{k=0.5} P + S$$

Let's make 3 Concentration dependent measurements

	A_0	B_0
# 1	*1*	*1*
# 2	*0.5*	*1*
# 3	*1*	*0.5*

Simulated absorption spectra



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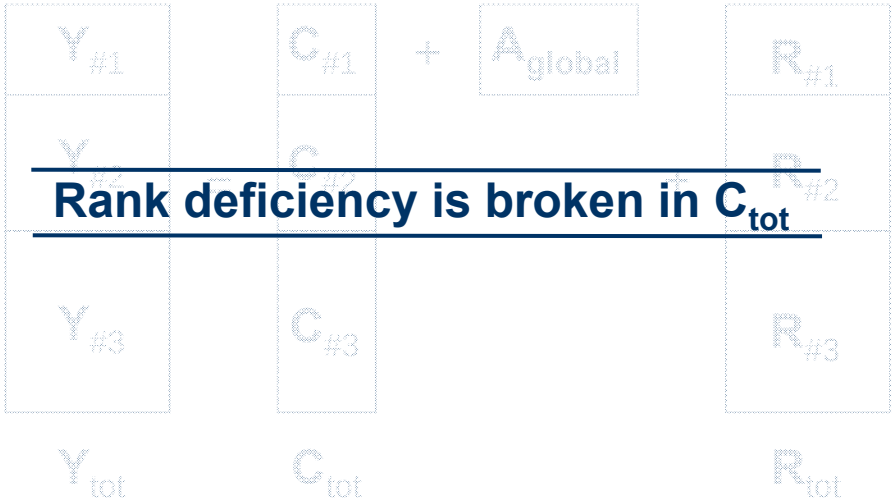
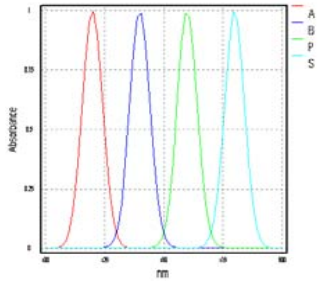


Rank deficiency is broken in C_{tot}

Fitted absorption spectra :



$k_{fitted} = 0.5$

Second Order Global Analysis

$$A + B \xrightarrow{k=0.5} P + S$$

Let's make 3 Concentration dependent measurements

	A_0	B_0
# 1	*1*	*1*
# 2	*0.5*	*1*
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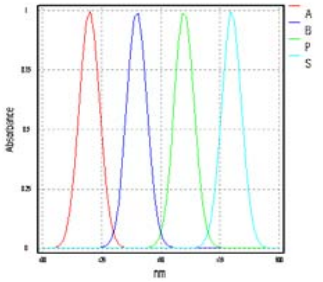
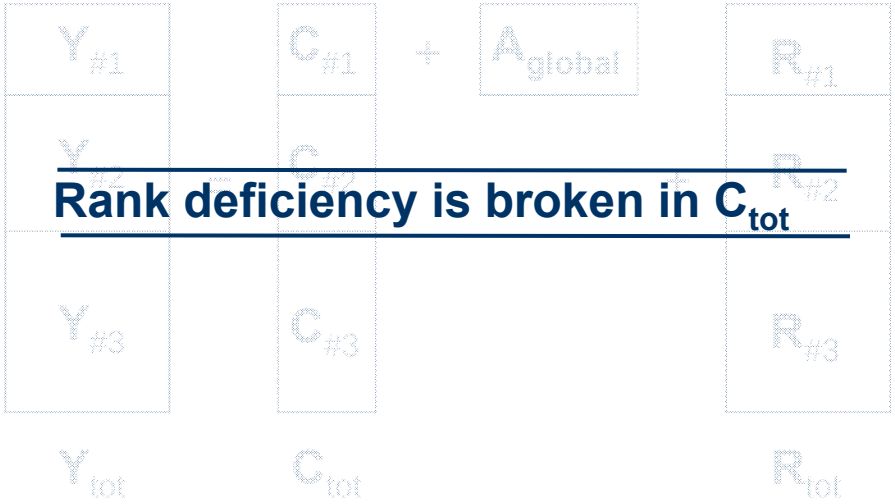
Simulated absorption spectra

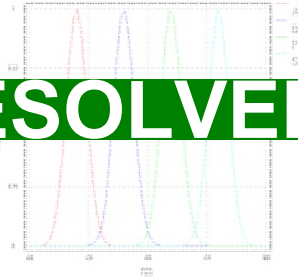


Hypothesis of Global Spectra :
The 3 sets of experiment share the same pure spectra



Rank deficiency is broken in C_{tot}

Fitted absorption spectra :



RESOLVED

$k_{fitted} = 0.5$

End of the Tutorial

That is « already » the end
of this Tutorial

Thank you for your attention !

Case study overview

The aims of these case studies are :

- Identify the number of detectable species
- Know the phenomenon of parameter interchange
- Identify and break rank deficiency
- Use residuals for model validation

Using the following techniques :

- PCA
- EFA
- ALS
- Hard-modeling

On simulated data from different types :

- Spectroscopy
- Calorimetry

Under batch and semi-batch conditions

- $A \xrightarrow{k=0.5} P$
- $A \xrightarrow{k_1=0.5} B \xrightarrow{k_2=0.3} P$
- $A + B \xrightarrow{k=0.5} P$
- $A + B(dosed) \xrightarrow{k=0.5} P$
- $A + B(dosed) \xrightarrow{k_1=10} C$

  $B(dosed) + C \xrightarrow{k_2=5} P$

Case study 1

Mechanism	:	$A \xrightarrow{k=0.5} P$
Signal	:	Spectroscopy
Process	:	Batch
Conversion	:	99%

- PCA/EFA : 2 species $\Rightarrow$ Full rank
- ALS : Easily resolved
  with non-negativity constraint
- Hard-modeling : *k* does not depend on $c_0(A)$ for a 1st order reaction

Case study 2

Mechanism	: $A \xrightarrow{k=0.5} P$
Signal	: Spectroscopy
Process	: Batch
Conversion	: 63%

- PCA/EFA : 2 species $\Rightarrow$ Full rank
- ALS : Resolved under constraints
  with non-negativity constraint and known spectrum of *P*
- Hard-modeling : *k* does not depend on $c_0(A)$ for a 1st order reaction

Case study 3

Mechanism	:	$A \xrightarrow{k=0.5} P$
Signal	:	Calorimetry
Process	:	Batch
Conversion	:	99%

- PCA/EFA/ALS : NA

- Hard-modeling : *k* does not depend on $c_0(A)$ for a 1st order reaction

Case study 4

Mechanism : $A + B \xrightarrow{k=0.5} P$

Signal : Spectroscopy

Process : Batch

Fitted mechanism : $A \longrightarrow P$

- PCA/EFA : 2 species $\Rightarrow$ Full rank
- ALS : Apparently resolved under constraints
  with non-negativity constraint and known spectrum of *P*
- Hard-modeling : 1st order mechanism is wrong !

Case study 5

Mechanism : $A \xrightarrow{k_1=0.5} B \xrightarrow{k_2=0.3} P$

Signal : Spectroscopy

Process : Batch

Conversion : 99%

- PCA/EFA : 3 species $\Rightarrow$ Full rank
- ALS : Resolved under constraints
  with non-negativity constraint and known spectrum of *B*
- Hard-modeling : *Parameter interchange*
  k_1 and k_2 swap depending on the initial guess

Case study 6

Mechanism : $A \xrightarrow{k_1=0.5} B \xrightarrow{k_2=0.3} P$

Signal : Spectroscopy

Process : Batch

Conversion : 77%

- PCA/EFA : 3 species $\Rightarrow$ Full rank
- ALS : Hardly resolved

  with non-negativity constraint and known spectrum of *B* and *P*
- Hard-modeling : *Parameter interchange*

  k_1 and k_2 swap depending on the initial guess

Case study 7

Mechanism	:	$A \xrightarrow{k_1=0.5} B \xrightarrow{k_2=0.3} P$
Signal	:	Calorimetry
Process	:	Batch
Conversion	:	99%

- PCA/EFA/ALS : NA

- Hard-modeling : *1.* Less robust than multivariate fitting
  2. Parameter interchange

Case study 8

Mechanism	:	$A \xrightarrow{k_1=0.5} B \xrightarrow{k_2=0.3} P$
Signal	:	Spectroscopy
Process	:	Batch
Fitting	:	Univariate (λ = 400)

- PCA/EFA/ALS : NA

- Hard-modeling : *1.* Less robust than multivariate fitting
  2. Parameter interchange

Case study 9

Mechanism : $A \xrightarrow{k_1=0.5} B \xrightarrow{k_2=0.3} P$

Signal : Spectroscopy

Fitting : Univariate (λ = 400)

Fitted mechanism : $A \longrightarrow P$

- PCA/EFA/ALS : NA

- Hard-modeling : *1.* Less robust than multivariate fitting
  2. Parameter interchange
  3. Structured residuals

  $\Rightarrow$ The model is slightly wrong but hard to validate at this single wavelength

Case study 10

Mechanism : $A + B \xrightarrow{k=0.5} P$

Signal : Spectroscopy

Process : Batch

Colored species : *A* and *P* (model reduction)

- PCA/EFA : 2 species ⇒ Rank deficiency
- ALS : NA
- Hard-modeling : Species *B* set as non-absorbing

  ⇒ *1. k* is correct

  2. The fitted pure spectra are wrong
  (linear combinations of the true pure spectra)

Case study 11

Mechanism : $A + B \xrightarrow{k=0.5} P$

Signal : Spectroscopy

Process : Batch

Known spectrum : Species *B*

- PCA/EFA : 2 species $\Rightarrow$ Rank deficiency
- ALS : NA
- Hard-modeling : The pure spectrum of B is provided

  $\Rightarrow$ *1.* *k* is correct

  2. The fitted pure spectra are resolved

Case study 12

Mechanism : $A + B(dosed) \xrightarrow{k=0.5} P$

Signal : Spectroscopy

Process : Semibatch

Conversion : 68%

- PCA/EFA : 3 species $\Rightarrow$ Full rank
- ALS : Resolved under strong constraints
  non-negativity constraint and known spectrum of *B* and *P* !
- Hard-modeling : The pure spectra are resolved

Case study 13

Mechanism	:	$A + B(dosed) \xrightarrow{k=0.5} P$
Signal	:	Calorimetry
Process	:	Semibatch
Conversion	:	68%

- PCA/EFA/ALS : NA

- Hard-modeling : Fitting calorimetric data is more robust in semibatch than in batch conditions !

Case study 14

Mechanism : $A + B(dosed) \xrightarrow{k_1=10} C$

$B(dosed) + C \xrightarrow{k_2=5} P$

Signal : Spectroscopy

Process : Semibatch

- PCA/EFA : 4 species $\Rightarrow$ Full rank
- ALS : Resolved under very strong constraints
  non-negativity constraint and known spectrum of *B*, *C* and *P* !
- Hard-modeling : The pure spectra are resolved

Case study 15

Mechanism : $A + B(dosed) \xrightarrow{k_1=10} C$

$B(dosed) + C \xrightarrow{k_2=5} P$

Signal : Calorimetry

Process : Semibatch

- PCA/EFA/ALS : NA
- Hard-modeling : Fitting calorimetric data conditions is more robust in semibatch than in batch conditions !

Summary on the Case studies (1)

- PCA/EFA
  - These two techniques provide information on the number of observable species and therefore the maximum rank of **Y**

- ALS
  - Non-negativity constraint alone only resolves $A \rightarrow P$
  - Pure spectra of the products are required if they are not fully formed
  - Pure spectra of the intermediates are generally required for complete resolution
  - Without *a priori* knowledge on mechanisms and/or spectra, rank deficiency is undetectable

Summary on the Case studies (2)

- HARD-MODELING

BATCH CONDITIONS

- Multivariate fitting of spectrocopy data is more reliable than univariate fitting
- Fitting of calorimetric data is not very stable in batch conditions
- Rank deficiency due to the model can be easily broken by :

  - **Model reduction** (some species are set non-absorbing)
    Fitted pure spectra are wrong (linear combination of the true ones)
    but nonlinear parameters are correct !

  - **Known spectra provided**
    Fitted pure spectra and nonlinear parameters are in this case both correct

SEMIBATCH CONDITIONS

- Fitting of calorimetric data is more robust in semibatch conditions
- The dosing completely breaks simple rank deficient problems and partially highly complex mechanisms (eg. 2 intermediates, 2 products …)

FIRST ORDER MECHANISMS

- With 1st order mechanisms, rates are independent on initial concentrations and rate constants can swap without differences in fitting (*Parameter Interchange*)

Appendix 1 : List of the Matlab files

#	Simulated Mechanism	Fitted mechanism	Process	Model rank	Rank deficiency	Conversion	Fitted signal	Fitting	PCA / EFA	ALS	Hard-modeling	File
1	A -> P	same	Batch	2	No	99.3%	Spec	Multivariate	2	[> 0]	C_0(A) = 1 and 10	main_AtoP_X99
2	A -> P	same	Batch	2	No	63.2%	Spec	Multivariate	2	[> 0 , P]	NA	main_AtoP_X60
3	A -> P	same	Batch	2	No	99.3%	Cal	Univariate	NA	NA	NA	main_AtoP_cal
4	A + B -> P	A -> P	Batch	2	Yes	90.9%	Spec	Multivariate	2	NA	Structured residuals	main_ApBtoP_1st_order
5	A -> B -> P	same	Batch	3	No	99.9%	Spec	Multivariate	3	[> 0 , B]	Swap of k's	main_AtoBtoP_X99
6	A -> B -> P	same	Batch	3	No	77.7%	Spec	Multivariate	3	[> 0 , B , P]	NA	main_AtoBtoP_X75
7	A -> B -> P	same	Batch	3	No	99.9%	Cal	Univariate	NA	NA	Less robust than Case 5	main_AtoBtoP_cal
8	A -> B -> P	same	Batch	3	No	99.9%	Spec	Univariate	NA	NA	Less robust than Case 5	main_AtoBtoP_univar1
9	A -> B -> P	A -> B	Batch	3	No	99.9%	Spec	Univariate	NA	NA	Structure in the residuals	main_AtoBtoP_univar2
10	A + B -> P	same	Batch	2	Yes	90.9%	Spec	Multivariate	2	NA	B not absorbing	main_ApBtoP_batch
11	A + B -> P	same	Batch	2	Yes	90.9%	Spec	Multivariate	2	NA	Pure spectrum of B provided	main_ApBtoP_batch_Bknown
12	A + B(dosed) -> P	same	Semibatch	3	No	68.2%	Spec	Multivariate	3	[> 0 , B , P]	NA	main_ApBtoP_semi_Y
13	A + B(dosed) -> P	same	Semibatch	3	No	68.2%	Cal	Univariate	3	NA	NA	main_ApBtoP_semi_cal
14	A + B(dosed) -> C B(dosed) + C -> P	same	Semibatch	4	No	99.8%	Spec	Multivariate	4	[> 0 , B , C , P]	NA	main_ApBtoC_BpCtoP_semi_Y
15	A + B(dosed) -> C B(dosed) + C -> P	same	Semibatch	4	No	99.8%	Cal	Univariate	4	NA	NA	main_ApBtoC_BpCtoP_semi_cal