binding. Furthermore, as acetylation occurs only after microtubule polymerization, the enzyme responsible for this modification should probably localize to the microtubule lumen.

The preference of ER sliding for acetylated microtubules may indicate a functional difference between this type of ER motility and that mediated by the tip attachment complex. Intermembrane contacts are important for many functions of the ER and other membranous compartments [9]. The authors' results led them to hypothesize that the sliding mechanism may be used as a way for ER tubules to find and contact other organelles along a subpopulation of microtubules.

To explore this possibility, Friedman et al. [8] tracked the movements of two other organelles, mitochondria and endosomes, with respect to both ER and acetylated microtubules. A majority of both organelles remained in persistent contact with the ER. However, only mitochondria appeared to also localize preferentially to acetvlated microtubules, suggesting that ER contacts with mitochondria. but not endosomes, are enriched along acetvlated microtubules. Consistent with this finding is the fact that both ER and mitochondria are cargoes of kinesin-1, while endosomes are moved by KIF16B, a member of the kinesin-3 family [15]. Further, work by Cai et al. [6] showed that another kinesin-3 family member, KIF1A, displayed no preference for acetylated microtubules in COS-7 cells.

By biasing protein association with a particular subset of microtubules, acetylation allows for subpopulations of microtubules to act as compartments along which specific cargoes can find each other, and be found in return. In a sense, these microtubule compartments can be thought of as cellular pubs that attract a specific cargo crowd for mingling. Microtubule acetylation is also probably involved in the regulation of cell migration both in fibroblasts [16] and neurons [17]. It is attractive to speculate that polarization of moving cells requires kinesin-dependent recruitment of selective cargoes to the leading edge of migrating cells.

Additional investigation is necessary to determine whether this selective motor recruitment applies to other microtubule modifications and other motor proteins. It is possible that at least some other microtubule motors have a higher affinity for either a specific tubulin isoform or a particular post-translational modification, similar to the preferential recruitment of kinesin-1 to acetylated microtubules. Such selectivity could create multiple microtubule compartments to recruit particular cargoes - different pubs for different crowds (Figure 1). Once bound to microtubules, the cargo can undergo bidirectional transport to facilitate its movement through the crowded cytoplasm. Cargoes recruited to the same subset of microtubules will interact with much higher efficiency. thus promoting exchange of molecules between particular cell compartments. If this simple model is correct, it indicates that, in addition to their role as tracks for long-distance transport, microtubules serve an important role as scaffolds for the organization and compartmentalization of the otherwise randomly distributed cellular components.

## References

- Westermann, S., and Weber, K. (2003). Post-translational modifications regulate microtubule function. Nat. Rev. Mol. Cell Biol. 4, 938–947.
- Liao, G., and Gundersen, G.G. (1998). Kinesin is a candidate for cross-bridging microtubules and intermediate filaments. Selective binding of kinesin to detyrosinated tubulin and vimentin. J. Biol. Chem. 273, 9797–9803.
- Reed, N.A., Cai, D., Blasius, T.L., Jih, G.T., Meyhofer, E., Gaertig, J., and Verhey, K.J. (2006). Microtubule acetylation promotes kinesin-1 binding and transport. Curr. Biol. 16, 2166–2172.
- Dompierre, J.P., Godin, J.D., Charrin, B.C., Cordelieres, F.P., King, S.J., Humbert, S., and Saudou, F. (2007). Histone deacetylase 6 inhibition compensates for the transport deficit in Huntington's disease by increasing tubulin acetylation. J. Neurosci. 27, 3571–3583.
- Konishi, Y., and Setou, M. (2009). Tubulin tyrosination navigates the kinesin-1 motor domain to axons. Nat. Neurosci. 12, 559–567.

- Cai, D., McEwen, D.P., Martens, J.R., Meyhofer, E., and Verhey, K.J. (2009). Single molecule imaging reveals differences in microtubule track selection between Kinesin motors. PLoS Biol. 7, e1000216.
- Hammond, J.W., Huang, C.F., Kaech, S., Jacobson, C., Banker, G., and Verhey, K.J. (2010). Posttranslational modifications of tubulin and the polarized transport of kinesin-1 in neurons. Mol. Biol. Cell *21*, 572–583.
- Friedman, J.R., Webster, B.M., Mastronarde, D.N., Verhey, K.J., and Voeltz, G.K. (2010). ER sliding dynamics and ER-mitochondrial contacts occur on acetylated microtubules. J. Cell Biol. 190, 363–375.
- Voeltz, G.K., Rolls, M.M., and Rapoport, T.A. (2002). Structural organization of the endoplasmic reticulum. EMBO Rep. 3, 944–950.
- Terasaki, M., Chen, L.B., and Fujiwara, K. (1986). Microtubules and the endoplasmic reticulum are highly interdependent structures. J. Cell Biol. *103*, 1557–1568.
- Waterman-Storer, C.M., and Salmon, E.D. (1998). Endoplasmic reticulum membrane tubules are distributed by microtubules in living cells using three distinct mechanisms. Curr. Biol. 8, 798–806.
- Allan, V., and Vale, R. (1994). Movement of membrane tubules along microtubules in vitro: evidence for specialised sites of motor attachment. J. Cell Sci. 107, 1885–1897.
- Wozniak, M.J., Bola, B., Brownhill, K., Yang, Y.C., Levakova, V., and Allan, V.J. (2009). Role of kinesin-1 and cytoplasmic dynein in endoplasmic reticulum movement in VERO cells. J. Cell Sci. 122, 1979–1989.
- Nogales, E., Whittaker, M., Milligan, R.A., and Downing, K.H. (1999). High-resolution model of the microtubule. Cell 96, 79–88.
- Hoepfner, S., Severin, F., Cabezas, A., Habermann, B., Runge, A., Gillooly, D., Stenmark, H., and Zerial, M. (2005). Modulation of receptor recycling and degradation by the endosomal kinesin KIF16B. Cell *121*, 437–450.
- Gundersen, G.G., and Bulinski, J.C. (1988). Selective stabilization of microtubules oriented toward the direction of cell migration. Proc. Natl. Acad. Sci. USA 85, 5946–5950.
- Creppe, C., Malinouskaya, L., Volvert, M.L., Gillard, M., Close, P., Malaise, O., Laguesse, S., Cornez, I., Rahmouni, S., Ormenese, S., *et al.* (2009). Elongator controls the migration and differentiation of cortical neurons through acetylation of alpha-tubulin. Cell *136*, 551–564.

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DOI: 10.1016/j.cub.2010.08.058

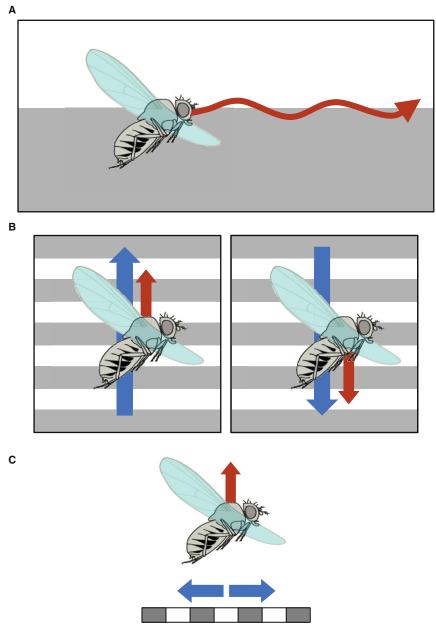
## Insect Vision: A Few Tricks to Regulate Flight Altitude

A recent study sheds new light on the visual cues used by *Drosophila* to regulate flight altitude. The striking similarity with previously identified steering mechanisms provides a coherent basis for novel models of vision-based flight control in insects and robots.

Dario Floreano and Jean-Christophe Zufferey

Insects predominantly use vision to steer, to regulate their flight speed

and height, to avoid impending obstacles, to chase moving targets, and to land on objects. Their evolutionary success, ability to fly in complex environments, and the



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Figure 1. Visual regulation of altitude in Drosophila.

Novel findings suggest that *Drosophila* regulate altitude (A) by aligning with static horizontal edges, (B) by increasing or decreasing lift in accordance with the vertical displacement of horizontal structure, and (C) by increasing lift in the presence of optic flow expansion in the ventral field when flying at low altitude.

relatively compact size of their brains and bodies make flying insects not only an appealing subject of research for biologists, but also a compelling source of inspiration for computer scientists and engineers aiming at creating vision-based flying machines [1].

Many experimental studies performed so far on flying insects have concentrated on visual cues for horizontal steering behaviors. Decades of observations of insects in arenas with controlled visual stimulation have shown that they selectively respond to specific patterns of retinal image motion (optic flow) to stabilize flight [2], regulate speed [3], and avoid collisions [4]. However, little is known about the visual cues used to control flight altitude. A recent computational model [5], which was validated in experiments with a miniature helicopter, predicts that insects may regulate flight altitude by maintaining a constant angular speed of the ventral optic flow — lower flight altitude generates higher image speed in the ventral area of the compound eyes.

In contrast to this prediction, the novel experiments performed by Straw et al. [6], recently published in Current Biology, suggest that Drosophila does not regulate altitude by monitoring image speed in the ventral area. The authors used an advanced flight arena with computer-controlled speed and direction of images projected on the ground and walls. In a first series of experiments, they cancelled the natural motion of the ground image caused by the insect flight and imposed a controlled set of ventral optic-flow speeds and directions. Because insects did not react to this optical stimulation by changing altitude, the authors ruled out the hypothesis that ventral optic flow is used to maintain altitude and searched for alternative ways in which Drosophila could regulate altitude.

In the first set of experiments, Straw et al. [6] showed that insects roughly align their flight altitude with the height of horizontal edges that appear near the eye equator, independently of the presence, speed, or direction of a ventral image. They also showed that insects change flight altitude in order to maintain alignment with vertically moving horizontal edges (Figure 1A). This behavior is remarkably similar to previously identified behaviors of horizontal stabilization [7] and tracking of vertical edges [8], suggesting the duplication or use of the same neuronal circuitry for horizontal steering and altitude regulation.

In a second set of experiments, the authors showed that insects increased altitude when presented with lateral, upward moving horizontal gratings and decreased altitude when presented with downward moving gratings (Figure 1B). This behavior, which cannot be explained by edge alignment, could be mediated by a separate mechanism to compensate for unintended altitude changes — dropping altitude would generate coherent upward motion of the image and thus trigger increased lift. Once again, this behavior is reminiscent of the fly's optomotor response to wide-field horizontal shifts of the visual scene in order to compensate for unintended deviations from straight flight [9,10].

In a third set of experiments, they showed that insects are also sensitive to optic flow on the ventral area, but only when the optic flow corresponded to a pattern of radial expansion centered under the insect at a height of less than 15 cm (Figure 1C). This behavior, which could serve as emergency collision avoidance, is analogous to the well-documented behavior of steering away from areas of expanding fields when flying on a horizontal plane [2].

These results not only provide a novel explanation for flight altitude regulation, but also suggest that insects may use the same few sensory-motor reflexes to control flight in the three dimensions. This hypothesis is very appealing because it provides the basis for the formulation of a novel, coherent theory of vision-based flight regulation that abstracts from the division between flight in the horizontal and vertical dimensions. Such a theory could also generate new predictions about the function and architecture of the neural circuitries mapping optical stimulation of the compound eyes into motor control and drive novel experiments to elucidate the behavioral, neuronal, and genetic basis of vision-based flight control.

Nonetheless, this study raises a few questions. For example, it is not clear to what extent the findings apply also to other insect species. Experimental evidence indicates that honeybees do regulate their ventral optic flow, which makes them react to varying ground speeds by changing their altitude [11], as predicted by the computational model of Franceschini *et al.* [5]. Furthermore, there seems to be a precise correlation between the magnitude of the ventral optic flow and the altitude of honeybees during landing [12]. Finally, it is not clear to what extent horizontal edges are present in the natural habitat of these insects, at what distance they must be to be used by the animal, and what happens to altitude regulation when no such edges are present.

The work of Straw et al. [6] also shows the increasing relevance of computational models developed and tested on robots in order to understand biological principles of behavioral systems. Although the model of ventral optic flow regulation [5] was not confirmed by experimental observations of this study (notwithstanding the caveats mentioned above), it did provide a precise working hypothesis that motivated this novel study. In the future, robotics computational models could also be useful for precisely formulating, refining, and validating in an artificial behavioral system the new mechanisms suggested by Straw et al. [6].

Finally, these novel findings can also guide the development of novel control strategies and optoelectronic devices for miniature robots autonomously flying in urban environments, in buildings, or in forests. Although engineers and computer scientists have successfully used ventral optic flow to regulate altitude both in simulated [13] and real flying robots [14-17], those solutions assume the presence of detectable texture in the ventral area of the robot, which in reality is not always the case. Horizontal contrast, which is often prominent in built environments and forests, could replace or complement other strategies for altitude control. The capability and need of detecting horizontal contrasts near the equator line of the eyes may also affect the design of artificial compound eyes, which are attracting growing interest in the engineering community [18-20] because they could offer a field of view, size, and response speed that is better than conventional single-aperture cameras for motion-related tasks.

## References

 Floreano, D., Zufferey, J.-C., Srinivasan, M.V., and Ellington, C. (2009). Flying Insects and Robots (Berlin: Springer).

- Egelhaaf, M., and Borst, A. (1993). A look into the cockpit of the fly: visual orientation, algorithms, and identified neurons. J. Neurosci. 13, 4563–4574.
- Srinivasan, M., Lehrer, M., Kirchner, W., and Zhang, S. (1991). Range perception through apparent image speed in freelyflying honeybees. Visual Neurosci. 6, 519–535.
- Tammero, L., and Dickinson, M. (2002). Collision-avoidance and landing responses are mediated by separate pathways in the fruit fly. J. Exp. Biol. 205, 2785–2798.
- Franceschini, N., Ruffier, F., and Serres, J. (2007). A Bio-inspired flying robot sheds light on insect piloting abilities. Curr. Biol. 17, 1–7
- Straw, A.D., Lee, S., and Dickinson, M.H. (2010). Visual control of altitude in flying *Drosophila*. Curr. Biol. 20, 1550–1556.
- Buchner, E. (1976). Elementary movement detectors in an insect visual system. Biol. Cybernet. 24, 85–101.
- Horn, E., and Wehner, R. (1975). The mechanism of pattern fixation in the walking fly, *Drosophila Melanogaster*. J. Comp. Physiol. 101, 39–56.
- Götz, K. (1975). The optomotor equilibrium of the *Drosophila* navigation system. J. Comp. Physiol. 99, 187–210.
- Srinivasan, M., Poteser, M., and Kral, K. (1999). Motion detection in insect orientation and navigation. Vision Res. 39, 2749–2766.
- Portelli, G., Ruffier, F., and Franceschini, N. (2010). Honeybees change their height to restore their optic flow. J. Comp. Physiol. A Neuroethol. Sens. Neural. Behav. Physiol. 196, 307–313.
- Srinivasan, M., Zhang, S., Chahl, J., Barth, E., and Venkatesh, S. (2000). How honeybees make grazing landings on flat surfaces. Biol. Cybernet. 83, 171–183.
- Neumann, T., Huber, S., and Bülthoff, H.H. (2001). Artificial systems as models in biological cybernetics. Behavioral Brain Sci. 24, 1071–1072.
- Ruffier, F., and Franceschini, N. (2005). Optic flow regulation: the key to aircraft automatic guidance. Robotics Autonomous Systems 50, 177–194.
- Barber, D.B., Griffiths, S.R., McLain, T.W., and Beard, R.W. (2007). Autonomous landing of miniature aerial vehicles. J. Aerosp. Comp. Inf. Commun. 4, 770–784.
- Beyeler, A., Zufferey, J.-C., and Floreano, D. (2009). Vision-based control of near-obstacle flight. Auton. Robots 27, 201–219.
- Zufferey, J.-C., Beyeler, A., and Floreano, D. (2010). Autonomous flight at low altitude using light sensors and little computational power. Int. J. Micro. Air Veh. 2, 107–117.
- Duparré, J., Dannberg, P., Schreiber, P., Bräuer, A., and Tünnermann, A. (2005). Thin compound eye camera. Appl. Optics 44, 2949–2956.
- Radtke, D., Duparré, J., Zeitner, U., and Tünnermann, A. (2007). Laser lithographic fabrication and characterization of a spherical artificial compound eye. Opt. Exp. 15, 3067–3077.
- Jeong, K.-H., Kim, J., and Lee, L.P. (2006). Biologically inspired artificial compound eyes. Science 312, 557–561.

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